

# Overview over the DKFZ kidney data package

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```
> library(kidpack)
```

The package contains five data objects: two for the processed data, including sample information (`phenoData`) and probe (genes) information, and three for the raw data, including spotting information and array processing information.

The data was measured at the German Cancer Research Centre in 2002 by Holger Sültmann [1]. He hybridized labeled cDNA from around 85 renal cell cancer biopsies that had been obtained at the University of Göttingen to cDNA arrays that he had produced himself. The cDNA arrays use the two-color Stanford-type spotted cDNA technology, with 4224 different clones spotted in duplicate. About half of the clones were selected for being expressed in kidney according to a previous study on whole genome arrays, and the other half are from Bernd Korn's (RZPD) 'onco collection'. Each sample was hybridized twice. 175 chips were scanned and digitized. After quality control, we selected one representative (good) chip for each sample, resulting in a set of 74. These are presented in the *exprSet* named `eset`.

## 1 What is it good for?

There were three different subtypes of renal cell cancer (RCC): clear cell (`cc`), papillary (`p`), and chromophobe (`ch`). These pheno-variables may be used for classification or differential expression. The gene expression is quite strongly associated with the subtype.

Other interesting phenovariabls are the survival variables (`progress`, `rf.survival`) and (`died`, `survival.time`). Obviously, the two are highly correlated. The binary variable `m` indicates whether metastases were present (and known) at the time of surgery. The association of the gene expression data with these variables is more subtle. Perhaps only wishful thinking.

The manuscript has been submitted. As soon as it is accepted, final, and public, the preprint will be made available in the `doc` directory of the package. Until then, please contact me (WH) directly and I can send you the most current version by email.

## 2 Processed data

```
> data(eset)
> data(cloneanno)
```

For later use, we define some plot colors for the `type` variable:

```
> unique(pData(eset)$type)

[1] "ccRCC" "pRCC" "chRCC"

> cols <- c("red", "blue", "darkgreen")
> names(cols) <- c("ccRCC", "pRCC", "chRCC")
```

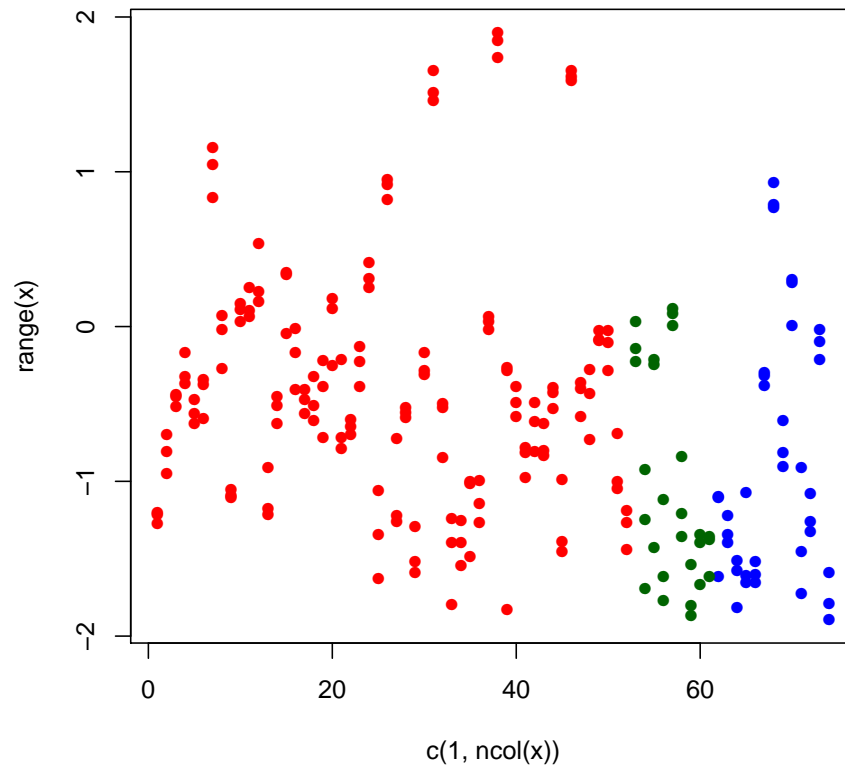
The chips contained three different clones that all probed for Fibronectin 1:

```
> sel <- grep("fibronectin 1", cloneanno$description)
> cloneanno[sel, ]
```

	plate	SrcRow	SrcCol	imageid	AccNumber	spot1	spot2	
2119	6	11	3	136798 r36450,"r36451"		1612	5964	
2626	7	14	15	324061 w46461,"w46530"		1730	6082	
2627	7	14	19	324997 w48576,"w48721"		1731	6083	
								description
2119				human mrna for fibronectin (fn precursor),"fibronectin 1 : fn1","fn1"				
2626				human mrna for fibronectin (fn precursor),"fibronectin 1 : fn1","fn1"				
2627				human mrna for fibronectin (fn precursor),"fibronectin 1 : fn1","fn1"				
				vendor				
2119				IMAGp950				
2626				IMAGp950				
2627				IMAGp950				

Let's plot the expression values:

```
> eo <- eset[sel, order(pData(eset)$type)]
> x <- exprs(eo)
> plot(c(1, ncol(x)), range(x), type = "n")
> for (i in 1:nrow(x)) points(x[i, ], col = cols[pData(eo)$type],
+   pch = 16)
```



### 3 Raw data

Let's have a look at the raw data

```
> data(qua)
> data(hybanno)
> data(spotanno)
> s1 <- cloneanno$spot1[sel]
> s2 <- cloneanno$spot2[sel]
> s1
```

```
[1] 1612 1730 1731
```

```
> qua[s1, "fg.green", 1:3]
```

	1	2	3
1612	23.2745	7.4989	12.4166
1730	18.4937	10.3098	16.4911
1731	17.2570	8.1597	12.4474

```
> hybanno[1:3, ]
```

	filename	patientid	slideid
1	00-P09206_E44-1.txt	87	E44-1
2	00-P09206_E63-3.txt	87	E63-3
3	00-U00363_E34-1.txt	86	E34-1

The columns `cloneanno$spot1`, `cloneanno$spot2` are of class `numeric`, with values from 1 to 8704. They refer to the rows of `spotannoanno`.

The column `spotanno$probe` is of class `numeric`, with values from 1 to 4224, referring to the rows of `cloneanno`.

## References

- [1] Gene expression in kidney cancer is associated with novel tumor subtypes, cytogenetic abnormalities and metastasis formation. Holger Suelthmann, Anja von Heydebreck, Wolfgang Huber, Ruprecht Kuner, Andreas Buness, Markus Vogt, Bastian Gunawan, Martin Vingron, Laszlo Fuzesi, and Annemarie Poustka (Division of Molecular Genome Analysis, German Cancer Research Center, Heidelberg). *Submitted 2004*.