

# Package ‘miRcomp’

May 14, 2024

**Version** 1.35.0

**Date** 2017-03-04

**Title** Tools to assess and compare miRNA expression estimation methods

**Description** Based on a large miRNA dilution study, this package provides tools to read in the raw amplification data and use these data to assess the performance of methods that estimate expression from the amplification curves.

**Author** Matthew N. McCall <mccallm@gmail.com>, Lauren Kemperman <lkemperm@u.rochester.edu>

**Maintainer** Matthew N. McCall <mccallm@gmail.com>

**Depends** R (>= 3.2), Biobase (>= 2.22.0), miRcompData

**Imports** utils, methods, graphics, KernSmooth, stats

**VignetteBuilder** knitr

**Suggests** BiocStyle, knitr, rmarkdown, RUnit, BiocGenerics, shiny

**License** GPL-3 | file LICENSE

**biocViews** Software, qPCR, Preprocessing, QualityControl

**NeedsCompilation** no

**git\_url** <https://git.bioconductor.org/packages/miRcomp>

**git\_branch** devel

**git\_last\_commit** 54a639a

**git\_last\_commit\_date** 2024-04-30

**Repository** Bioconductor 3.20

**Date/Publication** 2024-05-13

## Contents

accuracy . . . . .	2
completeFeatures . . . . .	3
expressionComp . . . . .	4

lifetech . . . . .	5
limitOfDetection . . . . .	5
miRcompShinyApp . . . . .	6
precision . . . . .	7
qpcRb4 . . . . .	8
qpcRb5 . . . . .	8
qpcRdefault . . . . .	9
qpcR15 . . . . .	9
qpcRlinexp . . . . .	10
qualityAssessment . . . . .	10
titrationResponse . . . . .	11

## Index 13

---

accuracy	<i>Assess the accuracy of the expression estimates</i>
----------	--

---

### Description

This function assesses the signal detect slope (a measure of accuracy) for each feature.

### Usage

```
accuracy(object1, qcThreshold1, object2=NULL, qcThreshold2=NULL,
commonFeatures=TRUE, bins=3, label1=NULL, label2=NULL)
```

### Arguments

object1	a list containing two elements: ct (the expression estimates) and qc (quality scores)
qcThreshold1	a numeric threshold corresponding to object1\$qc below which values are considered low quality.
object2	an optional second list of the same format as object1, used to compare two methods.
qcThreshold2	a numeric threshold corresponding to object2\$qc below which values are considered low quality.
commonFeatures	if TRUE and object2 is non-NULL, only high quality non-NA features in common between both objects are used.
bins	the number of bins to divide the data into.
label1	optional label corresponding to object 1 to be used in plotting.
label2	optional label corresponding to object 2 to be used in plotting.

### Value

A plot of signal detect slopes stratified by difference in pure sample expression is produced, and a summary (median and MAD) of the signal detect slopes in each bin is returned.

**Author(s)**

Matthew N. McCall

**Examples**

```
data(lifetech)
accuracy(object1=lifetech, qcThreshold1=1.25)
data(qpcRdefault)
accuracy(object1=lifetech, qcThreshold1=1.25,
         object2=qpcRdefault, qcThreshold2=0.99)
```

---

completeFeatures

*Determine the Number of Completely Observed Features*

---

**Description**

This function determines the number of features that are good quality and non-NA across all samples using a given quality threshold.

**Usage**

```
completeFeatures(object1, qcThreshold1, object2=NULL, qcThreshold2=NULL,
                 label1=NULL, label2=NULL)
```

**Arguments**

object1	a list containing two elements: ct (the expression estimates) and qc (quality scores)
qcThreshold1	a numeric threshold corresponding to object1\$qc below which values are considered low quality.
object2	an optional second list of the same format as object1, used to compare two methods.
qcThreshold2	a numeric threshold corresponding to object2\$qc below which values are considered low quality.
label1	optional label corresponding to object 1 to be used in plotting.
label2	optional label corresponding to object 2 to be used in plotting.

**Value**

The function generates a table of the number of complete, partial, and absent features.

**Author(s)**

Matthew N. McCall

**Examples**

```

data(lifetech)
completeFeatures(object1=lifetech, qcThreshold1=1.25)
data(qpcRdefault)
completeFeatures(object1=lifetech, qcThreshold1=1.25,
                 object2=qpcRdefault, qcThreshold2=0.99)

```

---

expressionComp

*Compare expression estimates between two methods*


---

**Description**

This function allows one to easily compare the expression estimates generated by two different algorithms. Feature / sample combinations that differ above a given threshold can be easily highlighted.

**Usage**

```

expressionComp(object1, qcThreshold1,
               object2, qcThreshold2,
               label1=NULL, label2=NULL,
               diffThreshold=2,
               plotType=c("scatterplot", "MAplot"))

```

**Arguments**

object1	a list containing two elements: ct (the expression estimates) and qc (quality scores)
qcThreshold1	a numeric threshold corresponding to object1\$qc below which values are considered low quality.
object2	a list containing two elements: ct (the expression estimates) and qc (quality scores)
qcThreshold2	a numeric threshold corresponding to object2\$qc below which values are considered low quality.
label1	an optional label for plotting on the x-axis
label2	an optional label for plotting on the y-axis
diffThreshold	feature / sample combinations for which the expression estimates differ by more than this value are returned by the function and highlighted in the plot.
plotType	scatterplot or MA-plot

**Value**

This function plots the expression estimates produced by two different methods. Feature / sample combinations for which the expression estimates differ by more than diffThreshold are returned by the function and highlighted in the plot.

**Author(s)**

Matthew N. McCall

**Examples**

```
data(lifetech)
data(qpcRdefault)
tmp <- expressionComp(object1=lifetech, qcThreshold1=1.25,
object2=qpcRdefault, qcThreshold2=0.99, plotType="scatter")
```

---

lifetech

*The processed data generated using the LifeTech software.*

---

**Description**

These data are also used to demonstrate the functionality of the miRcomp package.

**Usage**

```
data(lifetech)
```

**Format**

A list with two elements.

```
ct  a miRNA x sample matrix of expression estimates
qc  a miRNA x sample matrix of quality scores (here AmpScores)
```

**Examples**

```
data(lifetech)
```

---

limitOfDetection

*Assess the limit of detection of a given method*

---

**Description**

This function assesses the limit of detection in one of two ways: (1) the distribution of expression estimates stratified by the proportion of poor quality values within replicates, (2) the average vs expected expression for the two most diluted sample types.

**Usage**

```
limitOfDetection(object, qcThreshold, plotType=c("boxplot", "scatterplot", "MAplot"))
```

**Arguments**

object	a list containing two elements: ct (the expression estimates) and qc (quality scores)
qcThreshold	a numeric threshold corresponding to object1\$qc below which values are considered low quality.
plotType	the desired output type – boxplot is option (1); scatterplot is option (2), an MAplot is option (3).

**Value**

This function assesses the limit of detection in several ways. If plotType is boxplot, then boxplots of expression estimates stratified by the proportion of poor quality values within replicates is displayed. The function also outputs a list with the values plotted in each box of the boxplot. If plotType is scatterplot, then the average within replicate expression vs expected expression (based on pure sample expression) is displayed for the 0.1/0.1 dilution and 0.01/0.01 dilution. If plotType is MAplot, then the difference in expression (average within replicate expression - expected expression) is displayed for the 0.1/0.1 dilution and 0.01/0.01 dilution. For both plotTypes, scatterplot and MAplot, the function outputs a matrix containing estimates of the limit of detection for four different tolerances. Specifically, the two columns correspond to the two dilutions (0.1/0.1 and 0.01/0.01) and rows correspond to the median difference between the observed and expected values. The values in the matrix are the expected expression values such that the median absolute difference of all larger expected expression values is approximately equal to the given tolerance.

**Author(s)**

Matthew N. McCall

**Examples**

```
data(lifetech)
tmp <- limitOfDetection(object=lifetech, qcThreshold=1.25)
data(qpcRdefault)
limitOfDetection(object=qpcRdefault, qcThreshold=0.99, plotType="scatter")
```

---

miRcompShinyApp

*Launch a Shiny app to use the miRcomp package interactively*

---

**Description**

This function launches the miRcomp Shiny app locally.

**Usage**

```
miRcompShinyApp()
```

**Author(s)**

Matthew N. McCall and Lauren Kemperman

**Examples**

```
## miRcompShinyApp()
```

---

precision	<i>Assess the precision of the expression estimates</i>
-----------	---

---

**Description**

This function assesses the within-replicate precision for each feature.

**Usage**

```
precision(object1, qcThreshold1, object2=NULL, qcThreshold2=NULL,
          commonFeatures=TRUE, statistic=c("sd", "cv"),
          scale=c("none", "log", "log10"), bins=3,
          label1=NULL, label2=NULL)
```

**Arguments**

object1	a list containing two elements: ct (the expression estimates) and qc (quality scores)
qcThreshold1	a numeric threshold corresponding to object1\$qc below which values are considered low quality.
object2	an optional second list of the same format as object1, used to compare two methods.
qcThreshold2	a numeric threshold corresponding to object2\$qc below which values are considered low quality.
commonFeatures	if TRUE and object2 is non-NULL, only high quality non-NA features in common between both objects are used.
statistic	whether to compute the standard deviation (sd) or coefficient of variation (cv).
scale	optional scaling of the values. This can help with visualizing the distributions.
bins	the number of bins to divide the data into.
label1	optional label corresponding to object 1 to be used in plotting.
label2	optional label corresponding to object 2 to be used in plotting.

**Value**

A boxplot of either the standard deviation or coefficient of variation stratified by expression is produced. The values plotted in each box of the boxplot are returned.

**Author(s)**

Matthew N. McCall

**Examples**

```
data(lifetech)
tmp1 <- precision(object1=lifetech,qcThreshold1=1.25)
data(qpcRdefault)
tmp2 <- precision(object1=lifetech,qcThreshold1=1.25,
                  object2=qpcRdefault,qcThreshold2=0.99)
```

---

qpcRb4

*The processed data generated using the 4 parameter sigmoidal method from the qpcR software.*

---

**Description**

These data are also used to demonstrate the functionality of the miRcomp package.

**Usage**

```
data(qpcRb4)
```

**Format**

A list with two elements.

```
ct  a miRNA x sample matrix of expression estimates
qc  a miRNA x sample matrix of quality scores (here R-squared values)
```

**Examples**

```
data(qpcRb4)
```

---

qpcRb5

*The processed data generated using the 5 parameter sigmoidal method from the qpcR software.*

---

**Description**

These data are also used to demonstrate the functionality of the miRcomp package.

**Usage**

```
data(qpcRb5)
```



**Format**

A list with two elements.

- ct a miRNA x sample matrix of expression estimates
- qc a miRNA x sample matrix of quality scores (here R-squared values)

**Examples**

```
data(qpcRb5)
```

---

qpcRdefault	<i>The processed data generated using the default method (4 parameter log-logistic) implemented in the qpcR software package.</i>
-------------	---

---

**Description**

These data are also used to demonstrate the functionality of the miRcomp package.

**Usage**

```
data(qpcRdefault)
```

**Format**

A list with two elements.

- ct a miRNA x sample matrix of expression estimates
- qc a miRNA x sample matrix of quality scores (here R-squared values)

**Examples**

```
data(qpcRdefault)
```

---

qpcR15	<i>The processed data generated using the 5 parameter log-logistic method from the qpcR software.</i>
--------	---

---

**Description**

These data are also used to demonstrate the functionality of the miRcomp package.

**Usage**

```
data(qpcR15)
```

**Format**

A list with two elements.

```
ct  a miRNA x sample matrix of expression estimates
qc  a miRNA x sample matrix of quality scores (here R-squared values)
```

**Examples**

```
data(qpcR15)
```

---

qpcR1inexp	<i>The processed data generated using the linear-exponential method implemented in the qpcR software package.</i>
------------	---

---

**Description**

These data are also used to demonstrate the functionality of the miRcomp package.

**Usage**

```
data(qpcR1inexp)
```

**Format**

A list with two elements.

```
ct  a miRNA x sample matrix of expression estimates
qc  a miRNA x sample matrix of quality scores (here R-squared values)
```

**Examples**

```
data(qpcR1inexp)
```

---

qualityAssessment	<i>Quality Assessment for MicroRNA Expression Estimates</i>
-------------------	---

---

**Description**

This function produces several figures to examine quality scores.

**Usage**

```
qualityAssessment(object1, object2=NULL, cloglog1=FALSE, cloglog2=FALSE,
  na.rm=FALSE, plotType=c("scatterplot", "boxplot"),
  label1=NULL, label2=NULL)
```

**Arguments**

object1	a list containing two elements: ct (the expression estimates) and qc (quality scores)
object2	an optional second list of the same format as object1, used to compare two methods.
cloglog1	if TRUE, the $-\log(-\log(\text{object1}\$qc))$ is plotted. This is useful to visualize certain quality scores such as R-squared.
cloglog2	if TRUE, the $-\log(-\log(\text{object2}\$qc))$ is plotted. This is useful to visualize certain quality scores such as R-squared.
na.rm	if TRUE, quality scores corresponding to NA expression estimates are removed from analysis. This is only used if plotType=="boxplot".
plotType	type of plot to return. Options are a scatterplot or a boxplot.
label1	optional label corresponding to object 1 to be used in plotting.
label2	optional label corresponding to object 2 to be used in plotting.

**Value**

The function generates a plot of the desired type.

**Author(s)**

Matthew N. McCall

**Examples**

```
data(lifetech)
qualityAssessment(lifetech, plotType="boxplot")

data(qpcRdefault)
qualityAssessment(object1=lifetech,object2=qpcRdefault,cloglog2=TRUE)
```

---

titrationResponse      *Assess monotone signal across titration*

---

**Description**

This function determines

**Usage**

```
titrationResponse(object1, qcThreshold1, object2=NULL, qcThreshold2=NULL,
                  commonFeatures=TRUE, label1=NULL, label2=NULL)
```

**Arguments**

object1	a list containing two elements: ct (the expression estimates) and qc (quality scores)
qcThreshold1	a numeric threshold corresponding to object1\$qc below which values are considered low quality.
object2	an optional second list of the same format as object1, used to compare two methods.
qcThreshold2	a numeric threshold corresponding to object2\$qc below which values are considered low quality.
commonFeatures	if TRUE and object2 is non-NULL, only high quality non-NA features in common between both objects are used.
label1	optional label corresponding to object 1 to be used in plotting.
label2	optional label corresponding to object 2 to be used in plotting.

**Value**

A table listing the number of features showing a monotone titration response and a figure showing the proportion of features showing a monotone titration response vs the difference in pure sample expression.

**Author(s)**

Matthew N. McCall

**Examples**

```
data(lifetech)
titrationResponse(object1=lifetech, qcThreshold1=1.25)
data(qpcRdefault)
titrationResponse(object1=lifetech, qcThreshold1=1.25,
                  object2=qpcRdefault, qcThreshold2=0.99)
```

# Index

## \* datasets

- lifetech, [5](#)
- qpcRb4, [8](#)
- qpcRb5, [8](#)
- qpcRdefault, [9](#)
- qpcRl5, [9](#)
- qpcRlinexp, [10](#)

## \* manip

- accuracy, [2](#)
- completeFeatures, [3](#)
- expressionComp, [4](#)
- limitOfDetection, [5](#)
- miRcompShinyApp, [6](#)
- precision, [7](#)
- qualityAssessment, [10](#)
- titrationResponse, [11](#)

[accuracy, 2](#)

[completeFeatures, 3](#)

[expressionComp, 4](#)

[lifetech, 5](#)

[limitOfDetection, 5](#)

[miRcompShinyApp, 6](#)

[precision, 7](#)

[qpcRb4, 8](#)

[qpcRb5, 8](#)

[qpcRdefault, 9](#)

[qpcRl5, 9](#)

[qpcRlinexp, 10](#)

[qualityAssessment, 10](#)

[titrationResponse, 11](#)