

# Package ‘iCheck’

May 12, 2024

**Type** Package

**Title** QC Pipeline and Data Analysis Tools for High-Dimensional  
Illumina mRNA Expression Data

**Version** 1.34.0

**Date** 2016-11-17

**Author** Weiliang Qiu [aut, cre],  
Brandon Guo [aut, ctb],  
Christopher Anderson [aut, ctb],  
Barbara Klanderman [aut, ctb],  
Vincent Carey [aut, ctb],  
Benjamin Raby [aut, ctb]

**Maintainer** Weiliang Qiu <stwxq@channing.harvard.edu>

**Depends** R (>= 3.2.0), Biobase, lumi, gplots

**Imports** stats, graphics, preprocessCore, grDevices, randomForest,  
affy, limma, parallel, GeneSelectMMD, rgl, MASS, lmtest,  
scatterplot3d, utils

**biocViews** GeneExpression, DifferentialExpression, Microarray,  
Preprocessing, DNAMethylation, OneChannel, TwoChannel,  
QualityControl

**Description** QC pipeline and data analysis tools for high-dimensional  
Illumina mRNA expression data.

**License** GPL (>= 2)

**NeedsCompilation** no

**git\_url** <https://git.bioconductor.org/packages/iCheck>

**git\_branch** RELEASE\_3\_19

**git\_last\_commit** 4e9fb55

**git\_last\_commit\_date** 2024-04-30

**Repository** Bioconductor 3.19

**Date/Publication** 2024-05-12

## Contents

|                                  |           |
|----------------------------------|-----------|
| boxPlots . . . . .               | 2         |
| densityPlots . . . . .           | 4         |
| genExprSet . . . . .             | 6         |
| genSimData.BayesNormal . . . . . | 7         |
| getPCAFunc . . . . .             | 9         |
| glmWrapper . . . . .             | 10        |
| lkhrrWrapper . . . . .           | 13        |
| lmFitPaired . . . . .            | 15        |
| lmFitWrapper . . . . .           | 18        |
| LumiBatch2Table . . . . .        | 21        |
| pca2DPlot . . . . .              | 22        |
| pca3DPlot . . . . .              | 24        |
| plotCurves . . . . .             | 27        |
| plotQCCurves . . . . .           | 28        |
| plotSamplep95p05 . . . . .       | 31        |
| quantilePlot . . . . .           | 34        |
| R2PlotFunc . . . . .             | 36        |
| scatterPlots . . . . .           | 38        |
| sortExpressionSet . . . . .      | 41        |
| <b>Index</b>                     | <b>43</b> |

---

boxPlots

*Draw parallel plots for top results in whole-genome-wide analysis*


---

## Description

Draw scatter plots for top results in whole-genome-wide analysis to test for the association of probes to a continuous-type phenotype variable.

## Usage

```
boxPlots(
  resFrame,
  es,
  col.resFrame = c("probeIDs", "stats", "pval", "p.adj"),
  var.pheno = "sex",
  var.probe = "TargetID",
  var.gene = "Symbol",
  var.chr = "Chr",
  nTop = 20,
  myylab = "expression level",
  datExtrFunc = exprs,
  fileFlag = FALSE,
  fileFormat = "ps",
  fileName = "boxPlots.ps")
```

**Arguments**

|                           |  |
|---------------------------|--|
| <code>resFrame</code>     | A data frame stores testing results, which must contain columns that indicate probe id, test statistic, p-value and optionally adjusted p-value.   |
| <code>es</code>           | An ExpressionSet object that used to run the whole genome-wide tests.  |
| <code>col.resFrame</code> | A vector of characters indicating column names of <code>resFrame</code> corresponding to probe id, test statistic, p-value and optionally adjusted p-value.  |
| <code>var.pheno</code>    | character. the name of continuous-type phenotype variable that is used to test the association of this variable to probes.   |
| <code>var.probe</code>    | character. the name of feature variable indicating probe id.   |
| <code>var.gene</code>     | character. the name of feature variable indicating gene symbol.  |
| <code>var.chr</code>      | character. the name of feature variable indicating chromosome number.  |
| <code>nTop</code>         | integer. indicating how many top tests will be used to draw the scatter plot.  |
| <code>myylab</code>       | character. indicating y-axis label.  |
| <code>datExtrFunc</code>  | name of the function to extract genomic data. For an ExpressionSet object, you should set <code>datExtrFunc=exprs</code> ; for a MethyLumiSet object, you should set <code>datExtrFunc=betas</code> .                        |
| <code>fileFlag</code>     | logic. indicating if plot should be saved to an external figure file.  |
| <code>fileFormat</code>   | character. indicating the figure file type. Possible values are “ps”, “pdf”, or “jpeg”. All other values will produce “png” file.  |
| <code>fileName</code>     | character. indicating figure file name (file extension should be specified). For example, you set <code>fileFormat="pdf"</code> , then you can set <code>fileName="test.pdf"</code> , but not <code>fileName="test"</code> . |

**Value**

Value 0 will be returned if no error occurs.

**Author(s)**

Weiliang Qiu <stwxq@channing.harvard.edu>, Brandon Guo <brandowonder@gmail.com>, Christopher Anderson <christopheranderson84@gmail.com>, Barbara Klanderman <BKLANDERMAN@partners.org>, Vincent Carey <stvj@channing.harvard.edu>, Benjamin Raby <rebar@channing.harvard.edu>

**Examples**

```
# generate simulated data set from conditional normal distribution
set.seed(1234567)
es.sim = genSimData.BayesNormal(nCpGs = 100,
  nCases = 20, nControls = 20,
  mu.n = -2, mu.c = 2,
  d0 = 20, s02 = 0.64, s02.c = 1.5, testPara = "var",
  outlierFlag = FALSE,
  eps = 1.0e-3, applier = lapply)
print(es.sim)

res.limma = lmFitWrapper(
```

```

es = es.sim,
formula = ~as.factor(memSubj),
pos.var.interest = 1,
pvalAdjMethod = "fdr",
alpha = 0.05,
probeID.var = "probe",
gene.var = "gene",
chr.var = "chr",
verbose = TRUE)

boxPlots(
  resFrame=res.limma$frame,
  es=es.sim,
  col.resFrame = c("probeIDs", "stats", "pval"),
  var.pheno = "memSubj",
  var.probe = "probe",
  var.gene = "gene",
  var.chr = "chr",
  nTop = 20,
  myylab = "expression level",
  datExtrFunc = exprs,
  fileFlag = FALSE,
  fileFormat = "ps",
  fileName = "boxPlots.ps")

```

---

densityPlots

*Draw estimated density plots for all arrays*


---

## Description

Draw estimated density plots for all arrays.

## Usage

```

densityPlots(
  es,
  requireLog2 = TRUE,
  myxlab = "expression level",
  mymain = "density plots",
  datExtrFunc = exprs,
  fileFlag = FALSE,
  fileFormat = "ps",
  fileName = "densityPlots.ps")

```

## Arguments

|             |   |
|-------------|---|
| es          | An ExpressionSet object that used to run the whole genome-wide tests.             |
| requireLog2 | logic. indicating if log2 transformation is required before estimating densities. |

|             |  |
|-------------|--|
| myxlab      | character. indicating x-axis label.  |
| mymain      | character. indicating title of the plot.   |
| datExtrFunc | name of the function to extract genomic data. For an ExpressionSet object, you should set datExtrFunc=exprs; for a MethyLumiSet object, you should set datExtrFunc=betas.          |
| fileFlag    | logic. indicating if plot should be saved to an external figure file.  |
| fileFormat  | character. indicating the figure file type. Possible values are “ps”, “pdf”, or “jpeg”. All other values will produce “png” file.  |
| fileName    | character. indicating figure file name (file extension should be specified). For example, you set fileFormat=“pdf”, then you can set fileName=“test.pdf”, but not fileName=“test”. |

### Value

A list object, the  $i$ -th element is the object returned by function density for the  $i$ -th array.

### Author(s)

Weiliang Qiu <stwxq@channing.harvard.edu>, Brandon Guo <brandowonder@gmail.com>, Christopher Anderson <christopheranderson84@gmail.com>, Barbara Klanderman <BKLANDERMAN@partners.org>, Vincent Carey <stvjc@channing.harvard.edu>, Benjamin Raby <rebar@channing.harvard.edu>

### Examples

```
# generate simulated data set from conditional normal distribution
set.seed(1234567)
es.sim = genSimData.BayesNormal(nCpGs = 100,
  nCases = 20, nControls = 20,
  mu.n = -2, mu.c = 2,
  d0 = 20, s02 = 0.64, s02.c = 1.5, testPara = "var",
  outlierFlag = FALSE,
  eps = 1.0e-3, applier = lapply)
print(es.sim)

densityPlots(
  es = es.sim,
  requireLog2 = FALSE,
  myxlab = "expression level",
  mymain = "density plots",
  datExtrFunc = exprs,
  fileFlag = FALSE,
  fileFormat = "ps",
  fileName = "densityPlots.ps")
```

---

|            |   |
|------------|---|
| genExprSet | <i>Generate an ExpressionSet object</i> |
|------------|---|

---

## Description

Generate a simple ExpressionSet object.

## Usage

```
genExprSet(  
  ex,  
  pDat,  
  fDat = NULL,  
  annotation = "lumiHumanAll.db")
```

## Arguments

|            |  |
|------------|--|
| ex         | A matrix of expression levels. Rows are gene probes and columns are arrays.  |
| pDat       | A data frame describing arrays. Rows are arrays and columns are variables describing arrays. The row names of pDat must be the same as the column of ex.         |
| fDat       | A data frame describing gene probes. Rows are gene probes and columns are variables describing gene probes. The rownames of fDat must be the same as that of ex. |
| annotation | character string. Indicating the annotation library (e.g. lumiHumanAll.db for the gene probes.   |

## Value

an ExpressionSet object.

## Author(s)

Weiliang Qiu <stwxq@channing.harvard.edu>, Brandon Guo <brandowonder@gmail.com>, Christopher Anderson <christopheranderson84@gmail.com>, Barbara Klanderman <BKLANDERMAN@partners.org>, Vincent Carey <stvjc@channing.harvard.edu>, Benjamin Raby <rebar@channing.harvard.edu>

---

genSimData.BayesNormal

*Generating simulated data set from conditional normal distributions*


---

## Description

Generating simulated data set from conditional normal distributions.

## Usage

```
genSimData.BayesNormal(
  nCpGs,
  nCases,
  nControls,
  mu.n = -2,
  mu.c = 2,
  d0 = 20,
  s02 = 0.64,
  s02.c = 1.5,
  testPara = "var",
  outlierFlag = FALSE,
  eps = 0.001,
  applier = lapply)
```

## Arguments

|             |   |
|-------------|---|
| nCpGs       | integer. Number of genes.   |
| nCases      | integer. Number of cases.   |
| nControls   | integer. Number of controls.  |
| mu.n        | numeric. mean of the conditional normal distribution for controls. See details.   |
| mu.c        | numeric. mean of the conditional normal distribution for cases. See details.  |
| d0          | integer. degree of freedom for scale-inverse chi squared distribution. See details.   |
| s02         | numeric. scaling parameter for scale-inverse chi squared distribution for controls. See details.  |
| s02.c       | numeric. scaling parameter for scale-inverse chi squared distribution for cases. See details.   |
| testPara    | character string. indicating if the test is for testing equal mean, equal variance, or both.  |
| outlierFlag | logical. indicating if outliers would be generated. If outlierFlag=TRUE, then we followed Phipson and Oshlack's (2014) simulation studies to generate one outlier for each CpG site by replacing the DNA methylation level of one diseased subject by the maximum of the DNA methylation levels of all CpG sites. |

|         |  |
|---------|--|
| eps     | numeric. if $ mean0 - mean1  < eps$ then we regard $mean0 = mean1$ . Similarly, if $ var0 - var1  < eps$ then we regard $var0 = var1$ . $mean0$ and $var0$ are the mean and variance of the chi squared distribution for controls. $mean1$ and $var1$ are the mean and variance of the chi squared distribution for cases. |
| applier | function name to do apply operation.   |

## Details

Based on Phipson and Oshlack's (2014) simulation algorithm. For each CpG site, variance of the DNA methylation was first sampled from a scaled inverse chi-squared distribution with degree of freedom  $d_0$  and scaling parameter  $s_0^2$ :  $\sigma_i^2 \text{ scale} - inv\chi^2(d_0, s_0^2)$ . M value for each CpG was then sampled from a normal distribution with mean  $\mu_n$  and variance equal to the simulated variance  $\sigma_i^2$ . For cases, the variance was first generated from  $\sigma_{i,c}^2 \text{ scale} - inv\chi^2(d_0, s_{0,c}^2)$ . M value for each CpG was then sampled from a normal distribution with mean  $\mu_c$  and variance equal to the simulated variance  $\sigma_{i,c}^2$ .

## Value

An ExpressionSet object. The phenotype data of the ExpressionSet object contains 2 columns: arrayID (array id) and memSubj (subject membership, i.e., case (memSubj=1) or control (memSubj=0)). The feature data of the ExpressionSet object contains 4 elements: probe (probe id), gene (psuedo gene symbol), chr (psuedo chromosome number), and memGenes (indicating if a gene is differentially expressed (when testPara="mean") or indicating if a gene is differentially variable (when testPara="var") ).

## Author(s)

Weiliang Qiu <stwxq@channing.harvard.edu>, Brandon Guo <brandowonder@gmail.com>, Christopher Anderson <christopheranderson84@gmail.com>, Barbara Klanderman <BKLANDERMAN@partners.org>, Vincent Carey <stvjc@channing.harvard.edu>, Benjamin Raby <rebar@channing.harvard.edu>

## References

Phipson B, Oshlack A. DiffVar: A new method for detecting differential variability with application to methylation in cancer and aging. *Genome Biol* 2014; 15:465

## Examples

```
# generate simulated data set from conditional normal distribution
set.seed(1234567)
es.sim = genSimData.BayesNormal(nCpGs = 100,
  nCases = 20, nControls = 20,
  mu.n = -2, mu.c = 2,
  d0 = 20, s02 = 0.64, s02.c = 1.5, testPara = "var",
  outlierFlag = FALSE,
  eps = 1.0e-3, applier = lapply)
print(es.sim)
```



getPCAFunc

*Get principal components of arrays***Description**

Get principal components of arrays.

**Usage**

```
getPCAFunc(es,
            labelVariable = "subjID",
            hybName = "Hybridization_Name",
            requireLog2 = TRUE,
            corFlag = FALSE
          )
```

**Arguments**

|               |  |
|---------------|--|
| es            | An ExpressionSet object  |
| labelVariable | A character string. The name of a phenotype data variable use to label the arrays in the boxplots. By default, labelVariable = "subjID" which is equivalent to labelVariable = "Hybridization_Name". |
| hybName       | character string. indicating the phenotype variable Hybridization_Name.  |
| requireLog2   | logical. requiredLog2=TRUE indicates probe expression levels will be log2 transformed. Otherwise, no transformation will be performed.   |
| corFlag       | logical. Indicating if correlation matrix (corFlag=TRUE) or covariance (corFlag=FALSE) is used to obtain principal components.   |

**Value**

A list with 3 elements:

|             |   |
|-------------|---|
| es.s        | An ExpressionSet object with the arrays sorted according to Batch_Run_Date, Chip_Barcode, and Chip_Address  |
| pcs         | An object returned by the function prcomp of the R package stats. It contains the following components. sdev (the square roots of the eigenvalues of the covariance/correlation matrix); rotation (a matrix whose columns contain the eigenvectors); x (a matrix whose columns contain principal components); center (the centering used or FALSE); scale (the scale used or FALSE) |
| requireLog2 | logical. The same value as the input requireLog2.   |

**Author(s)**

Weiliang Qiu <stwxq@channing.harvard.edu>, Brandon Guo <brandowonder@gmail.com>, Christopher Anderson <christopheranderson84@gmail.com>, Barbara Klanderman <BKLANDERMAN@partners.org>, Vincent Carey <stvjc@channing.harvard.edu>, Benjamin Raby <rebar@channing.harvard.edu>

## Examples

```
# generate simulated data set from conditional normal distribution
set.seed(1234567)
es.sim = genSimData.BayesNormal(nCpGs = 100,
  nCases = 20, nControls = 20,
  mu.n = -2, mu.c = 2,
  d0 = 20, s02 = 0.64, s02.c = 1.5, testPara = "var",
  outlierFlag = FALSE,
  eps = 1.0e-3, applier = lapply)
print(es.sim)

pca.obj = getPCAFunc(es = es.sim,
  labelVariable = "subjID",
  hybName = "memSubj",
  requireLog2 = FALSE,
  corFlag = FALSE
)
```

---

glmWrapper

---

*Perform glm test for all gene probes*


---

## Description

Perform glm test for all gene probes.

## Usage

```
glmWrapper(es,
  formula = FEV1 ~ xi + age + gender,
  pos.var.interest = 1,
  family = gaussian,
  logit = FALSE,
  pvalAdjMethod = "fdr",
  alpha = 0.05,
  probeID.var = "ProbeID",
  gene.var = "Symbol",
  chr.var = "Chromosome",
  applier = lapply,
  verbose = TRUE)
```

## Arguments

|         |  |
|---------|--|
| es      | An LumiBatch object. fData(es) should contains information about probe ID, chromosome number and gene symbol.  |
| formula | An object of class formula. The left handside of ~ is the response variable. Gene probe must be represented by the variable xi. For example, xi~age+gender (gene probe is the response variable); Or FEV1~xi+age+gender (gene probe is the predictor). |

|                               |   |
|-------------------------------|---|
| <code>pos.var.interest</code> | integer. Indicates which covariate in the right-hand-side of <code>~</code> of formula is of the interest. <code>pos.var.interest = 0</code> means the intercept is of the interest. If the covariate of the interest is an factor or interaction term with more than 2 levels, the smallest p-value will represent the pvalue for the covariate of the interest. |
| <code>family</code>           | By default is gaussian. refer to <a href="#">glm</a> .  |
| <code>logit</code>            | logical. Indicate if the gene probes will be logit transformed. For example, for DNA methylation data, one might want to logit transformation for the beta-value ( $methyalted/(methyalted + unmethyalted)$ ).  |
| <code>pvalAdjMethod</code>    | One of p-value adjustment methods provided by the R function <code>p.adjust</code> in R package <code>stats</code> : “holm”, “hochberg”, “hommel”, “bonferroni”, “BH”, “BY”, “fdr”, “none”.   |
| <code>alpha</code>            | Significance level. A test is claimed to be significant if the adjusted p-value < alpha.  |
| <code>probeID.var</code>      | character string. Name of the variable indicating probe ID in feature data set.   |
| <code>gene.var</code>         | character string. Name of the variable indicating gene symbol in feature data set.  |
| <code>chr.var</code>          | character string. Name of the variable indicating chromosome number in feature data set.  |
| <code>applier</code>          | By default, it is <code>lapply</code> . If the library <code>multicore</code> is available, can use <code>mclapply</code> to replace <code>lapply</code> .  |
| <code>verbose</code>          | logical. Determine if intermediate output need to be suppressed. By default <code>verbose=TRUE</code> , intermediate output will be printed.  |

## Details

This function applies R function `glm` for each gene probe.

## Value

A list with the following elements:

|                      |  |
|----------------------|--|
| <code>n.sig</code>   | Number of significant tests after p-value adjustment.  |
| <code>frame</code>   | A data frame containing test results sorted according to the ascending order of unadjusted p-values for the covariate of the interest. The data frame contains 7 columns: <code>probeIDs</code> , <code>geneSymbols</code> (gene symbols of the genes where the probes come from), <code>chr</code> (numbers of chromosomes where the probes locate), <code>stats</code> (z-value), <code>pval</code> (p-values of the tests for the covariate of the interest), <code>p.adj</code> (adjusted p-values), <code>pos</code> (row numbers of the probes in the expression data matrix). |
| <code>statMat</code> | A matrix containing test statistics for all covariates and for all probes. Rows are probes and columns are covariates. The rows are ordered according to the ascending order of unadjusted p-values for the covariate of the interest.   |
| <code>pvalMat</code> | A matrix containing pvalues for all covariates and for all probes. Rows are probes and columns are covariates. The rows are ordered according to the ascending order of unadjusted p-values for the covariate of the interest.   |

|                               |   |
|-------------------------------|---|
| <code>pval.quantile</code>    | Quantiles (minimum, 25 for each covariate including intercept provided in the input argument formula).  |
| <code>frame.unsorted</code>   | A data frame containing test results. The data frame contains 7 columns: <code>probeIDs</code> , <code>geneSymbols</code> (gene symbols of the genes where the probes come from), <code>chr</code> (numbers of chromosomes where the probes locate), <code>stats</code> (z-value for the covariate of the interest), <code>pval</code> (p-values of the tests for the covariate of the interest), <code>p.adj</code> (adjusted p-values), <code>pos</code> (row numbers of the probes in the expression data matrix). |
| <code>statMat.unsorted</code> | A matrix containing test statistics for all covariates and for all probes. Rows are probes and columns are covariates.  |
| <code>pvalMat.unsorted</code> | A matrix containing pvalues for all covariates and for all probes. Rows are probes and columns are covariates.  |
| <code>memGenes</code>         | A numeric vector indicating the cluster membership of probes (unsorted). <code>memGenes[i]=1</code> if the <i>i</i> -th probe is significant (adjusted pvalue < alpha) with positive z-value for the covariate of the interest; <code>memGenes[i]=2</code> if the <i>i</i> -th probe is nonsignificant ; <code>memGenes[i]=3</code> if the <i>i</i> -th probe is significant with negative z-value for the covariate of the interest;   |
| <code>memGenes2</code>        | A numeric vector indicating the cluster membership of probes (unsorted). <code>memGenes2[i]=1</code> if the <i>i</i> -th probe is significant (adjusted pvalue < alpha). <code>memGenes2[i]=0</code> if the <i>i</i> -th probe is nonsignificant.   |
| <code>mu1</code>              | Mean expression levels for arrays for probe cluster 1 (average taking across all probes with <code>memGenes</code> value equal to 1.  |
| <code>mu2</code>              | Mean expression levels for arrays for probe cluster 2 (average taking across all probes with <code>memGenes</code> value equal to 2.  |
| <code>mu3</code>              | Mean expression levels for arrays for probe cluster 3 (average taking across all probes with <code>memGenes</code> value equal to 3.  |
| <code>resMat</code>           | A matrix with $2p$ columns, where $p$ is the number of covariates (including intercept; for a nominal variable with 3 levels say, there were 2 dummy covariates). The first $p$ columns are p-values. The remaining $p$ columns are test statistics.  |

### Note

If the covariate of the interest is a factor or interaction term with more than 2 levels, then the p-value of the likelihood ratio test might be more appropriate than the smallest p-value for the covariate of the interest.

### Author(s)

Weiliang Qiu <stwxq@channing.harvard.edu>, Brandon Guo <brandowonder@gmail.com>, Christopher Anderson <christopheranderson84@gmail.com>, Barbara Klanderma <BKLANDERMAN@partners.org>, Vincent Carey <stvjc@channing.harvard.edu>, Benjamin Raby <rebar@channing.harvard.edu>

**Examples**

```
# generate simulated data set from conditional normal distribution
set.seed(1234567)
es.sim = genSimData.BayesNormal(nCpGs = 100,
  nCases = 20, nControls = 20,
  mu.n = -2, mu.c = 2,
  d0 = 20, s02 = 0.64, s02.c = 1.5, testPara = "var",
  outlierFlag = FALSE,
  eps = 1.0e-3, applier = lapply)
print(es.sim)

res.glm = glmWrapper(
  es = es.sim,
  formula = xi~as.factor(memSubj),
  pos.var.interest = 1,
  family = gaussian,
  logit = FALSE,
  pvalAdjMethod = "fdr",
  alpha = 0.05,
  probeID.var = "probe",
  gene.var = "gene",
  chr.var = "chr",
  applier = lapply,
  verbose = TRUE)
```

---

lkhRWrapper

---

*Perform glm test for all gene probes*


---

**Description**

Perform glm test for all gene probes.

**Usage**

```
lkhRWrapper(es,
  formulaReduced = FEV1 ~ xi + gender,
  formulaFull = FEV1 ~ xi + age + gender,
  family = gaussian,
  logit = FALSE,
  pvalAdjMethod = "fdr",
  alpha = 0.05,
  probeID.var = "ProbeID",
  gene.var = "Symbol",
  chr.var = "Chromosome",
  applier = lapply,
  verbose = TRUE)
```

## Arguments

|                             |   |
|-----------------------------|---|
| <code>es</code>             | An LumiBatch object. <code>fData(es)</code> should contains information about probe ID, chromosome number and gene symbol.  |
| <code>formulaReduced</code> | An object of class formula. Formula for reduced model. The left handside of <code>~</code> is the response variable. Gene probe must be represented by the variable <code>xi</code> . For example, <code>xi~gender</code> (gene probe is the response variable); Or <code>FEV1~xi+gender</code> (gene probe is the predictor).      |
| <code>formulaFull</code>    | An object of class formula. Formula for Full model. The left handside of <code>~</code> is the response variable. Gene probe must be represented by the variable <code>xi</code> . For example, <code>xi~age+gender</code> (gene probe is the response variable); Or <code>FEV1~xi+age+gender</code> (gene probe is the predictor). |
| <code>family</code>         | By default is gaussian. refer to <a href="#">glm</a> .  |
| <code>logit</code>          | logical. Indicate if the gene probes will be logit transformed. For example, for DNA methylation data, one might want to logit transformation for the beta-value ( $methy\!l\!a\!t\!e\!d/(methy\!l\!a\!t\!e\!d + unmethy\!l\!a\!t\!e\!d)$ ).  |
| <code>pvalAdjMethod</code>  | One of p-value adjustment methods provided by the R function <code>p.adjust</code> in R package <code>stats</code> : “holm”, “hochberg”, “hommel”, “bonferroni”, “BH”, “BY”, “fdr”, “none”.   |
| <code>alpha</code>          | Significance level. A test is claimed to be significant if the adjusted p-value < alpha.  |
| <code>probeID.var</code>    | character string. Name of the variable indicating probe ID in feature data set.   |
| <code>gene.var</code>       | character string. Name of the variable indicating gene symbol in feature data set.  |
| <code>chr.var</code>        | character string. Name of the variable indicating chromosome number in feature data set.  |
| <code>applier</code>        | By default, it is <code>lapply</code> . If the library <code>multicore</code> is available, can use <code>mclapply</code> to replace <code>lapply</code> .  |
| <code>verbose</code>        | logical. Determine if intermediate output need to be suppressed. By default <code>verbose=TRUE</code> , intermediate output will be printed.  |

## Details

This function applies R functions `lrtest` in R package `lmtest` and `glm` for each gene probe.

## Value

A list with the following elements:

|                             |   |
|-----------------------------|---|
| <code>frame</code>          | A data frame containing test results sorted according to the ascending order of unadjusted p-values for the covariate of the interest. The data frame contains 8 columns: <code>probeIDs</code> , <code>geneSymbols</code> (gene symbols of the genes where the probes come from), <code>chr</code> (numbers of chromosomes where the probes locate), <code>Chisq</code> (chi square test statistic), <code>Df</code> (degree of freedom of the chisquare test statistic), <code>pval</code> (p-values of the tests for the covariate of the interest), <code>p.adj</code> (adjusted p-values), <code>pos</code> (row numbers of the probes in the expression data matrix). The rows are ordered based on the descending order of chisquare test statistic. |
| <code>frame.unsorted</code> | A data frame containing test results. unordered frame.  |

**Author(s)**

Weiliang Qiu <stwxq@channing.harvard.edu>, Brandon Guo <brandowonder@gmail.com>, Christopher Anderson <christopheranderson84@gmail.com>, Barbara Klanderman <BKLANDERMAN@partners.org>, Vincent Carey <stvjc@channing.harvard.edu>, Benjamin Raby <rebar@channing.harvard.edu>

**Examples**

```
# generate simulated data set from conditional normal distribution
set.seed(1234567)
es.sim = genSimData.BayesNormal(nCpGs = 100,
  nCases = 20, nControls = 20,
  mu.n = -2, mu.c = 2,
  d0 = 20, s02 = 0.64, s02.c = 1.5, testPara = "var",
  outlierFlag = FALSE,
  eps = 1.0e-3, applier = lapply)
print(es.sim)

set.seed(1234567)
es.sim$age = rnorm(ncol(es.sim), mean=50, sd=5)
res.lkh = lkhrWrapper(
  es = es.sim,
  formulaReduced = xi ~ memSubj,
  formulaFull = xi ~ memSubj + age,
  family = gaussian(),
  logit = FALSE,
  pvalAdjMethod = "fdr",
  alpha = 0.05,
  probeID.var = "probe",
  gene.var = "gene",
  chr.var = "chr",
  applier = lapply,
  verbose = TRUE)
```

---

lmFitPaired

*A wrapper function for the function 'lmFit' of the R Bioconductor package 'limma' for paired data*

---

**Description**

A wrapper function for the function 'lmFit' of the R Bioconductor package 'limma' for paired data.

**Usage**

```
lmFitPaired(
  esDiff,
  formula = ~1,
  pos.var.interest = 0,
  pvalAdjMethod = "fdr",
```

```
alpha = 0.05,
probeID.var="ProbeID",
gene.var = "Symbol",
chr.var = "Chromosome",
verbose = TRUE)
```

## Arguments

|                               |  |
|-------------------------------|--|
| <code>esDiff</code>           | An <code>LumiBatch</code> object containing log2 difference between cases and controls. <code>fData(esDiff)</code> should contains information about probe ID, chromosome number and gene symbol.  |
| <code>formula</code>          | An object of class <code>formula</code> . The intercept measures the effect of treatment. Other covariates measure the effects of their interaction and treatment. The p-values for the intercept will be output. No left handside of <code>~</code> should be specified since the response variable will be the expression level. |
| <code>pos.var.interest</code> | integer. Indicates which covariate on the right-hand-side of <code>~</code> in <code>formula</code> is the covariate of the interest. By default, it is the intercept <code>pos.var.interest=0</code> .  |
| <code>pvalAdjMethod</code>    | One of p-value adjustment methods provided by the R function <code>p.adjust</code> in R package <code>stats</code> : "holm", "hochberg", "hommel", "bonferroni", "BH", "BY", "fdr", "none".  |
| <code>alpha</code>            | Significance level. A test is claimed to be significant if the adjusted p-value < alpha.   |
| <code>probeID.var</code>      | character string. Name of the variable indicating probe ID in feature data set.  |
| <code>gene.var</code>         | character string. Name of the variable indicating gene symbol in feature data set.   |
| <code>chr.var</code>          | character string. Name of the variable indicating chromosome number in feature data set.   |
| <code>verbose</code>          | logical. Determine if intermediate output need to be suppressed. By default <code>verbose=TRUE</code> , intermediate output will be printed.   |

## Details

This is a wrapper function of R Bioconductor functions `lmFit` and `eBayes` for paired data to make it easier to input design and output list of significant results.

## Value

A list with the following elements:

|                    |   |
|--------------------|---|
| <code>n.sig</code> | Number of significant tests after p-value adjustment.   |
| <code>frame</code> | A data frame containing test results sorted according to the ascending order of unadjusted p-values for the intercept. The data frame contains 7 columns: <code>probeIDs</code> , <code>geneSymbols</code> (gene symbols of the genes where the probes come from), <code>chr</code> (numbers of chromosomes where the probes locate), <code>stats</code> (moderated t-statistics for the intercept), <code>pval</code> (p-values of the tests for the intercept), <code>p.adj</code> (adjusted p-values), <code>pos</code> (row numbers of the probes in the expression data matrix). |



|                               |  |
|-------------------------------|--|
| <code>statMat</code>          | A matrix containing test statistics for all covariates and for all probes. Rows are probes and columns are covariates. The rows are ordered according to the ascending order of unadjusted p-values for the intercept.   |
| <code>pvalMat</code>          | A matrix containing pvalues for all covariates and for all probes. Rows are probes and columns are covariates. The rows are ordered according to the ascending order of unadjusted p-values for the intercept.   |
| <code>pval.quantile</code>    | Quantiles (minimum, 25 for all covariates including intercept provided in the input argument formula.  |
| <code>frame.unsorted</code>   | A data frame containing test results. The data frame contains 7 columns: probeIDs, geneSymbols (gene symbols of the genes where the probes come from), chr (numbers of chromosomes where the probes locate), stats (moderated t-statistics for the intercept), pval (p-values of the tests for the intercept), p.adj (adjusted p-values), pos (row numbers of the probes in the expression data matrix). |
| <code>statMat.unsorted</code> | A matrix containing test statistics for all covariates and for all probes. Rows are probes and columns are covariates.   |
| <code>pvalMat.unsorted</code> | A matrix containing pvalues for all covariates and for all probes. Rows are probes and columns are covariates.   |
| <code>memGenes</code>         | A numeric vector indicating the cluster membership of probes (unsorted). <code>memGenes[i]=1</code> if the <i>i</i> -th probe is significant (adjusted pvalue < alpha) with positive moderated t-statistic; <code>memGenes[i]=2</code> if the <i>i</i> -th probe is nonsignificant ; <code>memGenes[i]=3</code> if the <i>i</i> -th probe is significant with negative moderated t-statistic;            |
| <code>memGenes2</code>        | A numeric vector indicating the cluster membership of probes (unsorted). <code>memGenes2[i]=1</code> if the <i>i</i> -th probe is significant (adjusted pvalue < alpha). <code>memGenes2[i]=0</code> if the <i>i</i> -th probe is nonsignificant.  |
| <code>mu1</code>              | Mean expression levels for arrays for probe cluster 1 (average taking across all probes with <code>memGenes</code> value equal to 1.   |
| <code>mu2</code>              | Mean expression levels for arrays for probe cluster 2 (average taking across all probes with <code>memGenes</code> value equal to 2.   |
| <code>mu3</code>              | Mean expression levels for arrays for probe cluster 3 (average taking across all probes with <code>memGenes</code> value equal to 3.   |
| <code>ebFit</code>            | object returned by R Bioconductor function <code>eBayes</code> .   |

### Author(s)

Weiliang Qiu <stwxq@channing.harvard.edu>, Brandon Guo <brandowonder@gmail.com>, Christopher Anderson <christopheranderson84@gmail.com>, Barbara Klanderman <BKLANDERMAN@partners.org>, Vincent Carey <stvjc@channing.harvard.edu>, Benjamin Raby <rebar@channing.harvard.edu>

### Examples

```
# generate simulated data set from conditional normal distribution
set.seed(1234567)
es.sim = genSimData.BayesNormal(nCpGs = 100,
  nCases = 20, nControls = 20,
```

```

mu.n = -2, mu.c = 2,
d0 = 20, s02 = 0.64, s02.c = 1.5, testPara = "var",
outlierFlag = FALSE,
eps = 1.0e-3, applier = lapply)
print(es.sim)

# although the generated data is not from
# paired design, we use it to illustrate the
# usage of the function lmFitPaired

res.limma = lmFitPaired(
  es = es.sim,
  formula = ~as.factor(memSubj),
  pos.var.interest = 0, # the intercept is what we are interested
  pvalAdjMethod = "fdr",
  alpha = 0.05,
  probeID.var = "probe",
  gene.var = "gene",
  chr.var = "chr",
  verbose = TRUE)

```

---

lmFitWrapper

*A wrapper function for the function 'lmFit' of the R Bioconductor package 'limma'*


---

## Description

A wrapper function for the function 'lmFit' of the R Bioconductor package 'limma'.

## Usage

```

lmFitWrapper(
  es,
  formula = ~as.factor(gender),
  pos.var.interest = 1,
  pvalAdjMethod = "fdr",
  alpha = 0.05,
  probeID.var = "ProbeID",
  gene.var = "Symbol",
  chr.var = "Chromosome",
  verbose = TRUE)

```

## Arguments

|         |   |
|---------|---|
| es      | An LumiBatch object. fData(es) should contains information about chromosome number and gene symbol.                             |
| formula | An object of class formula. No left handside of ~ should be specified since the response variable will be the expression level. |

|                               |  |
|-------------------------------|--|
| <code>pos.var.interest</code> | integer. Indicates which covariate on the right-hand-side of <code>~</code> in formula is the covariate of the interest. By default, it is the first covariate <code>pos.var.interest=1</code> . |
| <code>pvalAdjMethod</code>    | One of p-value adjustment methods provided by the R function <code>p.adjust</code> in R package <code>stats</code> : “holm”, “hochberg”, “hommel”, “bonferroni”, “BH”, “BY”, “fdr”, “none”.      |
| <code>alpha</code>            | Significance level. A test is claimed to be significant if the adjusted p-value $<$ alpha.   |
| <code>probeID.var</code>      | character string. Name of the variable indicating probe ID in feature data set.  |
| <code>gene.var</code>         | character string. Name of the variable indicating gene symbol in feature data set.   |
| <code>chr.var</code>          | character string. Name of the variable indicating chromosome number in feature data set.   |
| <code>verbose</code>          | logical. Determine if intermediate output need to be suppressed. By default <code>verbose=TRUE</code> , intermediate output will be printed.   |

## Details

This is a wrapper function of R Bioconductor functions `lmFit` and `eBayes` to make it easier to input design and output list of significant results.

## Value

A list with the following elements:

|                             |  |
|-----------------------------|--|
| <code>n.sig</code>          | Number of significant tests after p-value adjustment.  |
| <code>frame</code>          | A data frame containing test results sorted according to the ascending order of unadjusted p-values for the covariate of the interest. The data frame contains 7 columns: <code>probeIDs</code> , <code>geneSymbols</code> (gene symbols of the genes where the probes come from), <code>chr</code> (numbers of chromosomes where the probes locate), <code>stats</code> (moderated t-statistics for the covariate of interest, i.e. the first covariate), <code>\codepval</code> (p-values of the tests for the covariate of interest, i.e. the first covariate), <code>p.adj</code> (adjusted p-values), <code>pos</code> (row numbers of the probes in the expression data matrix). |
| <code>statMat</code>        | A matrix containing test statistics for all covariates and for all probes. Rows are probes and columns are covariates. The rows are ordered according to the ascending order of unadjusted p-values for the covariate of the interest.   |
| <code>pvalMat</code>        | A matrix containing pvalues for all covariates and for all probes. Rows are probes and columns are covariates. The rows are ordered according to the ascending order of unadjusted p-values for the covariate of the interest.   |
| <code>pval.quantile</code>  | Quantiles (minimum, 25 for all covariates including intercept provided in the input argument formula.  |
| <code>frame.unsorted</code> | A data frame containing test results. The data frame contains 7 columns: <code>probeIDs</code> , <code>geneSymbols</code> (gene symbols of the genes where the probes come from), <code>chr</code> (numbers of chromosomes where the probes locate), <code>stats</code> (moderated t-statistics for the covariate of the interest), <code>pval</code> (p-values of the tests for the covariate of  |

|                               |   |
|-------------------------------|---|
|                               | the interest), <code>p.adj</code> (adjusted p-values), <code>pos</code> (row numbers of the probes in the expression data matrix).  |
| <code>statMat.unsorted</code> | A matrix containing test statistics for all covariates and for all probes. Rows are probes and columns are covariates.  |
| <code>pvalMat.unsorted</code> | A matrix containing pvalues for all covariates and for all probes. Rows are probes and columns are covariates.  |
| <code>memGenes</code>         | A numeric vector indicating the cluster membership of probes (unsorted). <code>memGenes[i]=1</code> if the <i>i</i> -th probe is significant (adjusted pvalue < alpha) with positive moderated t-statistic; <code>memGenes[i]=2</code> if the <i>i</i> -th probe is nonsignificant ; <code>memGenes[i]=3</code> if the <i>i</i> -th probe is significant with negative moderated t-statistic; |
| <code>memGenes2</code>        | A numeric vector indicating the cluster membership of probes (unsorted). <code>memGenes2[i]=1</code> if the <i>i</i> -th probe is significant (adjusted pvalue < alpha). <code>memGenes2[i]=0</code> if the <i>i</i> -th probe is nonsignificant.   |
| <code>mu1</code>              | Mean expression levels for arrays for probe cluster 1 (average taking across all probes with <code>memGenes</code> value equal to 1.  |
| <code>mu2</code>              | Mean expression levels for arrays for probe cluster 2 (average taking across all probes with <code>memGenes</code> value equal to 2.  |
| <code>mu3</code>              | Mean expression levels for arrays for probe cluster 3 (average taking across all probes with <code>memGenes</code> value equal to 3.  |
| <code>ebFit</code>            | object returned by R Bioconductor function <code>eBayes</code> .  |

### Author(s)

Weiliang Qiu <stwxq@channing.harvard.edu>, Brandon Guo <brandowonder@gmail.com>, Christopher Anderson <christopheranderson84@gmail.com>, Barbara Klanderma <BKLANDERMAN@partners.org>, Vincent Carey <stvj@channing.harvard.edu>, Benjamin Raby <rebar@channing.harvard.edu>

### Examples

```
# generate simulated data set from conditional normal distribution
set.seed(1234567)
es.sim = genSimData.BayesNormal(nCpGs = 100,
  nCases = 20, nControls = 20,
  mu.n = -2, mu.c = 2,
  d0 = 20, s02 = 0.64, s02.c = 1.5, testPara = "var",
  outlierFlag = FALSE,
  eps = 1.0e-3, applier = lapply)
print(es.sim)

res.limma = lmFitWrapper(
  es = es.sim,
  formula = ~as.factor(memSubj),
  pos.var.interest = 1,
  pvalAdjMethod = "fdr",
  alpha = 0.05,
```

```

probeID.var = "probe",
gene.var = "gene",
chr.var = "chr",
verbose = TRUE)

```

---

|                 |  |
|-----------------|--|
| LumiBatch2Table | <i>Output slots (exprs, pData, fData) of an LumiBatch object into 3 text files</i> |
|-----------------|--|

---

## Description

Output slots (exprs, pData, fData) of an LumiBatch object into 3 text files.

## Usage

```

LumiBatch2Table(
  es,
  probeID.var="ProbeID",
  gene.var="Symbol",
  chr.var="Chromosome",
  sep = ",",
  quote = FALSE,
  filePrefix = "test",
  fileExt = "csv")

```

## Arguments

|             |  |
|-------------|--|
| es          | An LumiBatch object  |
| probeID.var | character string. Name of the variable indicating probe ID in feature data set.          |
| gene.var    | character string. Name of the variable indicating gene symbol in feature data set.       |
| chr.var     | character string. Name of the variable indicating chromosome number in feature data set. |
| sep         | Field delimiter for the output text files  |
| quote       | logical. Indicating if any character or factor. See also <a href="#">write.table</a> .   |
| filePrefix  | Prefix of the names of the output files.   |
| fileExt     | File extension of the names of the output files.   |

## Details

Suppose filePrefix="test" and fileExt=".csv". Then, the file names of the 3 output files are: "test\_exprs.csv", "test\_pDat.csv", and "test\_fDat.csv", respectively.

## Value

None.

**Author(s)**

Weiliang Qiu <stwxq@channing.harvard.edu>, Brandon Guo <brandowonder@gmail.com>, Christopher Anderson <christopheranderson84@gmail.com>, Barbara Klanderman <BKLANDERMAN@partners.org>, Vincent Carey <stvjc@channing.harvard.edu>, Benjamin Raby <rebar@channing.harvard.edu>

**Examples**

```
# generate simulated data set from conditional normal distribution
set.seed(1234567)
es.sim = genSimData.BayesNormal(nCpGs = 100,
  nCases = 20, nControls = 20,
  mu.n = -2, mu.c = 2,
  d0 = 20, s02 = 0.64, s02.c = 1.5, testPara = "var",
  outlierFlag = FALSE,
  eps = 1.0e-3, applier = lapply)
print(es.sim)

LumiBatch2Table(
  es = es.sim,
  probeID.var="probe",
  gene.var="gene",
  chr.var="chr",
  sep = ",",
  quote = FALSE,
  filePrefix = "test",
  fileExt = "csv")
```

pca2DPlot

*Scatter plot of first 2 principal components***Description**

Scatter plot of first 2 principal components.

**Usage**

```
pca2DPlot(pcaObj,
  plot.dim = c(1,2),
  labelVariable = "subjID",
  hybName = "Hybridization_Name",
  outFileName = "test_pca_raw.pdf",
  title = "Scatter plot of pcas",
  plotOutPutFlag = FALSE,
  mar = c(5, 4, 4, 2) + 0.1,
  lwd = 1.5,
  equalRange = TRUE,
  xlab = NULL,
```

```

ylab = NULL,
xlim = NULL,
ylim = NULL,
cex.legend = 1.5,
cex = 1.5,
cex.lab = 1.5,
cex.axis = 1.5,
legendPosition = "topright",
...)

```

### Arguments

|                |  |
|----------------|--|
| pcaObj         | An object returned by the function <code>pca</code> of the R package <code>pcaMethods</code> .   |
| plot.dim       | A vector of 2 positive-integer-value integer specifying which 2 pcas will be plot.   |
| labelVariable  | The name of a column of the phenotype data matrix. The elements of the column will replace the column names of the expression data matrix.   |
| hybName        | character string. indicating the phenotype variable <code>Hybridization_Name</code> .  |
| outFileName    | Name of the figure file to be created.   |
| title          | Title of the scatter plot.   |
| plotOutPutFlag | logical. <code>plotOutPutFlag=TRUE</code> indicates the plots will be output to pdf format files. Otherwise, the plots will not be output to external files.   |
| mar            | A numerical vector of the form <code>'c(bottom, left, top, right)'</code> which gives the number of lines of margin to be specified on the four sides of the plot. The default is <code>'c(5, 4, 4, 2) + 0.1'</code> . see <a href="#">par</a> . |
| lwd            | The line width, a <code>_positive_</code> number, defaulting to <code>'1'</code> . see <a href="#">par</a> .   |
| equalRange     | logical. Indicating if the x-axis and y-axis have the same range.  |
| xlab           | Label of x axis.   |
| ylab           | Label of y axis.   |
| xlim           | Range of x axis.   |
| ylim           | Range of y axis.   |
| cex.legend     | Font size for legend.  |
| cex            | numerical value giving the amount by which plotting text and symbols should be magnified relative to the default. see <a href="#">par</a> .  |
| cex.lab        | The magnification to be used for x and y labels relative to the current setting of <code>cex</code> .  |
| cex.axis       | The magnification to be used for axis annotation relative to the current setting of <code>cex</code> .<br>see <a href="#">par</a> .  |
| legendPosition | Position of legend. Possible values are "bottomright", "bottom", "bottomleft", "left", "topleft", "top", "topright", "right" and "center".   |
| ...            | Arguments to be passed to <a href="#">plot</a> .   |

**Value**

A matrix of PCA scores. Each column corresponds to a principal component.

**Author(s)**

Weiliang Qiu <stwxq@channing.harvard.edu>, Brandon Guo <brandowonder@gmail.com>, Christopher Anderson <christopheranderson84@gmail.com>, Barbara Klanderman <BKLANDERMAN@partners.org>, Vincent Carey <stvjc@channing.harvard.edu>, Benjamin Raby <rebar@channing.harvard.edu>

**Examples**

```
# generate simulated data set from conditional normal distribution
set.seed(1234567)
es.sim = genSimData.BayesNormal(nCpGs = 100,
  nCases = 20, nControls = 20,
  mu.n = -2, mu.c = 2,
  d0 = 20, s02 = 0.64, s02.c = 1.5, testPara = "var",
  outlierFlag = FALSE,
  eps = 1.0e-3, applier = lapply)
print(es.sim)

pca.obj = getPcaFunc(es = es.sim,
  labelVariable = "subjID",
  hybName = "memSubj",
  requireLog2 = FALSE,
  corFlag = FALSE
)

pca2DPlot(pcaObj = pca.obj,
  plot.dim = c(1,2),
  labelVariable = "subjID",
  hybName = "memSubj",
  plotOutPutFlag = FALSE,
  cex.legend = 0.5,
  legendPosition = "topright")
```

---

pca3DPlot

*Scatter plot of 3 specified principal components*


---

**Description**

Scatter plot of 3 specified principal components.

**Usage**

```
pca3DPlot(pcaObj,
  plot.dim = c(1,2, 3),
  labelVariable = "subjID",
```



```

    hybName = "Hybridization_Name",
    outFileName = "test_pca_raw.pdf",
    title = "Scatter plot of pcas",
    plotOutPutFlag = FALSE,
    mar = c(5, 4, 4, 2) + 0.1,
    lwd = 1.5,
    equalRange = TRUE,
    xlab = NULL,
    ylab = NULL,
    zlab = NULL,
    xlim = NULL,
    ylim = NULL,
    zlim = NULL,
    cex.legend = 1.5,
    cex = 1.5,
    cex.lab = 1.5,
    cex.axis = 1.5,
    legendPosition = "topright",
    ...)

```

### Arguments

|                             |  |
|-----------------------------|--|
| <code>pcaObj</code>         | An object returned by the function <code>pca</code> of the R package <code>pcaMethods</code> .   |
| <code>plot.dim</code>       | A vector of 3 positive-integer-value integer specifying which 3 pcas will be plot.   |
| <code>labelVariable</code>  | The name of a column of the phenotype data matrix. The elements of the column will replace the column names of the expression data matrix.   |
| <code>hybName</code>        | character string. indicating the phenotype variable <code>Hybridization_Name</code> .  |
| <code>outFileName</code>    | Name of the figure file to be created.   |
| <code>title</code>          | Title of the scatter plot.   |
| <code>plotOutPutFlag</code> | logical. <code>plotOutPutFlag=TRUE</code> indicates the plots will be output to pdf format files. Otherwise, the plots will not be output to external files.   |
| <code>mar</code>            | A numerical vector of the form <code>'c(bottom, left, top, right)'</code> which gives the number of lines of margin to be specified on the four sides of the plot. The default is <code>'c(5, 4, 4, 2) + 0.1'</code> . see <a href="#">par</a> . |
| <code>lwd</code>            | The line width, a <code>_positive_</code> number, defaulting to <code>'1'</code> . see <a href="#">par</a> .   |
| <code>equalRange</code>     | logical. Indicating if the x-axis and y-axis have the same range.  |
| <code>xlab</code>           | Label of x axis.   |
| <code>ylab</code>           | Label of y axis.   |
| <code>zlab</code>           | Label of z axis.   |
| <code>xlim</code>           | Range of x axis.   |
| <code>ylim</code>           | Range of y axis.   |
| <code>zlim</code>           | Range of z axis.   |
| <code>cex.legend</code>     | Font size for legend.  |

|                             |   |
|-----------------------------|---|
| <code>cex</code>            | numerical value giving the amount by which plotting text and symbols should be magnified relative to the default. see <a href="#">par</a> . |
| <code>cex.lab</code>        | The magnification to be used for x and y labels relative to the current setting of <code>cex</code> .                                       |
| <code>cex.axis</code>       | The magnification to be used for axis annotation relative to the current setting of <code>cex</code> .<br>see <a href="#">par</a> .         |
| <code>legendPosition</code> | Position of legend. Possible values are “bottomright”, “bottom”, “bottomleft”, “left”, “topleft”, “top”, “topright”, “right” and “center”.  |
| <code>...</code>            | Arguments to be passed to <a href="#">plot</a> .  |

### Value

A matrix of PCA scores. Each column corresponds to a principal component.

### Author(s)

Weiliang Qiu <stwxq@channing.harvard.edu>, Brandon Guo <brandowonder@gmail.com>, Christopher Anderson <christopheranderson84@gmail.com>, Barbara Klanderman <BKLANDERMAN@partners.org>, Vincent Carey <stvjc@channing.harvard.edu>, Benjamin Raby <rebar@channing.harvard.edu>

### Examples

```
# generate simulated data set from conditional normal distribution
set.seed(1234567)
es.sim = genSimData.BayesNormal(nCpGs = 100,
  nCases = 20, nControls = 20,
  mu.n = -2, mu.c = 2,
  d0 = 20, s02 = 0.64, s02.c = 1.5, testPara = "var",
  outlierFlag = FALSE,
  eps = 1.0e-3, applier = lapply)
print(es.sim)

pca.obj = getPcaFunc(es = es.sim,
  labelVariable = "subjID",
  hybName = "memSubj",
  requireLog2 = FALSE,
  corFlag = FALSE
)

pca3DPlot(pcaObj = pca.obj,
  plot.dim = c(1,2,3),
  labelVariable = "subjID",
  hybName = "memSubj",
  plotOutPutFlag = FALSE,
  cex.legend = 0.5,
  legendPosition = "topright")
```

---

plotCurves

---

*Plot trajectories of probe profiles across arrays*


---

## Description

Plot trajectories of probe profiles across arrays

## Usage

```
plotCurves(
  dat,
  curveNames,
  fileName,
  plotOutPutFlag=FALSE,
  requireLog2 = FALSE,
  cex = 1,
  ylim = NULL,
  xlab = "",
  ylab = "intensity",
  lwd = 3,
  main = "Trajectory plot",
  mar = c(10, 4, 4, 2) + 0.1,
  las = 2,
  cex.axis=1,
  ...)
```

## Arguments

|                |   |
|----------------|---|
| dat            | Numeric data matrix. Rows are probes; columns are arrays.   |
| curveNames     | Probe names.  |
| fileName       | file name of output figure.   |
| plotOutPutFlag | logical. plotOutPutFlag=TRUE indicates the plots will be output to pdf format files. Otherwise, the plots will not be output to external files. |
| requireLog2    | logical. requiredLog2=TRUE indicates probe expression levels will be log2 transformed. Otherwise, no transformation will be performed.          |
| cex            | numerical value giving the amount by which plotting text and symbols should be magnified relative to the default. see <a href="#">par</a> .     |
| ylim           | Range of y axis.  |
| xlab           | Label of x axis.  |
| ylab           | Label of y axis.  |
| lwd            | The line width, a <code>_positive_</code> number, defaulting to '1'. see <a href="#">par</a> .  |
| main           | Main title of the plot.   |

|                       |   |
|-----------------------|---|
| <code>mar</code>      | A numerical vector of the form 'c(bottom, left, top, right)' which gives the number of lines of margin to be specified on the four sides of the plot. The default is 'c(5, 4, 4, 2) + 0.1'. see <a href="#">par</a> . |
| <code>las</code>      | 'las' numeric in 0,1,2,3; the style of axis labels. 0 - always parallel to the axis, 1 - always horizontal, 2 - always perpendicular to the axis, or 3 - always vertical. see <a href="#">par</a> .                   |
| <code>cex.axis</code> | The magnification to be used for axis annotation relative to the current setting of <code>cex</code> .<br>see <a href="#">par</a> .   |
| <code>...</code>      | Arguments to be passed to <a href="#">plot</a> .  |

**Value**

no return value.

**Author(s)**

Weiliang Qiu <stwxq@channing.harvard.edu>, Brandon Guo <brandowonder@gmail.com>, Christopher Anderson <christopheranderson84@gmail.com>, Barbara Klanderman <BKLANDERMAN@partners.org>, Vincent Carey <stvjc@channing.harvard.edu>, Benjamin Raby <rebar@channing.harvard.edu>

**Examples**

```
# generate simulated data set from conditional normal distribution
set.seed(1234567)
es.sim = genSimData.BayesNormal(nCpGs = 100,
  nCases = 20, nControls = 20,
  mu.n = -2, mu.c = 2,
  d0 = 20, s02 = 0.64, s02.c = 1.5, testPara = "var",
  outlierFlag = FALSE,
  eps = 1.0e-3, applier = lapply)
print(es.sim)

# plot trajectories of the first 5 genes
plotCurves(
  dat = exprs(es.sim)[1:5,],
  curveNames = featureNames(es.sim)[1:5],
  plotOutPutFlag=FALSE,
  cex = 0.5,
  requireLog2 = FALSE)
```

---

plotQCCurves

*Plot trajectories of specific QC probes (e.g., biotin, cy3\_hyb, house-keeping gene probes, low stringency probes, etc.) across arrays*

---

## Description

Plot trajectories of specific QC probes (e.g., biotin, cy3\_hyb, housekeeping gene probes, low stringency probes, etc.) across arrays

## Usage

```
plotQCCurves(
  esQC,
  probes = c("biotin", "cy3_hyb", "housekeeping",
    "low_stringency_hyb", "signal", "p95p05"),
  labelVariable = "subjID",
  hybName = "Hybridization_Name",
  reporterGroupName = "Reporter_Group_Name",
  requireLog2 = TRUE,
  projectName = "test",
  plotOutPutFlag = FALSE,
  cex = 1,
  ylim = NULL,
  xlab = "",
  ylab = "intensity",
  lwd = 3,
  mar = c(10, 4, 4, 2) + 0.1,
  las = 2,
  cex.axis = 1,
  sortFlag = TRUE,
  varSort = c("Batch_Run_Date", "Chip_Barcode", "Chip_Address"),
  timeFormat = c("%m/%d/%Y", NA, NA),
  ...)
```

## Arguments

|                   |  |
|-------------------|--|
| esQC              | ExpressionSet object of QC probe profiles. fData(esQC) should contains the variable Reporter_Group_Name.   |
| probes            | A character vectors of QC probe names. By default, it includes the following probe names "biotin", "cy3_hyb", "housekeeping", "low_stringency_hyb", "signal", "p95p05". For "signal", trajectories of 5th, 25th, 50th, 75th, and 95th percentiles of the expression levels of all QC probes will be plotted. For "p95p05", the trajectory of the ratio of 95th percentile to 5th percentile of the expression levels of all QC probes will be plotted. |
| labelVariable     | A character string. The name of a phenotype data variable use to label the arrays in the boxplots. By default, labelVariable = "subjID" which is equivalent to labelVariable = "Hybridization_Name".   |
| hybName           | character string. indicating the phenotype variable Hybridization_Name.  |
| reporterGroupName | character string. indicating feature variable Reporter_Group_Name (QC probe's name).   |

|                |   |
|----------------|---|
| requireLog2    | logical. requiredLog2=TRUE indicates probe expression levels will be log2 transformed. Otherwise, no transformation will be performed.  |
| projectName    | A character string. Name of the project. The plots will be saved as pdf format files, the names of which have the format projectName_probeName_traj_plot.pdf.   |
| plotOutPutFlag | logical. plotOutPutFlag=TRUE indicates the plots will be output to pdf format files. Otherwise, the plots will not be output to external files.   |
| cex            | numerical value giving the amount by which plotting text and symbols should be magnified relative to the default. see <a href="#">par</a> .   |
| ylim           | Range of y axis.  |
| xlab           | Label of x axis.  |
| ylab           | Label of y axis.  |
| lwd            | The line width, a <code>_positive_</code> number, defaulting to '1'. see <a href="#">par</a> .  |
| mar            | A numerical vector of the form 'c(bottom, left, top, right)' which gives the number of lines of margin to be specified on the four sides of the plot. The default is 'c(5, 4, 4, 2) + 0.1'. see <a href="#">par</a> .     |
| las            | 'las' numeric in 0,1,2,3; the style of axis labels. 0 - always parallel to the axis, 1 - always horizontal, 2 - always perpendicular to the axis, or 3 - always vertical. see <a href="#">par</a> .                       |
| cex.axis       | The magnification to be used for axis annotation relative to the current setting of cex. see <a href="#">par</a> .  |
| sortFlag       | logical. Indicates if arrays need to be sorted according to Batch_Run_Date, Chip_Barcode, and Chip_Address.   |
| varSort        | A vector of phenotype variable names to be used to sort the samples of es.  |
| timeFormat     | A vector of time format for the possible time variables in varSort. The length of timeFormat should be the same as that of varSort. For non-time variable, the corresponding time format should be set to be equal to NA. |
| ...            | Arguments to be passed to <a href="#">plot</a> .  |

**Value**

no return value.

**Author(s)**

Weiliang Qiu <stwxq@channing.harvard.edu>, Brandon Guo <brandowonder@gmail.com>, Christopher Anderson <christopheranderson84@gmail.com>, Barbara Klanderman <BKLANDERMAN@partners.org>, Vincent Carey <stvjc@channing.harvard.edu>, Benjamin Raby <rebar@channing.harvard.edu>

**Examples**

```
# generate simulated data set from conditional normal distribution
set.seed(1234567)
esQC.sim = genSimData.BayesNormal(nCpGs = 10,
  nCases = 20, nControls = 20,
```

```

mu.n = -2, mu.c = 2,
d0 = 20, s02 = 0.64, s02.c = 1.5, testPara = "var",
outlierFlag = FALSE,
eps = 1.0e-3, applier = lapply)

print(esQC.sim)

fDat = fData(esQC.sim)
esQC.sim$Hybridization_Name = sampleNames(esQC.sim)
fDat$Reporter_Group_Name = c( rep("biotin", 5),
  rep("housekeeping", 5))
fData(esQC.sim)=fDat

# plot trajectories of the QC probes
plotQCCurves(
  esQC = esQC.sim,
  probes = c("biotin", "housekeeping"),
  labelVariable = "subjID",
  hybName = "Hybridization_Name",
  reporterGroupName = "Reporter_Group_Name",
  requireLog2 = FALSE,
  plotOutPutFlag = FALSE,
  sortFlag = FALSE)

```

---

plotSamplep95p05

---

*Plot trajectories of the ratio of 95th percentile to 5th percentile of sample probe profiles across arrays*


---

## Description

Plot trajectories of the ratio of 95th percentile to 5th percentile of sample probe profiles across arrays.

## Usage

```

plotSamplep95p05(
  es,
  labelVariable = "subjID",
  hybName = "Hybridization_Name",
  requireLog2 = FALSE,
  projectName = "test",
  plotOutPutFlag = FALSE,
  cex = 1,
  ylim = NULL,
  xlab = "",
  ylab = "",
  lwd = 1.5,
  mar = c(10, 4, 4, 2) + 0.1,

```

```

las = 2,
cex.axis=1.5,
title = "Trajectory of p95/p05",
cex.legend = 1.5,
cex.lab = 1.5,
legendPosition = "topright",
cut1 = 10,
cut2 = 6,
sortFlag = TRUE,
varSort = c("Batch_Run_Date", "Chip_Barcode", "Chip_Address"),
timeFormat = c("%m/%d/%Y", NA, NA),
verbose = FALSE,
...)

```

### Arguments

|                             |   |
|-----------------------------|---|
| <code>es</code>             | ExpressionSet object of Sample probe profiles.  |
| <code>labelVariable</code>  | A character string. The name of a phenotype data variable use to label the arrays in the boxplots. By default, <code>labelVariable = "subjID"</code> which is equivalent to <code>labelVariable = "Hybridization_Name"</code> .                     |
| <code>hybName</code>        | character string. indicating the phenotype variable <code>Hybridization_Name</code> .   |
| <code>requireLog2</code>    | logical. <code>requiredLog2=TRUE</code> indicates probe expression levels will be log2 transformed. Otherwise, no transformation will be performed.   |
| <code>projectName</code>    | A character string. Name of the project. The plots will be saved as pdf format files, the names of which have the format <code>projectName_probeName_traj_plot.pdf</code> .   |
| <code>plotOutPutFlag</code> | logical. <code>plotOutPutFlag=TRUE</code> indicates the plots will be output to pdf format files. Otherwise, the plots will not be output to external files.  |
| <code>cex</code>            | numerical value giving the amount by which plotting text and symbols should be magnified relative to the default. see <a href="#">par</a> .   |
| <code>ylim</code>           | Range of y axis.  |
| <code>xlab</code>           | Label of x axis.  |
| <code>ylab</code>           | Label of y axis.  |
| <code>lwd</code>            | The line width, a <code>_positive_</code> number, defaulting to '1'. see <a href="#">par</a> .  |
| <code>mar</code>            | A numerical vector of the form ' <code>c(bottom, left, top, right)</code> ' which gives the number of lines of margin to be specified on the four sides of the plot. The default is ' <code>c(5, 4, 4, 2) + 0.1</code> '. see <a href="#">par</a> . |
| <code>las</code>            | 'las' numeric in 0,1,2,3; the style of axis labels. 0 - always parallel to the axis, 1 - always horizontal, 2 - always perpendicular to the axis, or 3 - always vertical. see <a href="#">par</a> .   |
| <code>cex.axis</code>       | The magnification to be used for axis annotation relative to the current setting of <code>cex</code> .<br>see <a href="#">par</a> .   |
| <code>title</code>          | Figure title.   |
| <code>cex.legend</code>     | Font size of legend text.   |



|                |  |
|----------------|--|
| cex.lab        | The magnification to be used for x and y labels relative to the current setting of cex.  |
| legendPosition | Position of legend. Possible values are “bottomright”, “bottom”, “bottomleft”, “left”, “topleft”, “top”, “topright”, “right” and “center”.   |
| cut1           | second horizontal line setting the cutoff for the ratio p95/p05. A ratio above this line indicates the corresponding array is good.  |
| cut2           | second horizontal line setting the cutoff for the ratio p95/p05. A ratio below this line indicates the corresponding array is bad.   |
| sortFlag       | logical. Indicates if arrays need to be sorted according to Batch_Run_Date, Chip_Barcode, and Chip_Address.  |
| varSort        | A vector of phenotype variable names to be used to sort the samples of es.   |
| timeFormat     | A vector of time format for the possible time variables in varSort. The length of timeFormat should be the same as that of varSort. For non-time variable, the corresponding time format should be set to be equal to NA. The details of the time format for time variable can be found in the R function <a href="#">strptime</a> . |
| verbose        | logical. Determine if intermediate output need to be suppressed. By default verbose=FALSE, intermediate output will not be printed.  |
| ...            | Arguments to be passed to <a href="#">plot</a> .   |

## Details

The trajectory of the ratio of 95 to 5

## Value

A list of 2 elements. The first element is the 2 x n matrix, where n is the number of arrays. The first row of the matrix is the 5-th percentile and the second row of the matrix is the 95-th percentile.

The second element is the ratio of the 95-th percentile to the 5-th percentile.

## Author(s)

Weiliang Qiu <stwxq@channing.harvard.edu>, Brandon Guo <brandowonder@gmail.com>, Christopher Anderson <christopheranderson84@gmail.com>, Barbara Klanderman <BKLANDERMAN@partners.org>, Vincent Carey <stvjc@channing.harvard.edu>, Benjamin Raby <rebar@channing.harvard.edu>

## Examples

```
# generate simulated data set from conditional normal distribution
set.seed(1234567)
es.sim = genSimData.BayesNormal(nCpGs = 100,
  nCases = 20, nControls = 20,
  mu.n = -2, mu.c = 2,
  d0 = 20, s02 = 0.64, s02.c = 1.5, testPara = "var",
  outlierFlag = FALSE,
  eps = 1.0e-3, applier = lapply)
print(es.sim)
```

```

es.sim$Batch_Run_Date = 1:ncol(es.sim)
es.sim$Chip_Barcode = 1:ncol(es.sim)
es.sim$Chip_Address = 1:ncol(es.sim)

plotSamplep95p05(
  es = es.sim,
  labelVariable = "subjID",
  hybName = "memSubj",
  requireLog2 = FALSE,
  projectName = "test",
  plotOutPutFlag = FALSE,
  title = "Trajectory of p95/p05",
  cex.legend = 0.5,
  legendPosition = "topright",
  sortFlag = TRUE,
  varSort = c("Batch_Run_Date", "Chip_Barcode", "Chip_Address"),
  timeFormat = c("%m/%d/%Y", NA, NA),
  verbose = FALSE)

```

---

quantilePlot

*Plot trajectories of quantiles across arrays*


---

## Description

Plot trajectories of quantiles across arrays.

## Usage

```

quantilePlot(
  dat,
  fileName,
  probs = c(0, 0.05, 0.25, 0.5, 0.75, 0.95, 1),
  plotOutPutFlag = FALSE,
  requireLog2 = FALSE,
  sortFlag = TRUE,
  cex = 1,
  ylim = NULL,
  xlab = "",
  ylab = "intensity",
  lwd = 3,
  main = "Trajectory plot of quantiles",
  mar = c(15, 4, 4, 2) + 0.1,
  las = 2,
  cex.axis = 1)

```

**Arguments**

|                             |   |
|-----------------------------|---|
| <code>dat</code>            | Expression data. Rows are gene probes; columns are arrays.  |
| <code>fileName</code>       | File name of output figure.   |
| <code>probs</code>          | quantiles (any real values between the interval $[0, 1]$ ).   |
| <code>plotOutPutFlag</code> | logical. <code>plotOutPutFlag=TRUE</code> indicates the plots will be output to pdf format files. Otherwise, the plots will not be output to external files.  |
| <code>requireLog2</code>    | logical. <code>requiredLog2=TRUE</code> indicates probe expression levels will be log2 transformed. Otherwise, no transformation will be performed.   |
| <code>sortFlag</code>       | logical. <code>sortFlag=TRUE</code> indicates arrays will be sorted by the ascending order of MAD (median absolute deviation)   |
| <code>cex</code>            | numerical value giving the amount by which plotting text and symbols should be magnified relative to the default. see <a href="#">par</a> .   |
| <code>ylim</code>           | Range of y axis.  |
| <code>xlab</code>           | Label of x axis.  |
| <code>ylab</code>           | Label of y axis.  |
| <code>lwd</code>            | The line width, a <code>_positive_</code> number, defaulting to '1'. see <a href="#">par</a> .  |
| <code>main</code>           | Character string. main title of the plot.   |
| <code>mar</code>            | A numerical vector of the form ' <code>c(bottom, left, top, right)</code> ' which gives the number of lines of margin to be specified on the four sides of the plot. The default is ' <code>c(5, 4, 4, 2) + 0.1</code> '. see <a href="#">par</a> . |
| <code>las</code>            | 'las' numeric in 0,1,2,3; the style of axis labels. 0 - always parallel to the axis, 1 - always horizontal, 2 - always perpendicular to the axis, or 3 - always vertical. see <a href="#">par</a> .   |
| <code>cex.axis</code>       | The magnification to be used for axis annotation relative to the current setting of <code>cex</code> . see <a href="#">par</a> .  |

**Value**

The quantile matrix with row quantiles and column array.

**Author(s)**

Weiliang Qiu <stwxq@channing.harvard.edu>, Brandon Guo <brandowonder@gmail.com>, Christopher Anderson <christopheranderson84@gmail.com>, Barbara Klanderman <BKLANDERMAN@partners.org>, Vincent Carey <stvjc@channing.harvard.edu>, Benjamin Raby <rebar@channing.harvard.edu>

**Examples**

```
# generate simulated data set from conditional normal distribution
set.seed(1234567)
es.sim = genSimData.BayesNormal(nCpGs = 100,
  nCases = 20, nControls = 20,
  mu.n = -2, mu.c = 2,
```

```

    d0 = 20, s02 = 0.64, s02.c = 1.5, testPara = "var",
    outlierFlag = FALSE,
    eps = 1.0e-3, applier = lapply)
print(es.sim)

png(file="qplot.png")

quantilePlot(
  dat = exprs(es.sim),
  probs = c(0, 0.05, 0.25, 0.5, 0.75, 0.95, 1),
  plotOutPutFlag = FALSE,
  requireLog2 = FALSE,
  sortFlag = TRUE)

dev.off()

```

---

R2PlotFunc

---

*Draw heatmap of square of correlations among arrays*


---

## Description

Draw heatmap of square of correlations among arrays.

## Usage

```

R2PlotFunc(
  es,
  hybName = "Hybridization_Name",
  arrayType = c("all", "replicates", "GC"),
  GCid = c("128115", "Hela", "Brain"),
  probs = seq(0, 1, 0.25),
  col = gplots::greenred(75),
  labelVariable = "subjID",
  outFileName = "test_R2_raw.pdf",
  title = "Raw Data R^2 Plot",
  requireLog2 = FALSE,
  plotOutPutFlag = FALSE,
  las = 2,
  keysize = 1,
  margins = c(10, 10),
  sortFlag = TRUE,
  varSort=c("Batch_Run_Date", "Chip_Barcode", "Chip_Address"),
  timeFormat=c("%m/%d/%Y", NA, NA),
  ...)

```

**Arguments**

|                             |  |
|-----------------------------|--|
| <code>es</code>             | ExpressionSet object of QC probe profiles.   |
| <code>hybName</code>        | character string. indicating the phenotype variable Hybridization_Name.  |
| <code>arrayType</code>      | A character string indicating if the correlations are calculated based on all arrays, arrays with replicates, or genetic control arrays.   |
| <code>GCid</code>           | A vector of character string. symbols for genetic control samples. The symbols can be more than one.   |
| <code>probs</code>          | A vector of probabilities specify the quantiles of correlations to be output.  |
| <code>col</code>            | colors used for the image. see the function <a href="#">heatmap.2</a> in R package gplots.   |
| <code>labelVariable</code>  | A character string indicating how to label the arrays.   |
| <code>outFileName</code>    | A character string. The name of output file.   |
| <code>title</code>          | Title of the plot.   |
| <code>requireLog2</code>    | logical. <code>requiredLog2=TRUE</code> indicates probe expression levels will be log2 transformed. Otherwise, no transformation will be performed.  |
| <code>plotOutPutFlag</code> | logical. <code>plotOutPutFlag=TRUE</code> indicates the plots will be output to pdf format files. Otherwise, the plots will not be output to external files.   |
| <code>las</code>            | 'las' numeric in 0,1,2,3; the style of axis labels. 0 - always parallel to the axis, 1 - always horizontal, 2 - always perpendicular to the axis, or 3 - always vertical. see <a href="#">par</a> .  |
| <code>keysize</code>        | numeric value indicating the size of the key. see the function <a href="#">heatmap.2</a> in R package gplots.  |
| <code>margins</code>        | numeric vector of length 2 containing the margins. see the function <a href="#">heatmap.2</a> in R package gplots.   |
| <code>sortFlag</code>       | logical. Indicates if arrays need to be sorted according to Batch_Run_Date, Chip_Barcode, and Chip_Address.  |
| <code>varSort</code>        | A vector of phenotype variable names to be used to sort the samples of es.   |
| <code>timeFormat</code>     | A vector of time format for the possible time variables in varSort. The length of timeFormat should be the same as that of varSort. For non-time variable, the corresponding time format should be set to be equal to NA. The details of the time format for time variable can be found in the R function <a href="#">strptime</a> . |
| <code>...</code>            | Arguments to be passed to <a href="#">heatmap.2</a> .  |

**Value**

A list with 3 elments. The first element R2Mat is the matrix of squared correlation. The second element R2vec is the vector of the upper triangle of the matrix of squared correlation (diagonal elements are excluded). The third element R2vec.within.req is the vector of within-replicate  $R^2$ , that is, any element in R2vec.within.req is the squared correlation coefficient between two arrays/replicates for a subject.

**Author(s)**

Weiliang Qiu <stwxq@channing.harvard.edu>, Brandon Guo <brandowonder@gmail.com>, Christopher Anderson <christopheranderson84@gmail.com>, Barbara Klanderman <BKLANDERMAN@partners.org>, Vincent Carey <stvjc@channing.harvard.edu>, Benjamin Raby <rebar@channing.harvard.edu>

## Examples

```
# generate simulated data set from conditional normal distribution
set.seed(1234567)
es.sim = genSimData.BayesNormal(nCpGs = 100,
  nCases = 20, nControls = 20,
  mu.n = -2, mu.c = 2,
  d0 = 20, s02 = 0.64, s02.c = 1.5, testPara = "var",
  outlierFlag = FALSE,
  eps = 1.0e-3, applier = lapply)
print(es.sim)

es.sim$Batch_Run_Date = 1:ncol(es.sim)
es.sim$Chip_Barcode = 1:ncol(es.sim)
es.sim$Chip_Address = 1:ncol(es.sim)

# draw heatmap for the first 5 subjects
png(file="r2plot.png")
R2PlotFunc(
  es = es.sim[, 1:5],
  hybName = "memSubj",
  arrayType = c("all", "replicates", "GC"),
  GCid = c("128115", "Hela", "Brain"),
  probs = seq(0, 1, 0.25),
  col = gplots::greenred(75),
  labelVariable = "subjID",
  outFileName = "test_R2_raw.pdf",
  title = "Raw Data R^2 Plot",
  requireLog2 = FALSE,
  plotOutPutFlag = FALSE,
  las = 2,
  keysize = 1,
  margins = c(10, 10),
  sortFlag = TRUE,
  varSort=c("Batch_Run_Date", "Chip_Barcode", "Chip_Address"),
  timeFormat=c("%m/%d/%Y", NA, NA))
dev.off()
```

---

scatterPlots

---

*Draw scatter plots for top results in whole-genome-wide analysis*


---

## Description

Draw scatter plots for top results in whole-genome-wide analysis to test for the association of probes to a continuous-type phenotype variable.

## Usage

```
scatterPlots(
```

```

resFrame,
es,
col.resFrame = c("probeIDs", "stats", "pval", "p.adj"),
var.pheno = "bmi",
outcomeFlag = FALSE,
fitLineFlag = TRUE,
var.probe = "TargetID",
var.gene = "Symbol",
var.chr = "Chr",
nTop = 20,
myylab = "expression level",
datExtrFunc = exprs,
fileFlag = FALSE,
fileFormat = "ps",
fileName = "scatterPlots.ps")

```

### Arguments

|                           |  |
|---------------------------|--|
| <code>resFrame</code>     | A data frame stores testing results, which must contain columns that indicate probe id, test statistic, p-value and optionally adjusted p-value.   |
| <code>es</code>           | An ExpressionSet object that used to run the whole genome-wide tests.  |
| <code>col.resFrame</code> | A vector of characters indicating column names of <code>resFrame</code> corresponding to probe id, test statistic, p-value and optionally adjusted p-value.  |
| <code>var.pheno</code>    | character. the name of continuous-type phenotype variable that is used to test the association of this variable to probes.   |
| <code>outcomeFlag</code>  | logic. indicating if <code>var.pheno</code> is the outcome variable in regression analysis.  |
| <code>fitLineFlag</code>  | logic. indicating if a fitted line $y = a + bx$ should be plotted. If <code>outcomeFlag=TRUE</code> , then $y$ is <code>var.pheno</code> and $x$ is the top probe. If <code>outcomeFlag=FALSE</code> , then $y$ is the top probe and $x$ is <code>var.pheno</code> . |
| <code>var.probe</code>    | character. the name of feature variable indicating probe id.   |
| <code>var.gene</code>     | character. the name of feature variable indicating gene symbol.  |
| <code>var.chr</code>      | character. the name of feature variable indicating chromosome number.  |
| <code>nTop</code>         | integer. indicating how many top tests will be used to draw the scatter plot.  |
| <code>myylab</code>       | character. indicating y-axis label.  |
| <code>datExtrFunc</code>  | name of the function to extract genomic data. For an ExpressionSet object, you should set <code>datExtrFunc=exprs</code> ; for a MethyLumiSet object, you should set <code>datExtrFunc=betas</code> .  |
| <code>fileFlag</code>     | logic. indicating if plot should be saved to an external figure file.  |
| <code>fileFormat</code>   | character. indicating the figure file type. Possible values are "ps", "pdf", or "jpeg". All other values will produce "png" file.  |
| <code>fileName</code>     | character. indicating figure file name (file extension should be specified). For example, you set <code>fileFormat="pdf"</code> , then you can set <code>fileName="test.pdf"</code> , but not <code>fileName="test"</code> .   |

**Value**

Value 0 will be returned if no error occurs.

**Author(s)**

Weiliang Qiu <stwxq@channing.harvard.edu>, Brandon Guo <brandowonder@gmail.com>, Christopher Anderson <christopheranderson84@gmail.com>, Barbara Klanderman <BKLANDERMAN@partners.org>, Vincent Carey <stvjc@channing.harvard.edu>, Benjamin Raby <rebar@channing.harvard.edu>

**Examples**

```
# generate simulated data set from conditional normal distribution
set.seed(1234567)
es.sim = genSimData.BayesNormal(nCpGs = 100,
  nCases = 20, nControls = 20,
  mu.n = -2, mu.c = 2,
  d0 = 20, s02 = 0.64, s02.c = 1.5, testPara = "var",
  outlierFlag = FALSE,
  eps = 1.0e-3, applier = lapply)
print(es.sim)

# generate phenotype age
es.sim$age = rnorm(ncol(es.sim), mean=50, sd=5)

res.limma = lmFitWrapper(
  es = es.sim,
  formula = ~age,
  pos.var.interest = 1,
  pvalAdjMethod = "fdr",
  alpha = 0.05,
  probeID.var = "probe",
  gene.var = "gene",
  chr.var = "chr",
  verbose = TRUE)

scatterPlots(
  resFrame=res.limma$frame,
  es=es.sim,
  col.resFrame = c("probeIDs", "stats", "pval"),
  var.pheno = "age",
  outcomeFlag = FALSE,
  fitLineFlag = TRUE,
  var.probe = "probe",
  var.gene = "gene",
  var.chr = "chr",
  nTop = 20,
  myylab = "expression level",
  datExtrFunc = exprs,
  fileFlag = FALSE,
  fileFormat = "ps",
  fileName = "scatterPlots.ps")
```



---

|                   |  |
|-------------------|--|
| sortExpressionSet | <i>Sort the order of samples for an ExpressionSet object</i> |
|-------------------|--|

---

## Description

Sort the order of samples for an ExpressionSet object.

## Usage

```
sortExpressionSet(
  es,
  varSort = c("Batch_Run_Date", "Chip_Barcode", "Chip_Address"),
  timeFormat = c("%m/%d/%Y", NA, NA)
)
```

## Arguments

|            |  |
|------------|--|
| es         | An ExpressionSet.  |
| varSort    | A vector of phenotype variable names to be used to sort the samples of es.   |
| timeFormat | A vector of time format for the possible time variables in varSort. The length of timeFormat should be the same as that of varSort. For non-time variable, the corresponding time format should be set to be equal to NA. Please refer to function <a href="#">strptime</a> of the base package. |

## Value

An ExpressionSet object with samples sorted based on the variables indicated in varSort.

## Author(s)

Weiliang Qiu <stwxq@channing.harvard.edu>, Brandon Guo <brandowonder@gmail.com>, Christopher Anderson <christopheranderson84@gmail.com>, Barbara Klanderman <BKLANDERMAN@partners.org>, Vincent Carey <stvjc@channing.harvard.edu>, Benjamin Raby <rebar@channing.harvard.edu>

## Examples

```
# generate simulated data set from conditional normal distribution
set.seed(1234567)
es.sim = genSimData.BayesNormal(nCpGs = 100,
  nCases = 20, nControls = 20,
  mu.n = -2, mu.c = 2,
  d0 = 20, s02 = 0.64, s02.c = 1.5, testPara = "var",
  outlierFlag = FALSE,
  eps = 1.0e-3, applier = lapply)
print(es.sim)

es.sim$Batch_Run_Date = 1:ncol(es.sim)
es.sim$Chip_Barcode = 1:ncol(es.sim)
```

```
es.sim$Chip_Address = 1:ncol(es.sim)

es.sim2 = sortExpressionSet(
  es = es.sim,
  varSort = c("Batch_Run_Date", "Chip_Barcode", "Chip_Address"),
  timeFormat = c("%m/%d/%Y", NA, NA)
)
```

# Index

## \* **methods**

- genExprSet, 6
- getPCAFunc, 9
- glmWrapper, 10
- lkhrrWrapper, 13
- lmFitPaired, 15
- lmFitWrapper, 18
- LumiBatch2Table, 21
- pca2DPlot, 22
- pca3DPlot, 24
- plotCurves, 27
- plotQCCurves, 28
- plotSamplep95p05, 31
- quantilePlot, 34
- R2PlotFunc, 36
- sortExpressionSet, 41

## \* **method**

- boxPlots, 2
- densityPlots, 4
- genSimData.BayesNormal, 7
- scatterPlots, 38

boxPlots, 2

densityPlots, 4

genExprSet, 6  
genSimData.BayesNormal, 7  
getPCAFunc, 9  
glm, 11, 14  
glmWrapper, 10

heatmap.2, 37

lkhrrWrapper, 13  
lmFitPaired, 15  
lmFitWrapper, 18  
LumiBatch2Table, 21

par, 23, 25–28, 30, 32, 35, 37  
pca2DPlot, 22

pca3DPlot, 24  
plot, 23, 26, 28, 30, 33  
plotCurves, 27  
plotQCCurves, 28  
plotSamplep95p05, 31

quantilePlot, 34

R2PlotFunc, 36

scatterPlots, 38  
sortExpressionSet, 41  
strptime, 33, 37, 41

write.table, 21