Package ‘BPRMeth’

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Type Package
Title Model higher-order methylation profiles
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Description BPRMeth package uses the Binomial Probit Regression
  likelihood to model methylation profiles and extract higher
  order features. These features quantitate precisely notions of
  shape of a methylation profile. Using these higher order
  features across promoter-proximal regions, we construct a
  powerful predictor of gene expression. Also, these features are
  used to cluster proximal-promoter regions using the EM
  algorithm.
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R topics documented:

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**Description**

`boxplot_cluster_gex` creates a boxplot of clustered gene expression levels which depend on the clustered methylation profiles. Each colour denotes a different cluster.

**Usage**

```r
boxplot_cluster_gex(bpr_cluster_obj, gex, main_lab = "Gene expression levels")
```

**Arguments**

- `bpr_cluster_obj`
  - The output of the `bpr_cluster_wrap` function.
- `gex`
  - The vector of gene expression data for each promoter region.
- `main_lab`
  - The title of the plot

**Value**

The figure to be plotted in the device.

**Author(s)**

C.A.Kapourani <C.A.Kapourani@ed.ac.uk>

**See Also**

`plot_cluster_prof`, `plot_scatter_gex`, `plot_fitted_profiles`
Examples

# Cluster methylation profiles using 4 RBFs
obs <- meth_data
basis <- create_rbf_object(M = 4)
res <- bpr_cluster_wrap(x = obs, K = 3, em_max_iter = 5, opt_itnmax = 4,
init_opt_itnmax = 5, is_parallel = FALSE)

# Create the plot
boxplot_cluster_gex(bpr_cluster_obj = res, gex = gex_data)

BPRMeth

Description

Higher order methylation features for clustering and prediction in epigenomic studies

Usage

.datatable.aware

Format

An object of class logical of length 1.

Value

BPRMeth main package documentation.

Author(s)

C.A.Kapourani <C.A.Kapourani@ed.ac.uk>

bpr_cluster_wrap

Cluster methylation profiles

Description

bpr_cluster_wrap is a wrapper function that clusters methylation profiles using the EM algorithm. Initially, it performs parameter checking, and initializes main parameters, such as mixing proportions, basis function coefficients, then the EM algorithm is applied and finally model selection metrics are calculated, such as BIC and AIC.

Usage

bpr_cluster_wrap(x, K = 3, pi_k = NULL, w = NULL, basis = NULL,
em_max_iter = 100, epsilon_conv = 1e-04, opt_method = "CG",
opt_itnmax = 100, init_opt_itnmax = 100, is_parallel = TRUE,
no_cores = NULL, is_verbose = FALSE)
Arguments

- **x**: The binomial distributed observations, which has to be a list of elements of length N, where each element is an L x 3 matrix of observations, where 1st column contains the locations. The 2nd and 3rd columns contain the total trials and number of successes at the corresponding locations, respectively. See `process_haib_caltech_wrap` on a possible way to get this data structure.

- **K**: Integer denoting the number of clusters K.

- **pi_k**: Vector of length K, denoting the mixing proportions.

- **w**: A MxK matrix, where each column consists of the basis function coefficients for each corresponding cluster.

- **basis**: A 'basis' object. E.g. see `create_rbf_object`.

- **em_max_iter**: Integer denoting the maximum number of EM iterations.

- **epsilon_conv**: Numeric denoting the convergence parameter for EM.

- **opt_method**: The optimization method to be used. See `optim` for possible methods. Default is "CG".

- **opt_itnmax**: Optional argument giving the maximum number of iterations for the corresponding method. See `optim` for details.

- **init_opt_itnmax**: Optimization iterations for obtaining the initial EM parameter values.

- **is_parallel**: Logical, indicating if code should be run in parallel.

- **no_cores**: Number of cores to be used, default is max_no_cores - 2.

- **is.verbose**: Logical, print results during EM iterations.

Value

A 'bpr_cluster' object which, in addition to the input parameters, consists of the following variables:

- **pi_k**: Fitted mixing proportions.

- **w**: A MxK matrix with the fitted coefficients of the basis functions for each cluster k.

- **NLL**: The Negative Log Likelihood after the EM algorithm has finished.

- **post_prob**: Posterior probabilities of each promoter region belonging to each cluster.

- **labels**: Hard clustering assignments of each observation/promoter region.

- **BIC**: Bayesian Information Criterion metric.

- **AIC**: Akaike Information Criterion metric.

- **ICL**: Integrated Complete Likelihood criterion metric.

Author(s)

C.A.Kapourani <C.A.Kapourani@ed.ac.uk>

Examples

```r
ex_data <- meth_data
data_clust <- bpr_cluster_wrap(x = ex_data, em_max_iter = 3, opt_itnmax = 5, init_opt_itnmax = 10, is_parallel = FALSE)
```
bpr_optimize

Optimize BPR negative log likelihood function

Description

The function bpr_optimize minimizes the negative log likelihood of the BPR function. Since it cannot be evaluated analytically, an optimization procedure is used. The optim packages is used for performing optimization.

Usage

bpr_optim(x, ...)

## S3 method for class 'list'
bpr_optim(x, w = NULL, basis = NULL, fit_feature = "RMSE", cpg_dens_feat = TRUE, opt_method = "CG", opt_itnmax = 100, is_parallel = TRUE, no_cores = NULL, ...)

## S3 method for class 'matrix'
bpr_optim(x, w = NULL, basis = NULL, fit_feature = "RMSE", cpg_dens_feat = TRUE, opt_method = "CG", opt_itnmax = 100, ...)

Arguments

x The input object, either a matrix or a list.
...
Additional parameters.
w A vector of parameters (i.e. coefficients of the basis functions)
basis A 'basis' object. E.g. see create_rbf_object.
fit_feature Return additional feature on how well the profile fits the methylation data. Either NULL for ignoring this feature or one of the following: 1) "RMSE" for returning the fit of the profile using the RMSE as measure of error or 2) "NLL" for returning the fit of the profile using the Negative Log Likelihood as measure of error.
cpg_dens_feat Logical, whether to return an additional feature for the CpG density across the promoter region.
opt_method The optimization method to be used. See optim for possible methods. Default is "CG".
opt_itnmax Optional argument giving the maximum number of iterations for the corresponding method. See optim for details.
is_parallel Logical, indicating if code should be run in parallel.
no_cores Number of cores to be used, default is max_no_cores - 2.

Value

Depending on the input object x:

- If x is a list: An object containing the following elements:
bpr_predict_wrap

Predict gene expression from methylation profiles

Description

bpr_predict_wrap is a function that wraps all the necessary subroutines for performing prediction on gene expression levels. Initially, it optimizes the parameters of the basis functions so as to learn the methylation profiles. Then, uses the learned parameters / coefficients of the basis functions as input features for performing regression in order to predict the corresponding gene expression levels.
bpr_predict_wrap

Usage

bpr_predict_wrap(formula = NULL, x, y, model_name = "svm", w = NULL, basis = NULL, train_ind = NULL, train_perc = 0.7, fit_feature = "RMSE", cpg_dens_feat = TRUE, opt_method = "CG", opt_itnmax = 100, is_parallel = TRUE, no_cores = NULL, is_summary = TRUE)

Arguments

formula An object of class formula, e.g. see lm function. If NULL, the simple linear regression model is used.
x The binomial distributed observations, which has to be a list of elements of length N, where each element is an L x 3 matrix of observations, where 1st column contains the locations. The 2nd and 3rd columns contain the total trials and number of successes at the corresponding locations, respectively. See process_haib_caltech_wrap on a possible way to get this data structure.
y Corresponding gene expression data for each element of the list x.
model_name A string denoting the regression model. Currently, available models are: "svm", "randomForest", "rlm", "mars" and "lm".
w Optional vector of initial parameter / coefficient values.
basis Optional basis function object, default is an 'rbf' object, see create_rbf_object.
train_ind Optional vector containing the indices for the train set.
train_perc Optional parameter for defining the percentage of the dataset to be used for training set, the remaining will be the test set.
fit_feature Return additional feature on how well the profile fits the methylation data. Either NULL for ignoring this feature or one of the following: 1) "RMSE" for returning the fit of the profile using the RMSE as measure of error or 2) "NLL" for returning the fit of the profile using the Negative Log Likelihood as measure of error.
cpg_dens_feat Logical, whether to return an additional feature for the CpG density across the promoter region.
opt_method The optimization method to be used. See optim for possible methods. Default is "CG".
opt_itnmax Optional argument giving the maximum number of iterations for the corresponding method. See optim for details.
is_parallel Logical, indicating if code should be run in parallel.
no_cores Number of cores to be used, default is max_no_cores - 2.
is_summary Logical, print the summary statistics.

Value

A 'bpr_predict' object which, in addition to the input parameters, consists of the following variables:

- \( W_{\text{opt}} \): An \( N \times (M+1) \) matrix with the optimized parameter values. Each row of the matrix corresponds to each element of the list \( x \). The columns are of the same length as the parameter vector \( w \) (i.e. number of basis functions).
- \( \mu_s \): An \( N \times M \) matrix with the RBF centers if basis object is create_rbf_object, otherwise NULL.
create_basis

- train: The training data.
- test: The test data.
- gex_model: The fitted regression model.
- train_pred: The predicted values for the training data.
- test_pred: The predicted values for the test data.
- train_errors: The training error metrics.
- test_errors: The test error metrics.

Author(s)

C.A.Kapourani <C.A.Kapourani@ed.ac.uk>

See Also

bpr_optimize, create_basis, eval_functions, train_model_gex, predict_model_gex

Examples

```
obsv <- meth_data
y <- gex_data
basis <- create_rbf_object(M = 5)
out <- bpr_predict_wrap(x = obs, y = y, basis = basis,
                        is_parallel = FALSE, opt_itnmax = 10)
```

create_basis

Create basis objects

Description

These functions create different basis objects. These objects can be used as input to complex functions in order to perform computations depending on the class of the basis function.

Usage

```
create_rbf_object(M = 2, gamma = NULL, mus = NULL, eq_spaced_mus = TRUE,
                  whole_region = TRUE)
create_polynomial_object(M = 1)
```

Arguments

- M: The number of the basis functions.
- gamma: Inverse width of radial basis function.
- mus: Optional centers of the RBF.
- eq_spaced_mus: Logical, if TRUE, equally spaced centers are created, otherwise centers are created using kmeans algorithm.
- whole_region: Logical, indicating if the centers will be evaluated equally spaced on the whole region, or between the min and max of the observation values.
create_methyl_region

Value
A basis object of class 'rbf' or 'polynomial'.

Author(s)
C.A.Kapourani <C.A.Kapourani@ed.ac.uk>

See Also
eval_functions, bpr_optimize

Examples
(obj <- create_rbf_object(M = 2))
#---------------------------------
(obj <- create_polynomial_object(M = 2))

create_methyl_region

Create methylation regions for each gene promoter.

Description
create_methyl_region creates methylation regions using BS-Seq and annotated gene promoter regions. BS-Seq data give information for the methylation of CpGs individually, and annotated data are used to locate the TSS of each gene and its promoter region.

Usage
create_methyl_region(bs_data, prom_region, cpg_density = 10, sd_thresh = 0.1, ignore_strand = TRUE, fmin = -1, fmax = 1)

Arguments
bs_data GRanges object containing the BS-Seq data. The GRanges object should also have two additional metadata columns named total_reads and meth_reads. A GRanges object used in this function can be the output of read_bs_encode_haib or its wrapper function preprocess_bs_seq.
prom_region GRanges object containing promoter regions, i.e. N bp upstream and M bp downstream of TSS location. The GRanges object should also have one additional metadata column named tss. A GRanges object used in this function can be the output of create_prom_region.
cpg_density Optional integer defining the minimum number of CpGs that have to be in a methylated region. Regions with less than n CpGs are discarded.
sd_thresh Optional numeric defining the minimum standard deviation of the methylation change in a region. This is used to filter regions with no methylation change.
ignore_strand Logical, whether or not to ignore strand information.
fmin Optional minimum range value for region location scaling. Under this version, this parameter should be left to its default value.
create_prom_region

fmax

Optional maximum range value for region location scaling. Under this version, this parameter should be left to its default value.

Value

A methyl_region object containing the following information:

- **meth_data**: A list containing methylation data, where each entry in the list is an \( L_i \times 3 \) dimensional matrix, where \( L_i \) denotes the number of CpGs found in region \( i \). The columns contain the following information:
  1. 1st column:Contains the locations of CpGs relative to TSS. Note that the actual locations are scaled to the \((f_{\text{min}}, f_{\text{max}})\) region.
  2. 2nd column:Contains the total reads of each CpG in the corresponding location.
  3. 3rd column:Contains the methylated reads each CpG in the corresponding location.

- **prom_ind**: A vector storing the corresponding promoter indices, so as to map each methylation region with its corresponding gene promoter.

The lengths of \texttt{meth_data} and \texttt{prom_ind} should be the same.

Author(s)

C.A.Kapourani <C.A.Kapourani@ed.ac.uk>

See Also

preprocess_bs_seq, create_prom_region

Examples

```r
# Obtain the path to the BS file and then read it
bs_file <- system.file("extdata", "rrbs.bed", package = "BPRMeth")
between_data <- read_bs_encode_haib(bs_file)

# Create promoter regions
rnaseq_file <- system.file("extdata", "rnaseq.bed", package = "BPRMeth")
annot_data <- read_rna_encode_caltech(rnaseq_file)
prom_region <- create_prom_region(annot_data)

# Finally, create methylation regions
meth_region <- create_methyl_region(between_data, prom_region)
```

create_prom_region

Create promoter regions from gene annotation data.

Description

create_prom_region creates promoter region from gene annotation data. Using the TSS of gene annotation data as ground truth labels we create promoter regions \( N \) bp upstream and \( M \) bp downstream of TSS.
Usage

create_prom_region(annot_data, chrom_size = NULL, upstream = -7000, downstream = 7000)

Arguments

annot_data A GRanges object containing the gene annotation data. This for example can be RNA-Seq data output from read_rna_encode_caltech.

chrom_size Optional data.table containing chromosome sizes, e.g. using the read_chrom_size function.

upstream Integer defining the length of bp upstream of TSS.

downstream Integer defining the length of bp downstream of TSS.

Value

A GRanges object containing the promoter regions data.

The GRanges object contains one additional metadata column:

• tss: TSS of each gene promoter.

This column can be accessed as follows: granges_object$tss

Author(s)

C.A.Kapourani <C.A.Kapourani@ed.ac.uk>

See Also

create_methyl_region, read_chrom_size, read_rna_encode_caltech

Examples

# Obtain the path to the file and then read it
rnaseq_file <- system.file("extdata", "rnaseq.bed", package = "BPRMeth")
annot_data <- read_rna_encode_caltech(rnaseq_file)
prom_region <- create_prom_region(annot_data)

# Extract the TSS
tss <- prom_region$tss

eval_functions

Evaluate basis functions

Description

Method for evaluating an M basis function model with observation data obs and coefficients w.
Usage

eval_probit_function(x, ...)

eval_function(x, ...)

## S3 method for class 'rbf'
eval_function(x, obs, w, ...)

## S3 method for class 'polynomial'
eval_function(x, obs, w, ...)

Arguments

x

The basis function object.

...

Optional additional parameters

obs

Observation data.

w

Vector of length M, containing the coefficients of an Mth-order basis function.

Value

The evaluated function values.

NOTE that the eval_probit_function computes the probit transformed basis function values.

Author(s)

C.A.Kapourani <C.A.Kapourani@ed.ac.uk>

See Also

create_basis

Examples

# Evaluate the probit transformed basis function values
x <- create_rbf_object(M=2)
obs <- c(1,2,3)
w <- c(0.1, 0.3, -0.6)
out <- eval_probit_function(x, obs, w)

# Evaluate the RBF basis function values
x <- create_rbf_object(M=2, mus = c(2,2.5))
obs <- c(1,2,3)
w <- c(0.1, 0.3, -0.6)
out <- eval_function(x, obs, w)

# Evaluate the Polynomial basis function values
x <- create_polynomial_object(M=2)
obs <- c(1,2,3)
w <- c(0.1, 0.3, -0.6)
out <- eval_function(x, obs, w)
**gex_data**

*Synthetic data for mpgex package*

**Description**

Corresponding gene expression data for the `meth_data`

**Usage**

```r
gex_data
```

**Format**

A vector of length 600

**Value**

Synthetic gene expression data

---

**meth_data**

*Synthetic data for BPRMeth package*

**Description**

A synthetic dataset containing 600 entries.

**Usage**

```r
meth_data
```

**Format**

A list with 600 elements, where each element is a 3x3 matrix of observations, where:

1. **1st column** locations of observations
2. **2nd column** total trials at corresponding locations
3. **3rd column** number of successes at corresponding locations

**Value**

Synthetic methylation data
plot_cluster_prof  
Plot of clustered methylation profiles

Description

plot_cluster_prof creates a plot of cluster methylation profiles, where each colour denotes a different cluster.

Usage

plot_cluster_prof(bpr_cluster_obj, 
                 main_lab = "Clustered methylation profiles")

Arguments

bpr_cluster_obj
  The output of the bpr_cluster_wrap function.
main_lab
  The title of the plot

Value

The figure to be plotted in the device.

Author(s)

C.A.Kapourani <C.A.Kapourani@ed.ac.uk>

See Also

plot_scatter_gex, plot_fitted_profiles, boxplot_cluster_gex

Examples

# Cluster methylation profiles using 4 RBFs
obs <- meth_data
basis <- create_rbf_object(M = 4)
res  <- bpr_cluster_wrap(x = obs, K = 3, em_max_iter = 5, opt_itnmax = 4, 
                         init_opt_itnmax = 5, is_parallel = FALSE)

# Create the plot
plot_cluster_prof(bpr_cluster_obj = res)
plot_fitted_profiles  

Plot the fit of methylation profiles across a region

Description

plot_fitted_profiles is a simple function for plotting the methylation data across a given region, together with the fit of the methylation profiles.

Usage

plot_fitted_profiles(region, X, fit_prof, fit_mean = NULL, title = "Gene promoter", ...)

Arguments

region  
Promoter region number
X  
Methylation data observations
fit_prof  
Fitted profile
fit_mean  
Fitted mean function
title  
Title of the plot
...  
Additional parameters

Value

The figure to be plotted in the device.

Author(s)

C.A.Kapurani <C.A.Kapurani@ed.ac.uk>

See Also

plot_cluster_prof, plot_scatter_gex, boxplot_cluster_gex

Examples

# Fit methylation profiles using 8 RBFs
obs <- meth_data
y <- gex_data
basis <- create_rbf_object(M = 8)
out <- bpr_predict_wrap(x = obs, y = y, basis = basis,
is_parallel = FALSE, opt_itnmax = 10)

# Create the plot
plot_fitted_profiles(region = 16, X = meth_data, fit_prof = out)
plot_scatter_gex

Scatter plot of predicted vs measured gene expression levels

Description

plot_scatter_gex creates a scatter plot of predicted gene expression values on the x-axis versus the measured gene expression values on the y-axis.

Usage

```r
plot_scatter_gex(bpr_predict_obj, main_lab = "Methylation Profile", isMargins = TRUE)
```

Arguments

- `bpr_predict_obj` - The output of the `bpr_predict_wrap` function.
- `main_lab` - The title of the plot.
- `isMargins` - Use specific margins or not.

Value

The figure to be plotted in the device.

Author(s)

C.A.Kapourani <C.A.Kapourani@ed.ac.uk>

See Also

`plot_cluster_prof, plot_fitted_profiles, boxplot_cluster_gex`

Examples

```r
# Fit methylation profiles using 8 RBFs
obs <- meth_data
y <- gex_data
basis <- create_rbf_object(M = 8)
res <- bpr_predict_wrap(x = obs, y = y, basis = basis,
                        is_parallel = FALSE, opt_itnmax = 10)

# Create the scatter plot
plot_scatter_gex(bpr_predict_obj = res)
```
pool_bs_seq_rep  Read and pool replicates from BS-Seq data

Description
pool_bs_seq_rep reads and pools replicate methylation data from BS-Seq experiments that are either in Encode RRBS or Bismark Cov format. Read the Important section below on when to use this function.

Usage
pool_bs_seq_rep(files, file_format = "encode_rrbs", chr_discarded = NULL)

Arguments
files A vector of filenames containing replicate experiments. This can also be just a single replicate.
file_format A string denoting the file format that the BS-Seq data are stored. Current version allows "encode_rrbs" or "bismark_cov" formats.
chr_discarded A vector with chromosome names to be discarded.

Value
A GRanges object. The GRanges object contains two additional metadata columns:

• total_reads: total reads mapped to each genomic location.
• meth_reads: methylated reads mapped to each genomic location.

These columns can be accessed as follows: granges_object$total_reads

Important
Unless you want to create a different workflow when processing the BS-Seq data, you should NOT call this function, since this is a helper function. Instead you should call the preprocess_bs_seq function.

Information about the file formats can be found in the following links:

Encode RRBS format: http://rohsdb.cmb.usc.edu/GBshape/cgi-bin/hgTables?db=hg19&hgta_group=regulation&hgta_track=wgEncodeHaibMethylRrbs&hgta_table=wgEncodeHaibMethylRrbsBcBreast0203015BiochainSitesRep2&hgta_doSchema=describe+table+schema

Bismark Cov format: http://rnbeads.mpi-inf.mpg.de/data/RnBeads.pdf

Author(s)
C.A.Kapourani <C.A.Kapourani@ed.ac.uk>

See Also
read_bs_bismark_cov, read_bs_encode_haib, preprocess_bs_seq
Examples

# Obtain the path to the file
bs_file1 <- system.file("extdata", "rrbs.bed", package = "BPRMeth")
bs_file2 <- system.file("extdata", "rrbs.bed", package = "BPRMeth")

# Concatenate the files
bs_files <- c(bs_file1, bs_file2)
# Pool the replicates
pooled_data <- pool_bs_seq_rep(bs_files)

predict_model_gex    Predict gene expression model from methylation profiles

Description

predict_model_gex makes predictions of gene expression levels using a model trained on higher order methylation features extracted from specific genomic regions.

Usage

predict_model_gex(model, test, is_summary = TRUE)

Arguments

model    The fitted regression model, i.e. the output of train_model_gex.
test     The testing data.
is_summary Logical, print the summary statistics.

Value

A list containing the following elements:

- test_pred: The predicted values for the test data.
- test_errors: The test error metrics.

Author(s)

C.A.Kapourani <C.A.Kapourani@ed.ac.uk>

See Also

train_model_gex
Examples

# Create synthetic data
train_data <- data.frame(x = rnorm(20), y=rnorm(20, 1, 4))
test_data <- data.frame(x = rnorm(20), y=rnorm(20, 1, 3))

# Train the model
train_model <- train_model_gex(formula = y~., train = train_data)

# Make predictions
res <- predict_model_gex(model = train_model$gex_model, test = test_data)

preprocess_bs_seq

Pre-process BS-Seq data in any given format

Description

preprocess_bs_seq is a general function for reading and preprocessing BS-Seq data. If a vector of files is given, these are considered as replicates and are pooled together. Finally, noisy reads are discarded.

Usage

preprocess_bs_seq(files, file_format = "encode_rrbs", chr_discarded = NULL, min_bs_cov = 4, max_bs_cov = 1000)

Arguments

files A vector of filenames containing replicate experiments. This can also be just a single replicate.

file_format A string denoting the file format that the BS-Seq data are stored. Current version allows "encode_rrbs" or "bismark_cov" formats.

chr_discarded A vector with chromosome names to be discarded.

min_bs_cov The minimum number of reads mapping to each CpG site. CpGs with less reads will be considered as noise and will be discarded.

max_bs_cov The maximum number of reads mapping to each CpG site. CpGs with more reads will be considered as noise and will be discarded.

Value

A GRanges object. The GRanges object contains two additional metadata columns:

- total_reads: total reads mapped to each genomic location.
- meth_reads: methylated reads mapped to each genomic location.

These columns can be accessed as follows: granges_object$total_reads
Additional Info

Information about the file formats can be found in the following links:

Encode RRBS format: [http://rohsdb.cmb.usc.edu/GBshape/cgi-bin/hgTables?db=hg19&hgta_group=regulation&hgta_track=wgEncodeHaibMethylRrbs&hgta_table=wgEncodeHaibMethylRrbsBcbreast0203015BiochainSitesRep2&hgta_doSchema=describe+table+schema](http://rohsdb.cmb.usc.edu/GBshape/cgi-bin/hgTables?db=hg19&hgta_group=regulation&hgta_track=wgEncodeHaibMethylRrbs&hgta_table=wgEncodeHaibMethylRrbsBcbreast0203015BiochainSitesRep2&hgta_doSchema=describe+table+schema)


Author(s)

C.A. Kapourani <C.A.Kapourani@ed.ac.uk>

See Also

read_bs_bismark_cov, read_bs_encode_haib
pool_bs_seq_rep

Examples

```
# Obtain the path to the files
bs_file <- system.file("extdata", "rrbs.bed", package = "BPRMeth")
bs_data <- preprocess_bs_seq(bs_file, file_format = "encode_rrbs")

# Extract the total reads and methylated reads
total_reads <- bs_data$total_reads
meth_reads <- bs_data$meth_reads
```

---

`preprocess_final_HTS_data`

*Pre-process final HTS data for downstream analysis*

Description

`preprocess_final_HTS_data` performs a final filtering and preprocessing on the data for use in downstream analysis. These include, removing noisy gene expression data, removing or not unexpressed genes and log2-transforming of the FPKM values.

Usage

```
preprocess_final_HTS_data(methyl_region, prom_reg, rna_data,
  gene_log2_transf = TRUE, gene_outl_thresh = TRUE, gex_outlier = 300)
```

Arguments

- `methyl_region`: Methylation region data, which are the output of the "create_methyl_region" function.
- `prom_reg`: A `GRanges` object containing corresponding annotated promoter regions for each entry of the `methyl_region` list.
- `rna_data`: A `GRanges` object containing corresponding RNA-Seq data for each entry of the `methyl_region` list. This is the output of the "read_rna_encode_caltech" function.
process_haib_caltech_wrap

**gene_log2_transf**
Logical, whether or not to log2 transform the gene expression data.

**gene_outl_thresh**
Logical, whether or not to remove outlier gene expression data.

**gex_outlier**
Numeric, denoting the threshold above of which the gene expression data (before the log2 transformation) are considered as noise.

**Value**
An object which contains following information:

- **methyl_region**: The subset of promoter methylation region data after the filtering process.
- **gex**: A vectoring storing only the corresponding gene expression values for each promoter region.
- **rna_data**: The corresponding gene expression data stored as a GRanges object.

**Author(s)**
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**See Also**
read_rna_encode_caltech process_haib_caltech_wrap

**Examples**

```r
# Obtain the path to the BS file and then read it
bs_file <- system.file("extdata", "rrbs.bed", package = "BPRMeth")
bs_data <- read_bs_encode_haib(bs_file)

# Create promoter regions
rnaseq_file <- system.file("extdata", "rnaseq.bed", package = "BPRMeth")
annot_data <- read_rna_encode_caltech(rnaseq_file)
prom_region <- create_prom_region(annot_data)

# Create methylation regions
methyl_region <- create_methyl_region(bs_data, prom_region)

# Finally preprocess the HTS data
res <- preprocess_final_HTS_data(methyl_region, prom_region, annot_data)
```

---

**Description**

process_haib_caltech_wrap is a wrapper method for processing HTS data and returning the methylation promoter regions and the corresponding gene expression data for those promoter regions. Note that the format of BS-Seq data should be in the Encode Haib bed format and for the RNA-Seq data in Encode Caltech bed format.
process_haib_caltech_wrap

Usage

process_haib_caltech_wrap(bs_files, rna_files, chrom_size_file = NULL,
chr_discarded = NULL, upstream = -7000, downstream = 7000,
min_bs_cov = 4, max_bs_cov = 1000, cpg_density = 10, sd_thresh = 0.1,
ignore_strand = TRUE, gene_log2_transf = TRUE, gene_outl_thresh = TRUE,
gex_outlier = 300, fmin = -1, fmax = 1)

Arguments

bs_files Filename (or vector of filenames if there are replicates) of the BS-Seq `.bed` formatted data to read values from.

rna_files Filename of the RNA-Seq `.bed` formatted data to read values from. Currently, this version does not support pooling RNA-Seq replicates.

chrom_size_file Optional filename containing genome chromosome sizes.

chr_discarded A vector with chromosome names to be discarded.

upstream Integer defining the length of bp upstream of TSS for creating the promoter region.

downstream Integer defining the length of bp downstream of TSS for creating the promoter region.

min_bs_cov The minimum number of reads mapping to each CpG site. CpGs with less reads will be considered as noise and will be discarded.

max_bs_cov The maximum number of reads mapping to each CpG site. CpGs with more reads will be considered as noise and will be discarded.

cpg_density Optional integer defining the minimum number of CpGs that have to be in a methylated region. Regions with less than n CpGs are discarded.

sd_thresh Optional numeric defining the minimum standard deviation of the methylation change in a region. This is used to filter regions with no methylation change.

ignore_strand Logical, whether or not to ignore strand information.

gene_log2_transf Logical, whether or not to log2 transform the gene expression data.

gene_outl_thresh Logical, whether or not to remove outlier gene expression data.

gex_outlier Numeric, denoting the threshold above of which the gene expression data (before the log2 transformation) are considered as noise.

fmin Optional minimum range value for region location scaling. Under this version, this parameter should be left to its default value.

fmax Optional maximum range value for region location scaling. Under this version, this parameter should be left to its default value.

Value

A `processHTS` object which contains following information:

- methyl_region: A list containing methylation data, where each entry in the list is an $L_i \times 3$ dimensional matrix, where $L_i$ denotes the number of CpGs found in region $i$. The columns contain the following information:
1. 1st column: Contains the locations of CpGs relative to TSS. Note that the actual locations are scaled to the (fmin, fmax) region.
2. 2nd column: Contains the total reads of each CpG in the corresponding location.
3. 3rd column: Contains the methylated reads each CpG in the corresponding location.

- prom_region: A GRanges object containing corresponding annotated promoter regions for each entry of the methyl_region list. The GRanges object has one additional metadata column named tss, which stores the TSS of each promoter.
- rna_data: A GRanges object containing the corresponding RNA-Seq data for each entry of the methyl_region list. The GRanges object has three additional metadata columns which are explained in read_rna_encode_caltech
- upstream: Integer defining the length of bp upstream of TSS.
- downstream: Integer defining the length of bp downstream of TSS.
- cpg_density: Integer defining the minimum number of CpGs that have to be in a methylated region. Regions with less than n CpGs are discarded.
- sd_thresh: Numeric defining the minimum standard deviation of the methylation change in a region. This is used to filter regions with no methylation change.
- fmin: Minimum range value for region location scaling.
- fmax: Maximum range value for region location scaling.

Author(s)

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Examples

# Obtain the path to the files
rrbs_file <- system.file("extdata", "rrbs.bed", package = "BPRMeth")
rnaseq_file <- system.file("extdata", "rnaseq.bed", package = "BPRMeth")
proc_data <- process_haib_caltech_wrap(rrbs_file, rnaseq_file)

---

**Description**

read_bs_bismark_cov reads a file containing methylation data from BS-Seq experiments using the fread function. The BS-Seq file should be in Bismark Cov format. Read the Important section below on when to use this function.

**Usage**

read_bs_bismark_cov(file, chr_discarded = NULL, is_GRanges = TRUE)

**Arguments**

- **file**: The name of the file to read data values from.
- **chr_discarded**: A vector with chromosome names to be discarded.
- **is_GRanges**: Logical: if TRUE a GRanges object is returned, otherwise a data.frame object is returned.
Value

A `GRanges` object if `is_GRanges` is TRUE, otherwise a `data.table` object. The `GRanges` object contains two additional metadata columns:

- `total_reads`: total reads mapped to each genomic location.
- `meth_reads`: methylated reads mapped to each genomic location.

These columns can be accessed as follows: `granges_object$total_reads`

Important

Unless you want to create a different workflow when processing the BS-Seq data, you should NOT call this function, since this is a helper function. Instead you should call the `preprocess_bs_seq` function.

Author(s)

C.A. Kapourani <C.A.Kapourani@ed.ac.uk>

References

http://rnbeads.mpi-inf.mpg.de/data/RnBeads.pdf

See Also

`pool_bs_seq_rep`, `preprocess_bs_seq`

Examples

## Not run:
# Download the files and change the working directory to that location
file <- "name_of_bismark_file"
bs_data <- read_bs_bismark_cov(file)
# Extract the total reads and methylated reads
total_reads <- bs_data$total_reads
meth_reads <- bs_data$meth_reads

## End(Not run)
**Arguments**

- **file** The name of the file to read data values from.
- **chr_discarded** A vector with chromosome names to be discarded.
- **is_GRanges** Logical: if TRUE a GRanges object is returned, otherwise a data.frame object is returned.

**Value**

A GRanges object if is_GRanges is TRUE, otherwise a data.table object. The GRanges object contains two additional metadata columns:

- **total_reads**: total reads mapped to each genomic location.
- **meth_reads**: methylated reads mapped to each genomic location.

These columns can be accessed as follows: `granges_object$total_reads`

**Important**

Unless you want to create a different workflow when processing the BS-Seq data, you should NOT call this function, since this is a helper function. Instead you should call the `preprocess_bs_seq` function.

**Author(s)**

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**References**

http://rohsdb.cmb.usc.edu/GShape/cgi-bin/hgTables?db=hg19&hgta_group=regulation&hgta_track=wgEncodeHaibMethylRrbs&hgta_table=wgEncodeHaibMethylRrbsBcbreast0203015BiochainSitesRep2&hgta_doSchema=describe+table+schema

**See Also**

- `pool_bs_seq_rep`
- `preprocess_bs_seq`

**Examples**

```r
# Obtain the path to the file and then read it
bs_file <- system.file("extdata", "rrbs.bed", package = "BPRMeth")
bs_data <- read_bs_encode_haib(bs_file)

# Extract the total reads and methylated reads
total_reads <- bs_data$total_reads
meth_reads <- bs_data$meth_reads
```
**Description**

read_chrom_size reads a file containing genome chromosome sizes using the fread function.

**Usage**

read_chrom_size(file)

**Arguments**

file

The name of the file to read data values from.

**Value**

A data.table object.

**Author(s)**

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**See Also**

read_rna_encode_caltech, read_bs_encode_haib

**Examples**

```r
chr_file <- system.file("extdata", "hg19.chr.sizes", package = "BPRMeth")
chr.data <- read_chrom_size(chr_file)

# Extract the size of the chr1
chr.data[1]
```

---

**Description**

read_encode_cgi reads a file containing CpG island (CGI) locations in the human genome using the fread function.

**Usage**

read_encode_cgi(file, is_GRanges = TRUE)
Arguments

- **file**: The name of the file to read data values from.
- **is_GRanges**: Logical: if TRUE a GRanges object is returned, otherwise a data.frame object is returned.

Value

A GRanges object if is_GRanges is TRUE, otherwise a data.table object.

The GRanges object contains one additional metadata column:

- **cgi_id**: Unique ID of the CpG Island.

This column can be accessed as follows: `granges_object$cgi_id`

Author(s)

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See Also

- `read_rna_encode_caltech`
- `read_bs_encode_haib`

Examples

```r
## Not run:
# Download the file and change the working directory to that location
file <- "name_of_CGI_file"
.cgi_data <- read_encode_cgi(file)

# Extract the CGI ID
cgi_id <- .cgi_data$cgi_id

## End(Not run)
```

---

read_rna_encode_caltech

*Read ENCODE Caltech bed formatted RNA-Seq file*

Description

read_rna_encode_caltech reads a file containing promoter annotation data together with gene expression levels from RNA-Seq experiments using the `scan` function. The RNA-Seq file should be in ENCODE Caltech bed format, e.g. use gtf2bed tool if your initial file is in gtf format.

Usage

```r
read_rna_encode_caltech(file, chr_discarded = NULL, is_GRanges = TRUE)
```
Arguments

- **file**: The name of the file to read data values from.
- **chr_discarded**: A vector with chromosome names to be discarded.
- **is_GRanges**: Logical: if TRUE a GRanges object is returned, otherwise a data.frame object is returned.

Value

A GRanges object if is_GRanges is TRUE, otherwise a data.table object.

The GRanges object contains three additional metadata columns:

- **ensembl_id**: Ensembl IDs of each gene promoter.
- **gene_name**: Gene name.
- **gene_fpkm**: Expression level in FPKM.

These columns can be accessed as follows: granges_object$ensembl_id

Author(s)

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See Also

read_chrom_size, read_bs_encode_haib

Examples

```r
# Obtain the path to the file and then read it
rnaseq_file <- system.file("extdata", "rnaseq.bed", package = "BPRMeth")
rna_data <- read_rna_encode_caltech(rnaseq_file)

# Extract the gene name and gene expression in fpkm
gene_name <- rna_data$gene_name
gene_fpkm <- rna_data$gene_fpkm
```

Description

train_model_gex trains a regression model for predicting gene expression levels by taking as input the higher order methylation features extracted from specific genomic regions.

Usage

```r
train_model_gex(formula = NULL, model_name = "svm", train, is_summary = TRUE)
```
train_model_gex

Arguments

- **formula**: An object of class formula, e.g. see `lm` function. If NULL, the simple linear regression model is used.
- **model_name**: A string denoting the regression model. Currently, available models are: "svm", "randomForest", "rlm", "mars" and "lm".
- **train**: The training data.
- **is_summary**: Logical, print the summary statistics.

Value

A list containing the following elements:

- **formula**: The formula that was used.
- **gex_model**: The fitted model.
- **train_pred**: The predicted values for the training data.
- **train_errors**: The training error metrics.

Author(s)

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See Also

- `predict_model_gex`

Examples

```r
# Create synthetic data
train_data <- data.frame(x = rnorm(20), y=rnorm(20, 1, 4))
res <- train_model_gex(formula = y~, train = train_data)

# Using a different model
res <- train_model_gex(model_name = "randomForest", train = train_data)
```
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