

# Package ‘MLP’

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**Title** Mean Log P Analysis

**Type** Package

**Description** Pathway analysis based on p-values associated to genes from a genes expression analysis of interest. Utility functions enable to extract pathways from the Gene Ontology Biological Process (GOBP), Molecular Function (GOMF) and Cellular Component (GOCC), Kyoto Encyclopedia of Genes of Genomes (KEGG) and Reactome databases. Methodology, and helper functions to display the results as a table, barplot of pathway significance, Gene Ontology graph and pathway significance are available.

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**biocViews** Genetics, GeneExpression, Pathways, Reactome, KEGG, GO

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**Imports** AnnotationDbi, gplots, graphics, stats, utils

**Suggests** GO.db, org.Hs.eg.db, org.Mm.eg.db, org.Rn.eg.db, org.Cf.eg.db, org.Mmu.eg.db, KEGGREST, annotate, Rgraphviz, GOstats, graph, limma, mouse4302.db, reactome.db

**Collate** 'addGeneSetDescription.R' 'getGeneSets.R' 'mlpBarplot.R' 'MLP.R' 'plotGeneSetSignificance.R' 'plotGOgraph.R' 'plot.MLP.R' 'plotQuantileCurves.R' 'utils.R'

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addGeneSetDescription *Utility function which adds the biological description of the gene sets as a column to the return value of the MLP function (data frame)*

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## Description

Utility function which adds the biological description of the gene sets as a column to the return value of the MLP function (data frame)

## Usage

```
addGeneSetDescription(object, geneSetSource = NULL)
```

## Arguments

object	object of class 'MLP' as produced by the 'MLP' function
geneSetSource	source to be used to construct the list of pathway categories; for public data sources, the user can specify a string (one of 'GOBP', 'GOMF', 'GOCC', 'KEGG' or 'REACTOME') and BioC packages will be used to construct the list of pathway categories; for non-public data sources, the user can pass the pathway data as a dataframe with (at least) the following four columns: PATHWAYID, TAXID, PATHWAYNAME and GENEID. It is assumed all columns are of type character. The 'geneSetSource' argument should be the same as the argument provided to the getGeneSets function; defaults to NULL

## Value

the data frame as returned by MLP enriched with an additional column geneSetDescription, providing a concise description of the gene set

## See Also

[MLP](#)

**Examples**

```

if (require(GO.db)){
  pathExamplePValues <- system.file("exampleFiles", "examplePValues.rda", package = "MLP")
  load(pathExamplePValues)
  geneSet <- getGeneSets(species = "Mouse", geneSetSource = "GOBP", entrezIdentifiers = names(examplePValues))
  mlpResult <- MLP(geneSet = geneSet, geneStatistic = examplePValues, addGeneSetDescription = FALSE)
  head(mlpResult)
  mlpResultsWithGSDescr <- addGeneSetDescription(object = mlpResult, geneSetSource = "GOBP")
  head(mlpResultsWithGSDescr)
}

```

---

formatKEGGDescription *Format KEGG descriptions*

---

**Description**

This includes the removal of the species

**Usage**

```
formatKEGGDescription(descriptions, species = "Mouse")
```

**Arguments**

descriptions    character vector with descriptions of KEGG pathways

species         character vector of length one indicating the species, one of 'Mouse', 'Human', 'Rat', 'Dog' or 'Rhesus'; defaults to 'Mouse'.

**Value**

formatted descriptions

---

getGeneSets                    *Prepare Pathway Data for the MLP Function*

---

**Description**

The return value of the getGeneSets function has as primary use to serve as geneSet argument for the MLP function

**Usage**

```
getGeneSets(species = "Mouse", geneSetSource = NULL, entrezIdentifiers)
```

## Arguments

species	character vector of length one indicating the species, one of 'Mouse', 'Human', 'Rat', 'Dog' or 'Rhesus'; defaults to 'Mouse'.
geneSetSource	source to be used to construct the list of pathway categories; for public data sources, the user can specify a string (one of 'GOBP', 'GOMF', 'GOCC', 'KEGG' or 'REACTOME') and BioC packages will be used to construct the list of pathway categories; for non-public data sources, the user can pass the pathway data as a dataframe with (at least) the following four columns: PATHWAYID, TAXID, PATHWAYNAME and GENEID. It is assumed all columns are of type character.
entrezIdentifiers	Entrez Gene identifiers used to subset the relevant gene set

## Value

object of class `geneSetMLP` which is essentially a named list of pathway categories. Each list component is named with the pathway ID and contains a vector of Entrez Gene identifiers related to that particular pathway.

The object contains additionally the attributes:

- 'species' and 'geneSetSource': species and geneSetSource (as provided as input)
- 'descriptions': named character vector with pathway descriptions. The vector is named with the pathway ID.

## Examples

```
if (require(GO.db) && require(org.Mm.eg.db)){
  pathExamplePValues <- system.file("exampleFiles", "examplePValues.rda", package = "MLP")
  load(pathExamplePValues)
  geneSet <- getGeneSets(species = "Mouse", geneSetSource = "GOBP", entrezIdentifiers = names(examplePValues)[
  geneSet <- getGeneSets(species = "Mouse", geneSetSource = "KEGG", entrezIdentifiers = names(examplePValues)[
}
```

---

MLP

*This function calculates p-values for each gene set based on row permutations of the gene p values or column permutations of the expression matrix; the p values can be obtained either as individual gene set p values or p values based on smoothing across gene sets of similar size.*

---

## Description

This function calculates p-values for each gene set based on row permutations of the gene p values or column permutations of the expression matrix; the p values can be obtained either as individual gene set p values or p values based on smoothing across gene sets of similar size.

**Usage**

```
MLP(
  geneSet,
  geneStatistic,
  minGenes = 5,
  maxGenes = 100,
  rowPermutations = TRUE,
  nPermutations = 100,
  smoothPValues = TRUE,
  probabilityVector = c(0.5, 0.9, 0.95, 0.99, 0.999, 0.9999, 0.99999),
  df = 9,
  addGeneSetDescription = TRUE
)
```

**Arguments**

<code>geneSet</code>	is the input list of gene sets (components) and gene IDs (character vectors). A gene set can, for example, be a GO category with for each category Entrez Gene identifiers; The <a href="#">getGeneSets</a> function can be used to construct the <code>geneSet</code> argument for different pathway sources.
<code>geneStatistic</code>	is either a named numeric vector (if <code>rowPermutations</code> is TRUE) or a numeric matrix of pvalues (if <code>rowPermutations</code> is FALSE). The names of the numeric vector or row names of the matrix should represent the gene IDs.
<code>minGenes</code>	minimum number of genes in a gene set for it to be considered (lower threshold for gene set size)
<code>maxGenes</code>	maximum number of genes in a gene set for it to be considered (upper threshold for gene set size)
<code>rowPermutations</code>	logical indicating whether to use row permutations (TRUE; default) or column permutations (FALSE)
<code>nPermutations</code>	is the number of simulations. By default 100 permutations are conducted.
<code>smoothPValues</code>	logical indicating whether one wants to calculate smoothed cut-off thresholds (TRUE; default) or not (FALSE).
<code>probabilityVector</code>	vector of quantiles at which p values for each gene set are desired
<code>df</code>	degrees of freedom for the <code>smooth.spline</code> function used in <code>getSmoothedPValues</code>
<code>addGeneSetDescription</code>	logical indicating whether a column with the gene set description be added to the output data frame; defaults to TRUE.

**Value**

data frame with four (or five) columns: `totalGeneSetSize`, `testedGeneSetSize`, `geneSetStatistic` and `geneSetPValue` and (if `addDescription` is set to TRUE) `geneSetDescription`; the rows of the data frame are ordered by ascending `geneSetPValue`.

**References**

Raghavan, Nandini et al. (2007). The high-level similarity of some disparate gene expression measures, *Bioinformatics*, 23, 22, 3032-3038.

**Examples**

```

if (require(GO.db)){
  pathExampleGeneSet <- system.file("exampleFiles", "exampleGeneSet.rda", package = "MLP")
  pathExamplePValues <- system.file("exampleFiles", "examplePValues.rda", package = "MLP")
  load(pathExampleGeneSet)
  load(pathExamplePValues)
  head(examplePValues)
  head(exampleGeneSet)
  mlpResult <- MLP(geneSet = exampleGeneSet, geneStatistic = examplePValues)
  head(mlpResult)
}

```

mlpBarplot

*Draw a Barplot for MLP Results***Description**

Draw a Barplot for MLP Results

**Usage**

```

mlpBarplot(
  object,
  nRow = 20,
  barColors = NULL,
  main = NULL,
  ylab = "",
  cex = 1
)

```

**Arguments**

object	object of class MLP
nRow	number of rows of the MLP data frame to depict in the barplot; defaults to 20.
barColors	vector of colors to use for the bars of the barplot; defaults to NULL; if NULL, three gray shades are used reflecting the proportion of tested genes of a gene set versus the total number of genes in a geneset. If the proportion exceeds 75%, the darkest shade is used; between 50 and 75% a moderately dark shade is used; below 50% a lighter gray shade is used.
main	main title; if NULL (default) "Effect of the treatment on <geneSetSource> gene sets" will be used
ylab	string with label for the y-axis
cex	numeric, cex used in par

**Value**

the midpoints of all the bars are returned invisibly (using the conventions of barplot); an MLP-specific barplot is drawn to the current device;

**See Also**

barplot

**Examples**

```
pathExampleMLPResult <- system.file("exampleFiles", "exampleMLPResult.rda", package = "MLP")
load(pathExampleMLPResult)
dev.new(width = 10, height = 10)
op <- par(mar = c(30, 10, 6, 2))
mlpBarplot(exampleMLPResult)
par(op)
```

plot.MLP

*Plot the Results of an MLP Run***Description**

Plot the Results of an MLP Run

**Usage**

```
## S3 method for class 'MLP'
plot(x, y = NULL, type = c("barplot", "GOgraph", "quantileCurves"), ...)
```

**Arguments**

x	object of class 'MLP'
y	argument added to comply with generic; not used, defaults to NULL
type	character of length one; one of 'barplot', 'GOgraph' or 'quantileCurves'
...	further arguments for the plot functions for each type

**Value**

for type = "barplot", the midpoints of the barplot

**Examples**

```
pathExampleMLPResult <- system.file("exampleFiles", "exampleMLPResult.rda", package = "MLP")
load(pathExampleMLPResult)
dev.new(width = 10, height = 10)
op <- par(mar = c(30, 10, 6, 2))
plot(exampleMLPResult, type = "barplot")
par(op)
plot(exampleMLPResult, type = "quantileCurves")
if (require(GO.db) && require(Rgraphviz)){
  plot(exampleMLPResult, type = "GOgraph")
}
```

---

plotGeneSetSignificance

*Plot the Significance for the Genes of a Given Gene Set*


---

### Description

Plot the Significance for the Genes of a Given Gene Set

### Usage

```
plotGeneSetSignificance(
  geneSet,
  geneSetIdentifier,
  geneStatistic,
  annotationPackage,
  barColors = NULL,
  descriptionInMainTitle = TRUE
)
```

### Arguments

geneSet	object of class 'geneSetMLP' as produced by function getGeneSets
geneSetIdentifier	identifier of the gene set for which a significance plot should be produced; character of length one
geneStatistic	named vector of gene statistics (e.g. p values); the names of the vector are Entrez Gene identifiers
annotationPackage	name of the annotation package to be used (without .db extension); character of length one
barColors	named vector of colors to use for the bars of the barplot; the names of the vector are Entrez Gene identifiers and the vector should be of length equal to the length of the geneStatistic vector defaults to NULL in which case 'grey50' is used
descriptionInMainTitle	Boolean whether or not to use the gene set description in the main title of the plot

### Value

no return value

### Examples

```
pathExamplePValues <- system.file("exampleFiles", "examplePValues.rda", package = "MLP")
pathExampleGeneSet <- system.file("exampleFiles", "exampleGeneSet.rda", package = "MLP")
pathExampleMLPResult <- system.file("exampleFiles", "exampleMLPResult.rda", package = "MLP")
load(pathExampleGeneSet)
load(pathExamplePValues)
load(pathExampleMLPResult)
# annotationPackage <- if (require(mouse4302mmentrezg.db)) "mouse4302mmentrezg" else "mouse4302"
annotationPackage <- "mouse4302"
```



```
geneSetID <- rownames(exampleMLPResult)[1]
dev.new(width = 10, height = 10)
op <- par(mar = c(25, 10, 6, 2))
plotGeneSetSignificance(
  geneSet = exampleGeneSet,
  geneSetIdentifier = geneSetID,
  geneStatistic = examplePValues,
  annotationPackage = annotationPackage
)
par(op)
```

---

plotGOgraph

*Graphical Representation of GO Based MLP Results*

---

### Description

Graphical Representation of GO Based MLP Results

### Usage

```
plotGOgraph(object, nRow = 5, main = NULL, nCutDescPath = 30)
```

### Arguments

object	object of class MLP (as produced by the MLP function)
nRow	number of GO IDs for which to produce the plot
main	main title of the graph; if NULL (default) the main title is set to 'GO graph'
nCutDescPath	number of characters at which the pathway description should be cut (inserted in a new line), 30 by default

### Value

GO graph is plotted to the current device

### Examples

```
if (require(GO.db) && require(Rgraphviz)){
  pathExampleMLPResult <- system.file("exampleFiles", "exampleMLPResult.rda", package = "MLP")
  load(pathExampleMLPResult)
  plotGOgraph(exampleMLPResult, main = "GO Graph")
}
```

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