

# Package ‘branchpointer’

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**Type** Package

**Title** Prediction of intronic splicing branchpoints

**Version** 1.23.0

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**Description** Predicts branchpoint probability for sites in intronic branchpoint windows. Queries can be supplied as intronic regions; or to evaluate the effects of mutations, SNPs.

**License** BSD\_3\_clause + file LICENSE

**LazyData** FALSE

**Depends** caret, R(>= 3.4)

**Imports** plyr, kernlab, gbm, stringr, cowplot, ggplot2, biomaRt,  
Biostrings, parallel, utils, stats,  
BSgenome.Hsapiens.UCSC.hg38, rtracklayer, GenomicRanges,  
GenomeInfoDb, IRanges, S4Vectors, data.table

**Suggests** knitr, BiocStyle

**RoxygenNote** 6.0.1

**VignetteBuilder** knitr

**biocViews** Software, GenomeAnnotation, GenomicVariation,  
MotifAnnotation

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gtfToExons	<i>Convert GTF file to exon location file</i>
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### Description

Converts a GTF annotation to exon locations

### Usage

```
gtfToExons(gtf)
```

### Arguments

gtf                   file containing the gtf annotation.

### Value

exon annotation GRanges

### Author(s)

Beth Signal

### Examples

```
smallExons <- system.file("extdata", "gencode.v26.annotation.small.gtf",
  package = "branchpointer")
exons <- gtfToExons(smallExons)
```

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`makeBranchpointWindowForExons`*Make branchpoint window regions*

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**Description**

Generate branchpoint window regions corresponding to annotated exon(s) within a queried gene, transcript or exon id

**Usage**

```
makeBranchpointWindowForExons(id, idType, exons, forceClosestExon = FALSE)
```

**Arguments**

<code>id</code>	identifier(s) for the query gene/transcript/exon id
<code>idType</code>	type of id to match in the exon annotation file ("gene_id", "transcript_id", or "exon_id")
<code>exons</code>	GRanges containing exon co-ordinates.
<code>forceClosestExon</code>	Force branchpointer to find the closest exon and not the exon annotated as 5' to the query

**Value**

Granges with formatted query

**Author(s)**

Beth Signal

**Examples**

```
smallExons <- system.file("extdata", "gencode.v26.annotation.small.gtf", package = "branchpointer")
exons <- gtfToExons(smallExons)
windowquery <- makeBranchpointWindowForExons("ENSG00000139618.14", "gene_id", exons)
windowquery <- makeBranchpointWindowForExons("ENST00000357654.7", "transcript_id", exons)
windowquery <- makeBranchpointWindowForExons("ENSE000003518965.1", "exon_id", exons)
```

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`makeBranchpointWindowForSNP`*Makes a branchpointer formatted GRanges object from refsnp ids*

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**Description**

Searches Biomart for refsnp ids, and pulls genomic location and sequence identity information  
Reformats alleles so each query has only one alternative allele

**Usage**

```
makeBranchpointWindowForSNP(refSNP, mart.snp, exons, maxDist = 50,  
  filter = TRUE)
```

**Arguments**

<code>refSNP</code>	Vector of refsnp ids
<code>mart.snp</code>	biomaRt mart object specifying the BioMart database and dataset to be used
<code>exons</code>	GRanges containing exon co-ordinates. Should be produced by <code>gtfToExons()</code>
<code>maxDist</code>	maximum distance a SNP can be from an annotated 3' exon.
<code>filter</code>	remove SNP queries prior to finding finding nearest exons?

**Value**

formatted SNP query GRanges

**Author(s)**

Beth Signal

**Examples**

```
smallExons <- system.file("extdata", "gencode.v26.annotation.small.gtf", package = "branchpointer")  
exons <- gtfToExons(smallExons)
```

```
mart.snp <- biomaRt::useMart("ENSEMBL_MART_SNP", dataset="hsapiens_snp", host="www.ensembl.org")  
query <- makeBranchpointWindowForSNP("rs587776767", mart.snp, exons)
```

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plotBranchpointWindow *Plots branchpointer predictions*

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## Description

Plots branchpointer predictions

## Usage

```
plotBranchpointWindow(queryName, predictions, probabilityCutoff = 0.52,  
  plotMutated = FALSE, plotStructure = TRUE, exons)
```

## Arguments

queryName	query id used to identify the SNP or region
predictions	Granges object generated by predictBranchpoints()
probabilityCutoff	probability score cutoff value for displaying U2 binding energy
plotMutated	plot alternative sequence predicitions alongside reference sequence predictions
plotStructure	plot structures for gene and 3' exon containing and skipping isoforms
exons	Granges containing exon co-ordinates. Should be produced by gtfToExons()

## Value

ggplot2 plot with branchpoint features in the specified intronic region

## Author(s)

Beth Signal

## Examples

```
smallExons <- system.file("extdata", "gencode.v26.annotation.small.gtf",  
  package = "branchpointer")  
exons <- gtfToExons(smallExons)  
g <- BSgenome.Hsapiens.UCSC.hg38::BSgenome.Hsapiens.UCSC.hg38  
  
querySNPFile <- system.file("extdata", "SNP_example.txt", package = "branchpointer")  
querySNP <- readQueryFile(querySNPFile, queryType = "SNP", exons = exons, filter = FALSE)  
predictionsSNP <- predictBranchpoints(querySNP, queryType = "SNP", BSgenome = g)  
plotBranchpointWindow(querySNP$id[1], predictionsSNP,  
  plotMutated = TRUE, exons = exons)
```

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predictBranchpoints    *Predict branchpoint probability scores*

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### Description

predicts branchpoint probability scores for each query site.

### Usage

```
predictBranchpoints(query, uniqueId = "test", queryType,
  workingDirectory = ".", genome = NA, bedtoolsLocation = NA,
  BSgenome = NULL, useParallel = FALSE, cores = 1, rmChr = FALSE)
```

### Arguments

query	branchpointer query GenomicRanges
uniqueId	unique string identifier for intermediate .bed and .fa files.
queryType	type of branchpointer query. "SNP" or "region".
workingDirectory	directory where intermediate .bed and .fa are located
genome	.fa genome file location
bedtoolsLocation	bedtools binary location (which bedtools)
BSgenome	BSgenome object
useParallel	use parallelisation to speed up code?
cores	number of cores to use in parallelisation (default = 1)
rmChr	remove "chr" before chromosome names before writing bed file. Required if genome sequence names do not contain "chr"

### Value

GenomicRanges object with branchpoint probability scores for each site in query

### Author(s)

Beth Signal

### Examples

```
smallExons <- system.file("extdata", "gencode.v26.annotation.small.gtf",
  package = "branchpointer")
exons <- gtfToExons(smallExons)
g <- BSgenome.Hsapiens.UCSC.hg38::BSgenome.Hsapiens.UCSC.hg38

querySNPFile <- system.file("extdata", "SNP_example.txt", package = "branchpointer")
querySNP <- readQueryFile(querySNPFile, queryType = "SNP", exons = exons, filter = FALSE)
predictionsSNP <- predictBranchpoints(querySNP, queryType = "SNP", BSgenome = g)
```

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predictionsToSummary *Convert SNP branchpoint predictions across the branchpoint window to an intronic summary*

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### Description

Takes predictions of branchpoint probabilities from a reference and alternative SNP and summarises the effect(s) of the SNP.

### Usage

```
predictionsToSummary(query, predictions, probabilityCutoff = 0.52,  
  probabilityChange = 0.15)
```

### Arguments

query            query GRanges containing all SNP ids to be summarised

predictions    site-wide branchpoint probability predictions produced from predictBranchpoints()

probabilityCutoff            Value to be used as the cutoff for discriminating branchpoint sites from non-branchpoint sites (default = 0.52)

probabilityChange            Minimum probability score change required to call a branchpoint site as deleted or created by a SNP (default = 0.15)

### Value

GRanges with summarised branchpoint changes occurring within the intron due to a SNP.

### Author(s)

Beth Signal

### Examples

```
smallExons <- system.file("extdata", "gencode.v26.annotation.small.gtf", package = "branchpointer")  
exons <- gtfToExons(smallExons)  
g <- BSgenome.Hsapiens.UCSC.hg38::BSgenome.Hsapiens.UCSC.hg38  
  
querySNPFile <- system.file("extdata", "SNP_example.txt", package = "branchpointer")  
querySNP <- readQueryFile(querySNPFile, queryType = "SNP", exons = exons, filter = FALSE)  
predictionsSNP <- predictBranchpoints(querySNP, queryType = "SNP", BSgenome = g)  
  
summarySNP <- predictionsToSummary(querySNP, predictionsSNP)
```

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readQueryFile	<i>Read a query file</i>
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### Description

Reads and formats a manually generated query file, and finds relative locations of the closest annotated exons Converts unstranded SNPs to two entries for each strand. Checks for duplicate names and replaces if found.

### Usage

```
readQueryFile(queryFile, queryType, exons, maxDist = 50, filter = TRUE)
```

### Arguments

queryFile	tab delimited file containing query information. For intronic regions should be in the format: region id, chromosome name, region start, region end, strand. For SNP variants should be in the format: SNP id, chromosome name, SNP position, strand, reference allele (A/T/C/G), alternative allele (A/T/C/G)
queryType	type of query file ("SNP" or "region")
exons	GRanges containing exon co-ordinates. Should be produced by gtfToExons()
maxDist	maximum distance a SNP can be from an annotated 3' exon.
filter	remove SNP queries prior to finding finding nearest exons.

### Value

Formatted query GRanges

### Author(s)

Beth Signal

### Examples

```
smallExons <- system.file("extdata", "gencode.v26.annotation.small.gtf", package = "branchpointer")
exons <- gtfToExons(smallExons)

querySNPFile <- system.file("extdata", "SNP_example.txt", package = "branchpointer")
querySNP <- readQueryFile(querySNPFile, queryType = "SNP", exons)

queryIntronFile <- system.file("extdata", "intron_example.txt", package = "branchpointer")
queryIntron <- readQueryFile(queryIntronFile, queryType = "region", exons)
```



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