

# Package ‘sangerseqR’

November 9, 2021

**Type** Package

**Title** Tools for Sanger Sequencing Data in R

**Version** 1.31.0

**Date** 2014-07-17

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**VignetteBuilder** knitr

**Imports** methods, shiny

**Depends** R (>= 3.0.2), Biostrings

**Suggests** BiocStyle, knitr, RUnit, BiocGenerics

**Description** This package contains several tools for analyzing Sanger Sequencing data files in R, including reading .scf and .ab1 files, making basecalls and plotting chromatograms.

**License** GPL-2

**biocViews** Sequencing, SNP, Visualization

**git\_url** <https://git.bioconductor.org/packages/sangerseqR>

**git\_branch** master

**git\_last\_commit** 26fe818

**git\_last\_commit\_date** 2021-10-26

**Date/Publication** 2021-11-09

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## Description

This package contains several tools for analyzing Sanger Sequencing data files in R, including reading .scf and .ab1 files, making basecalls and plotting chromatograms.

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## References

Jonathon T Hill, Bradley L Demarest, Brent W Bisgrove, Yi-chu Su, Megan Smith and H. Joseph Yost (2014). Poly peak parser: Method and software for identification of unknown indels using sanger sequencing of polymerase chain reaction products. *Developmental Dynamics* 243:1632-1636

## See Also

Biostrings

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abif-class	<i>ABIF Class Objects</i>
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## Description

S4 object returned by `read.abif` containing all fields in the ABIF file format (see [http://home.appliedbiosystems.com/support/software\\_community/ABIF\\_File\\_Format.pdf](http://home.appliedbiosystems.com/support/software_community/ABIF_File_Format.pdf)). Data fields vary by machine and basecaller versions. Must be converted to `sangerseq` to be used in other functions from this package.

**Slots**

header Header information from the file.

directory Directory information from file containing field names and information for reading binary data.

data List object containing all data fields and values in file. Included fields vary by machine and basecaller versions.

**See Also**

[read.abif](#), [scf](#), [sangerseq](#)

**Examples**

```
hetab1 <- read.abif(system.file("extdata",
                              "heterozygous.ab1",
                              package = "sangerseqR"))
str(hetab1)
```

---

chromatogram

*Generate Chromatogram*

---

**Description**

Generates a chromatogram from a [sangerseq](#) class object.

**Usage**

```
chromatogram(obj, trim5 = 0, trim3 = 0, showcalls = c("primary",
  "secondary", "both", "none"), width = 500, height = NA, cex.mtext = 1,
  cex.base = 1, ylim = 2, filename = NULL, showtrim = FALSE,
  showhets = TRUE)
```

```
## S4 method for signature 'sangerseq'
chromatogram(obj, trim5 = 0, trim3 = 0,
  showcalls = c("primary", "secondary", "both", "none"), width = 100,
  height = 2, cex.mtext = 1, cex.base = 1, ylim = 3, filename = NULL,
  showtrim = FALSE, showhets = TRUE)
```

**Arguments**

obj	<a href="#">sangerseq</a> class object
trim5	Number of bases to trim from the beginning of the sequence.
trim3	Number of bases to trim from the end of the sequence.
showcalls	Which basecall sequence to show. Any value other than "primary", "secondary" or "both" will result in basecalls not being shown.

<code>width</code>	Approximate number of bases per line.
<code>height</code>	Height of each plot row. Ignored by some devices.
<code>cex.mtext</code>	Size factor for the text in the margins.
<code>cex.base</code>	Size factor for the basecall text.
<code>ylim</code>	Peaks larger than this many times the IQR larger than the median will be cutoff.
<code>filename</code>	Name of PDF file to save to. A "NULL" value outputs the chromatogram to the current device.
<code>showtrim</code>	If true, highlights trimmed region instead of hiding it.
<code>showhets</code>	Whether or not to highlight heterozygous positions.

### Details

This function outputs a chromatogram to the current device or to a PDF file (`filename` is not NULL). Primary, Secondary or both basecalls can be shown if they are contained in the `sangerseq` object provided. What is shown will depend on how they were generated. If generated and provided by the ABIF or SCF file, then it will show the primary calls on the first line and the secondary calls on the second line. If generated by `makeBaseCalls`, then they will show the maximum and alternate basecalls only for heterozygous peaks. Finally, if the `setAllelePhase` has been run on the object, then the first line is the reference sequence and the second line is the alternate allele.

The range of the trace data shown is trimmed to the called sequence by default. Setting `trim5` and `trim3` to NULL will show the complete trace 5' and 3' of the called bases, respectively. This will generally create a very long trace. Conversely, setting `trim5` and `trim3` to an integer  $> 0$  will hide the data corresponding to that number of bases at each end. For example, `trim5=10` and `trim3=20` will remove 10 bases from the 5' end and 20 bases from the 3' end.

Several output parameters can also be set to control how the figure appears. However, it should be noted that if the current device is too small, R will return an error and not show the chromatogram. This is common with long sequence reads. To bypass this error, we recommend that the user set `filename`. This will cause the chromatogram to be saved to a PDF file in the current working directory.

### Value

A plot showing the chromatogram `tracedata` and, optionally, basecalls.

### See Also

[makeBaseCalls](#), [setAllelePhase](#), [sangerseq](#)

### Examples

```
hetsangerseq <- readsangerseq(system.file("extdata",
                                         "heterozygous.ab1",
                                         package = "sangerseqR"))
hetcalls <- makeBaseCalls(hetsangerseq, ratio = 0.33)
chromatogram(hetcalls, width = 100, height = 2, trim5 = 50, trim3 = 100,
             showcalls = "both", filename = "chromatogram.pdf")
```

---

makeBaseCalls	<i>Make Basecalls</i>
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---

## Description

Updates a [sangerseq](#) class object to contain primary and secondary peak calls.

## Usage

```
makeBaseCalls(obj, ratio = 0.33)

## S4 method for signature 'sangerseq'
makeBaseCalls(obj, ratio = 0.33)
```

## Arguments

<code>obj</code>	<a href="#">sangerseq</a> class object
<code>ratio</code>	cutoff ratio for separating signal and noise. Ratio is relative to maximum peak in basecall window.

## Details

Scf files do not contain secondary basecalls and ABIF files sometimes (but not always) contain secondary peak calls that show the secondary peak even if clearly a negative peak. This is problematic in sequence reads where heterozygous sequence data is contained in the chromatogram data. `makeBaseCalls` identifies basecall windows containing more than one peak using the provided cutoff ratio and then makes heterozygous calls for those windows. The `primarySeq` will always contain the base corresponding to the maximum peak amplitude within the window. The `secondaryPeak` will have the same base if the peak was classified as a homozygous peak, the base corresponding to the second tallest peak if two peaks were above the cutoff, or an ambiguous base if more than two peaks were identified in the window that are higher than the cutoff ratio.

## Value

[sangerseq](#) s4 object with new basecalls in the `primarySeq` and `secondarySeq` fields. Matrix values are also updated to reflect newly called base positions and amplitudes of all peaks within window.

## See Also

[chromatogram](#), [setAllelePhase](#), [sangerseq](#)

## Examples

```
hetsangerseq <- readsangerseq(system.file("extdata",
                                         "heterozygous.ab1",
                                         package = "sangerseqR"))
hetcalls <- makeBaseCalls(hetsangerseq, ratio = 0.33)
```

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PolyPeakParser	<i>Run Poly Peak Parser</i>
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**Description**

Runs the Poly Peak Parser shiny ([shiny.rstudio.com](http://shiny.rstudio.com)) app in the system's default browser. Poly Peak Parser is a web front end that reads, plots and parses double peaks from chromatogram files. Instructions can be found on the webpage once it launches.

**Usage**

```
PolyPeakParser()
```

**Value**

[scf](#) s4 object

**See Also**

[read.abif](#), [readsangerseq](#), [scf](#)

**Examples**

```
## Not run:  
PolyPeakParser()  
  
## End(Not run)
```

---

primarySeqID	<i>Sangerseq Accessor Functions</i>
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---

**Description**

Accessor Functions allow the user to retrieve results from and assign values to [sangerseq-class](#) objects.

**Usage**

```
primarySeqID(obj)  
  
primarySeqID(obj) <- value  
  
primarySeq(obj, string = FALSE)  
  
primarySeq(obj) <- value
```

```
secondarySeqID(obj)
secondarySeqID(obj) <- value
secondarySeq(obj, string = FALSE)
secondarySeq(obj) <- value
traceMatrix(obj)
traceMatrix(obj) <- value
peakPosMatrix(obj)
peakPosMatrix(obj) <- value
peakAmpMatrix(obj)
peakAmpMatrix(obj) <- value
## S4 method for signature 'sangerseq'
primarySeq(obj, string = FALSE)
## S4 method for signature 'sangerseq'
secondarySeq(obj, string = FALSE)
## S4 method for signature 'sangerseq'
traceMatrix(obj)
## S4 method for signature 'sangerseq'
peakPosMatrix(obj)
## S4 method for signature 'sangerseq'
peakAmpMatrix(obj)
## S4 method for signature 'sangerseq'
primarySeqID(obj)
## S4 method for signature 'sangerseq'
secondarySeqID(obj)
## S4 replacement method for signature 'sangerseq'
primarySeq(obj) <- value
## S4 replacement method for signature 'sangerseq'
secondarySeq(obj) <- value
## S4 replacement method for signature 'sangerseq'
```

```
traceMatrix(obj) <- value

## S4 replacement method for signature 'sangerseq'
peakPosMatrix(obj) <- value

## S4 replacement method for signature 'sangerseq'
peakAmpMatrix(obj) <- value

## S4 replacement method for signature 'sangerseq'
primarySeqID(obj) <- value

## S4 replacement method for signature 'sangerseq'
secondarySeqID(obj) <- value
```

### Arguments

obj	sangerseq object to be manipulated
value	The value to set the slot to.
string	TRUE/FALSE. FALSE (default) returns a <a href="#">DNAString</a> class object. TRUE returns the DNA sequence as a character string.

### See Also

[sangerseq-class](#)

### Examples

```
hetsangerseq <- readsangerseq(system.file("extdata",
                                          "heterozygous.ab1",
                                          package = "sangerseqR"))

primarySeq(hetsangerseq)
secondarySeq(hetsangerseq, string=TRUE)
primarySeqID(hetsangerseq)
primarySeqID(hetsangerseq) <- "Some string"
primarySeqID(hetsangerseq)
```

---

read.abif

*Read ABIF Files*

---

### Description

Reads ABIF sanger sequencing data files. ABIF files are a proprietary binary sanger sequencing chromatogram data file created by Applied Biosystems (see [http://home.appliedbiosystems.com/support/software\\_community/ABIF\\_File\\_Format.pdf](http://home.appliedbiosystems.com/support/software_community/ABIF_File_Format.pdf)). The file is read and parsed into an [abif](#) class object. This method is based on the read.abif function in the seqinr package available on CRAN.



**Usage**

```
read.abif(filename)
```

**Arguments**

filename            Location of the file.

**Value**

`abif` s4 object

**References**

Charif, D. and Lobry, J.R. (2007) SeqinR 1.0-2: a contributed package to the R project for statistical computing devoted to biological sequences retrieval and analysis. Structural approaches to sequence evolution: Molecules, networks, populations. pp. 207-232.

**See Also**

[read.scf](#), [readsangerseq](#), [abif](#)

**Examples**

```
hetab1 <- read.abif(system.file("extdata",
                               "heterozygous.ab1",
                               package = "sangerseqR"))
str(hetab1)
```

---

read.scf

*Read Scf Files*

---

**Description**

Reads Scf sanger sequencing data files. Scf files are an open source binary sanger sequencing chromatogram data file (see [http://staden.sourceforge.net/manual/formats\\_unix\\_2.html](http://staden.sourceforge.net/manual/formats_unix_2.html)). The file is read and parsed into an `scf` class object.

**Usage**

```
read.scf(filename)
```

**Arguments**

filename            Location of the file.

**Value**

`scf` s4 object

**See Also**

[read.abif](#), [readsangerseq](#), [scf](#)

**Examples**

```
homoscf <- read.scf(system.file("extdata",
                               "homozygous.scf",
                               package = "sangerseqR"))
str(homoscf)
```

---

readsangerseq

*Read Scf or ABIF Files*

---

**Description**

This is a convenience function for reading Scf or ABIF files into a sangerseq object, which can be used by the other sangerseq package functions. It is equivalent to calling [read.scf](#) or [read.abif](#) as appropriate and then calling [sangerseq](#).

**Usage**

```
readsangerseq(filename)
```

**Arguments**

filename      Location of the file.

**Value**

[sangerseq](#) s4 object

**See Also**

[read.abif](#), [read.scf](#), [abif](#), [scf](#), [sangerseq](#)

**Examples**

```
hetsangerseq <- readsangerseq(system.file("extdata",
                                          "heterozygous.ab1",
                                          package = "sangerseqR"))
str(hetsangerseq)
#same for scf files
homosangerseq <- readsangerseq(system.file("extdata",
                                          "homozygous.scf",
                                          package = "sangerseqR"))
str(homosangerseq)
```

---

sangerseq-class      *Sangerseq Class Objects*

---

## Description

Sangerseq Class Objects contain data necessary for using sangerseq package functions (e.g. [chromatogram](#), [makeBaseCalls](#)). The exact content will depend on the source of the data (for example, scf files do not have secondary Basecalls).

## Usage

```
sangerseq(obj)

## S4 method for signature 'abif'
sangerseq(obj)

## S4 method for signature 'scf'
sangerseq(obj)
```

## Arguments

obj                    Can be either an [abif](#) or [scf](#) object.

## Slots

**primarySeqID** Source of the primary basecalls. Functions that modify these calls, such as [makeBaseCalls](#) and [setAllelePhase](#) will also change this value.

**secondarySeqID** Source of the secondary basecalls. See above.

**primarySeq** The primary Basecalls formatted as a [DNAStrng](#) object.

**secondarySeq** The secondary Basecalls formatted as a [DNAStrng](#) object.

**traceMatrix** A numerical matrix containing 4 columns corresponding to the normalized signal values for the chromatogram traces. Column order = A,C,G,T.

**peakPosMatrix** A numerical matrix containing the position of the maximum peak values for each base within each Basecall window. If no peak was detected for a given base in a given window, then "NA". Column order = A,C,G,T.

**peakAmpMatrix** A numerical matrix containing the maximum peak amplitudes for each base within each Basecall window. If no peak was detected for a given base in a given window, then 0. Column order = A,C,G,T.

## Accessor methods

[primarySeqID](#), [primarySeq](#), [secondarySeqID](#), [secondarySeq](#), [traceMatrix](#), [peakPosMatrix](#), [peakAmpMatrix](#)



---

setAllelePhase	<i>Set Reference and Alternate Alleles</i>
----------------	--

---

### Description

Parses the Primary and Secondary Sequences into Reference and Alternate Alleles

### Usage

```
setAllelePhase(obj, refseq, trim5 = 0, trim3 = 0)

## S4 method for signature 'sangerseq'
setAllelePhase(obj, refseq, trim5 = 0, trim3 = 0)
```

### Arguments

obj	<a href="#">sangerseq</a> class object
refseq	DNASTring for character string of reference allele sequence.
trim5	Number of bases to trim from the beginning of the sequence.
trim3	Number of bases to trim from the end of the sequence.

### Details

When multiple heterozygous basecalls are made, it is generally unclear which calls are in phase with each other. This function takes a reference sequence for one of the alleles to match the primary and secondary basecalls as reference or alternate allele.

### Value

A [sangerseq](#) object with the Reference Allele in the primarySeq slot and the Alternate Allele in the secondarySeq slot.

### See Also

[makeBaseCalls](#), [chromatogram](#), [sangerseq](#)

### Examples

```
#Load Sequences
hetsangerseq <- readsangerseq(system.file("extdata",
                                         "heterozygous.ab1",
                                         package = "sangerseqR"))
homosangerseq <- readsangerseq(system.file("extdata",
                                         "homozygous.scf",
                                         package = "sangerseqR"))

#Make calls on heterozygous sequence to be parsed
hetcalls <- makeBaseCalls(hetsangerseq, ratio = 0.33)
#Need a reference sequence to set phase. Can get from annotation
```

```
 #(e.g. Refseq) or another sanger sequencing file
 ref <- subseq(primarySeq(homosangerseq, string = TRUE),
               start = 30,
               width = 500)
 #Set the phase
 hetseqalleles <- setAllelePhase(hetcalls, ref, trim5 = 50, trim3 = 100)
 #Align to compare alleles
 pa <- pairwiseAlignment(primarySeq(hetseqalleles),
                          secondarySeq(hetseqalleles),
                          type = "global-local")

 writePairwiseAlignments(pa)
```

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