

Overview of the *DMRcaller* package

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1 Introduction

DNA methylation is an epigenetic modification of the DNA where a methyl group is added to the cytosine nucleotides. This modification is heritable, able to control the gene regulation and, in general, is associated with transcriptional gene silencing. While in mammals the DNA is predominantly methylated in CG context, in plants non-CG methylation (CHG and CHH, where H can be any of the A, C or T nucleotides) is also present and is important for the epigenetic regulation of transcription. Sequencing of bisulfite converted DNA has become the method of choice to determine genome wide methylation distribution. The *DMRcaller* package computes the set of Differentially Methylated Regions (DMRs) between two samples. *DMRcaller* will compute the differentially methylated regions from Whole Genome Bisulfite Sequencing (WGBS) or Reduced Representation Bisulfite Sequencing (RRBS) data. There are several tools able to call DMRs, but most work has been done in mammalian systems and, thus, they were designed to primarily call CG methylation.

2 Methods

The package computes the DMRs using the CX report files generated by Bismark (Krueger and Andrews, 2011), which contain the number of methylated and unmethylated reads for each cytosine in the genome. The coverage at each position on the genome is not homogeneous and this makes it difficult to compute the differentially methylated cytosines. Here, we implemented three methods:

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- **noise.filter** where we use a kernel (Hebestreit et al., 2013) to smooth the number of methylated reads and the total number of reads (the *DMRcaller* package provides four kernels: "uniform", "triangular", "gaussian" and "epanechnikov")
- **neighbourhood** where individual cytosines in a specific context are considered in the analysis without any smoothing
- **bins** where the genome is split into equal bins where all the reads are pooled together

The DMRs are then computed by performing a statistical test between the number of methylated reads and the total number of reads in the two conditions for each position, cytosine or bin. In particular, we implemented two statistical tests: (i) Fisher's exact test and (ii) the Score test. The former (Fisher's exact test) uses the `fisher.test` in the *stats* package.

The Score test is a statistical test of a simple null hypothesis that a parameter of interest is equal to some particular value. In our case, we are interested if the methylation levels in the two samples are equal or different. Given that m_1 is the number of methylated reads in condition 1, m_2 is the number of methylated reads in condition 2, n_1 is the total number of reads in condition 1 and n_2 is the total number of reads in condition 2, the Z-score of the Score test is

$$Z = \frac{(p_1 - p_2) \nu}{\sqrt{p(1-p)}} \quad (1)$$

where $p_1 = m_1/n_1$, $p_2 = m_2/n_2$,

$$p = \frac{m_1 + m_2}{n_1 + n_2} \quad \text{and} \quad \nu = \sqrt{\frac{n_1 n_2}{n_1 + n_2}} \quad (2)$$

We then convert the Z-score to the p-value assuming a normal distribution and a two sided test.

```
pValue <- 2*pnorm(-abs(zScore))
```

Finally, for both statistical tests (Fisher's exact test and Score test), we adjust the p-values for multiple testing using Benjamini and Hochberg's method (Benjamini and Hochberg, 1995) to control the false discovery

```
pValue <- p.adjust(pValue, method="fdr")
```

The algorithm performs the statistical test for each position, cytosine or bin and then marks as DMRs all positions/cytosines/bins that satisfy the following three conditions:

- the difference in methylation levels between the two conditions is statistically significant according to the statistical test;
- the difference in methylation proportion between the two conditions is higher than a threshold value;
- the mean number of reads per cytosine is higher than a threshold.

To group adjacent DMRs, we run an iterative process, where neighbouring DMRs (within a certain distance of each other) are joined only if these three conditions are still met after joining the DMRs.

Finally, we filter the DMRs as follow

- Remove DMRs whose lengths are less than a minimum size.
- Remove DMRs with fewer cytosines than a threshold value.

For a set of potential DMRs (e.g. genes, transposable elements or CpG islands) the user can call the function `filterDMRs` where all reads in a set of provided regions are pooled together and then the algorithm performs the statistical test for each region.

3 Description

3.1 Data

Bismark (Krueger and Andrews, 2011) is a popular tool for methylation call on WGBS or RRBS data. *DMRcaller* takes as inputs the CX report files generated by Bismark and stores this data in a **GRanges** object. In the package, we included two CX report files that contain the methylation calls of WT and *met1-3 Arabidopsis thaliana* (Stroud et al., 2013). *MET1* gene encodes for the main DNA methyltransferase in *Arabidopsis thaliana* and the *met1-3* mutation results in a genome-wide loss of DNA methylation (mainly in CG context). Due to running time, we restricted the data and analysis to the first 1 *Mb* of the third chromosome of *A. thaliana*.

```
library(DMRcaller)

#load presaved data
data(methylationDataList)
```

To load a different dataset, one can use `readBismark` function, which takes as input the filename of the CX report file to be loaded.

```
# specify the Bismark CX report files
saveBismark(methylationDataList[["WT"]],
            "chr3test_a_thaliana_wt.CX_report")
saveBismark(methylationDataList[["met1-3"]],
            "chr3test_a_thaliana_met13.CX_report")

# load the data
methylationDataWT <- readBismark("chr3test_a_thaliana_wt.CX_report")
methylationDataMet13 <- readBismark("chr3test_a_thaliana_met13.CX_report")
methylationDataList <- GRangesList("WT" = methylationDataWT,
                                   "met1-3" = methylationDataMet13)
```

`methylationDataList` is a **GRangesList** object, where the **GRanges** elements contain four metadata columns

- **context** - the context of the Cytosine (CG, CHG or CHH)
- **readsM** - the number of methylated reads
- **readsN** - the total number of reads
- **trinucleotide_context** - the specific context of the cytosine (H is replaced by the actual nucleotide)

If the data consists of two or more replicates, these can be pooled together using the function `poolMethylationDatasets` or `poolTwoMethylationDatasets` (in the case of pooling only two datasets). The latter function (`poolTwoMethylationDatasets`) is useful when the datasets are large and creating a **GRangesList** object is not possible (e.g. the **GRanges** objects are too large).

```
# load the data
methylationDataAll <- poolMethylationDatasets(methylationDataList)

# In the case of 2 elements, this is equivalent to
methylationDataAll <- poolTwoMethylationDatasets(methylationDataList[[1]],
                                                  methylationDataList[[2]])
```

Alternatively, one can use `readBismarkPool` to directly read a list of CX report files and pool them together.

```
# load the data
methylationDataAll <- readBismarkPool(c(file_wt, file_met13))
```

3.2 Low resolution profiles

The *DMRcaller* package also offers the possibility to visualise context specific global changes in the methylation profile. To achieve this, the user can call `plotMethylationProfileFromData` function, which computes the mean methylation proportion in tiling bins of fixed size; see Figure 1.

```
par(mar=c(4, 4, 3, 1)+0.1)
plotMethylationProfileFromData(methylationDataList[["WT"]],
                              methylationDataList[["met1-3"]],
                              conditionsNames = c("WT", "met1-3"),
                              windowSize = 10000,
                              autoscale = FALSE,
                              context = c("CG"))

## Recompute regions...
## Computing low resolution profiles...
## Calculating methylation profile for Chr3:101..999999 using a window of 10000 bp
## Calculating methylation profile for Chr3:101..999999 using a window of 10000 bp
```

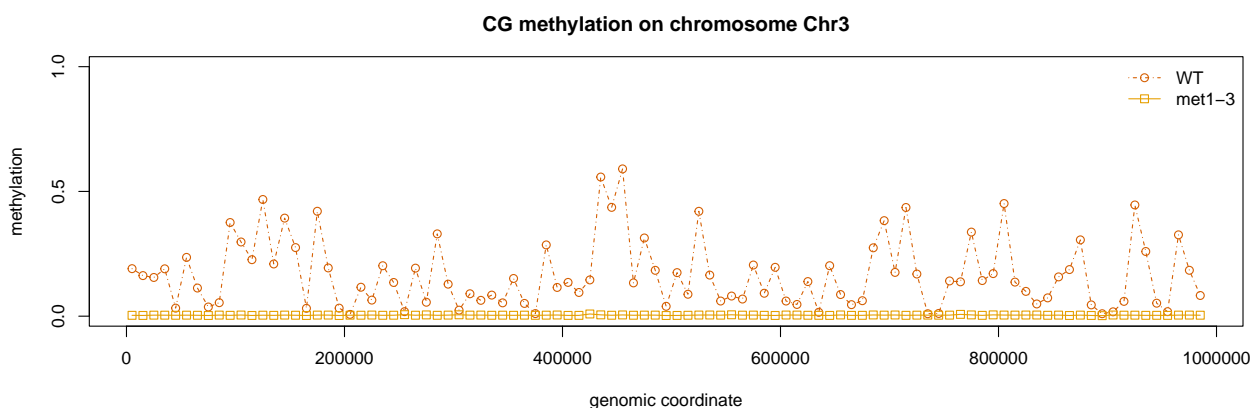


Figure 1: *Low resolution profile in CG context for WT and met1-3.*

Alternatively, for a finer control, the user can use `computeMethylationProfile` function to compute the methylation profile at certain locations on the genome. This function returns a `GRanges` object with four metadata columns

- **sumReadsM** - the number of methylated reads
- **sumReadsN** - the total number of reads
- **Proportion** - the proportion of methylated reads
- **context** - the context

One or more of these `GRanges` objects can be put in a `GRangesList` object which is then passed as a parameter to the `plotMethylationProfile` function.

```
regions <- GRanges(seqnames = Rle("Chr3"), ranges = IRanges(1,1E6))

# compute low resolution profile in 10 Kb windows
profileCGWT <- computeMethylationProfile(methylationDataList[["WT"]],
                                         regions,
                                         windowSize = 10000,
                                         context = "CG")

profileCGMet13 <- computeMethylationProfile(methylationDataList[["met1-3"]],
```

```

                                regions,
                                windowSize = 10000,
                                context = "CG")

profilesCG <- GRangesList("WT" = profileCGWT, "met1-3" = profileCGMet13)

#plot the low resolution profile
par(mar=c(4, 4, 3, 1)+0.1)
par(mfrow=c(1,1))
plotMethylationProfile(profilesCG,
                        autoscale = FALSE,
                        labels = NULL,
                        title="CG methylation on Chromosome 3",
                        col=c("#D55E00", "#E69F00"),
                        pch = c(1,0),
                        lty = c(4,1))

```

3.3 Coverage of the bisulfite sequencing data

The number of reads from the bisulfite sequencing can differ significantly between different locations on the genome in the sense that cytosines in the same context (including neighbouring cytosines) can display large variability in the coverage. To plot the coverage of the bisulfite sequencing datasets, one can use `plotMethylationDataCoverage` function which takes as input one or two datasets and the vector with the thresholds used to compute the proportion of cytosines with at least that many reads; see Figure 2.

Alternatively, the *DMRcaller* also provides the `computeMethylationDataCoverage` function which returns a numeric vector with the number or proportion of cytosines in a specific context that have at least a certain number of reads specified by the input vector `breaks`.

```

# compute the coverage in the two contexts
coverageCGWT <- computeMethylationDataCoverage(methylationDataList[["WT"]],
                                                context="CG",
                                                breaks = c(1,5,10,15))

```

3.4 Spatial correlation of methylation levels

Methylation levels are often spatially correlated and some methods to detect DMRs assume this spatial correlation. Nevertheless, different tissues, samples and even methylation context will display different levels of correlation. *DMRcaller* implements `plotMethylationDataSpatialCorrelation` function that plots the correlation of methylation levels as function of distance between cytosines. This function takes as input one or two datasets and the vector with the distances between cytosines; see Figure 2.

Alternatively, the *DMRcaller* also provides the `computeMethylationDataSpatialCorrelation` function which returns a numeric vector with the correlation of methylation levels of cytosines separated by certain distances in a specific context.

```

# compute the coverage in the two contexts
correlation_CG_wt <- computeMethylationDataSpatialCorrelation(methylationDataList[["WT"]],
                                                             context="CG",
                                                             distances=c(1,10,100,1000,10000))

```

3.5 Calling DMRs

DMRcaller package provides `computeDMRs` function to call DMRs. The output of this function is a `GRanges` with 11 metadata columns.

```

# plot the coverage in the two contexts
par(mar=c(4, 4, 3, 1)+0.1)
plotMethylationDataCoverage(methylationDataList[["WT"]],
  methylationDataList[["met1-3"]],
  breaks = c(1,5,10,15),
  regions = NULL,
  conditionsNames=c("WT","met1-3"),
  context = c("CHH"),
  proportion = TRUE,
  labels=LETTERS,
  contextPerRow = FALSE)

```

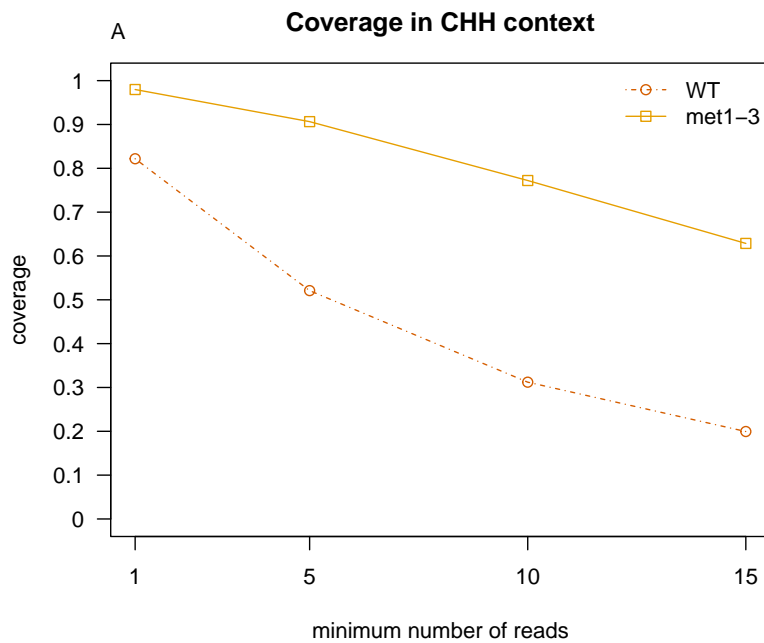


Figure 2: *Coverage*. For example, this figure shows that in WT only 30% of the cytosines in CHH context have at least 10 reads.

```

# compute the spatial correlation of methylation levels
plotMethylationDataSpatialCorrelation(methylationDataList[["WT"]],
    distances = c(1,100,10000), regions = NULL,
    conditionsNames = c("WT"),
    context = c("CG"),
    labels = LETTERS, col = NULL,
    pch = c(1,0,16,2,15,17), lty = c(4,1,3,2,6,5),
    contextPerRow = FALSE,
    log = "x")

## Computing methylation levels correlation for distances of 1 bp
## Computing methylation levels correlation for distances of 100 bp
## Computing methylation levels correlation for distances of 10000 bp

```

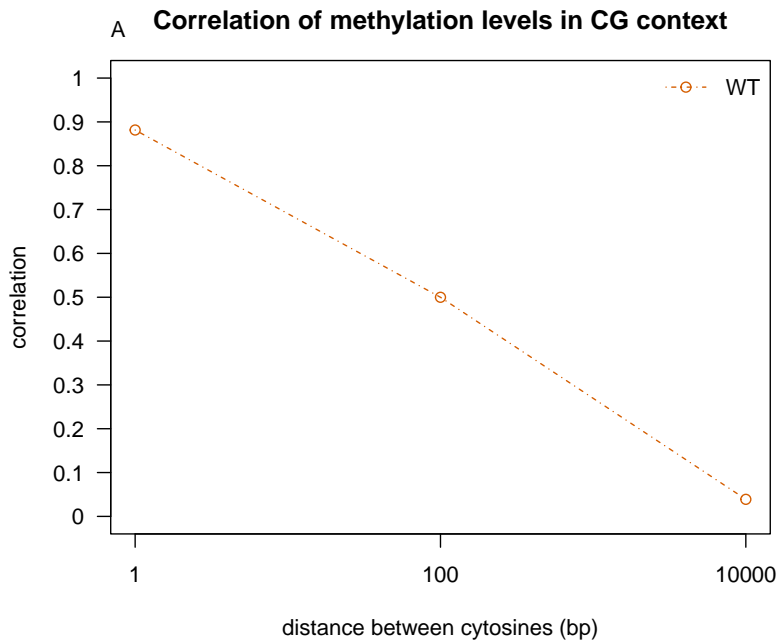


Figure 3: *Spatial correlation of methylation levels.*

- **direction** - a numeric value indicating whether the methylation was lost in the second condition compared to the first one (-1) or gained (+1)
- **context** - the context of the cytosine (CG, CHG or CHH)
- **sumReadsM1** - the number of methylated reads in the DMR in condition 1
- **sumReadsN1** - the total number of reads in the DMR in condition 1
- **proportion1** - the proportion of methylated reads in the DMR in condition 1
- **sumReadsM2** - the number of methylated reads in the DMR in condition 2
- **sumReadsN2** - the total number of reads in the DMR in condition 2
- **proportion2** - the proportion of methylated reads in the DMR in condition 2
- **cytosinesCount** - the number of cytosines in the DMR
- **pValue** - the adjusted p-value of the statistical test
- **regionType** - a character string indicating whether the methylation was lost in the second condition compared to the first one ("loss") or gained ("gain")

For predefined regions (e.g. genes, transposons or CpG islands) the user can call `filterDMRs` function to extract the list of regions that are differentially methylated. The output of this function is again a GRanges with the same 11 metadata columns.

Below we present examples of calling both functions.

```
chr_local <- GRanges(seqnames = Rle("Chr3"), ranges = IRanges(5E5,6E5))

# compute the DMRs in CG context with noise_filter method
DMRsNoiseFilterCG <- computeDMRs(methylationDataList[["WT"]],
  methylationDataList[["met1-3"]],
  regions = chr_local,
  context = "CG",
  method = "noise_filter",
  windowSize = 100,
  kernelFunction = "triangular",
  test = "score",
  pValueThreshold = 0.01,
  minCytosinesCount = 4,
  minProportionDifference = 0.4,
  minGap = 0,
  minSize = 50,
  minReadsPerCytosine = 4,
  cores = 1)

## Parameters checking ...
## Extract methylation in the corresponding context
## Computing DMRs at Chr3:500000..600000
## Calculating interpolations...
## Identifying DMRs...
## Analysed reads inside DMRs
## Merge DMRs iteratively
## Filter DMRs

print(DMRsNoiseFilterCG)
```



```
## GRanges object with 60 ranges and 11 metadata columns:
##      seqnames      ranges strand | direction      context sumReadsM1
##      <Rle>       <IRanges> <Rle> | <numeric> <character> <numeric>
## [1] Chr3 503043-503148 * | -1 CG 299
## [2] Chr3 503390-503542 * | -1 CG 158
## [3] Chr3 503612-503901 * | -1 CG 342
## [4] Chr3 504042-504093 * | -1 CG 59
## [5] Chr3 504255-504348 * | -1 CG 265
## ...
## [56] Chr3 593906-594076 * | -1 CG 216
## [57] Chr3 594128-594214 * | -1 CG 27
## [58] Chr3 594285-594385 * | -1 CG 128
## [59] Chr3 599027-599107 * | -1 CG 57
## [60] Chr3 599509-599634 * | -1 CG 168
##      sumReadsN1 proportion1 sumReadsM2 sumReadsN2 proportion2 cytosinesCount
##      <numeric> <numeric> <numeric> <numeric> <numeric> <numeric>
## [1] 365 0.819178 0 419 0.0000000 10
## [2] 198 0.797980 0 414 0.0000000 12
## [3] 442 0.773756 3 807 0.00371747 25
## [4] 86 0.686047 0 249 0.0000000 6
## [5] 351 0.754986 0 412 0.0000000 12
## ...
## [56] 253 0.853755 1 648 0.00154321 16
## [57] 45 0.600000 2 107 0.01869159 4
## [58] 149 0.859060 0 258 0.0000000 4
## [59] 111 0.513514 3 154 0.01948052 4
## [60] 219 0.767123 0 201 0.0000000 8
##      pValue regionType
##      <numeric> <character>
## [1] 4.18251e-122 loss
## [2] 1.86673e-98 loss
## [3] 4.84005e-185 loss
## [4] 7.28326e-47 loss
## [5] 3.75353e-105 loss
## ...
## [56] 2.23081e-158 loss
## [57] 8.30991e-17 loss
## [58] 5.30109e-72 loss
## [59] 2.60868e-21 loss
## [60] 1.19821e-57 loss
## -----
## seqinfo: 1 sequence from an unspecified genome; no seqlengths
```

```
# compute the DMRs in CG context with neighbourhood method
DMRsNeighbourhoodCG <- computeDMRs(methylationDataList[["WT"]],
                                   methylationDataList[["met1-3"]],
                                   regions = chr_local,
                                   context = "CG",
                                   method = "neighbourhood",
                                   test = "score",
                                   pValueThreshold = 0.01,
                                   minCytosinesCount = 4,
                                   minProportionDifference = 0.4,
                                   minGap = 200,
                                   minSize = 1,
```

```

minReadsPerCytosine = 4,
cores = 1)

## Parameters checking ...
## Extract methylation in the corresponding context
## Computing DMRs
## Merge DMRs iteratively
## Filter DMRs

print(DMRsNeighbourhoodCG)

## GRanges object with 34 ranges and 16 metadata columns:
##      seqnames      ranges strand | context trinucleotide_context  readsM1
##      <Rle>        <IRanges> <Rle> | <factor>                <factor> <integer>
## [1] Chr3 503058-503853 * | CG CGG 96
## [2] Chr3 504058-504069 * | CG CGG 22
## [3] Chr3 504292-504490 * | CG CGA 35
## [4] Chr3 506440-506776 * | CG CGT 28
## [5] Chr3 507119-507480 * | CG CGA 6
## ...
## [30] Chr3 588591-588633 * | CG CGC 11
## [31] Chr3 591681-591790 * | CG CGT 25
## [32] Chr3 593736-594337 * | CG CGA 65
## [33] Chr3 598934-599219 * | CG CGT 6
## [34] Chr3 599556-599586 * | CG CGA 46
##      readsN1  readsM2  readsN2      pValue sumReadsM1 sumReadsN1
##      <integer> <integer> <integer> <numeric> <numeric> <numeric>
## [1] 139 0 117 0.00000e+00 806 1217
## [2] 25 0 48 1.55222e-42 56 78
## [3] 39 0 42 6.07831e-180 389 584
## [4] 42 0 59 2.59969e-108 228 370
## [5] 9 0 21 1.10736e-102 225 415
## ...
## [30] 12 0 38 1.39683e-145 353 426
## [31] 31 2 59 1.27477e-143 268 296
## [32] 75 1 56 7.82820e-312 659 1068
## [33] 6 0 15 2.41968e-39 83 178
## [34] 55 0 63 4.16663e-57 166 217
##      proportion1 sumReadsM2 sumReadsN2 proportion2 cytosinesCount direction
##      <numeric> <numeric> <numeric> <numeric> <numeric> <numeric>
## [1] 0.662284 4 2205 0.00181406 65 -1
## [2] 0.717949 0 210 0.00000000 5 -1
## [3] 0.666096 0 911 0.00000000 24 -1
## [4] 0.616216 3 629 0.00476948 20 -1
## [5] 0.542169 1 687 0.00145560 22 -1
## ...
## [30] 0.828638 4 509 0.00785855 8 -1
## [31] 0.905405 4 486 0.00823045 14 -1
## [32] 0.617041 6 1817 0.00330215 41 -1
## [33] 0.466292 3 331 0.00906344 13 -1
## [34] 0.764977 0 200 0.00000000 7 -1
##      regionType
##      <character>
## [1] loss
## [2] loss
## [3] loss
## [4] loss

```

```
## [5] loss
## ...
## [30] loss
## [31] loss
## [32] loss
## [33] loss
## [34] loss
## -----
## seqinfo: 1 sequence from an unspecified genome; no seqlengths
```

```
# compute the DMRs in CG context with bins method
DMRsBinsCG <- computeDMRs(methylationDataList[["WT"]],
  methylationDataList[["met1-3"]],
  regions = chr_local,
  context = "CG",
  method = "bins",
  binSize = 100,
  test = "score",
  pValueThreshold = 0.01,
  minCytosinesCount = 4,
  minProportionDifference = 0.4,
  minGap = 200,
  minSize = 50,
  minReadsPerCytosine = 4,
  cores = 1)

## Parameters checking ...
## Extract methylation in the corresponding context
## Computing DMRs at Chr3:500000..600000
## Count inside each bin...
## Filter the bins...
## Identifying DMRs...
## Merge adjacent DMRs
## Merge DMRs iteratively
## Filter DMRs

print(DMRsBinsCG)

## GRanges object with 40 ranges and 11 metadata columns:
##      seqnames      ranges strand | sumReadsM1 sumReadsN1 proportion1
##      <Rle>       <IRanges> <Rle> | <numeric> <numeric> <numeric>
## [1] Chr3 503000-503199 * | 299 731 0.409029
## [2] Chr3 503100-503199 * | 13 28 0.464286
## [3] Chr3 503400-503499 * | 158 198 0.797980
## [4] Chr3 503400-504499 * | 959 1674 0.572879
## [5] Chr3 506400-506699 * | 182 321 0.566978
## ...
## [36] Chr3 593700-594399 * | 660 1151 0.573414
## [37] Chr3 599000-599299 * | 77 184 0.418478
## [38] Chr3 599200-599299 * | 20 35 0.571429
## [39] Chr3 599500-599599 * | 168 219 0.767123
## [40] Chr3 599500-599599 * | 168 219 0.767123
##      sumReadsM2 sumReadsN2 proportion2 cytosinesCount context direction
##      <numeric> <numeric> <numeric> <numeric> <character> <numeric>
## [1] 1 776 0.001288660 17 CG -1
## [2] 0 100 0.000000000 4 CG -1
```

```
## [3] 0 414 0.000000000 12 CG -1
## [4] 3 3183 0.000942507 90 CG -1
## [5] 3 546 0.005494505 18 CG -1
## ... ..
## [36] 7 1907 0.00367069 44 CG -1
## [37] 3 356 0.00842697 14 CG -1
## [38] 0 107 0.000000000 6 CG -1
## [39] 0 201 0.000000000 8 CG -1
## [40] 0 201 0.000000000 8 CG -1
## pValue regionType
## <numeric> <character>
## [1] 4.74723e-87 loss
## [2] 6.54241e-13 loss
## [3] 1.65932e-98 loss
## [4] 0.00000e+00 loss
## [5] 2.63625e-84 loss
## ... ..
## [36] 2.64404e-298 loss
## [37] 5.89785e-37 loss
## [38] 3.36772e-17 loss
## [39] 1.19821e-57 loss
## [40] 1.19821e-57 loss
## -----
## seqinfo: 1 sequence from an unspecified genome; no seqlengths
```

```
# load the gene annotation data
data(GEs)

#select the genes
genes <- GEs[which(GEs$type == "gene")]

# compute the DMRs in CG context over genes
DMRsGenesCG <- filterDMRs(methylationDataList[["WT"]],
                          methylationDataList[["met1-3"]],
                          potentialDMRs = genes[overlapsAny(genes, chr_local)],
                          context = "CG",
                          test = "score",
                          pValueThreshold = 0.01,
                          minCytosinesCount = 4,
                          minProportionDifference = 0.4,
                          minReadsPerCytosine = 3,
                          cores = 1)

## Parameters checking ...
## Extract methylation in the corresponding context
## Computing DMRs at Chr3:101..999999
## Selecting data...
## Identifying DMRs...

print(DMRsGenesCG)

## GRanges object with 3 ranges and 21 metadata columns:
## seqnames ranges strand | source type score phase
## <Rle> <IRanges> <Rle> | <factor> <factor> <numeric> <integer>
## [1] Chr3 576378-579559 + | TAIR10 gene NA <NA>
## [2] Chr3 528574-532582 - | TAIR10 gene NA <NA>
```

```
## [3] Chr3 570134-572345 - | TAIR10 gene NA <NA>
## ID Name Note Parent Index
## <character> <character> <CharacterList> <CharacterList> <character>
## [1] AT3G02680 AT3G02680 protein_coding_gene <NA>
## [2] AT3G02530 AT3G02530 protein_coding_gene <NA>
## [3] AT3G02660 AT3G02660 protein_coding_gene <NA>
## Derives_from Alias sumReadsM1 sumReadsN1 proportion1 sumReadsM2
## <character> <CharacterList> <numeric> <numeric> <numeric> <numeric>
## [1] <NA> 1106 2379 0.464901 14
## [2] <NA> 1150 2756 0.417271 30
## [3] <NA> 874 1848 0.472944 10
## sumReadsN2 proportion2 cytosinesCount pValue regionType direction
## <numeric> <numeric> <numeric> <numeric> <character> <numeric>
## [1] 4546 0.00307963 142 0 loss -1
## [2] 6207 0.00483325 171 0 loss -1
## [3] 3487 0.00286779 104 0 loss -1
## -----
## seqinfo: 7 sequences from an unspecified genome; no seqlengths
```

3.6 Merge DMRs

Finally, for merging adjacent DMRs, *DMRcaller* provides the function `mergeDMRsIteratively` which can be used as follows:

```
DMRsNoiseFilterCGMerged <- mergeDMRsIteratively(DMRsNoiseFilterCG,
                                                minGap = 200,
                                                respectSigns = TRUE,
                                                methylationDataList[["WT"]],
                                                methylationDataList[["met1-3"]],
                                                context = "CG",
                                                minProportionDifference = 0.4,
                                                minReadsPerCytosine = 4,
                                                pValueThreshold = 0.01,
                                                test="score")

## Parameters checking ...
## Merge DMRs iteratively ...

print(DMRsNoiseFilterCGMerged)

## GRanges object with 37 ranges and 11 metadata columns:
## seqnames ranges strand | direction context sumReadsM1
## <Rle> <IRanges> <Rle> | <numeric> <character> <numeric>
## [1] Chr3 503043-503148 * | -1 CG 299
## [2] Chr3 503390-504509 * | -1 CG 959
## [3] Chr3 506392-506723 * | -1 CG 182
## [4] Chr3 507286-507422 * | -1 CG 153
## [5] Chr3 514791-514891 * | -1 CG 560
## ... .. ... .. ... ..
## [33] Chr3 588556-588681 * | -1 CG 355
## [34] Chr3 591657-591828 * | -1 CG 268
## [35] Chr3 593709-594385 * | -1 CG 659
## [36] Chr3 599027-599107 * | -1 CG 57
## [37] Chr3 599509-599634 * | -1 CG 168
## sumReadsN1 proportion1 sumReadsM2 sumReadsN2 proportion2 cytosinesCount
## <numeric> <numeric> <numeric> <numeric> <numeric> <numeric>
```

```

## [1] 365 0.819178 0 419 0.000000000 10
## [2] 1674 0.572879 3 3183 0.000942507 90
## [3] 321 0.566978 3 546 0.005494505 18
## [4] 217 0.705069 0 322 0.000000000 8
## [5] 760 0.736842 1 680 0.001470588 10
## ... ... ... ... ...
## [33] 458 0.775109 5 605 0.00826446 10
## [34] 321 0.834891 4 540 0.00740741 16
## [35] 1068 0.617041 6 1827 0.00328407 42
## [36] 111 0.513514 3 154 0.01948052 4
## [37] 219 0.767123 0 201 0.00000000 8
## pValue regionType
## <numeric> <character>
## [1] 4.18251e-122 loss
## [2] 0.00000e+00 loss
## [3] 1.44994e-84 loss
## [4] 1.20961e-70 loss
## [5] 2.03906e-178 loss
## ... ... ...
## [33] 4.02207e-150 loss
## [34] 4.83924e-140 loss
## [35] 5.44838e-314 loss
## [36] 2.60868e-21 loss
## [37] 1.19821e-57 loss
## -----
## seqinfo: 1 sequence from an unspecified genome; no seqlengths

```

Note that two neighbouring DMRs will be merged if all the conditions below are met

- they are within a distance from each other smaller than minGap
- the difference in methylation levels between the two conditions is statistically significant according to the statistical test when the two DMRs are joined
- the difference in methylation proportion between the two conditions is higher than a threshold value when the two DMRs are joined
- the number of reads per cytosine is higher than a threshold when the two DMRs are joined

3.7 Extract methylation data in regions

`analyseReadsInsideRegionsForCondition` function can extract additional information in a set of genomic regions (including DMRs) from any `methylationData` object. For example, to establish a link between the CG and CHH methylation, one might want to extract the number of methylated reads and the total number of reads in CHH context inside DMRs called in CG context.

```

#retrive the number of reads in CHH context in WT in CG DMRs
DMRsNoiseFilterCGreadsCHH <- analyseReadsInsideRegionsForCondition(
  DMRsNoiseFilterCGMerged,
  methylationDataList[["WT"]], context = "CHH",
  label = "WT")

## Parameters checking ...
## Extract methylation levels in corresponding context ...
## Compute reads inside each region ...

print(DMRsNoiseFilterCGreadsCHH)

```

```

## GRanges object with 37 ranges and 15 metadata columns:
##      seqnames      ranges strand | direction      context sumReadsM1
##      <Rle>       <IRanges> <Rle> | <numeric> <character> <numeric>
## [1]   Chr3 503043-503148    * |      -1      CG          299
## [2]   Chr3 503390-504509    * |      -1      CG          959
## [3]   Chr3 506392-506723    * |      -1      CG          182
## [4]   Chr3 507286-507422    * |      -1      CG          153
## [5]   Chr3 514791-514891    * |      -1      CG          560
## ...
## [33]  Chr3 588556-588681    * |      -1      CG          355
## [34]  Chr3 591657-591828    * |      -1      CG          268
## [35]  Chr3 593709-594385    * |      -1      CG          659
## [36]  Chr3 599027-599107    * |      -1      CG           57
## [37]  Chr3 599509-599634    * |      -1      CG          168
##      sumReadsN1 proportion1 sumReadsM2 sumReadsN2 proportion2 cytosinesCount
##      <numeric> <numeric> <numeric> <numeric> <numeric> <numeric>
## [1]      365    0.819178         0         419 0.000000000         10
## [2]     1674    0.572879         3        3183 0.000942507         90
## [3]      321    0.566978         3         546 0.005494505         18
## [4]      217    0.705069         0         322 0.000000000          8
## [5]      760    0.736842         1         680 0.001470588         10
## ...
## [33]     458    0.775109         5         605 0.008264446         10
## [34]     321    0.834891         4         540 0.00740741         16
## [35]    1068    0.617041         6        1827 0.00328407         42
## [36]     111    0.513514         3         154 0.01948052          4
## [37]     219    0.767123         0         201 0.00000000         8
##      pValue      regionType sumReadsMWTCHH sumReadsNWTCHH proportionWTCHH
##      <numeric> <character> <numeric> <numeric> <numeric>
## [1] 4.18251e-122      loss         0          303    0.00000000
## [2] 0.00000e+00      loss         99        3323   0.02979236
## [3] 1.44994e-84      loss         10        1047   0.00955110
## [4] 1.20961e-70      loss         0          571   0.00000000
## [5] 2.03906e-178     loss         1          665   0.00150376
## ...
## [33] 4.02207e-150     loss         12         672   0.01785714
## [34] 4.83924e-140     loss         6          792   0.00757576
## [35] 5.44838e-314     loss         29        2560   0.01132812
## [36] 2.60868e-21      loss         0          193   0.00000000
## [37] 1.19821e-57      loss         1          206   0.00485437
##      cytosinesCountCHH
##      <numeric>
## [1]      27
## [2]     309
## [3]      90
## [4]      33
## [5]      32
## ...
## [33]      32
## [34]      52
## [35]     196
## [36]      23
## [37]      36
## -----
## seqinfo: 1 sequence from an unspecified genome; no seqlengths

```

3.8 Plotting the distribution of DMRs

Sometimes, it is useful to obtain the distribution of the DMRs over the chromosomes. The *DMRcaller* provides the `computeOverlapProfile` function, which computes this distribution. The `GRanges` object generated by this function can then be added to a `GRangesList` object, which can be plotted using `plotOverlapProfile` function; see Figure 4. Additionally, the `plotOverlapProfile` function allows the user to specify two `GRangesList`, thus, allowing the plotting of distributions of hypo or hyper methylated DMRs separately.

```
# compute the distribution of DMRs
hotspots <- computeOverlapProfile(DMRsNoiseFilterCGMerged, chr_local,
                                windowSize=5000, binary=TRUE)

## Calculating overlaps for Chr3:500000..600000 using a window of 5000 bp

# plot the distribution of DMRs
plotOverlapProfile(GRangesList("Chr3"=hotspots))
```

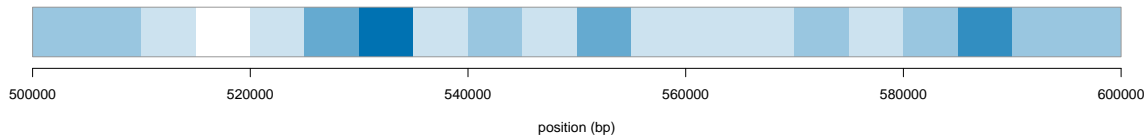


Figure 4: *Distribution of DMRs*. Darker colour indicates higher density, while lighter colour lower density.

3.9 Plotting profiles with DMRs

Finally, *DMRcaller* package also provides a function to plot methylation profiles at a specific location on the genome. To plot the methylation profile the user needs to call the `plotLocalMethylationProfile` function; see Figure 5.

4 Parallel computation

Computing the DMRs can be computationally intensive. For example, in the case of *A. thaliana* (with a genome of ≈ 130 Mb), it can take several hours to compute the DMRs depending on the method used and on the number of DMRs. To speed up computations, *DMRcaller* supports parallel computing of DMRs using the package *parallel*, but parallel computation is currently not supported on Windows.

The five functions used for computing and filtering the DMRs (`computeDMRs`, `filterDMRs`, `mergedDMRsIteratively` and `analyseReadsInsideRegionsForCondition`) accept the parameter `cores`, which specifies the number of cores that can be used when performing the corresponding computations. When using 10 cores, it can take between 10 and 30 minutes to compute the DMRs in *A. thaliana* depending on the selected parameters.

5 Analysis of biological replicates

The package also contains a set of functions for the analysis of multiple biological replicates.

The synthetic dataset is made by 300 different cytosines, extracted from those present in the *A. thaliana* dataset. The value for `readsN` are created using the function `rnorm`, while the values for `readsM` are generated using the function `rbinom`. The probabilities used are 0.1 in the external region and 0.8 in the central region. In this way a DMR should be detected in the central region of the synthetic dataset.

The difference in proportion is plotted in figure 6

The DMRs are computed using the function `computeDMRsReplicates`, which uses beta regression (Ferrari and Cribari-Neto, 2004) to detect differential methylation.


```

# select a 20 Kb location on the Chr3
chr3Reg <- GRanges(seqnames = Rle("Chr3"), ranges = IRanges(510000,530000))

# create a list with all DMRs
DMRsCGList <- list("noise filter" = DMRsNoiseFilterCGMerged,
                  "neighbourhood" = DMRsNeighbourhoodCG,
                  "bins" = DMRsBinsCG,
                  "genes" = DMRsGenesCG)

# plot the local profile
par(cex=0.9)
par(mar=c(4, 4, 3, 1)+0.1)
plotLocalMethylationProfile(methylationDataList[["WT"]],
                           methylationDataList[["met1-3"]],
                           chr3Reg,
                           DMRsCGList,
                           conditionsNames = c("WT", "met1-3"),
                           GEs,
                           windowSize = 300,
                           main="CG methylation")

```

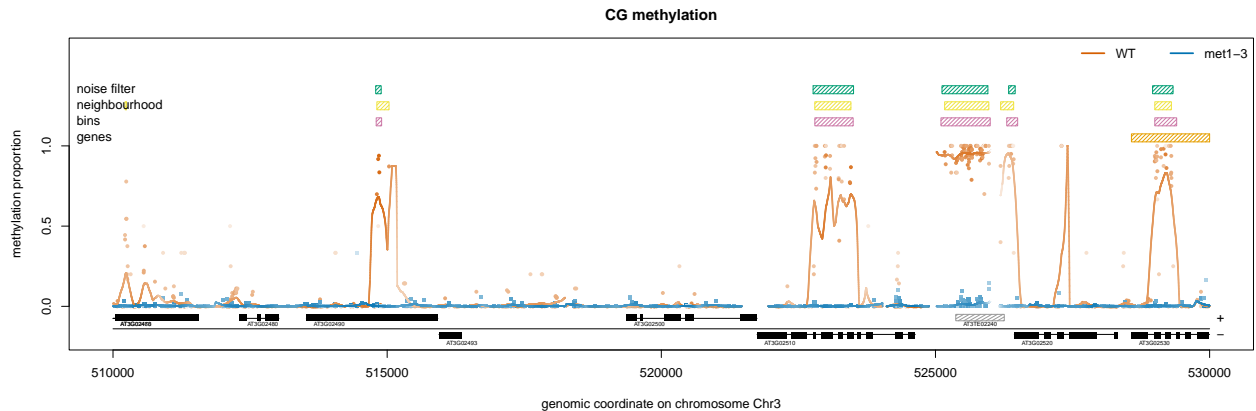


Figure 5: *Local methylation profile*. The points on the graph represent methylation proportion of individual cytosines, their colour (red or blue) which sample they belong to and the intensity of the the colour how many reads that particular cytosine had. This means that darker colours indicate stronger evidence that the corresponding cytosine has the corresponding methylation proportion, while lighter colours indicate a weaker evidence. The solid lines represent the smoothed profiles and the intensity of the colour the coverage at the corresponding position (darker colours indicate more reads while lighter ones less reads). The boxes on top represent the DMRs, where a filled box will represent a DMR which gained methylation while a box with a pattern represent a DMR that lost methylation. The DMRs need to have a metadata column `regionType` which can be either "gain" (where there is more methylation in condition 2 compared to condition 1) or "loss" (where there is less methylation in condition 2 compared to condition 1). In case this metadata column is missing all DMRs are drawn using filled boxes. Finally, we also allow annotation of the DNA sequence. We represent by black boxes all the exons, which are joined by a horizontal black line, thus, marking the full body of the gene. With grey boxes we mark the transposable elements. Both for genes and transposable elements we plot them over a mid line if they are on the positive strand and under the mid line if they are on the negative strand.

```

# loading synthetic data
data("syntheticDataReplicates")

# create vector with colours for plotting
cbbPalette <- c("#000000", "#E69F00", "#56B4E9", "#009E73", "#F0E442",
               "#0072B2", "#D55E00", "#CC79A7")

# plotting the difference in proportions
plot(start(methylationData), methylationData$readsM1/methylationData$readsN1,
     ylim=c(0,1), col=cbbPalette[2], xlab="Position in Chr3 (bp)",
     ylab="Methylation proportion")
points(start(methylationData), methylationData$readsM2/methylationData$readsN2,
       col=cbbPalette[7], pch=4)
points(start(methylationData), methylationData$readsM3/methylationData$readsN3,
       col=cbbPalette[3], pch=2)
points(start(methylationData), methylationData$readsM4/methylationData$readsN4,
       col=cbbPalette[6], pch=3)
legend(x = "topleft", legend=c("Treatment 1", "Treatment 2", "Control 1",
                              "Control 2"), pch=c(1,4,2,3),
       col=cbbPalette[c(2,7,3,6)], bty="n", cex=1.0)

```

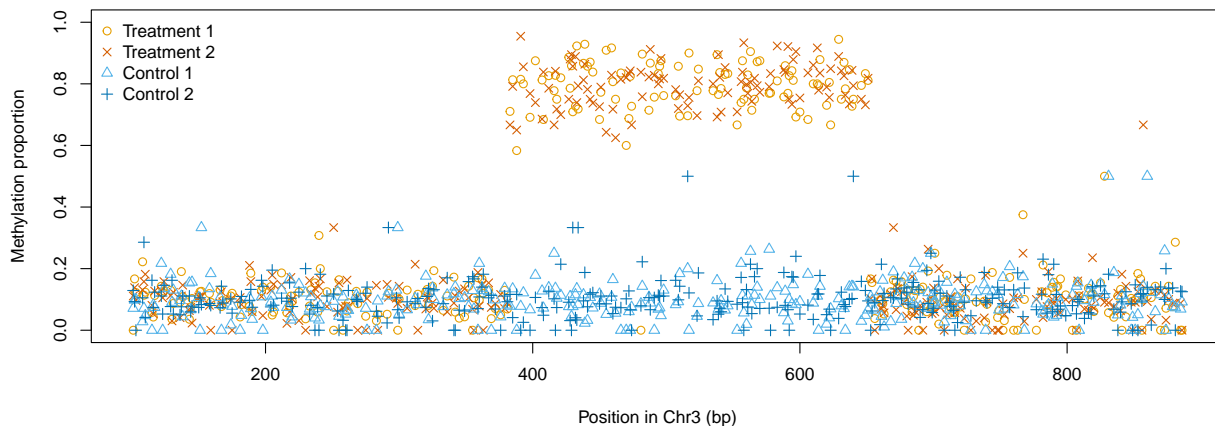


Figure 6: *Methylation proportions in the synthetic dataset.*

```

# loading betareg library to allow using computeDMRsReplicates function
library(betareg)

# creating condition vector
condition <- c("a", "a", "b", "b")

# computing DMRs using the neighbourhood method
DMRsReplicatesBins <- computeDMRsReplicates(methylationData = methylationData,
                                             condition = condition,
                                             regions = NULL,
                                             context = "CG",
                                             method = "bins",
                                             binSize = 100,
                                             test = "betareg",
                                             pseudocountM = 1,
                                             pseudocountN = 2,
                                             pValueThreshold = 0.01,
                                             minCytosinesCount = 4,
                                             minProportionDifference = 0.4,
                                             minGap = 0,
                                             minSize = 50,
                                             minReadsPerCytosine = 4,
                                             cores = 1)

## Parameters checking ...
## Extract methylation in the corresponding context
## Computing DMRs at Chr3:101..886
## Count inside each bin...
## Filter the bins...
## Identifying DMRs...
## Merge adjacent DMRs
## Merge DMRs iteratively
## Filter DMRs

print(DMRsReplicatesBins)

## GRanges object with 2 ranges and 11 metadata columns:
##      seqnames      ranges strand | sumReadsM1 sumReadsN1 proportion1 sumReadsM2
##      <Rle> <IRanges> <Rle> | <numeric> <numeric> <numeric> <numeric>
## [1] Chr3 401-500 * | 436 546 0.797445 61
## [2] Chr3 501-600 * | 419 521 0.803059 42
##      sumReadsN2 proportion2 cytosinesCount context direction pValue
##      <numeric> <numeric> <numeric> <character> <numeric> <numeric>
## [1] 596 0.103679 6 CG -1 0
## [2] 411 0.104116 4 CG -1 0
##      regionType
##      <character>
## [1] loss
## [2] loss
## -----
## seqinfo: 1 sequence from an unspecified genome; no seqlengths

```

6 Session information

```

sessionInfo()

## R version 4.5.0 beta (2025-04-02 r88102)
## Platform: x86_64-pc-linux-gnu
## Running under: Ubuntu 24.04.2 LTS
##
## Matrix products: default
## BLAS: /home/biocbuild/bbs-3.22-bioc/R/lib/libRblas.so
## LAPACK: /usr/lib/x86_64-linux-gnu/lapack/liblapack.so.3.12.0 LAPACK version 3.12.0
##
## locale:
## [1] LC_CTYPE=en_US.UTF-8 LC_NUMERIC=C
## [3] LC_TIME=en_GB LC_COLLATE=C
## [5] LC_MONETARY=en_US.UTF-8 LC_MESSAGES=en_US.UTF-8
## [7] LC_PAPER=en_US.UTF-8 LC_NAME=C
## [9] LC_ADDRESS=C LC_TELEPHONE=C
## [11] LC_MEASUREMENT=en_US.UTF-8 LC_IDENTIFICATION=C
##
## time zone: America/New_York
## tzcode source: system (glibc)
##
## attached base packages:
## [1] stats4 stats graphics grDevices utils datasets methods
## [8] base
##
## other attached packages:
## [1] betareg_3.2-2 DMRcaller_1.41.0 GenomicRanges_1.61.0
## [4] GenomeInfoDb_1.45.0 IRanges_2.43.0 S4Vectors_0.47.0
## [7] BiocGenerics_0.55.0 generics_0.1.3
##
## loaded via a namespace (and not attached):
## [1] httr_1.4.7 knitr_1.50 xfun_0.52
## [4] Formula_1.2-5 highr_0.11 UCSC.utils_1.5.0
## [7] jsonlite_2.0.0 zoo_1.8-14 RcppRoll_0.3.1
## [10] nnet_7.3-20 lmtest_0.9-40 modeltools_0.2-23
## [13] grid_4.5.0 evaluate_1.0.3 compiler_4.5.0
## [16] sandwich_3.1-1 Rcpp_1.0.14 XVector_0.49.0
## [19] lattice_0.22-7 flexmix_2.3-20 R6_2.6.1
## [22] parallel_4.5.0 GenomeInfoDbData_1.2.14 tools_4.5.0

```

References

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- Ferrari, S. and Cribari-Neto, F. (2004). Beta Regression for Modelling Rates and Proportions. *Journal of Applied Statistics*, 31(7):799–815.
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- Krueger, F. and Andrews, S. R. (2011). Bismark: a flexible aligner and methylation caller for Bisulfite-Seq applications. *Bioinformatics*, 27(11):1571–1572.
- Stroud, H., Greenberg, M. V., Feng, S., Bernatavichute, Y., and Jacobsen, S. E. (2013). Comprehensive analysis of silencing mutants reveals complex regulation of the *Arabidopsis* methylome. *Cell*, 152(1-2):352–364.