Package ‘ASpli’

March 22, 2017

Type Package
Title Analysis of alternative splicing using RNA-Seq
Version 1.0.0
Date 2016-08-22
Author Estefania Mancini, Marcelo Yanovsky and Ariel Chernomoretz
License GPL

biocViews GeneExpression, Transcription, AlternativeSplicing, Coverage, DifferentialExpression, DifferentialSplicing, TimeCourse, RNASeq, GenomeAnnotation, Sequencing, Alignment

Depends methods,GenomicRanges,GenomicFeatures,edgeR,BiocGenerics,IRanges,GenomicAlignments,DESeq2,DEXSeq,Gviz,grDevices,stats,utils,S4Vectors,AnnotationDbi,parallel

Suggests RNAseqData,HNRNPC.bam.chr14,BiocStyle

Maintainer Estefania Mancini <emancini@leloir.org.ar>

Description Integrative pipeline for the analysis of alternative splicing using RNAseq.

NeedsCompilation no

R topics documented:

ASpli-package ..................................................... 2
AS accesors ....................................................... 3
AsDiscover .......................................................... 4
ASpliAS-class ...................................................... 5
ASpliCounts ....................................................... 6
ASpliCounts-class ............................................... 7
ASpliDU-class ..................................................... 8
ASpliFeatures-class ............................................. 9
binGenome ........................................................ 10
binGenome-methods ............................................ 10
Counts accesors ................................................. 10
DU accesors ....................................................... 11
DUreport .......................................................... 12
Dureport_DEXSeq ................................................. 13
features accesors ............................................... 14
loadBAM .......................................................... 15
**Description**

ASpli is an integrative and flexible package that facilitates the characterization of genome-wide changes in AS under different experimental conditions. ASpli analyzes the differential usage of introns, exons, and splice junctions using read counts, and estimates the magnitude of changes in AS by calculating differences in the percentage of exon inclusion or intron retention using splice junctions. This integrative approach allows the identification of changes in both annotated and novel AS events. ASpli allows users to produce self-explanatory intermediate outputs, based on the aim of their analysis. A typical workflow involves parsing the genome annotation into new features called bins, overlapping read alignments against those bins, and inferring differential bin usage based on the number of reads aligning to the bins and junctions.

**Details**

- **Package**: ASpli
- **Type**: Package
- **Version**: 0.99.0
- **Date**: 2016-05-25
- **License**: GPL
- **Depends**: methods, GenomicRanges, GenomicFeatures, edgeR, methods, BiocGenerics, IRanges, GenomicAlignments, Gviz

**Author(s)**

Estefania Mancini, Marcelo Yanovsky and Ariel Chernomoretz

**References**


Examples

library(RNAseqData.HNRNPC.bam.chr14)
chr14 <- system.file("extdata","chr14.sqlite", package="ASpli")
genome <- loadDb(chr14)
features <- binGenome(genome)
targets <- data.frame(bam=RNAseqData.HNRNPC.bam.chr14_BAMFILES,
  condition=c(rep("CT",4),rep("KD",4)))
bam <- loadBAM(targets)
counts <- readCounts(features, bam, l=100L, maxISize=50000);
group <- factor(c(rep("CT",4),rep("KD",4)))
pair <- c("CT","KD")
du <- DUreport(counts, targets, pair, group)
as <- AsDiscover(counts, targets, features, bam, threshold=5, l=100, pair=pair)

<table>
<thead>
<tr>
<th>AS accesors</th>
<th>Accessors for ASpliAS object</th>
</tr>
</thead>
</table>

Description

Accessors for ASpliAS object

Usage

altPSI(x)
esPSI(x)
irPIR(x)
joint(x)
junctionsPIR(x)
junctionsPSI(x)

Arguments

x An ASpliAS object

Value

Returns dataframes with genomic metadata and PSI and PIR metrics

Author(s)

Estefania Mancini, Marcelo Yanovsky, Ariel Chernomoretz
Examples

```r
chr14 <- system.file("extdata","chr14.sqlite", package="ASpli")
genome <- loadDb(chr14)
features <- binGenome(genome)
library(RNAseqData.HNRNPC.bam.chr14)
targets <- data.frame(bam=RNAseqData.HNRNPC.bam.chr14_BAMFILES,
                     condition=c(rep("CT",4),rep("KD",4)))
bam <- loadBAM(targets)
counts <- readCounts(features, bam, l=100L, maxISize=50000);
group <- factor(c(rep("CT",4),rep("KD",4)))
pair <- c("CT","KD")
as <- AsDiscover(counts, targets, features, bam, threshold=5, l=100, pair=pair)
altPSI(as)
esPSI(as)
irPIR(as)
joint(as)
junctionsPIR(as)
junctionsPSI(as)
```

AsDiscover

Report PSI and PIR using experimental junctions

Description

Given a bin, it is possible to calculate PSI/PIR metric using junctions to estimate changes in the use of it along different conditions.

Usage

```r
AsDiscover(counts, targets, features, bam, l, pair, threshold, cores)
```

Arguments

- `counts`: An object of class ASpliCounts.
- `targets`: A dataframe containing sample, bam and condition columns.
- `features`: An object of class ASpliFeatures.
- `bam`: A list with BAM files.
- `l`: Read length of sequenced read. Default 100L.
- `pair`: Vector of length two, either numeric or character, providing the pair of groups to be compared.
- `threshold`: Minimum number of reads supporting junctions. Default=5.
- `cores`: Number of processors to use.
ASpliAS-class

Value

An object of class ASpliAS

<table>
<thead>
<tr>
<th>Function</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>irPIR</td>
<td>reports: event, e1i counts (J1), ie1 counts (J2), j_within (J3), PIR by condition. J1, J2, J3 sum of junctions (J1, J2, J3) by condition.</td>
</tr>
<tr>
<td>altPSI</td>
<td>reports: event, J1 (start), J2 (end), J3 (exclusion), PSI. J1, J2, J3 sum of junctions (J1, J2, J3) by condition.</td>
</tr>
<tr>
<td>esPSI</td>
<td>reports: event, J1 (start), J2 (end), J3 (exclusion), PSI. J1, J2, J3 sum of junctions (J1, J2, J3) by condition.</td>
</tr>
<tr>
<td>junctionsPIR</td>
<td>PIR metric for each experimental junction using e1i and ie2 counts. Exclusion junction is the junction itself. This output helps to discover new introns as well as new retention events</td>
</tr>
<tr>
<td>junctionsPSI</td>
<td>Given a junction, it is possible to analyze if it shares start, end or both with another junction. If so, it is because there is more than one way for/of splicing. Using strand information it is possible to classify those pair of junctions into Alt5’ss, Alt3’ss or ES. Ratio between them along samples is reported.</td>
</tr>
</tbody>
</table>

Author(s)

Estefania Mancini, Marcelo Yanovsky and Ariel Chernomoretz

See Also

Accessors: irPIR, altPSI, esPSI, junctionsPIR, junctionsPSI Export: writeAS

Examples

```r
library(RNAseqData.HNRNPC.bam.chr14)
chr14 <- system.file("extdata","chr14.sqlite", package="ASpli")
genome <- loadDb(chr14)
features <- binGenome(genome)
targets <- data.frame(bam=RNAseqData.HNRNPC.bam.chr14_BAMFILES, 
condition=c(rep("CT",4),rep("KD",4)))
bam <- loadBAM(targets)
counts <- readCounts(features, bam, l=100L, maxISize=50000)
group <- factor(c(rep("CT",4),rep("KD",4)))
pair <- c("CT","KD")
as <- AsDiscover(counts, targets, features, bam, l=100L, pair=pair)
writeAS(as=as, output.dir="only_as")
```

---

**ASpliAS-class**

**Class** "ASpliAS"

**Description**

Results of PSI and PIR using experimental junctions
Slots

irPIR: Reports: event, eli counts (J1), ie1 counts (J2), j_within (J3), PIR by condition. J1, J2, J3 sum of junctions (J1, J2, J3) by condition.

altPSI: Reports: event, J1 (start), J2 (end), J3 (exclusion), PSI. J1, J2, J3 sum of junctions (J1, J2, J3) by condition.

esPSI: Reports: event, J1 (start), J2 (end), J3 (exclusion), PSI. J1, J2, J3 sum of junctions (J1, J2, J3) by condition.

join: It is a combination of irPIR, altPSI and esPSI tables

junctionsPIR: PIR metric for each experimental junction using eli and ie2 counts. Exclusion junction is the junction itself. This output helps to discover new introns as well as new retention events

junctionsPSI: Given a junction, it is possible to analyze if it shares start, end or both with another junction. If so, is because there is more than one way for/of splicing. Using strand information it is possible to classify those pair of junctions into Alt5’ss, Alt3’ss or ES. Ratio between them along samples is reported.

Author(s)

Estefania Mancini, Marcelo Yanovsky, Ariel Chernomoretz

See Also

Methods: AsDiscover, Accesors: irPIR,esPSI,junctionsPIR,junctionsPSI

ASpliCounts

Class "ASpliCounts"

Description

Contains results of read overlaps against all feature levels summarization

Slots

gene.counts
exon.intron.counts
junction.counts
eli.counts
ie2.counts
gene.rd
bin.rd

Author(s)

Estefania Mancini, Marcelo Yanovsky, Ariel Chernomoretz
**Description**

Contains results of read overlaps against all feature levels summarization

**Slots**

- gene.counts: Object of class "data.frame"
- exon.intron.counts: Object of class "data.frame"
- junction.counts: Object of class "data.frame"
- e1i.counts: Object of class "data.frame"
- ie2.counts: Object of class "data.frame"
- gene.rd: Object of class "data.frame"
- bin.rd: Object of class "data.frame"

**Methods**

- AsDiscover psi and pir metrics
- countsb bin counts accessor
- countseli e1i counts accessor
- countsdg gene counts accessor
- countsie2 ie2 counts accessor
- countsj junction counts accessor
- DUreport_DEXSeq differential expression and usage estimation using DEXSeq
- DUreport differential expression and usage estimation using DEXSeq
- rdsb bin read densities accessor
- rdsg gen read densities accessor
- rds compute read densities on genes and bins
- writeCounts Export count tables
- writeRds Export read density tables

**Author(s)**

Estefania Mancini, Marcelo Yanovsky, Ariel Chernomoretz
Description

Contains results of differential expression at gene level and differential usage at bin and junction level estimation using DEreport method.

Slots

genes
bins
junctions

Author(s)

Estefania Mancini, Marcelo Yanovsky, Ariel Chernomoretz
binGenome

Description

Exons and introns are subdivided into new features called exon and intron bins and are then classified into exclusively exonic bins, exclusively intronic bins or alternative splicing (AS) bins.

Usage

binGenome(genome, md = NULL)

Arguments

genome An object of class transcriptDb (TxDb)
md A dataframe with symbol (common names) of TxDb genes. If there isn’t md file, gene name will be repeated

Details

Exon and intron coordinates are extracted from gene annotation, only those from multi-exonic genes are saved for further evaluation. In case more than one isoform exist, some exons and introns will overlap. Exons and introns are then disjoint into new features called exon and intron bins, and then they are classified into exclusively exonic bins, exclusively intronic bind or alternative splicing bins (AS-bins), which are labeled according to which alternative splicing event are assumed to came from:

- ES: exon skipping
- IR: intron retention
- Alt5'3'ss: alternative five/three prime splicing site
- "*" (ES*, IR*, AltSS*) means this AS bin/region is involved simultaneously in more than one AS event type
- external: from the beginning or the end of a transcript

Subgenic features are labeled as follow (hypothetical GeneAAA):

- GeneAAA:E001: defines first exonic bin
- GeneAAA:I001: defines first intronic bin
- GeneAAA:Io001: defines first intron before disjoint into bins
- GeneAAA:J001: defines first junction

Junctions are defined as the last position of five prime exon (donor position) and first position of three prime exon (acceptor position). Using TxDb object, it is possible to extract annotated/known junctions. This information will be useful for the analysis of "experimental" junctions (reads aligned with gaps). Bins and junctions are labelled always in 5’ to 3’ sense. This notation is strand independent. It implies that bin / junction with lower numbering is always at 5’.

Value

An ASpliFeatures object. It is a list of features using GRanges format.
Counts accesors

Author(s)
Estefania Mancini, Marcelo Yanovsky, Ariel Chernomoretz

See Also
featuresg, featuresb, featuresj

Examples
chr14 <- system.file("extdata","chr14.sqlite", package="ASpli")
genome <- loadDb(chr14)
features <- binGenome(genome)
GeneCoord <- featuresg(features)
BinCoord <- featuresb(features)
JunctionCoord <- featuresj(features)

Description
Feature coordinates extraction from a Transcript Db Database

Methods
signature(genome = "TxDb") An object of class transcriptDb (TxDb)

Author(s)
Estefania Mancini, Marcelo Yanovsky, Ariel Chernomoretz

See Also
featuresg, featuresb, featuresj

Counts accesors
Accessors for ASpliCounts object

Description
Accessors for ASpliCounts object

Usage
countsb(x)
countse1i(x)
countsg(x)
countsie2(x)
countsj(x)
rdsg(x)
rdsb(x)
## DU acceors

### Arguments

- **x**: An ASpliCounts object

### Value

Returns dataframes with counts by sample and genomic metadata

### Author(s)

Estefania Mancini, Marcelo Yanovsky, Ariel Chernomoretz

### Examples

```r
chr14 <- system.file("extdata","chr14.sqlite", package="ASpli")
genome <- loadDb(chr14)
features <- binGenome(genome)
library(RNAseqData.HNRNPC.bam.chr14)
targets <- data.frame(bam=RNAseqData.HNRNPC.bam.chr14_BAMFILES,
                      condition=c(rep("CT",4),rep("KD",4)))
bam <- loadBAM(targets)
counts <- readCounts(features, bam, l=100L, maxISize=50000);
countsb(counts)
countsels(counts)
counts1g(counts)
countsle2(counts)
countsj(counts)
rdsg(counts)
rdsb(counts)
```

---

## DU acceors

### Description

Accessors for ASpliDU object

### Usage

- genesDE(x)
- binsDU(x)
- junctionsDU(x)

### Arguments

- **x**: An ASpliDU object

### Value

Returns dataframes with genomic metadata and logFC and pvalue

### Author(s)

Estefania Mancini, Marcelo Yanovsky, Ariel Chernomoretz
Examples

```r
chr14 <- system.file("extdata","chr14.sqlite", package="ASpli")
genome <- loadDb(chr14)
features <- binGenome(genome)
library(RNAseqData.HNRNPC.bam.chr14)
targets <- data.frame(bam=RNAseqData.HNRNPC.bam.chr14_BAMFILES,
condition=c(rep("CT",4),rep("KD",4)))
bam <- loadBAM(targets)
counts <- readCounts(features, bam, l=100L, maxISize=50000);
group <- factor(c(rep("CT",4),rep("KD",4)))
pair <- c("CT","KD")
Du <- DUreport(counts, targets, pair, group)
genesDU(Du)
binsDU(Du)
junctionsDU(Du)
```

---

**DUreport**

Differential gene expression and differential bin/junction usage estimation

**Description**

Estimate differential expression at gene level and differential usage at bin and junction level.

**Usage**

`DUreport(counts, targets, pair, group, minGenReads, minBinReads, minRds,ignoreExternal,threshold)`

**Arguments**

- **counts** An object of class ASpliCounts
- **targets** A dataframe containing sample, bam and condition columns
- **pair** vector of length two, either numeric or character, providing the pair of groups to be compared
- **group** Factorial vector with tags for each sample
- **minGenReads** Default 10 reads
- **minBinReads** Default 5 reads
- **minRds** Default 0.05
- **ignoreExternal** Ignore Exon Bins at the beginning or end of the transcript. Default TRUE
- **threshold** Minimum number of junction. Default 5

**Value**

An ASpliDU object with results at genes, bins and junctions level

**Author(s)**

Estefania Mancini, Marcelo Yanovsky, Ariel Chernomoretz
See Also

DEXSeq, edgeR, Accessors: genesDE, binsDU, junctionsDU, Export: writeDU

Examples

library(RNAseqData.HNRNPC.bam.chr14)
chr14 <- system.file("extdata" ,"chr14.sqlite", package="ASpli")
genome <- loadDb(chr14)
features <- binGenome(genome)
targets <- data.frame(bam=RNAseqData.HNRNPC.bam.chr14_BAMFILES,
                      condition=c(rep("CT",4), rep("KD",4)))
bam <- loadBAM(targets)
counts <- readCounts(features, bam, l=100L, maxISize=50000)
group <- factor(c(rep("CT",4), rep("KD",4)))
pair <- c("CT","KD")
du <- DUreport(counts, targets, pair, group)
writeDU(du, output.dir="only_du")

---

**DUreport_DEXSeq**

**Differential gene expression and differential bin/junction usage estimation**

**Description**

Estimate differential expression at gene level and differential usage at bin and junction level.

**Usage**

DUreport_DEXSeq(counts, targets, pair, group, minGenReads, minBinReads, minRds, threshold)

**Arguments**

- **counts**: An object of class ASpliCounts
- **targets**: A dataframe containing sample, bam and condition columns
- **pair**: A vector of length two, either numeric or character, providing the pair of groups to be compared
- **group**: Factorial vector with tags for each sample
- **minGenReads**: Default 10 reads
- **minBinReads**: Default 5 reads
- **minRds**: Default 0.05
- **threshold**: Minimum number of junction. Default 5

**Value**

An ASpliDU object with results at genes, bins and junctions level

**Author(s)**

Estefania Mancini, Marcelo Yanovsky, Ariel Chernomoretz
See Also

DEXSeq, edgeR

Accesors: genesDE, binsDU, junctionsDU
Export: writeDU

Examples

```r
library(RNAseqData.HNRNPC.bam.chr14)
chr14 <- system.file("extdata","chr14.sqlite", package="ASpli")
genome <- loadDb(chr14)
features <- binGenome(genome)
targets <- data.frame(bam=RNAseqData.HNRNPC.bam.chr14_BAMFILES,
                      condition=c(rep("CT",4),rep("KD",4)))
bam <- loadBAM(targets)
counts <- readCounts(features, bam, l=100L, maxISize=50000)
group <- factor(c(rep("CT",4),
                  rep("KD",4)))
pair <- c("CT","KD")
du <- DUreport_DEXSeq(counts, targets, pair, group)
writeDU(du, output.dir="only_du")
```

---

**Description**

Accessors for ASpliFeatures object

**Usage**

```r
featuresg(x)
featuresb(x)
featuresj(x)
```

**Arguments**

- `x` An ASpliFeatures object

**Value**

Returns a GenomicRanges object

**Author(s)**

Estefania Mancini, Marcelo Yanovsky, Ariel Chernomoretz

**Examples**

```r
chr14 <- system.file("extdata","chr14.sqlite", package="ASpli")
genome <- loadDb(chr14)
features <- binGenome(genome)
featuresg(features)
featuresb(features)
featuresj(features)
```
loadBAM

**Description**

Load BAM files into R session using targets object specification

**Usage**

```r
loadBAM(targets, cores)
```

**Arguments**

- `targets`: A dataframe containing sample, bam and condition columns
- `cores`: Number of processors to use

**Value**

A list of GAlignments. Each element of the list correspond to a BAM file (or sample)

**Author(s)**

Estefania Mancini, Marcelo Yanovsky, Ariel Chernomoretz

**Examples**

```r
library(RNAseqData.HNRNPC.bam.chr14)
targets <- data.frame(bam=RNAseqData.HNRNPC.bam.chr14_BAMFILES,
                      condition=c(rep("CT",4),rep("KD",4)))
targets
bam <- loadBAM(targets)
```

---

plotTopTags

**Coverage plots**

**Description**

Using genomic coordinates and BAM files this function is useful for make coverage plots

**Usage**

```r
plotTopTags(auxdf, genome, targetsPlot, output.dir)
```

**Arguments**

- `auxdf`: A data frame: rowNamMes=bin names, gene coordinates, bin coordinates and event name columns
- `genome`: TxDb genome
- `targetsPlot`: A dataframe containing: bam files name, condition (y axe tag), color for each condition
- `output.dir`: Name of directory where plots are supposed to be exported
Value

Coverage plots in png format of selected events

Author(s)

Estefania Mancini, Marcelo Yanovsky, Ariel Chernomoretz

Examples

```r
library(RNAseqData.HNRNPC.bam.chr14)
chr14 <- system.file("extdata","chr14.sqlite", package="ASpli")
genome <- loadDb(chr14)
features <- binGenome(genome)
targets <- data.frame(bam=RNAseqData.HNRNPC.bam.chr14_BAMFILES,
                      condition=c(rep("CT",4),rep("KD",4)))
bam <- loadBAM(targets)
counts <- readCounts(features, bam, l=100L, maxISize = 50000)
pair <- c("CT","KD")
group <- c(rep("CT", 4),rep("KD", 4))
du_HNRNPC <- DUreport(counts, targets, pair, group)
bins <- binsDU(du_HNRNPC)
topTagsBins <- which(bins$bin.fdr <= 0.1 &
                      abs(bins$logFC) >=0.58)
targetsPlot <- data.frame(bam=targets$bam,
                          sample=targets$condition,
                          color=c(rep("blue", 4),rep("red", 4)),
                          stringsAsFactors=FALSE)

auxdf<-bins[topTagsBins,"]
#for simplicity, just one: LRR1:E005

plotTopTags(auxdf["LRR1:E005",],
genome,
targetsPlot,
output.dir="testPlots")
```

rds

Divides read counts by gene and bin length

Description

Divides read counts by gene and bin length

Usage

rds(counts, targets)

Arguments

counts An ASpliCounts object
targets Target dataframe
**readCounts**

*Value*

Read densities of genes and bins

*Author(s)*

Estefania Mancini, Marcelo Yanovsky, Ariel Chernomoretz

---

**readCounts**

*Summarize read overlaps*

**Description**

Summarize read overlaps against all feature levels

**Usage**

`readCounts(features, bam, cores, l, maxISize, minAnchor)`

**Arguments**

- `features`: An object of class ASpliFeatures. It is a list of GRanges at gene, bin and junction level
- `bam`: List of bam files
- `l`: Read length of sequenced library. It is used for compute E1I and IE2 read summarization
- `maxISize`: maximum intron expected size. Junctions longer than this size will be discarded
- `cores`: Number of cores to use. Default 1
- `minAnchor`: Percentage of read that should be aligned in exon-intron boundary

**Value**

An object of class ASpliCounts. Each slot is a dataframe containing features metadata and read counts. Summarization is reported at gene, bin, junction and intron flanking regions (E1I, IE2)

*Author(s)*

Estefania Mancini, Marcelo Yanovsky, Ariel Chernomoretz

**See Also**

Accessors: `countsg`, `countsb`, `countsj`, `counts1i`, `countsie2`, `rdsg`, `rdsb` Export: `writeCounts`

**Examples**

```r
library(RNAseqData.HNRNPC.bam.chr14)
chr14 <- system.file("extdata","chr14.sqlite", package="ASpli")
genome <- loadDb(chr14)
features <- binGenome(genome)
targets <- data.frame(bam=RNAseqData.HNRNPC.bam.chr14_BAMFILES,
                      condition=c(rep("CT",4),rep("KD",4)))
bam <- loadBAM(targets)
counts <- readCounts(features, bam, l=100L, maxISize=50000)#OK
writeCounts(counts, output.dir="only_counts")
```
show-methods

Display a summary of data contained in ASpliObjects

Description
Display a summary of data contained in ASpliObjects

Details
Display a summary of data contained in ASpliObjects

Author(s)
Estefania Mancini, Marcelo Yanovsky, Ariel Chernomoretz

write

Write results

Description
Export tab delimited files in structured output

Usage
writeCounts(counts, output.dir="counts")
writeRds(counts, output.dir="rds")
writeDU(du, output.dir="du")
writeAS(as, output.dir="as")
writeAll(counts, du, as, output.dir="output")

Arguments

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>counts</td>
<td>An ASpliCounts object</td>
</tr>
<tr>
<td>as</td>
<td>An ASpliAS object</td>
</tr>
<tr>
<td>du</td>
<td>An ASpliDU object</td>
</tr>
<tr>
<td>output.dir</td>
<td>Name of output folder (new or existing)</td>
</tr>
</tbody>
</table>

Value
Tab delimited files are exported in a tidy manner into output folder

Author(s)
Estefania Mancini, Marcelo Yanovsky, Ariel Chernomoretz

See Also
AsDiscover, binGenome, DUreport
### Description

Export tab delimited files in structured output

### Details

Tab delimited files are exported in a tidy manner into output folder

### Author(s)

Estefania Mancini, Marcelo Yanovsky, Ariel Chernomoretz

### See Also

- AsDiscover
- binGenome
- DUreport
Index

*Topic alternative splicing, RNA-seq, junctions

altPSI, 5
altPSI (AS accessors), 3
altPSI, ASpliAS-method (ASpliAS-class), 5
AS accessors, 3
AsDiscover, 4, 6, 18, 19
AsDiscover, ASpliCounts-method (ASpliCounts-class), 7
ASpli (ASpli-package), 2
ASpli-package, 2
ASpliAS-class, 5
ASpliCounts, 6
ASpliCounts-class, 7
ASpliDU-class, 8
ASpliFeatures-class, 8
binGenome, 9, 18, 19
binGenome, TxDB-method
  (binGenome-methods), 10
binGenome-methods, 10
binsDU, 13, 14
binsDU (DU accessors), 11
binsDU, ASpliDU-method (ASpliDU-class), 8
Counts accessors, 10
countsb, 17
countsb (Counts accessors), 10
countsb, ASpliCounts-method (ASpliCounts-class), 7
counts1, 17
counts1 (Counts accessors), 10
counts1, ASpliCounts-method (ASpliCounts-class), 7
countst, 17
countst (Counts accessors), 10
countst, ASpliCounts-method (ASpliCounts-class), 7
counts2, 17
counts2 (Counts accessors), 10
counts2, ASpliCounts-method (ASpliCounts-class), 7
countsj, 17
countsj (Counts accessors), 10
countsj, ASpliCounts-method (ASpliCounts-class), 7
DEXSeq, 13, 14
DU accessors, 11
DUreport, 12, 18, 19
DUreport, ASpliCounts-method (ASpliCounts-class), 7
DUreport_DEXSeq, 13
DUreport_DEXSeq, ASpliCounts-method (ASpliCounts-class), 7
dgeR, 13, 14
esPSI, 5, 6
esPSI (AS accessors), 3
esPSI, ASpliAS-method (ASpliAS-class), 5
features accessors, 14
featuresb, 10
featuresb (features accessors), 14
featuresb, ASpliFeatures-method (ASpliFeatures-class), 8
featuresg, 10
featuresg (features accessors), 14
featuresg, ASpliFeatures-method (ASpliFeatures-class), 8
featuresj, 10
featuresj (features accessors), 14
featuresj, ASpliFeatures-method (ASpliFeatures-class), 8
genesDE, 13, 14
genesDE (DU accessors), 11
genesDE, ASpliDU-method (ASpliDU-class), 8
irPIR, 5, 6
irPIR (AS accessors), 3
irPIR, ASpliAS-method (ASpliAS-class), 5
joint (AS accessors), 3
joint, ASpliAS-method (ASpliAS-class), 5
junctionsDU, 13, 14
INDEX

junctonsDU (DU accessors), 11
junctonsDU, ASpliDU-method
   (ASpliDU-class), 8
junctonsPIR, 5, 6
junctonsPIR (AS accessors), 3
junctonsPIR, ASpliAS-method
   (ASpliAS-class), 5
junctonsPSI, 5, 6
junctonsPSI (AS accessors), 3
junctonsPSI, ASpliAS-method
   (ASpliAS-class), 5

loadBAM, 15

plotTopTags, 15

rds, 16
rds, ASpliCounts-method
   (ASpliCounts-class), 7
rdsb, 17
rdsb (Counts accessors), 10
rdsb, ASpliCounts-method
   (ASpliCounts-class), 7
rdsg, 17
rdsg (Counts accessors), 10
rdsg, ASpliCounts-method
   (ASpliCounts-class), 7
readCounts, 17
readCounts, ASpliFeatures-method
   (ASpliFeatures-class), 8

show, ASpliAS-method (show-methods), 18
show, ASpliCounts-method (show-methods), 18
show, ASpliDU-method (show-methods), 18
show, ASpliFeatures-method
   (show-methods), 18
show-methods, 18

write, 18
write-methods, 19
writeAll (write), 18
writeAll, ANY-method (write-methods), 19
writeAS, 5
writeAS (write), 18
writeAS, ASpliAS-method (ASpliAS-class), 5
writeAS-methods (write-methods), 19
writeCounts, 17
writeCounts (write), 18
writeCounts, ASpliCounts-method
   (ASpliCounts-class), 7
writeCounts-methods (write-methods), 19

writeDU, 13, 14
writeDU (write), 18
writeDU, ASpliDU-method (ASpliDU-class), 8
writeDU-methods (write-methods), 19
writeRds (write), 18
writeRds, ASpliCounts-method
   (ASpliCounts-class), 7
writeRds-methods (write-methods), 19