Package ‘BicARE’

January 14, 2017

Version 1.32.0
Date 2008-06-05
Title Biclustering Analysis and Results Exploration
Depends R (>= 1.8.0), Biobase (>= 2.5.5), multtest, GSEABase
Author Pierre Gestraud
Maintainer Pierre Gestraud <pierre.gestraud@curie.fr>
Description Biclustering Analysis and Results Exploration
License GPL-2
URL http://bioinfo.curie.fr
biocViews Microarray, Transcription, Clustering
NeedsCompilation yes

R topics documented:

  BicARE-package .................................................. 1
  bicluster .......................................................... 2
  FLOC ................................................................. 3
  makeReport ........................................................ 5
  residue ............................................................. 5
  sample.bicData .................................................... 6
  sample.biclustering ............................................... 6
  testAnnot .......................................................... 7
  testSet ............................................................. 8

Index 9

BicARE-package  BicARE

Description
Biclustering Analysis and Results Exploration

Details
Package: BicARE
Version: 0.1.0
Date: 2008-06-05
Depends: R (>= 1.8.0), Biobase, multtest, GSEABase
License: GPL
biocViews: Microarray, Transcription, Statistics, Clustering
URL: http://bioinfo.curie.fr
Packaged: Tue Aug 19 14:21:38 2008; pgestraud
Built: R 2.8.0; i686-pc-linux-gnu; 2008-08-19 14:39:57; unix

Index:

FLOC Performs the FLOC algorithm
bicluster Extract a bicluster
makeReport Export the results as html files
residue Residue of a matrix
sample.bicData Example data set for BicARE
sample.biclustering Example biclustering object
testAnnot Find samples annotations over-represented in biclusters
testSet Find gene sets that are enriched in a bicluster

Further information is available in the following vignettes:

BicARE BicARE (source, pdf)

Author(s)
Pierre Gestraud
Maintainer: Pierre Gestraud, <pierre.gestraud@curie.fr>

bicluster Extract a bicluster

Description
Extract a bicluster from an object of class biclustering

Usage
bicluster(biclustering, k, graph=TRUE)

Arguments
biclustering an object of class "biclustering" created by function FLOC
k the number of the bicluster considered in the "biclustering" object
graph boolean, indicating whether the graph should be plotted or not
Value

Returns the bicluster as a matrix with the genes on rows and the samples on columns. Result matrix is of class "bicluster". The "graph" option allows to plot the expression profiles of the genes across the conditions in the bicluster.

Author(s)

Pierre Gestraud

Examples

```r
### extract the first bicluster
data(sample.biclustering)
sample.biclustering
bic <- bicluster(sample.biclustering, 1, graph=TRUE)
plot(bic)
```

FLOC

Performs the FLOC algorithm

Description

Find a given number of biclusters using the a modified version of the FLOC algorithm.

Usage

```r
FLOC(Data, k = 20, pGene = 0.5, pSample=pGene, r = NULL, N = 8, M = 6, t = 500, blocGene = NULL, blocSample = NULL)
```

Arguments

- **Data**: an `ExpressionSet` or a matrix (with genes on rows and conditions on columns)
- **k**: the number of biclusters searched
- **pGene**: genes initial probability of membership to the biclusters
- **pSample**: samples initial probability of membership to the biclusters
- **r**: the residue threshold
- **N**: minimal number of gene per bicluster
- **M**: minimal number of conditions per bicluster
- **t**: number of iterations
- **blocGene**: a matrix indicating the directed initialisation for the genes (see details)
- **blocSample**: a matrix indicating the directed initialisation for the conditions (see details)
FLOC

Details

This biclustering algorithm is based on the FLOC algorithm (FLexible Overlapped biClustering) defined by Yang et al. (see references). It can discover a set of \( k \), possibly overlapping, biclusters. If \( r \) is set to NULL, the residue threshold used in the analysis is the residue of data divided by 10.

\texttt{blocGene} and \texttt{blocSample} are matrix of 0 and 1 with the rows representing the features (gene or samples) and the columns the biclusters. A 1 on line \( i \) and column \( j \) indicates that the feature \( i \) (gene or sample) will be include in the bicluster \( j \) during the initialisation step and will not be removed from it during the analysis. If the number of columns in these matrices is different from the number of bicluster searched, \( k \) is set to the maximal value of these two.

See \texttt{bicluster} to extract a bicluster from the biclustering result.

Value

Returns an object of class “biclustering”, a list containing at least:

- \texttt{Call} the matched call.
- \texttt{ExpressionSet} the data used.
- \texttt{param} a data.frame with the algorithm parameters
- \texttt{bicRow} a matrix of boolean indicating the belonging of the genes to the biclusters.
- \texttt{bicCol} the same as for \texttt{bicRow} but for the conditions
- \texttt{mat.resvol.bic} a matrix describing the biclusters

Author(s)

Pierre Gestraud (<pierre.gestraud@curie.fr>)

References


Examples

data(sample.bicData) ## subset of sample.ExpressionSet from Biobase
residue(sample.bicData) ## 0.3401921
resBic <- FLOC(sample.bicData, k=10, pGene=0.5, r=0.05, N=8, M=10, t=500)
resBic

## initialising samples of 2 biclusters
iniSample <- matrix(0, ncol=2, nrow=26)
## first bicluster initialised around Female cases
iniSample[pData(sample.bicData)$sex="Female",1] <- 1
## second bicluster initialised around control cases
iniSample[pData(sample.bicData)$type="Control",2] <- 1
resBic <- FLOC(sample.bicData, k=10, pGene=0.5, r=0.05, N=8, M=10, t=500, blocSample=iniSample)
resBic
**makeReport**

*Export the results as html files*

**Description**

Creates a directory with html files containing the biclustering results.

**Usage**

```
makeReport(dirPath, dirName, resBic, browse=TRUE)
```

**Arguments**

- `dirPath`  
  path to the directory
- `dirName`  
  the name of the directory where the report will be created
- `resBic`  
  a biclustering result
- `browse`  
  logical. If TRUE the web browser will be opened

**Details**

`makeReport` produces a html report of biclustering results in a new directory named `dirName`. If the `browse` argument is set to TRUE the web browser will be opened on the "home.html" file.  
Make sure to have rights to create the result directory.

**Author(s)**

Pierre Gestraud <pierre.gestraud@curie.fr>

**Examples**

```
data(sample.biclustering)
dirPath <- getwd() ## report created in the current working directory
dirName <- "test"
makeReport(dirPath, dirName, sample.biclustering, browse=FALSE)
```

---

**residue**

*Residue of a matrix*

**Description**

Returns the residue of a matrix.

**Usage**

```
residue(Data)
```

**Arguments**

- `Data`  
  an `ExpressionSet-class` or a matrix
Details

This function computes the residue of a matrix as defined by Yang et al (see references).

Author(s)

Pierre Gestraud

References


See Also

FLOC

Examples

data(sample.bicData)
residue(sample.bicData)

---

**Description**

A subset of sample.ExpressionSet from package Biobase. The data for 26 cases, labeled A to Z and 350 genes. Each case has three covariates: sex (male/female), type (case/control) and score (testing score).

**Usage**

sample.bicData

**Format**

An ExpressionSet

---

**Description**

A biclustering object created by the FLOC function on the sample.bicData with the following options: k=10, pGene = 0.3, pSample = 0.5, r = 0.025, N = 8, M = 8, t = 1000.

**Usage**

sample.biclustering

**Format**

a biclustering object
**testAnnot**

*Find samples annotations over-represented covariates in biclusters*

**Description**

Characterisation of the biclusters in term of over-representation of sample covariates.

**Usage**

```r
testAnnot(resBic, annot=NULL, covariates="all")
```

**Arguments**

- `resBic`: a biclustering result from FLOC
- `annot`: annotation matrix, default value is set to NULL, then phenoData of the ExpressionSet is used
- `covariates`: the names of the covariates that should be tested, default value is set to "all"

**Details**

For each bicluster and each covariate a chi-squared test is performed to test the adequation between the distribution of the levels of the covariates in the bicluster and in the original dataset.

Multiple testing correction is performed by the Benjamini-Yekutieli procedure. The residuals of the tests indicate if the level is over or down represented in the bicluster.

Due to the amount of results it is advised to use the `makeReport` function to get a html report.

**Value**

A biclustering object containing `resBic` and updated with the results of the tests in `resBic$covar`.

The results are presented as a list with:

- `covar`: the samples covariates tested
- `pvalues`: a matrix with the p-values of the tests
- `adjpvalues`: a matrix with the p-values adjusted by the Benjamini Yekutieli procedure
- `index`: a list of matrices with the numbers of each level in each bicluster
- `residuals`: a list of matrices with the residuals of the tests for each modality in each bicluster

**Author(s)**

Pierre Gestraud

**Examples**

```r
data(sample.biclustering)
resBic <- testAnnot(sample.biclustering, annot=NULL, covariates=c("sex", "type"))
```
testSet

*Find gene sets that are enriched in a bicluster*

**Description**

Test of the over-representation of gene sets in the biclusters

**Usage**

```r
testSet(resBic, geneSetCol)
```

**Arguments**

- `resBic` a biclustering object created by `FLOC`
- `geneSetCol` a `GeneSetCollection-class`

**Details**

The over-representation of a gene set in a bicluster is evaluated by an hypergeometric test.

The genes identifiers of the gene sets will automatically be mapped to the same as those used in the data.

Due to the amount of results it is advised to use the `makeReport` function to get a html report.

**Value**

A biclustering object containing `resBic` and updated with the results of the tests in `resBic$geneSet`. The results are presented as a list with:

- `GeneSetCollection` the GeneSetCollection used
- `pvalues` a matrix containing the p-values of the tests for each geneSet and each bicluster
- `adjpvalue` a matrix containing the p-values adjusted by the Benjamini Yekutieli procedure

**Author(s)**

Pierre Gestraud <pierre.gestraud@curie.fr>

**Examples**

```r
data(sample.biclustering)
gss <- GeneSetCollection(sample.biclustering$ExpressionSet[1:50,], setType=GOCollection())
resBic <- testSet(sample.biclustering, gss)
```
Index

*Topic cluster
  bicluster, 2
  FLOC, 3
  makeReport, 5
  residue, 5
  testAnnot, 7
  testSet, 8

*Topic datasets
  sample.bicData, 6
  sample.biclustering, 6

*Topic package
  BicARE-package, 1

BicARE (BicARE-package), 1
BicARE-package, 1
bicluster, 2, 4

ExpressionSet, 3

FLOC, 2, 3, 6–8
makeReport, 5, 7, 8
residue, 5

sample.bicData, 6
sample.biclustering, 6

testAnnot, 7
testSet, 8