Package ‘BioNet’

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Description This package provides functions for the integrated analysis of protein-protein interaction networks and the detection of functional modules. Different datasets can be integrated into the network by assigning p-values of statistical tests to the nodes of the network. E.g. p-values obtained from the differential expression of the genes from an Affymetrix array are assigned to the nodes of the network. By fitting a beta-uniform mixture model and calculating scores from the p-values, overall scores of network regions can be calculated and an integer linear programming algorithm identifies the maximum scoring subnetwork.

License GPL (>= 2)

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BioNet-package

Routines for the functional analysis of biological networks

Description

This package provides functions for the integrated analysis of biological networks and the detection of functional modules. Different datasets can be integrated into the network by assigning p-values derived from statistical tests to the nodes of the network. E.g. p-values obtained from the differential expression of genes from an Affymetrix array are assigned to the nodes of a protein-protein interaction network. By fitting a beta-uniform mixture model and calculating scores from the p-values, overall scores of network regions can be calculated and an integer linear programming algorithm identifies the maximum scoring subnetwork.

Details

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Author(s)

Marcus Dittrich, Daniela Beisser
Maintainer: Marcus Dittrich <marcus.dittrich@biozentrum.uni-wuerzburg.de>

References


aggrPvals

Aggregate several p-values into one p-value

Description

The function aggregates several p-values into one p-value of p-values based on the order statistics of p-values. An overall p-value is given by the \( i \)th order statistic.

Usage

aggrPvals(pval.matrix, order, plot=TRUE)
Arguments

- `pval.matrix`  Numeric matrix of p-values, columns represent different sets of p-values
- `order`  Numeric constant, the order statistic that is used for the aggregation.
- `plot`  Boolean value whether to plot p-value distributions.

Value

Aggregated p-value of the given order.

Author(s)

Daniela Beisser

Examples

data(pvaluesExample)
aggrPvals(pval.matrix=pvaluesExample, order=2)

bumOptim  

Fitting a beta-uniform mixture model to p-value distribution

Description

The function fits a beta-uniform mixture model to a given p-value distribution.

Usage

`bumOptim(x, starts=1, labels=NULL)`

Arguments

- `x`  Numerical vector of p-values, has to be named with the gene names or the gene names can be given in the `labels` parameter.
- `starts`  Number of start points for the optimization.
- `labels`  Gene names for the p-values.

Value

List of class `fb` with the following elements:

- `lambda`  Fitted parameter $\lambda$ for the beta-uniform mixture model.
- `a`  Fitted parameter $a$ for the beta-uniform mixture model.
- `negLL`  Negative log-likelihood.
- `pvalues`  P-value vector.

Author(s)

Marcus Dittrich and Daniela Beisser
compareNetworks

References


See Also

*fitBumModel, plot.bum, hist.bum*

Examples

```r
data(pvaluesExample)
pvals <- pvaluesExample[,1]
bum <- bumOptim(x=pvals, starts=10)
bum
```

```r

compareNetworks(network1, network2, plot=TRUE)
```

Arguments

- **network1**: Network `graphNEL` or `igraph` format.
- **network2**: Second network in `graphNEL` or `igraph` format, or subnetwork drawn from first network.
- **plot**: Boolean value, whether to plot the cumulative degree distributions.

Value

A vector of network parameters is returned:

- **diam.network1**: Network diameter
- **diam.network2**: Diameter of the subnetwork
- **av.degree.network1**: Average degree of the network
- **av.degree.network2**: Average degree of the subnetwork

Description

The function compares the following parameters of two networks: diameter, average degree, degree exponent, average path length and plots the cumulative degree distributions. The networks have to be connected components.
consensusScores

Consensus Scores Calculation

The function calculates consensus scores for a network, given a list of replicate modules.

**Usage**

```r
consensusScores(modules, network, ro=length(modules)/2)
```

**Arguments**

- `modules`: Calculated modules from pseudo-replicates of expression values in `igraph` or `graphNEL` format.
- `network`: Interaction network, which should be scores. In `igraph` or `graphNEL` format.
- `ro`: Threshold which is subtracted from the scores to obtain positive and negative values. The default value is half of the number of replicates.

**Value**

A result list is returned, consisting of:

- `N.scores`: Numerical vector node scores.
- `E.scores`: Numerical vector edge scores.
- `N.frequencies`: Numerical vector node frequencies from the replicate modules.
- `E.frequencies`: Numerical vector edge frequencies from the replicate modules.

**Description**

The function calculates consensus scores for a network, given a list of replicate modules.

```r
library(DLBCL)
data(interactome)
subnet1 <- largestComp(subNetwork(nodes(interactome)[1:100], interactome))
subnet2 <- largestComp(subNetwork(nodes(interactome)[101:200], interactome))
compareNetworks(network1=subnet1, network2=subnet2)
```

Author(s)

Daniela Beisser
Examples

library(DLBCL)
data(interactome)

network <- interactome

# precomputed Heinz modules from pseudo-replicates
## Not run: lib <- file.path(.path.package("BioNet"), "extdata")
modules <- readHeinzGraph(node.file=file.path(datadir, "ALL_n_resample.txt.0.hnz"), network=network)
cons.scores <- consensusScores(modules, network)

## End(Not run)

fbum

Compute the density of the bum distribution

Description

Function to compute the density of the beta-uniform mixture model.

Usage

fbum(x, lambda, a)

Arguments

x A numeric value.
lambda Parameter lambda, mixture parameter, proportion of uniform component
a Parameter a, shape parameter of beta component

Value

Value of the density of the bum distribution for x.

Author(s)

Marcus Dittrich

References


See Also

bumOptim, fitBumModel

Examples

y <- fbum(x=0.5, lambda=0.1, a=0.1)
y
fbumLL

*Calculate log likelihood of BUM model*

**Description**

The function calculates the log likelihood of the BUM model.

**Usage**

```r
fbumLL(parms, x)
```

**Arguments**

- `parms`  
  Vector of parameters; lambda and a.
- `x`  
  Numerical vector of p-values.

**Value**

- Log likelihood.

**Author(s)**

Marcus Dittrich

**Examples**

```r
data(pvaluesExample)
pvals <- pvaluesExample[,1]
bum.mle <- fitBumModel(pvals, plot=FALSE)
fbumLL(parms=c(bum.mle$lambda, bum.mle$a), x=pvals)
```

fdrThreshold

*Calculate p-value threshold for given FDR*

**Description**

The function calculates the p-value threshold tau for a given false discovery rate. Tau is used for the scoring function.

**Usage**

```r
fdrThreshold(fdr, fb)
```

**Arguments**

- `fdr`  
  False discovery rate.
- `fb`  
  Model from the beta-uniform mixture fitting.

**Value**

- P-value threshold tau.
Author(s)
Marcus Dittrich

References

See Also
fbum, fitBumModel

Examples
```r
data(pvaluesExample)
pvals <- pvaluesExample[,1]
bum.mle <- fitBumModel(pvals, plot=FALSE)
tau <- fdrThreshold(fdr=0.001, fb=bum.mle)
tau
```

fitBumModel  
*Fit beta-uniform mixture model to a p-value distribution*

Description
The function fits a beta-uniform mixture model to a given p-value distribution. The BUM method was introduced by Stan Pounds and Steve Morris to model the p-value distribution as a signal-noise decompostion. The signal component is assumed to be B(a,1)-distributed, whereas the noise component is uniform-distributed under the null hypothesis.

Usage
```r
fitBumModel(x, plot = TRUE, starts=10)
```

Arguments
- **x**: Numeric vector of p-values.
- **plot**: Boolean value, whether to plot a histogram and qqplot of the p-values with the fitted model.
- **starts**: Numeric value giving the number of starts for the optimization.

Value
Maximum likelihood estimator object for the fitted bum model. List of class fb with the following elements:
- **lambda**: Fitted parameter *lambda* for the beta-uniform mixture model.
- **a**: Fitted parameter *a* for the beta-uniform mixture model.
- **negLL**: Negative log-likelihood.
- **pvalues**: P-value vector.
getCompScores

Author(s)

Daniela Beisser

References


Examples

```r
data(pvaluesExample)
pvals <- pvaluesExample[,1]
bum.mle <- fitBumModel(pvals, plot=TRUE)
bum.mle
```

```r
getCompScores
```

**Partition scores for subgraphs of the network**

Description

The function partitions the scores into scores for each subgraph of the network.

Usage

```r
getCompScores(network, score)
```

Arguments

- `network` A network in `graphNEL` or `igraph` format.
- `score` Vector of scores.

Value

A data frame with the components of the network and the score for each PPI identifier.

Author(s)

Marcus Dittrich

Examples

```r
library(DLBCL)
data(interactome)
data(dataLym)

# create random subgraph with 100 nodes and their direct neighbors
nodes <- nodes(interactome)[sample(length(nodes(interactome)), 100)]
subnet <- subNetwork(nodeList=nodes, network=interactome, neighbors="first")
score <- dataLym$score001
names(score) <- dataLym<label
getCompScores(score=score, network=subnet)
```
**getEdgeList**

- **Description**
  A network in *graphNEL* or *igraph* format is converted to an edgelist.

- **Usage**
  ```r
  getEdgeList(network)
  ```

- **Arguments**
  - `network` Network in *graphNEL* or *igraph* format.

- **Value**
  A matrix whose columns represent the connected edges.

- **Author(s)**
  Marcus Dittrich

- **Examples**
  ```r
  library(DLBCL)
  data(interactome)
  getEdgeList(interactome)[1:10,]
  ```

**hist.bum**

- **Description**
  The function plots a histogram of the p-values together with the fitted bum-model.

- **Usage**
  ```
  hist(x, breaks=50, main="Histogram of p-values", xlab="P-values", ylab="Density", ...)
  ```

- **Arguments**
  - `x` Maximum likelihood estimator object of the beta-uniform mixture fit.
  - `breaks` Breaks for the histogram.
  - `main` An overall title for the plot.
  - `xlab` A title for the x axis.
  - `ylab` A title for the y axis.
  - `...` Other graphic parameters for the plot.
largestComp

Author(s)

Daniela Beisser

See Also

fitBumModel, hist.bum, bumOptim

Examples

data(pvaluesExample)
pvals <- pvaluesExample[,1]
mle <- fitBumModel(pvals, plot=FALSE)
hist(mle)

largestComp Extract largest component of network

Description

The function extracts the largest component of a network.

Usage

largestComp(network)

Arguments

network A graph in graphNEL or igraph format.

Value

A new graph object that represents the largest component of the given network.

Author(s)

Marcus Dittrich

Examples

library(DLBCL)
data(interactome)
interactome
largestComp(interactome)
largestScoreComp

Component with largest score

Description
The function extracts the component of the network with the largest score. All nodes have to exceed the given level for the score.

Usage
largestScoreComp(network, score, level=0)

Arguments

- network: Network in graphNEL or igraph format.
- score: Vector of scores for the network.
- level: Cut-off level for the score for the component.

Value
Subgraph of the network with a score larger than the given level.

Author(s)
Marcus Dittrich

Examples
library(DLBCL)
data(interactome)
data(dataLym)
network <- rmSelfLoops(interactome)
score <- dataLym$score001
names(score) <- dataLym$label
lComp <- largestScoreComp(network=network, score=score, level=1)
## Not run: plotModule(lComp)

loadNetwork.sif

Load network from Cytoscape sif file

Description
The function loads a network from a Cytoscape sif file. Edge attributes are provided in the ea.file or vector of ea.files. The node attributes are provided the same way. For other formats see read.graph in the igraph package.

Usage
loadNetwork.sif(sif.file, na.file, ea.file, format=c("graphNEL", "igraph"), directed=FALSE)
loadNetwork.tab

Load network from tabular format

Description

The function loads a network from a tabular format.

Usage

loadNetwork.tab(file, header=TRUE, directed=FALSE, format=c("graphNEL", "igraph"))

Arguments

file       File with network to load.
header     Boolean value whether to include header or not.
directed   Boolean value whether the network is to be directed or not.
format     Output format of the network, either graphNEL or igraph

Author(s)

Marcus Dittrich

See Also

loadNetwork.sif
**makeNetwork**

*Create graph from source and target vectors*

**Description**

Function to create a graph in *graphNEL* or *igraph* format from a source and a target vector.

**Usage**

```r
makeNetwork(source, target, edgemode="undirected", format=c("graphNEL", "igraph"))
```

**Arguments**

- `source`: Vector of source nodes.
- `target`: Vector of corresponding target nodes.
- `edgemode`: For an "undirected" or "directed" network.
- `format`: Graph format, eiter *graphNEL* or *igraph*.

**Value**

A graph object.

**Author(s)**

Marcus Dittrich

**See Also**

`loadNetwork.sif`, `saveNetwork`

**Examples**

```r
source <- c("a", "b", "c", "d")
target <- c("b", "c", "a", "a")
graph <- makeNetwork(source, target, edgemode="undirected")
```

---

**mapByVar**

*Select probeset by variance and get PPI ID*

**Description**

The function selects for each gene the probeset with the highest variance and gets the PPI ID for each gene. The PPI identifier is: gene symbol(Entrez ID). Affymetrix identifiers are mapped to the ENTREZ ID.

**Usage**

```r
mapByVar(exprSet, network=NULL, attr="geneID", ignoreAFFX=TRUE)
```
Arguments

exprSet  Affymetrix ExpressionSet.
network  Network that is used to map the Affymetrix identifiers.
attr  The attribute of the network that is used to map the Affymetrix IDs. The IDs are mapped to the unique Entrez gene IDs, which are by default stored in the "geneID" attribute of the network.
ignoreAFFX  Boolean value, whether to ignore or leave AFFX control genes.

Value

Expression matrix with one gene (PPI ID) per probeset.

Author(s)

Daniela Beisser

Examples

## Not run: library(ALL);
data(ALL);
mapped.e.set <- mapByVar(ALL);
mapped.e.set[,1:10];
## End(Not run)

permutateNodes  

Permute node labels

Description

Function to permutate node labels of a given network.

Usage

permutateNodes(network)

Arguments

network  Network in graphNEL or igraph format.

Value

Network with permutated labels.

Author(s)

Marcus Dittrich
**Examples**

```r
library(DLBCL)
data(interactome)
# remove self-loops before permutating the labels
interactome <- rmSelfLoops(interactome)
perm.net <- permutateNodes(interactome)
perm.net
```

---

**piUpper**

*Upper bound π for the fraction of noise*

**Description**

The function calculates the upper bound π for the fraction of noise.

**Usage**

```r
piUpper(fb)
```

**Arguments**

- `fb` Fitted bum model, list with parameters a and lambda.

**Value**

Numerical value for the upper bound π.

**Author(s)**

Marcus Dittrich

**See Also**

`bumOptim`, `fitBumModel`

**Examples**

```r
data(pvaluesExample)
pvals <- pvaluesExample[,1]
bum <- bumOptim(pvals, starts=10)
piUpper(fb=bum)
```
plot.bum

Quantile-quantile plot for the beta-uniform mixture model

Description

The function plots the theoretical quantiles of the fitted bum model against the quantiles of the observed p-value distribution.

Usage

```r
## S3 method for class 'bum'
plot(x, main="QQ-Plot", xlab="Estimated p-value", ylab="Observed p-value", ...)
```

Arguments

- `x`: Maximum likelihood estimation object of the fitted bum model.
- `main`: An overall title for the plot.
- `xlab`: A title for the x axis.
- `ylab`: A title for the y axis.
- `...`: Other graphic parameters for the plot.

Author(s)

Daniela Beisser

See Also

`fitBumModel`, `plot.bum`, `bumOptim`

Examples

```r
data(pvaluesExample)
pvals <- pvaluesExample[,1]
mle <- fitBumModel(pvals, plot=FALSE)
plot(mle)
```

plot3dModule

3D plot of the network

Description

The function plots a network from `graphNEL` or `igraph` format in 3D using a modified function from the package igraph and requires the package rgl which uses openGL. The 3D plot can be zoomed, rotated, shifted on the canvas. This function is just used to visualize the modules. For further plotting options use the rglplot function of the igraph package. If a score attribute is provided in the graph this will be used for the coloring of the nodes. Otherwise a vector of values can be given by the `diff.or.score` argument. The vector has to contain positive and negative values, either scores or values for differential expression (fold changes). Labels for the nodes can be provided by the `labels` argument, otherwise it will be automatically looked for a `geneSymbol` attribute of the nodes.
plotLLSurface

Usage

plot3dModule(network, labels=NULL, windowSize = c(100,100,1500,1000), diff.or.scores=NULL, red=c("negative", "positive"), ...)

Arguments

network Network in graphNEL or igraph format.
labels Labels for the nodes of the network. Otherwise it will be automatically looked for a geneSymbol attribute of the nodes.
windowSize Numerical vector of size four to set the size of the rgl device.
diff.or.scores Named numerical vector of differential expression (fold changes) or scores of the nodes in the network. These will be used for node coloring. Otherwise a score attribute of the nodes will be automatically used.
red Either "negative" or "positive", to specify which values are to be colored red in the plot.
... Other graphic parameters for the plot.

Author(s)

Daniela Beisser

See Also

save3dModule, plotModule

Examples

library(DLBCL)
data(interactome)
data(dataLym)
interactome <- subNetwork(dataLym$label, interactome)
interactome <- rmSelfLoops(interactome)
fchange <- dataLym(diff
names(fchange) <- dataLym$label
subnet <- largestComp(subNetwork(nodes(interactome)[1:100], interactome))
diff <- fchange[nodes(subnet)]

## Not run: library(rgl);
plot3dModule(network=subnet, diff.or.score=diff)
## End(Not run)

plotLLSurface Log likelihood surface plot

Description

The function plots the log likelihood surface for all a and lambda parameter of the beta-uniform mixture model.

Usage

plotLLSurface(x, opt=NULL, main="Log-Likelihood Surface", color.palette = heat.colors, nlevels = 32)
Arguments

x     Numeric vector of p-values.
opt   List of optimal parameters for a and lambda from the beta-uniform mixture model.
main  The overall title of the plot.
color.palette  Color scheme of the image plot.
nlevels Number of color levels.

Author(s)

Marcus Dittrich

Examples

library(DLBCL)
data(dataLym)
pvals <- dataLym$t.pval
names(pvals) <- dataLym$label
mle <- fitBumModel(pvals, plot=FALSE)
plotLLSurface(x=pvals, opt=mle)

Description

The function plots a network from graphNEL or igraph format, adapted from an igraph plotting function. It is just used to visualize the modules. For further plotting options use the plot.igraph function of the igraph package. The shapes of the nodes can be changed according to the scores argument, then negative scores appear squared. The color of the nodes can be changed according to the diff.expr argument. Negative values lead to green nodes, positive values are colored in red. If the vectors are not provided, it will be automatically looked for nodes attributes with the name score and diff.expr.

Usage

plotModule(network, layout=layout.fruchterman.reingold, labels=NULL, diff.expr=NULL, scores=NULL, ...)

Arguments

network  Network in graphNEL or igraph format.
layout  Layout algorithm, e.g. layout.fruchterman.reingold or layout.kamada.kawai.
labels  Labels for the nodes of the network.
diff.expr  Named numerical vector of differential expression (fold changes) of the nodes in the network. These will be used for coloring of the nodes. It will be automatically looked for nodes attribute with the name diff.expr, if the argument is null.
scores  Named numerical vector of scores of the nodes in the network. These will be used for the shape of the nodes. It will be automatically looked for nodes attribute with the name score, if the argument is null.
print.bum

Main title of the plot.

Numerical value or vector for the size of the vertices.

Other graphic parameters for the plot.

Author(s)

Marcus Dittrich and Daniela Beisser

See Also

plot3dModule

Examples

library(DLBCL)
data(dataLym)
data(interactome)
interactome <- subNetwork(dataLym$label, interactome)
interactome <- rmSelfLoops(interactome)
fchange <- dataLym$diff
names(fchange) <- dataLym$label
# create random subnetwork
subnet <- largestComp(subNetwork(nodes(interactome)[1:100], interactome))
fchange <- fchange[nodes(subnet)]

# color random subnetwork by the fold change
## Not run: plotModule(network=subnet, diff.expr=fchange)

print.bum

Print information about bum model

Description

The function prints information about the bum model.

Usage

## S3 method for class 'bum'
print(x, ...)

Arguments

x

Maximum likelihood estimator object of the beta-uniform mixture fit.

... Other graphic parameters for print.

Author(s)

Marcus Dittrich

See Also

fitBumModel, summary.bum
Examples

```r
data(pvaluesExample)
pvals <- pvaluesExample[,1]
mle <- fitBumModel(pvals, plot=FALSE)
print(mle)
```

```
pvaluesExample  Example p-values for aggregation statistics
```

Description

Data example consisting of a matrix of p-values. Each gene has two corresponding p-values. These p-values can be aggregated into a p-value of p-values by the method `aggrPvals`.

Usage

```r
data(pvaluesExample)
```

Examples

```r
data(pvaluesExample)
pvaluesExample[1:10,]
```

```
readHeinzGraph  Convert HEINZ output to graph
```

Description

Function to convert the HEINZ output to a graph object, or if the output is in matrix form, it will create a list of graphs. The function needs the node and the original network, from which the module is calculated.

Usage

```r
readHeinzGraph(node.file, network, format=c("graphNEL", "igraph"))
```

Arguments

- `node.file`: Heinz node output file.
- `network`: Original network from which Heinz input was created.
- `format`: Graph format of output, either `igraph` or `graphNEL`.

Value

Graph object.

Author(s)

Daniela Beisser
**Examples**

```r
library(DLBCL)
data(interactome)
# precomputed Heinz output files
## Not run: lib <- file.path(.path.package("BioNet"), "extdata")
module <- readHeinzGraph(node.file=file.path(lib, "lymphoma_nodes_001.txt.0.hnz"), network=interactome, format="graphNEL");
plotModule(module);
## End(Not run)
```

---

**readHeinzTree**  
Convert HEINZ output to tree

**Description**

Converts the HEINZ output to a tree in graph format. If the output is in matrix form, it will create a list of graphs. The function needs the node and edge file and the original network from which the module is calculated.

**Usage**

```r
readHeinzTree(node.file, edge.file, network, format=c("graphNEL", "igraph"))
```

**Arguments**

- `node.file`: Heinz node output file.
- `edge.file`: Heinz edge output file.
- `network`: Original network from which Heinz input was created.
- `format`: Output format of the graph, either `igraph` or `graphNEL`.

**Value**

A graph object.

**Author(s)**

Daniela Beisser

**Examples**

```r
library(DLBCL)
data(interactome)
# precomputed Heinz output files
## Not run: lib <- file.path(.path.package("BioNet"), "extdata")
module <- readHeinzTree(node.file=file.path(lib, "lymphoma_nodes_001.txt.0.hnz"), edge.file=file.path(lib, "lymphoma_edges_001.txt.0.hnz");
plotModule(module);
## End(Not run)
```
resamplingPvalues  

Resampling of microarray expression values and test for differential expression.

Description

The function uses a 50% jackknife resampling to calculate a pseudo-replicate of the expression matrix. The resampled expression values are used thereupon to calculate p-values for the differential expression between the given groups. Only two-group comparisons are allowed for the performed t-test.

Usage

resamplingPvalues(exprMat, groups, alternative = c("two.sided", "less", "greater"), resampleMat=FALSE)

Arguments

exprMat  
Matrix with microarray expression values.

groups  
Factors for two groups that are tested for differential expression.

alternative  
Testing alternatives for the t-test: "two.sided", "less" or "greater".

resampleMat  
Boolean value, whether to retrieve the matrix of jackknife resamples or not.

Value

A result list is returned, consisting of:

p.values  
Numerical vector of p-values.

resampleMat  
Matrix of resampled expression values.

Author(s)

Daniela Beisser

Examples

library(ALL)
data(ALL)
mat <- exprs(ALL)
groups <- factor(c(rep("A", 64), rep("B", 64)))
results <- resamplingPvalues(mat, groups, alternative="greater")
rmSelfLoops

*Remove self-loops in a graph*

**Description**

The function removes self-loops, edges that start and end in the same node, from the network.

**Usage**

```r
rmsSelfLoops(network)
```

**Arguments**

- `network` A graph object, either in `graphNEL` or `igraph` format.

**Value**

The graph with the removed edges.

**Author(s)**

Marcus Dittrich

**Examples**

```r
graph <- makeNetwork(c("a", "b", "c", "d", "e", "a"), c("b", "c", "d", "e", "e", "e"))
graph2 <- rmSelfLoops(graph)
edges(graph)
edges(graph2)
```

runFastHeinz

*Calculate heuristically maximum scoring subnetwork*

**Description**

The function uses an heuristic approach to calculate the maximum scoring subnetwork. Based on the given network and scores the positive nodes are in the first step aggregated to meta-nodes between which minimum spanning trees are calculated. In regard to this, shortest paths yield the approximated maximum scoring subnetwork. This function can be used if a CPLEX license is not available to calculate the optimal solution.

**Usage**

```r
runFastHeinz(network, scores)
```

**Arguments**

- `network` A graph in `igraph` or `graphNEL` format.
- `scores` A named vector, containing the scores for the nodes of the network. All nodes need to be scored in order to run the algorithm.
runHeinz

Value

A subnetwork in the input network format.

Author(s)

Daniela Beisser

See Also

writeHeinzEdges, writeHeinzNodes, readHeinzTree, readHeinzGraph, runHeinz

Examples

library(DLBCL)
# load p-values
data(dataLym)
# load graph
data(interactome)
# get induced subnetwork for all genes contained on the chip
interactome <- subNetwork(dataLym$label, interactome)
p.values <- dataLym$t.pval
names(p.values) <- dataLym$label
bum <- fitBumModel(p.values, plot=TRUE)
scores <- scoreNodes(network=interactome, fb=bum, fdr=0.0001)
module <- runFastHeinz(network=interactome, scores=scores)
## Not run: plotModule(module)

Description

The function starts HEINZ from command line. The HEINZ folder has to include the heinz.py python script and the dhea file. CPLEX has to be installed and accessible from the computer R runs on.

Usage

runHeinz(heinz.folder="", heinz.e.file, heinz.n.file, N=TRUE, E=FALSE, diff=-1, n=1)

Arguments

heinz.folder The folder which contains the heinz.py python script and the dhea file.
heinz.e.file The HEINZ edge input file. See writeHeinzEdges
heinz.n.file The HEINZ node input file. See writeHeinzNodes
N Boolean value, whether to run HEINZ on nodes.
E Boolean value, whether to run HEINZ on edges. HEINZ can run on both with N and E set to TRUE.
diff Difference of suboptimal solutions to optimal solution in haming distance in percent. Parameter is set to -1 for optimal solution.
n Number of optimal and suboptimal solutions, the standard n=1 delivers only the optimal solution.
save3dModule

Details

This function starts the integer linear programming algorithm to calculate the optimal scoring sub-network. The algorithm might be started in the command line when the CPLEX is installed on another machine. To start it from command line use: heinz.py -e edge.file.txt -n node.file.txt -E False/True -N False/True. The results can be loaded with readHeinzTree, readHeinzGraph as a graph object.

Author(s)

Daniela Beisser

References


See Also

writeHeinzEdges, writeHeinzNodes, readHeinzTree, readHeinzGraph

save3dModule | Save a 3D plot of the network

Description

The function saves a 3D plot of a network to file, therefore it requires the plot to be open. A screenshot of the 3D plot can be saved in "pdf" format. Background of the device is changed to white for plotting. The screenshot can take several seconds for large plots.

Usage

save3dModule(file)

Arguments

file | File to save to.

Author(s)

Daniela Beisser

See Also

plot3dModule, plotModule
saveNetwork

Save undirected network in various formats

Description

The function saves a graph in a Cytoscape readable format: either in XGMML format, or as two tables, one for the nodes with attributes and one for the edges with attributes, or as .sif file. Or other standard formats like tab separated, .tgf, .net

Usage

saveNetwork(network, name="network", file, type=c("table", "XGMML", "sif", "tab", "tgf", "net"))

Arguments

network  Network to save.
name     Name of the network, only needed for the XGMML format.
file     File to save to.
type     Type in which graph shall be saved.

Details

The format types are "XGMML", "table", "sif", "tab", "tgf" and "net". XGMML (eXtensible Graph Markup and Modeling Language) is an XML format based on GML which is used for graph description. Edges, nodes and their affiliated attributes are all saved in one file. In the table format two tables are created, one for the nodes with attributes and one for the edges with attributes. The .sif format creates a .sif file for the network and an node attribute (.NA) or edge attribute (.EA) for each attribute. The name of the attribute is the filename. Tab writes only the edges of the network in a tabular format. Tgf save the network to simple .tgf format. The net format writes a Pajek readable file of the network and the ET type saves the edge tags to file.

Author(s)

Daniela Beisser and Marcus Dittrich
Examples

library(DLBCL)
# create small network
library(igraph)
data(interactome)
interactome <- igraph.from.graphNEL(interactome)
small.net <- subNetwork(V(interactome)[1:16]$name, interactome)
E(small.net)$e.weight <- rep(1, length(E(small.net)))
V(small.net)$n.weight <- rep(2, length(V(small.net)))
summary(small.net)
## Not run: saveNetwork(small.net, file="example_network", name="small.net", type="XGMML")

Description

The function generates a dataframe for a given range of FDRs.

Usage

scanFDR(fb, fdr, labels=names(fb$pvalues))

Arguments

fb Fitted bum model.
fdr Vector of FDRs.
labels Data frame labels.

Value

Dataframe of scores for given p-values and a range of FDRs.

Author(s)

Marcus Dittrich

See Also

bumOptim, fitBumModel

Examples

data(pvaluesExample)
pvals <- pvaluesExample[,1]
bum <- bumOptim(pvals, starts=10)
scores <- scanFDR(fb=bum, fdr=c(0.1, 0.001, 0.0001))
scores[1:10,]
scoreFunction

Scoring function for p-values

Description
The function calculates a score for each gene with a given FDR from the fitted beta-uniform mixture model.

Usage
scoreFunction(fb, fdr=0.01)

Arguments
fb
Model from the beta-uniform mixture fitting.
fdr
Numeric constant, from the false discovery rate a p-value threshold is calculated. P-values below this threshold are considered to be significant and will score positively, p-values above the threshold are supposed to arise from the null model. The FDR can be used to control the size of the maximum scoring subnetwork, by zooming in and out in the same region.

Value
Score vector for the given p-values.

Author(s)
Marcus Dittrich and Daniela Beisser

References

Examples
data(pvaluesExample)
pvals <- pvaluesExample[,1]
bum.mle <- fitBumModel(pvals, plot=FALSE)
scores <- scoreFunction(fdr=0.1, fb=bum.mle)
scores
**scoreNodes**

*Score the nodes of a network*

**Description**

The function derives scores from the p-values of the nodes of a network.

**Usage**

```r
scoreNodes(network, fb, fdr=0.05)
```

**Arguments**

- `network`: A network in `graphNEL` or `igraph` format.
- `fb`: Fitted bum model.
- `fdr`: False discovery rate.

**Value**

Ordered score vector for the nodes of the network.

**Author(s)**

Marcus Dittrich

**See Also**

`bumOptim`, `fitBumModel`

**Examples**

```r
catalogue <- library(DLBCL)
# load p-values
data(dataLym)
# load graph
data(interactome)
# get induced subnetwork for all genes contained on the chip
chipGraph <- subNetwork(dataLym$label, interactome)
p.values <- dataLym$t.pval
names(p.values) <- dataLym$label
bum <- fitBumModel(p.values, plot=TRUE)
scoreNodes(network=chipGraph, fb=bum, fdr=0.001)
```
scoreOffset  

*Change score offset for 2 FDRs*

**Description**

Function to change score offset from FDR1 to FDR2.

**Usage**

```r
scoreOffset(fb, fdr1, fdr2)
```

**Arguments**

- **fb** Model from the beta-uniform mixture fitting.
- **fdr1** First false discovery rate.
- **fdr2** Second false discovery rate.

**Value**

Offset for the score of the second FDR.

**Author(s)**

Marcus Dittrich

**See Also**

`bumOptim`, `fitBumModel`

**Examples**

```r
data(pvaluesExample)
pvals <- pvaluesExample[,1]
bum <- bumOptim(pvals, starts=10)
scoreOffset(bum, fdr1=0.001, fdr2=0.000001)
```

sortedEdgeList  

*Get a sorted edgelist*

**Description**

Function to get a sorted edgelist where the source protein is alphabetically smaller than the target protein from an undirected network.

**Usage**

```r
sortedEdgeList(network)
```

**Arguments**

- **network** Undirected network in *igraph* or *graphNEL* format.
Value

Vector of sorted edges, where the source protein is alphabetically smaller than the target protein.

Author(s)

Daniela Beisser

Examples

library(DLBCL)
data(interactome)
E.list <- sortedEdgeList(interactome)

subNetwork

Create a subGraph

Description

The function creates a subgraph with the nodes given in the nodeList or for these nodes including their direct neighbors.

Usage

subNetwork(nodeList, network, neighbors=c("none", "first"))

Arguments

nodeList Character vector of nodes, contained in the subgraph.
network Graph that is used for subgraph extraction.
neighbors Neighborhood, that is chosen for the subgraph extraction. "none" are only the selected nodes, "first" includes the direct neighbors of the selected nodes.

Value

A graph object.

Author(s)

Marcus Dittrich

Examples

library(igraph)
el <- cbind(c("a", "b", "c", "d", "e", "f", "d"), c("b", "c", "d", "e", "f", "a", "b"))
graph <- graph.edgelist(el, directed=TRUE)

node.list <- c("a", "b", "c")
graph2 <- subNetwork(nodeList=node.list, network=graph)
## Not run: par(mfrow=c(1,2));
plotModule(graph);
plotModule(graph2)
## End(Not run)
# or in graphNEL format:
graph3 <- igraph.to.graphNEL(graph)
graph4 <- subNetwork(nodeList=node.list, network=graph3)
graph3
graph4

**summary.bum**

*Print summary of informations about bum model*

### Description

The function summarizes information about the bum model.

### Usage

```r
## S3 method for class 'bum'
summary(object, ...)
```

### Arguments

- **object**: Maximum likelihood estimator object of the beta-uniform mixture fit.
- **...**: Other graphic parameters for summary.

### Author(s)

Daniela Beisser

### See Also

`fitBumModel`, `print.bum`

### Examples

```r
data(pvaluesExample)
pvals <- pvaluesExample[,1]
mle <- fitBumModel(pvals, plot=FALSE)
summary(mle)
```
writeHeinz

Write input files for HEINZ

Description

Function to write the input files with the node and edge scores for HEINZ. These files are used to calculate the maximum scoring subnetwork of the graph. The node scores are matched by their names to the nodes of the network, therefore if nodes.scores are provided as a vector or matrix, the vector has to be named, respectively the matrix has to be provided with rownames. If the network contains more nodes than the score vector, the nodes without a score are scored with the average over all nodes. If the nodes should not be scored and used for the calculation of the maximum scoring subnetwork, draw a subnetwork (subNetwork) first and use this for the argument network. The edge scores can be provided as a vector or matrix as the edge.scores argument. If no scores are provided in the arguments, but the use.node.scores or use.edge.scores argument is set to TRUE, it will be automatically looked for the "score" attribute of the nodes and edges of the network.

Usage

writeHeinz(network, file, node.scores=0, edge.scores=0, use.node.score=FALSE, use.edge.score=FALSE)

Arguments

- network: Network from which to calculate the maximum scoring subnetwork.
- file: File to write to.
- node.scores: Numeric vector or matrix of scores for the nodes of the network. Names of the vector or rownames of the matrix have to correspond to the PPI identifiers of the network. The scores can also be used from the node attribute "score", given one score for each node.
- edge.scores: Numeric vector of scores for the edges of the network. Edge scores have to be given in the order of the edges in the network. It is better to append the edge scores as the edge attribute "score" to the network: V(network)$score and set use.scores to TRUE.
- use.node.score: Boolean value, whether to use the node attribute "score" in the network as node scores.
- use.edge.score: Boolean value, whether to use the edge attribute "score" in the network as edge scores.

Author(s)

Daniela Beisser

See Also

writeHeinzNodes and writeHeinzEdges
Examples

```r
library(DLBCL)
data(interactome)
data(dataLym)
chipGraph <- subNetwork(dataLym$label, interactome)
score <- dataLym$score001
names(score) <- dataLym$label
## Not run: writeHeinz(network=chipGraph, file="lymphoma_001", node.scores=score, edge.scores=0)
```

Description

Function to write an input file for HEINZ with edge scores. If no edge scores are used, they are set to 0. In order to run HEINZ, a node input and edge input file are needed.

Usage

```r
writeHeinzEdges(network, file, edge.scores=0, use.score=FALSE)
```

Arguments

- `network`: Network from which to calculate the maximum scoring subnetwork.
- `file`: File to write to.
- `edge.scores`: Numeric vector of scores for the edges of the network. Edge scores have to be given in the order of the edges in the network. It is better to append the edge scores as the edge attribute "score" to the network: `V(network)$score` and set `use.score` to TRUE.
- `use.score`: Boolean value, whether to use the edge attribute "score" in the network as edge scores.

Author(s)

Daniela Beisser

See Also

- `writeHeinzNodes` and `writeHeinz`
## Not run: writeHeinzEdges(network=graph, file="lymphoma_edges_001", use.score=FALSE)
score <- dataLym$score001
names(score) <- dataLym$label
## Not run: writeHeinzNodes(network=graph, file="lymphoma_nodes_001", node.scores=score)

# write another edge file with edge scores
library(igraph)
data(interactome)
interactome <- igraph.from.graphNEL(interactome)
small.net <- subNetwork(V(interactome)[1:16]$name, interactome)
scores <- c(1:length(E(small.net)))
E(small.net)$score <- scores
## Not run: writeHeinzEdges(network=small.net, file="test_edges", use.score=TRUE)

writeHeinzNodes  Write node input file for HEINZ

Description

Function to write an input file with the node scores for HEINZ. This file is used together with the edge input file to calculate the maximum scoring subnetwork of the graph. The scores are matched by their names to the nodes of the network, therefore if nodes.scores are provided as a vector or matrix, the vector has to be named, respectively the matrix has to be provided with rownames. If the network contains more nodes than the score vector, the nodes without a score are scored with the average over all nodes. If the nodes should not be scored and used for the calculation of the maximum scoring subnetwork, draw a subnetwork subNetwork first and use this for the argument network.

Usage

writeHeinzNodes(network, file, node.scores=0, use.score=FALSE)

Arguments

- **network**: Network from which to calculate the maximum scoring subnetwork.
- **file**: File to write to.
- **node.scores**: Numeric vector or matrix of scores for the nodes of the network. Names of the vector or rownames of the matrix have to correspond to the PPI identifiers of the network. The scores can also be used from the node attribute "score", given one score for each node.
- **use.score**: Boolean value, whether to use the node attribute "score" in the network as node scores.

Details

Use scoreNodes or scoreFunction to derive scores from a vector of p-values.

Author(s)

Daniela Beisser
See Also

`writeHeinzEdges` and `writeHeinz`.

Examples

```r
# create small network
library(DLBCL)
data(interactome)
small.net <- subNetwork(nodes(interactome)[0:15], interactome)
scores <- c(1:length(nodes(small.net)))
names(scores) <- nodes(small.net)
## Not run: writeHeinzNodes(network=small.net, file="test_nodes", node.scores=scores)

# use Lymphoma data and graph to find module
library(DLBCL)
data(interactome)
data(dataLym)
# get induced subnetwork for all genes contained on the chip
chipGraph <- subNetwork(dataLym$label, interactome)
## Not run: writeHeinzEdges(network=chipGraph, file="lymphoma_edges_001", use.score=FALSE)
score <- dataLym$score001
names(score) <- dataLym$label
## Not run: writeHeinzNodes(network=chipGraph, file="lymphoma_nodes_001", node.scores=score)
```
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