Package ‘BioNet’

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Description This package provides functions for the integrated
analysis of protein-protein interaction networks and the
detection of functional modules. Different datasets can be
integrated into the network by assigning p-values of
statistical tests to the nodes of the network. E.g. p-values
obtained from the differential expression of the genes from an
Affymetrix array are assigned to the nodes of the network. By
fitting a beta-uniform mixture model and calculating scores
from the p-values, overall scores of network regions can be
calculated and an integer linear programming algorithm
identifies the maximum scoring subnetwork.
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Description

This package provides functions for the integrated analysis of biological networks and the detection of functional modules. Different datasets can be integrated into the network by assigning p-values derived from statistical tests to the nodes of the network. E.g. p-values obtained from the differential expression of genes from an Affymetrix array are assigned to the nodes of a protein-protein interaction network. By fitting a beta-uniform mixture model and calculating scores from the p-values, overall scores of network regions can be calculated and an integer linear programming algorithm identifies the maximum scoring subnetwork.

Details

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LazyLoad: yes

Author(s)

Marcus Dittrich, Daniela Beisser
Maintainer: Marcus Dittrich <marcus.dittrich@biozentrum.uni-wuerzburg.de>

References


Description

The function aggregates several p-values into one p-value based on the order statistics of p-values. An overall p-value is given by the $i$th order statistic.

Usage

```r
aggrPvals(pval.matrix, order, plot=TRUE)
```
Arguments

- **pval.matrix**  Numeric matrix of p-values, columns represent different sets of p-values
- **order**  Numeric constant, the order statistic that is used for the aggregation.
- **plot**  Boolean value whether to plot p-value distributions.

Value

Aggregated p-value of the given order.

Author(s)

Daniela Beisser

Examples

```r
data(pvaluesExample)
aggrPvals(pval.matrix=pvaluesExample, order=2)
```

---

**Fitting a beta-uniform mixture model to p-value distribution**

**Description**

The function fits a beta-uniform mixture model to a given p-value distribution.

**Usage**

```r
bumOptim(x, starts=1, labels=NULL)
```

Arguments

- **x**  Numerical vector of p-values, has to be named with the gene names or the gene names can be given in the labels parameter.
- **starts**  Number of start points for the optimization.
- **labels**  Gene names for the p-values.

Value

List of class fb with the following elements:

- **lambda**  Fitted parameter for the beta-uniform mixture model.
- **a**  Fitted parameter for the beta-uniform mixture model.
- **negLL**  Negative log-likelihood.
- **pvalues**  P-value vector.

Author(s)

Marcus Dittrich and Daniela Beisser
References


See Also

`fitBumModel`, `plot.bum`, `hist.bum`

Examples

```r
data(pvaluesExample)
pvals <- pvaluesExample[,1]
bum <- bumOptim(x=pvals, starts=10)
bum
```

```r
compareNetworks

Compare parameters of two networks

Description

The function compares the following parameters of two networks: diameter, average degree, degree exponent, average path length and plots the cumulative degree distributions. The networks have to be connected components.

Usage

```r
compareNetworks(network1, network2, plot=TRUE)
```

Arguments

- `network1` Network `graphNEL` or `igraph` format.
- `network2` Second network in `graphNEL` or `igraph` format, or subnetwork drawn from first network.
- `plot` Boolean value, whether to plot the cumulative degree distributions.

Value

A vector of network parameters is returned:

- `diam.network1` Network diameter
- `diam.network2` Diameter of the subnetwork
- `av.degree.network1` Average degree of the network
- `av.degree.network2` Average degree of the subnetwork
consensusScores

degree.exponent.network1
Degree exponent of the network
degree.exponent.network2
Degree exponent of the subnetwork
av.path.length.network1
Average path length of the network
av.path.length.network2
Average path length of the subnetwork

Author(s)
Daniela Beisser

Examples
library(DLBCL)
data(interactome)
subnet1 <- largestComp(subNetwork(nodes(interactome)[1:100], interactome))
subnet2 <- largestComp(subNetwork(nodes(interactome)[101:200], interactome))
compareNetworks(network1=subnet1, network2=subnet2)

consensusScores Calculation of a consensus score for a network

Description
The function calculates consensus scores for a network, given a list of replicate modules.

Usage
consensusScores(modules, network, ro=length(modules)/2)

Arguments
modules Calculated modules from pseudo-replicates of expression values in igraph or
        graphNEL format.
network Interaction network, which should be scores. In igraph or graphNEL format
ro Threshold which is subtracted from the scores to obtain positive and negative
      value. The default value is half of the number of replicates.

Value
A result list is returned, consisting of:
N.scores Numerical vector node scores.
E.scores Numerical vector edge scores.
N.frequencies Numerical vector node frequencies from the replicate modules.
E.frequencies Numerical vector edge frequencies from the replicate modules.

Author(s)
Daniela Beisser
Examples

```r
library(DLBCL)
data(interactome)
network <- interactome
# precomputed Heinz modules from pseudo-replicates
## Not run: lib <- file.path(.path.package("BioNet"), "extdata")
modules <- readHeinzGraph(node.file=file.path(datadir, "ALL_n_resample.txt.0.hnz"), network=network)
cons.scores <- consensusScores(modules, network)
## End(Not run)
```

---

fbum

**Compute the density of the bum distribution**

### Description

Function to compute the density of the beta-uniform mixture model.

### Usage

```r
fbum(x, lambda, a)
```

### Arguments

- `x` A numeric value.
- `lambda` Parameter lambda, mixture parameter, proportion of uniform component
- `a` Parameter a, shape parameter of beta component

### Value

Value of the density of the bum distribution for x.

### Author(s)

Marcus Dittrich

### References


### See Also

`bumOptim`, `fitBumModel`

### Examples

```r
y <- fbum(x=0.5, lambda=0.1, a=0.1)
y
```
**fbumLL**  
*Calculate log likelihood of BUM model*

**Description**

The function calculates the log likelihood of the BUM model.

**Usage**

```r
fbumLL(parms, x)
```

**Arguments**

- `parms` Vector of parameters; lambda and a.
- `x` Numerical vector of p-values.

**Value**

Log likelihood.

**Author(s)**

Marcus Dittrich

**Examples**

```r
data(pvaluesExample)
pvals <- pvaluesExample[,1]
bum.mle <- fitBumModel(pvals, plot=FALSE)
fbumLL(parms=c(bum.mle$lambda, bum.mle$a), x=pvals)
```

---

**fdrThreshold**  
*Calculate p-value threshold for given FDR*

**Description**

The function calculates the p-value threshold tau for a given false discovery rate. Tau is used for the scoring function.

**Usage**

```r
fdrThreshold(fdr, fb)
```

**Arguments**

- `fdr` False discovery rate.
- `fb` Model from the beta-uniform mixture fitting.

**Value**

P-value threshold tau.
fitBumModel

Author(s)
Marcus Dittrich

References

See Also
fbum, fitBumModel

Examples
```r
data(pvaluesExample)
pvals <- pvaluesExample[,1]
bum.mle <- fitBumModel(pvals, plot=FALSE)
tau <- fdrThreshold(fdr=0.001, fb=bum.mle)
tau```

Description
The function fits a beta-uniform mixture model to a given p-value distribution. The BUM method was introduced by Stan Pounds and Steve Morris to model the p-value distribution as a signal-noise decomposition. The signal component is assumed to be B(a,1)-distributed, whereas the noise component is uniform-distributed under the null hypothesis.

Usage
```r
fitBumModel(x, plot = TRUE, starts=10)```

Arguments
- `x`: Numeric vector of p-values.
- `plot`: Boolean value, whether to plot a histogram and qqplot of the p-values with the fitted model.
- `starts`: Numeric value giving the number of starts for the optimization.

Value
Maximum likelihood estimator object for the fitted bum model. List of class fb with the following elements:
- `lambda`: Fitted parameter `lambda` for the beta-uniform mixture model.
- `a`: Fitted parameter `a` for the beta-uniform mixture model.
- `negLL`: Negative log-likelihood.
- `pvalues`: P-value vector.
getCompScores

Author(s)
Daniela Beisser

References

Examples
data(pvaluesExample)
pvals <- pvaluesExample[,1]
bum.mle <- fitBumModel(pvals, plot=TRUE)
bum.mle

getCompScores

Partition scores for subgraphs of the network

Description
The function partitions the scores into scores for each subgraph of the network.

Usage
getCompScores(network, score)

Arguments
network A network in graphNEL or igraph format.
score Vector of scores.

Value
A data frame with the components of the network and the score for each PPI identifier.

Author(s)
Marcus Dittrich

Examples
library(DLBCL)
data(interactome)
data(dataLym)
# create random subgraph with 100 nodes and their direct neighbors
nodes <- nodes(interactome)[sample(length(nodes(interactome)), 100)]
subnet <- subNetwork(nodeList=nodes, network=interactome, neighbors="first")
score <- dataLym$score001
names(score) <- dataLym$label
getCompScores(score=score, network=subnet)
getEdgeList

Get representation of graph as edgelist

Description

A network in graphNEL or igraph format is converted to an edgelist.

Usage

getEdgeList(network)

Arguments

network Network in graphNEL or igraph format.

Value

A matrix whose columns represent the connected edges.

Author(s)

Marcus Dittrich

Examples

library(DLBCL)
data(interactome)
getEdgeList(interactome)[1:10,]

hist.bum

Histogram of the p-value distribution with the fitted bum model

Description

The function plots a histogram of the p-values together with the fitted bum-model.

Usage

## S3 method for class 'bum'
hist(x, breaks=50, main="Histogram of p-values", xlab="P-values", ylab="Density", ...)

Arguments

x Maximum likelihood estimator object of the beta-uniform mixture fit.
breaks Breaks for the histogram.
main An overall title for the plot.
xlab A title for the x axis.
ylab A title for the y axis.
... Other graphic parameters for the plot.
Author(s)
Daniela Beisser

See Also
fitBumModel, hist.bum, bumOptim

Examples
```
data(pvaluesExample)
pvals <- pvaluesExample[,1]
mle <- fitBumModel(pvals, plot=FALSE)
hist(mle)
```

---

largestComp  Extract largest component of network

Description
The function extracts the largest component of a network.

Usage
```
largestComp(network)
```

Arguments

network  A graph in graphNEL or igraph format.

Value
A new graph object that represents the largest component of the given network.

Author(s)
Marcus Dittrich

Examples
```
library(DLBCL)
data(interactome)
interactome
largestComp(interactome)
```
largestScoreComp  

**Component with largest score**

**Description**

The function extracts the component of the network with the largest score. All nodes have to exceed the given level for the score.

**Usage**

```r
largestScoreComp(network, score, level=0)
```

**Arguments**

- `network`: Network in `graphNEL` or `igraph` format.
- `score`: Vector of scores for the network.
- `level`: Cut-off level for the score for the component.

**Value**

Subgraph of the network with a score larger than the given level.

**Author(s)**

Marcus Dittrich

**Examples**

```r
library(DLBCL)
data(interactome)
data(dataLym)
network <- rmSelfLoops(interactome)
score <- dataLym$score001
names(score) <- dataLym$label
lComp <- largestScoreComp(network=network, score=score, level=1)
## Not run: plotModule(lComp)
```

loadNetwork.sif  

**Load network from Cytoscape sif file**

**Description**

The function loads a network from a Cytoscape sif file. Edge attributes are provided in the ea.file or vector of ea.files. The node attributes are provided the same way. For other formats see `read.graph` in the igraph package.

**Usage**

```r
loadNetwork.sif(sif.file, na.file, ea.file, format=c("graphNEL", "igraph"), directed=FALSE)
```
loadNetwork.tab

Load network from tabular format

Description

The function loads a network from a tabular format.

Usage

loadNetwork.tab(file, header=TRUE, directed=FALSE, format=c("graphNEL", "igraph"))

Arguments

- file: File with network to load.
- header: Boolean value whether to include header or not.
- directed: Boolean value whether the network is to be directed or not.
- format: Output format of the network, either graphNEL or igraph

Author(s)

Marcus Dittrich

See Also

loadNetwork.sif
makeNetwork

Create graph from source and target vectors

Description

Function to create a graph in graphNEL or igraph format from a source and a target vector.

Usage

makeNetwork(source, target, edgemode="undirected", format=c("graphNEL", "igraph"))

Arguments

- source: Vector of source nodes.
- target: Vector of corresponding target nodes.
- edgemode: For an "undirected" or "directed" network.
- format: Graph format, either graphNEL or igraph.

Value

A graph object.

Author(s)

Marcus Dittrich

See Also

loadNetwork.sif, saveNetwork

Examples

source <- c("a", "b", "c", "d")
target <- c("b", "c", "a", "a")
graph <- makeNetwork(source, target, edgemode="undirected")

mapByVar

Select probeset by variance and get PPI ID

Description

The function selects for each gene the probeset with the highest variance and gets the PPI ID for each gene. The PPI identifier is: gene symbol(Entrez ID). Affymetrix identifiers are mapped to the ENTREZ ID.

Usage

mapByVar(exprSet, network=NULL, attr="geneID", ignoreAFFX=TRUE)
**Arguments**

- **exprSet** Affymetrix ExpressionSet.
- **network** Network that is used to map the Affymetrix identifiers.
- **attr** The attribute of the network that is used to map the Affymetrix IDs. The IDs are mapped to the unique Entrez gene IDs, which are by default stored in the "geneID" attribute of the network.
- **ignoreAFFX** Boolean value, whether to ignore or leave AFFX control genes.

**Value**

Expression matrix with one gene (PPI ID) per probeset.

**Author(s)**

Daniela Beisser

**Examples**

```r
## Not run: library(ALL);
data(ALL);
mapped.e.set <- mapByVar(ALL);
mapped.e.set[1:10,];
## End(Not run)
```

**Description**

Function to permutate node labels of a given network.

**Usage**

```r
permutateNodes(network)
```

**Arguments**

- **network** Network in graphNEL or igraph format.

**Value**

Network with permutated labels.

**Author(s)**

Marcus Dittrich
Examples

```r
library(DLBCL)
data(interactome)
# remove self-loops before permutating the labels
interactome <- rmSelfLoops(interactome)
perm.net <- permutateNodes(interactome)
perm.net
```

---

**piUpper**

*Upper bound pi for the fraction of noise*

Description

The function calculates the upper bound pi for the fraction of noise.

Usage

```r
piUpper(fb)
```

Arguments

- **fb**
  
  Fitted bum model, list with parameters a and lambda.

Value

Numerical value for the upper bound pi.

Author(s)

Marcus Dittrich

See Also

`bumOptim`, `fitBumModel`

Examples

```r
data(pvaluesExample)
pvals <- pvaluesExample[,1]
bum <- bumOptim(pvals, starts=10)
piUpper(fb=bum)
```
plot.bum  

Quantile-quantile plot for the beta-uniform mixture model

Description

The function plots the theoretical quantiles of the fitted bum model against the quantiles of the observed p-value distribution.

Usage

```r
## S3 method for class 'bum'
plot(x, main="QQ-Plot", xlab="Estimated p-value", ylab="Observed p-value", ...)
```

Arguments

- `x`: Maximum likelihood estimation object of the fitted bum model.
- `main`: An overall title for the plot.
- `xlab`: A title for the x axis.
- `ylab`: A title for the y axis.
- `...`: Other graphic parameters for the plot.

Author(s)

Daniela Beisser

See Also

- `fitBumModel`, `plot.bum`, `bumOptim`

Examples

```r
data(pvaluesExample)
pvals <- pvaluesExample[,1]
mle <- fitBumModel(pvals, plot=FALSE)
plot(mle)
```

plot3dModule  

3D plot of the network

Description

The function plots a network from `graphNEL` or `igraph` format in 3D using a modified function from the package igraph and requires the package rgl which uses OpenGL. The 3D plot can be zoomed, rotated, shifted on the canvas. This function is just used to visualize the modules. For further plotting options use the rglplot function of the igraph package. If a score attribute is provided in the graph this will be used for the coloring of the nodes. Otherwise a vector of values can be given by the `diff.or.score` argument. The vector has to contain positive and negative values, either scores or values for differential expression (fold changes). Labels for the nodes can be provided by the `labels` argument, otherwise it will be automatically looked for a `geneSymbol` attribute of the nodes.
plotLLSurface

Usage

plot3dModule(network, labels=NULL, windowSize = c(100,100,1500,1000), diff.or.scores=NULL, red=c("negative", "positive"), ...)

Arguments

- **network**: Network in `graphNEL` or `igraph` format.
- **labels**: Labels for the nodes of the network. Otherwise it will be automatically looked for a `geneSymbol` attribute of the nodes.
- **windowSize**: Numerical vector of size four to set the size of the rgl device.
- **diff.or.scores**: Named numerical vector of differential expression (fold changes) or scores of the nodes in the network. These will be used for node coloring. Otherwise a `score` attribute of the nodes will be automatically used.
- **red**: Either "negative" or "positive", to specify which values are to be colored red in the plot.
- **...**: Other graphic parameters for the plot.

Author(s)

Daniela Beisser

See Also

`save3dModule`, `plotModule`

Examples

```r
library(DLBCL)
data(interactome)
data(dataLym)
interactome <- subNetwork(dataLym$label, interactome)
interactome <- rmSelfLoops(interactome)
fchange <- dataLym$diff
names(fchange) <- dataLym$label
subnet <- largestComp(subNetwork(nodes(interactome)[1:100], interactome))
diff <- fchange[nodes(subnet)]

## Not run: library(rgl);
plot3dModule(network=subnet, diff.or.score=diff)
## End(Not run)
```

---

plotLLSurface  

Log likelihood surface plot

Description

The function plots the log likelihood surface for all $a$ and lambda parameter of the beta-uniform mixture model.

Usage

plotLLSurface(x, opt=NULL, main="Log-Likelihood Surface", color.palette = heat.colors, nlevels = 32)
Arguments

- **x**: Numeric vector of p-values.
- **opt**: List of optimal parameters for a and lambda from the beta-uniform mixture model.
- **main**: The overall title of the plot.
- **color.palette**: Color scheme of the image plot.
- **nlevels**: Number of color levels.

Author(s)

Marcus Dittrich

Examples

```r
library(DLBCL)
data(dataLym)
pvals <- dataLym$t.pval
names(pvals) <- dataLym$label
mle <- fitBumModel(pvals, plot=FALSE)
plotLLSurface(x=pvals, opt=mle)
```

Description

The function plots a network from `graphNEL` or `igraph` format, adapted from an igraph plotting function. It is just used to visualize the modules. For further plotting options use the `plot.igraph` function of the igraph package. The shapes of the nodes can be changed according to the scores argument, then negative scores appear squared. The color of the nodes can be changed according to the `diff.expr` argument. Negative values lead to green nodes, positive values are colored in red. If the vectors are not provided, it will be automatically looked for nodes attributes with the name `score` and `diff.expr`.

Usage

```r
plotModule(network, layout=layout.fruchterman.reingold, labels=NULL, diff.expr=NULL, scores=NULL,)
```

Arguments

- **network**: Network in `graphNEL` or `igraph` format.
- **layout**: Layout algorithm, e.g. `layout.fruchterman.reingold` or `layout.kamada.kawai`.
- **labels**: Labels for the nodes of the network.
- **diff.expr**: Named numerical vector of differential expression (fold changes) of the nodes in the network. These will be used for coloring of the nodes. It will be automatically looked for nodes attribute with the name `diff.expr`, if the argument is null.
- **scores**: Named numerical vector of scores of the nodes in the network. These will be used for the shape of the nodes. It will be automatically looked for nodes attribute with the name `score`, if the argument is null.
**print.bum**

Main title of the plot.

**vertex.size**

Numerical value or vector for the size of the vertices.

... Other graphic parameters for the plot.

**Author(s)**

Marcus Dittrich and Daniela Beisser

**See Also**

plot3dModule

**Examples**

```r
library(DLBCL)
data(dataLym)
data(interactome)
interactome <- subNetwork(dataLym$label, interactome)
interactome <- rmSelfLoops(interactome)
fchange <- dataLym$diff
names(fchange) <- dataLym$label
# create random subnetwork
subnet <- largestComp(subNetwork(nodes(interactome)[1:100], interactome))
fchange <- fchange[nodes(subnet)]

# color random subnetwork by the fold change
## Not run: plotModule(network=subnet, diff.expr=fchange)
```

---

**print.bum**

Print information about bum model

**Description**

The function prints information about the bum model.

**Usage**

```r
## S3 method for class 'bum'
print(x, ...)
```

**Arguments**

**x**

Maximum likelihood estimator object of the beta-uniform mixture fit.

**...**

Other graphic parameters for print.

**Author(s)**

Marcus Dittrich

**See Also**

fitBumModel, summary.bum
data(pvaluesExample)
pvals <- pvaluesExample[,1]
mle <- fitBumModel(pvals, plot=FALSE)
print(mle)

pvaluesExample

Example p-values for aggregation statistics

Description
Data example consisting of a matrix of p-values. Each gene has two corresponding p-values. These p-values can be aggregated into a p-value of p-values by the method \texttt{aggrPvals}.

Usage
data(pvaluesExample)

Examples
data(pvaluesExample)
pvaluesExample[1:10,]

readHeinzGraph

Convert HEINZ output to graph

Description
Function to convert the HEINZ output to a graph object, or if the output is in matrix form, it will create a list of graphs. The function needs the node and the original network, from which the module is calculated.

Usage
readHeinzGraph(node.file, network, format=c("graphNEL", "igraph"))

Arguments
- \texttt{node.file} Heinz node output file.
- \texttt{network} Original network from which Heinz input was created.
- \texttt{format} Graph format of output, either \texttt{igraph} or \texttt{graphNEL}.

Value
Graph object.

Author(s)
Daniela Beisser
Examples

```r
library(DLBCL)
data(interactome)
# precomputed Heinz output files
## Not run: lib <- file.path(.path.package("BioNet"), "extdata")
module <- readHeinzGraph(node.file=file.path(lib, "lymphoma_nodes_001.txt.0.hnz"), network=interactome, format="graphNEL");
plotModule(module);
## End(Not run)
```

readHeinzTree

Convert HEINZ output to tree

Description

Converts the HEINZ output to a tree in graph format. If the output is in matrix form, it will create a list of graphs. The function needs the node and edge file and the original network from which the module is calculated.

Usage

```r
readHeinzTree(node.file, edge.file, network, format=c("graphNEL", "igraph"))
```

Arguments

- `node.file`: Heinz node output file.
- `edge.file`: Heinz edge output file.
- `network`: Original network from which Heinz input was created.
- `format`: Output format of the graph, either igraph or graphNEL.

Value

A graph object.

Author(s)

Daniela Beisser

Examples

```r
library(DLBCL)
data(interactome)
# precomputed Heinz output files
## Not run: lib <- file.path(.path.package("BioNet"), "extdata")
module <- readHeinzTree(node.file=file.path(lib, "lymphoma_nodes_001.txt.0.hnz"), edge.file=file.path(lib, "lymphoma_edges_001.txt.0.hnz"), network=interactome, format="graphNEL");
plotModule(module);
## End(Not run)
```
Resampling of microarray expression values and test for differential expression.

Description

The function uses a 50% jackknife resampling to calculate a pseudo-replicate of the expression matrix. The resampled expression values are used thereupon to calculate p-values for the differential expression between the given groups. Only two-group comparisons are allowed for the performed t-test.

Usage

resamplingPvalues(exprMat, groups, alternative = c("two.sided", "less", "greater"), resampleMat=FALSE)

Arguments

exprMat  Matrix with microarray expression values.
groups   Factors for two groups that are tested for differential expression.
alternative Testing alternatives for the t-test: "two.sided", "less" or "greater".
resampleMat Boolean value, whether to retrieve the matrix of jackknife resamples or not.

Value

A result list is returned, consisting of:

p.values VNumerical vector of p-values.
resampleMat   Matrix of resampled expression values.

Author(s)

Daniela Beisser

Examples

library(ALL)
data(ALL)
mat <- exprs(ALL)
groups <- factor(c(rep("A", 64), rep("B", 64)))
results <- resamplingPvalues(mat, groups, alternative="greater")
**rmSelfLoops**

**Remove self-loops in a graph**

**Description**

The function removes self-loops, edges that start and end in the same node, from the network.

**Usage**

```r
rmSelfLoops(network)
```

**Arguments**

- `network`: A graph object, either in `graphNEL` or `igraph` format.

**Value**

The graph with the removed edges.

**Author(s)**

Marcus Dittrich

**Examples**

```r
graph <- makeNetwork(c("a", "b", "c", "d", "e", "a"), c("b", "c", "d", "e", "e", "e"))
graph2 <- rmSelfLoops(graph)
edges(graph)
edges(graph2)
```

---

**runFastHeinz**

**Calculate heuristically maximum scoring subnetwork**

**Description**

The function uses an heuristic approach to calculate the maximum scoring subnetwork. Based on the given network and scores the positive nodes are in the first step aggregated to meta-nodes between which minimum spanning trees are calculated. In regard to this, shortest paths yield the approximated maximum scoring subnetwork. This function can be used if a CPLEX license is not available to calculate the optimal solution.

**Usage**

```r
runFastHeinz(network, scores)
```

**Arguments**

- `network`: A graph in `igraph` or `graphNEL` format.
- `scores`: A named vector, containing the scores for the nodes of the network. All nodes need to be scored in order to run the algorithm.
runHeinz

Start HEINZ

Value

A subnetwork in the input network format.

Author(s)

Daniela Beisser

See Also

writeHeinzEdges, writeHeinzNodes, readHeinzTree, readHeinzGraph, runHeinz

Examples

library(DLBCL)
# load p-values
data(dataLym)
# load graph
data(interactome)
# get induced subnetwork for all genes contained on the chip
interactome <- subNetwork(dataLym$label, interactome)
p.values <- dataLym$t.pval
names(p.values) <- dataLym$label
bum <- fitBumModel(p.values, plot=TRUE)
scores <- scoreNodes(network=interactome, fb=bum, fdr=0.0001)
module <- runFastHeinz(network=interactome, scores=scores)
## Not run: plotModule(module)

Description

The function starts HEINZ from command line. The HEINZ folder has to include the heinz.py python script and the dhea file. CPLEX has to be installed and accessible from the computer R runs on.

Usage

runHeinz(heinz.folder="", heinz.e.file, heinz.n.file, N=TRUE, E=FALSE, diff=-1, n=1)

Arguments

heinz.folder The folder which contains the heinz.py python script and the dhea file.
heinz.e.file The HEINZ edge input file. See writeHeinzEdges
heinz.n.file The HEINZ node input file. See writeHeinzNodes
N Boolean value, whether to run HEINZ on nodes.
E Boolean value, whether to run HEINZ on edges. HEINZ can run on both with N and E set to TRUE.
diff Difference of suboptimal solutions to optimal solution in haming distance in percent. Parameter is set to -1 for optimal solution.
n Number of optimal and suboptimal solutions, the standard n=1 delivers only the optimal solution.
Details

This function starts the integer linear programming algorithm to calculate the optimal scoring sub-network. The algorithm might be started in the command line when the CPLEX is installed on another machine. To start it from command line use: heinz.py -e edge.file.txt -n node.file.txt -E False/True -N False/True. The results can be loaded with readHeinzTree, readHeinzGraph as a graph object.

Author(s)

Daniela Beisser

References


See Also

writeHeinzEdges, writeHeinzNodes, readHeinzTree, readHeinzGraph

Description

The function saves a 3D plot of a network to file, therefore it requires the plot to be open. A screenshot of the 3D plot can be saved in "pdf" format. Background of the device is changed to white for plotting. The screenshot can take several seconds for large plots.

Usage

save3dModule(file)

Arguments

file File to save to.

Author(s)

Daniela Beisser

See Also

plot3dModule, plotModule
Examples

```r
library(DLBCL)
data(dataLym)
data(interactome)
interactome <- subNetwork(dataLym$label, interactome)
fchange <- dataLym$diff
names(fchange) <- dataLym$label
subnet <- largestComp(subNetwork(nodes(interactome)[1:100], interactome))
diff <- fchange[nodes(subnet)]
## Not run: library(rgl);
plot3dModule(network=subnet, diff.or.score=diff);
save3dModule(file="test")
## End(Not run)
```

### saveNetwork

#### Description

The function saves a graph in a Cytoscape readable format: either in XGMML format, or as two tables, one for the nodes with attributes and one for the edges with attributes, or as .sif file. Or other standard formats like tab separated, .tgf, .net

#### Usage

```r
saveNetwork(network, name="network", file, type=c("table", "XGMML", "sif", "tab", "tgf", "net"))
```

#### Arguments

- `network`: Network to save.
- `name`: Name of the network, only needed for the XGMML format.
- `file`: File to save to.
- `type`: Type in which graph shall be saved.

#### Details

The format types are "XGMML", "table", "sif", "tab", "tgf" and "net". XGMML (eXtensible Graph Markup and Modeling Language) is an XML format based on GML which is used for graph description. Edges, nodes and their affiliated attributes are all saved in one file. In the table format two tables are created, one for the nodes with attributes and one for the edges with attributes. The sif format creates a .sif file for the network and an node attribute (.NA) or edge attribute (.EA) for each attribute. The name of the attribute is the filename. Tab writes only the edges of the network in a tabular format. Tgf save the network to simple .tgf format. The net format writes a Pajek readable file of the network and the ET type saves the edge tags to file.

#### Author(s)

Daniela Beisser and Marcus Dittrich
Examples

library(DLBCL)
# create small network
library(igraph)
data(interactome)
interactome <- igraph.from.graphNEL(interactome)
small.net <- subNetwork(V(interactome)[1:16]$name, interactome)
E(small.net)$e.weight <- rep(1,length(E(small.net)))
V(small.net)$n.weight <- rep(2,length(V(small.net)))
summary(small.net)
## Not run: saveNetwork(small.net, file="example_network", name="small.net", type="XGMML")

scanFDR
Dataframe of scores over a given range of FDRs

Description

The function generates a dataframe for a given range of FDRs.

Usage

scanFDR(fb, fdr, labels=names(fb$pvalues))

Arguments

fb Fitted bum model.
fdr Vector of FDRs.
labels Data frame labels.

Value

Dataframe of scores for given p-values and a range of FDRs.

Author(s)

Marcus Dittrich

See Also

bumOptim, fitBumModel

Examples

data(pvaluesExample)
pvals <- pvaluesExample[,1]
bum <- bumOptim(pvals, starts=10)
scores <- scanFDR(fb=bum, fdr=c(0.1, 0.001, 0.0001))
scores[1:10,]
scoreFunction

Scoring function for p-values

Description

The function calculates a score for each gene with a given FDR from the fitted beta-uniform mixture model.

Usage

scoreFunction(fb, fdr=0.01)

Arguments

- fb: Model from the beta-uniform mixture fitting.
- fdr: Numeric constant, from the false discovery rate a p-value threshold is calculated. P-values below this threshold are considered to be significant and will score positively, p-values above the threshold are supposed to arise from the null model. The FDR can be used to control the size of the maximum scoring subnetwork, by zooming in and out in the same region.

Value

Score vector for the given p-values.

Author(s)

Marcus Dittrich and Daniela Beisser

References


Examples

data(pvaluesExample)
pvals <- pvaluesExample[,1]
bum.mle <- fitBumModel(pvals, plot=FALSE)
scores <- scoreFunction(fdr=0.1, fb=bum.mle)
scores
**scoreNodes**

**Score the nodes of a network**

**Description**

The function derives scores from the p-values of the nodes of a network.

**Usage**

```r
scoreNodes(network, fb, fdr=0.05)
```

**Arguments**

- `network`: A network in `graphNEL` or `igraph` format.
- `fb`: Fitted bum model.
- `fdr`: False discovery rate.

**Value**

Ordered score vector for the nodes of the network.

**Author(s)**

Marcus Dittrich

**See Also**

`bumOptim`, `fitBumModel`

**Examples**

```r
library(DLBCL)
# load p-values
data(dataLym)
# load graph
data(interactome)
# get induced subnetwork for all genes contained on the chip
chipGraph <- subNetwork(dataLym$label, interactome)
p.values <- dataLym$t.pval
names(p.values) <- dataLym$label
bum <- fitBumModel(p.values, plot=TRUE)
scoreNodes(network=chipGraph, fb=bum, fdr=0.001)
```
scoreOffset  

**Change score offset for 2 FDRs**

**Description**

Function to change score offset from FDR1 to FDR2.

**Usage**

```
scoreOffset(fb, fdr1, fdr2)
```

**Arguments**

- `fb`  
  Model from the beta-uniform mixture fitting.
- `fdr1`  
  First false discovery rate.
- `fdr2`  
  Second false discovery rate.

**Value**

Offset for the score of the second FDR.

**Author(s)**

Marcus Dittrich

**See Also**

`bumOptim`, `fitBumModel`

**Examples**

```
data(pvaluesExample)
pvals <- pvaluesExample[,1]
bum <- bumOptim(pvals, starts=10)
scoreOffset(bum, fdr1=0.001, fdr2=0.000001)
```

sortedEdgeList  

**Get a sorted edgelist**

**Description**

Function to get a sorted edgelist where the source protein is alphabetically smaller than the target protein from an undirected network.

**Usage**

```
sortedEdgeList(network)
```

**Arguments**

- `network`  
  Undirected network in `igraph` or `graphNEL` format.
Value

Vector of sorted edges, where the source protein is alphabetically smaller than the target protein.

Author(s)

Daniela Beisser

Examples

library(DLBCL)
data(interactome)
E.list <- sortedEdgeList(interactome)

subNetwork

Create a subGraph

Description

The function creates a subgraph with the nodes given in the nodeList or for these nodes including their direct neighbors.

Usage

subNetwork(nodeList, network, neighbors=c("none", "first"))

Arguments

nodeList Character vector of nodes, contained in the subgraph.
network Graph that is used for subgraph extraction.
neighbors Neighborhood, that is chosen for the subgraph extraction. "none" are only the selected nodes, "first" includes the direct neighbors of the selected nodes.

Value

A graph object.

Author(s)

Marcus Dittrich

Examples

library(igraph)
el <- cbind(c("a", "b", "c", "d", "e", "f", "d"), c("b", "c", "d", "e", "f", "a", "b"))
graph <- graph.edgelist(el, directed=TRUE)
node.list <- c("a", "b", "c")
graph2 <- subNetwork(nodeList=node.list, network=graph)
## Not run: par(mfrow=c(1,2));
plotModule(graph);
plotModule(graph2)
## End(Not run)
# or in graphNEL format:
graph3 <- igraph.to.graphNEL(graph)
graph4 <- subNetwork(nodeList=node.list, network=graph3)
graph3
graph4

## S3 method for class 'bum'
summary(object, ...)

object

... 

## S3 method for class 'bum'
summary(object, ...)

### Arguments

object  Maximum likelihood estimator object of the beta-uniform mixture fit.

... Other graphic parameters for summary.

### Author(s)

Daniela Beisser

### See Also

fitBumModel, print.bum

### Examples

data(pvaluesExample)
pvals <- pvaluesExample[,1]
mle <- fitBumModel(pvals, plot=FALSE)
summary(mle)
writeHeinz

Write input files for HEINZ

Description

Function to write the input files with the node and edge scores for HEINZ. These files are used to calculate the maximum scoring subnetwork of the graph. The node scores are matched by their names to the nodes of the network, therefore if nodes.scores are provided as a vector or matrix, the vector has to be named, respectively the matrix has to be provided with rownames. If the network contains more nodes than the score vector, the nodes without a score are scored with the average over all nodes. If the nodes should not be scored and used for the calculation of the maximum scoring subnetwork, draw a subnetwork (subNetwork) first and use this for the argument network. The edge scores can be provided as a vector or matrix as the edge.scores argument. If no scores are provided in the arguments, but the use.node.scores or use.edge.scores argument is set to TRUE, it will be automatically looked for the "score" attribute of the nodes and edges of the network.

Usage

writeHeinz(network, file, node.scores=0, edge.scores=0, use.node.score=FALSE, use.edge.score=FALSE)

Arguments

network Network from which to calculate the maximum scoring subnetwork.
file File to write to.
node.scores Numeric vector or matrix of scores for the nodes of the network. Names of the vector or rownames of the matrix have to correspond to the PPI identifiers of the network. The scores can also be used from the node attribute "score", given one score for each node.
edge.scores Numeric vector of scores for the edges of the network. Edge scores have to be given in the order of the edges in the network. It is better to append the edge scores as the edge attribute "score" to the network: V(network)$score and set use.scores to TRUE.
use.node.score Boolean value, whether to use the node attribute "score" in the network as node scores.
use.edge.score Boolean value, whether to use the edge attribute "score" in the network as edge scores.

Author(s)

Daniela Beisser

See Also

writeHeinzNodes and writeHeinzEdges
Examples

```r
code
```

Description

Function to write an input file for HEINZ with edge scores. If no edge scores are used, they are set to 0. In order to run HEINZ, a node input and edge input file are needed.

Usage

```r
writeHeinzEdges(network, file, edge.scores=0, use.score=FALSE)
```

Arguments

- `network`: Network from which to calculate the maximum scoring subnetwork.
- `file`: File to write to.
- `edge.scores`: Numeric vector of scores for the edges of the network. Edge scores have to be given in the order of the edges in the network. It is better to append the edge scores as the edge attribute “score” to the network: `V(network)$score` and set `use.score` to TRUE.
- `use.score`: Boolean value, whether to use the edge attribute "score" in the network as edge scores.

Author(s)

Daniela Beisser

See Also

- `writeHeinzNodes` and `writeHeinz`
writeHeinzNodes

## Not run: writeHeinzEdges(network=graph, file="lymphoma_edges_001", use.score=FALSE)
score <- dataLym$score001
names(score) <- dataLym$label
## Not run: writeHeinzNodes(network=graph, file="lymphoma_nodes_001", node.scores=score)

# write another edge file with edge scores
library(igraph)
data(interactome)
interactome <- igraph.from.graphNEL(interactome)
small.net <- subNetwork(V(interactome)[1:16]$name, interactome)
scores <- c(1:length(E(small.net)))
E(small.net)$score <- scores
## Not run: writeHeinzEdges(network=small.net, file="test_edges", use.score=TRUE)

---

writeHeinzNodes  Write node input file for HEINZ

### Description

Function to write an input file with the node scores for HEINZ. This file is used together with the edge input file to calculate the maximum scoring subnetwork of the graph. The scores are matched by their names to the nodes of the network, therefore if nodes.scores are provided as a vector or matrix, the vector has to be named, respectively the matrix has to be provided with rownames. If the network contains more nodes than the score vector, the nodes without a score are scored with the average over all nodes. If the nodes should not be scored and used for the calculation of the maximum scoring subnetwork, draw a subnetwork `subNetwork` first and use this for the argument `network`.

### Usage

```r
writeHeinzNodes(network, file, node.scores=0, use.score=FALSE)
```

### Arguments

- **network**: Network from which to calculate the maximum scoring subnetwork.
- **file**: File to write to.
- **node.scores**: Numeric vector or matrix of scores for the nodes of the network. Names of the vector or rownames of the matrix have to correspond to the PPI identifiers of the network. The scores can also be used from the node attribute "score", given one score for each node.
- **use.score**: Boolean value, whether to use the node attribute "score" in the network as node scores.

### Details

Use `scoreNodes` or `scoreFunction` to derive scores from a vector of p-values.

### Author(s)

Daniela Beisser
See Also

`writeHeinzEdges` and `writeHeinz`.

Examples

```r
# create small network
library(DLBCL)
data(interactome)
small.net <- subNetwork(nodes(interactome)[0:15], interactome)
scores <- c(1:length(nodes(small.net)))
names(scores) <- nodes(small.net)
## Not run: writeHeinzNodes(network=small.net, file="test_nodes", node.scores=scores)

# use Lymphoma data and graph to find module
library(DLBCL)
data(interactome)
data(dataLym)
# get induced subnetwork for all genes contained on the chip
chipGraph <- subNetwork(dataLym$label, interactome)
## Not run: writeHeinzEdges(network=chipGraph, file="lymphoma_edges_001", use.score=FALSE)
score <- dataLym$score001
names(score) <- dataLym$label
## Not run: writeHeinzNodes(network=chipGraph, file="lymphoma_nodes_001", node.scores=score)
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