Package ‘Clonality’

March 28, 2017

Type Package

Title Clonality testing

Version 1.22.0

Date 2012-20-04

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Depends R (>= 2.12.2), DNAcopy

Imports DNAcopy, grDevices, graphics, stats, utils

Suggests gdata, DNAcopy

Description Statistical tests for clonality versus independence of tumors from the same patient based on their LOH or genomewide copy number profiles

License GPL-3

LazyLoad yes

biocViews Microarray, CopyNumberVariation, Classification, aCGH

NeedsCompilation no

R topics documented:

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Clonality-package

**Clonality testing**

**Description**

Statistical tests for clonality versus independence of tumors from the same patient based on their LOH or genomewide copy number profiles.

**Details**

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**Author(s)**

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ave.adj.probes

**Averaging of adjacent probes in copy number arrays**

**Description**

For each sample the log-ratios at each consecutive K number of probes are averaged.

**Usage**

ave.adj.probes(data, K)

**Arguments**

<table>
<thead>
<tr>
<th>data</th>
<th>Copy Number Array object (output of function CNA() from the package DNA-copy). First column contains chromosomes, second column contains genomic locations. Each remaining column contains log-ratios from a particular tumor or sample.</th>
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<td>K</td>
<td>Number of markers to be averaged. Should be selected so that the final resolution of the averaged data would be 5,000-10,000 markers.</td>
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**Details**

Averages log-ratios in every K consecutive markers. The purpose of this step is to reduce the noise in the data, eliminate possible very small germline copy number variations, and get rid of a possible wave effect.
Value

Returns CNA object of reduced resolution

Examples

# Same example as in clonality.analysis()

set.seed(100)
chrom<rep(c(1:22),each=100)
maploc< runif(2200)* 200000
chromarm<splitChromosomes(chrom,maploc)

#Simulate the dataset with 10 pairs of tumors with 22 chromosomes, 100 markers each
#Simulated log-ratios are equal to signal + noise
#Signal: each chromosome has 50% chance to be normal, 30% to be whole-arm loss/gain, and 20% to be partial arm loss/gain
#There are no chromosomes with recurrent losses/gains
#Noise: drawn from normal distribution with mean 0, standard deviation 0.25
#First 9 patients have independent tumors, last patient has two tumors with identical signal, independent noise

set.seed(100)
chrom<paste("chr",rep(c(1:22),each=100),"p",sep="")
chrom[nchar(chrom)==5]<-paste("chr0",substr(chrom[nchar(chrom)==5] ,4,5),sep="")
maploc<- rep(c(1:100),22)
data<NULL
for (pt in 1:9) #first 9 patients have independent tumors
{
    tumor1<-tumor2<- NULL
    mean1<- rnorm(22)
    mean2<- rnorm(22)
    for (chr in 1:22)
    {
        r<runif(2)
        if (r[1]<0.5) tumor1<-c(tumor1,rep(0,100))
        else if (r[1]>0.7) tumor1<-c(tumor1,rep(mean1[chr],100))
        else { i<sort(sample(1:100,2))
            tumor1<-c(tumor1,mean1[chr]*c(rep(0, i[1]),rep(1, i[2]-i[1]), rep(0, 100-i[2])))
        }
        if (r[2]<0.5) tumor2<-c(tumor2,rep(0,100))
        else if (r[2]>0.7) tumor2<-c(tumor2,rep(mean2[chr],100))
        else { i<sort(sample(1:100,2))
            tumor2<-c(tumor2,mean2[chr]*c(rep(0, i[1]),rep(1, i[2]-i[1]), rep(0, 100-i[2])))
        }
    }
    data<cbind(data,tumor1,tumor2)
}
#last patient has identical profiles
    tumor1<- NULL
    mean1<- rnorm(22)
    for (chr in 1:22)
    {
        r<runif(1)
        if (r<0.4) tumor1<-c(tumor1,rep(0,100))
        else if (r>0.6) tumor1<-c(tumor1,rep(mean1[chr],100))
    }
else {  
  i<-sort(sample(1:100,2))
  tumor1<-c(tumor1,mean1[chr]*c(rep(0, i[1]),rep(1, i[2]-i[1]), rep(0, 100-i[2])))
}
}
data<-cbind(data,tumor1,tumor1)

data<-data+matrix(rnorm( 44000,mean=0,sd=0.4) ,nrow=2200,ncol=20)
dataCNA<-CNA(data,chrom=chrom,maploc=maploc,sampleid=paste("pt",rep(1:10,each=2),rep(1:2,10)))
dim(dataCNA)
dataCNA2<-ave.adj.probes(dataCNA, 2)
dim(dataCNA2)

---

**chromosomePlots**

Per-chromosome plots of the copy number arrays from a particular patient

**Description**

The function produces a sequence of plots for each chromosome with one-step segmented data of all samples of a particular patient.

**Usage**

`chromosomePlots(data.seg1, ptlist, ptname,nmad)`

**Arguments**

- `data.seg1`: Output of one-step segmentation - output OneStepSeg of clonality.analysis().
- `ptlist`: Vector of the patient IDs in the order of the samples appearing in the data. For example, if the first three tumors belong to patient A, and the following two belong to patient B, then `ptlist=c('ptA', 'ptA', 'ptA', 'ptB', 'ptB')`.
- `ptname`: Name of the patient from ptlist for which the data should be plotted.
- `nmad`: Number of MADs (median absolute deviations) that is used for Gain/Loss calls. Used to mark the Gain/Loss threshold on the plots.

**Details**

The function produces a sequence of plots for each chromosome with one-step segmented data of all samples of a particular patient. The dotted horizontal lines denote the gain and loss thresholds.

**Examples**

```r
# See example as in clonality.analysis()
```
Description

Function to test clonality of two tumors from the same patient based on their genomewide copy number profiles. This function calculates likelihood ratios and the reference distribution under the hypothesis of independence.

Usage

```r
clonality.analysis(data, ptlist, pfreq = NULL, refdata = NULL, nmad = 1.25, reference = TRUE, allpairs = TRUE, segmethod = "oneseg", segpar = NULL)
```

Arguments

- **data**: Copy Number Array object (output of function CNA() from package DNAcopy). First column contains chromosomes, second column contains genomic locations. Each remaining column contains log-ratios from a particular tumor or sample. Chromosomes X and Y should be removed prior to analysis, and chromosomes should be split into p and q arms to improve the power (use function splitChromosomes()).
- **ptlist**: Vector of the patient IDs in the order of the samples appearing in the data. For example, if the first three tumors (columns 3, 4, 5 of data) belong to patient A, and the following two (columns 6, 7 of data) belong to patient B, then `ptlist = c('ptA', 'ptA', 'ptA', 'ptB', 'ptB')`. Note that while sample names in data should be unique the ptlist should have repeated labels.
- **pfreq**: Marginal frequencies of Gains, Losses and Normals for all the chromosomes. If it is not known, pfreq should be set to NULL and frequencies will be estimated from all the samples in the dataset. If frequencies are known, pfreq should be a data frame with 4 columns: 1) chromosome arm in the format ‘chr01p’, probability of 2) gain, 3) loss and 4) normal.
- **refdata**: If available, additional cohort of patients with the same disease that should be used to estimate the marginal gain/loss frequencies. If NULL, the original set of tumors is used, otherwise, refdata should be a CNA object. It will be segmented with 1 step CBS and each chromosome will be classified as gain/loss as described in the manuscript, leading to frequency estimates. No averaging or chromosome splitting is done for this dataset, so users should make sure refdata has chromosomes in the format ‘chr01p’ and that its resolution is similar to the one of the original data.
- **nmad**: Number of MADs (median absolute deviations) that is used for Gain/Loss calls. For each array MAD of its residuals (that is, data minus segmentation means) is calculated. Residuals represent the array’s noise revel. Any segment of this array that has a mean at least nmad MADs above or below array’s median is called a gain or a loss. We use value of 1.25, while values in the range of 0.5 to 2 can also be admissible depending on the resolution and presence of artifacts.
- **reference**: If TRUE the reference distribution of likelihood ratios is created under hypothesis of independence by pairing (independent) tumors from different patients.
allpairs  If TRUE all possible pairs of tumors from different patients will be used for reference distribution. If two tumors in a pair are not exchangeable, for example primary tumor vs recurrence, or pre-cancerous lesion vs tumor, then allpairs should be set to FALSE and the 'first' tumor should always come earlier in the data before the 'second' tumor for all the patients. Then 'first' tumors of patients will only be paired with 'second' tumors of other patients for the reference distribution.

segmethod  The segmentation algorithm to be used. The default is "oneseg" which uses the built in function of the same name based on the CBS algorithm. An alternative segmentation algorithm can be used. A function should be created and the name passed as described in the vignette.

segpar  The parameters necessary for the segmentation algorithm as a list. For "oneseg" you can specify alpha (default = 0.01) and nperm (default = 2000) necessary for the CBS algorithm.

Details

The function implements the statistical procedure designed to distinguish whether the two tumors from the same patient are clonal (have the same progenitor cancer cell) or independent (developed from normal cells independently). At first data are segmented with one step CBS (Olshen, A. B., Venkatraman, E. S., Lucito, R., Wigler, M. (2004). Circular binary segmentation for the analysis of array-based DNA copy number data. Biostatistics 5: 557-572) that picks at most one copy number change per chromosome arm. Then each chromosome arm is classified as Gain/Loss/Normal based on a middle segment if there are 3 segments, or based on the most outstanding segment if there are 2 segments. The multinomial likelihood ratio comparing these classifications is computed (LR1). For each concordant partial arm gain or loss we also calculate likelihood ratio that this change is exactly the same in both tumors. These likelihood ratios are multiplied by LR1 to obtain our final statistic, LR2. If LR2 is much greater than 1, that indicates clonality. If LR2 is much smaller than 1, it indicates independence. The reference distribution of LR2 under the hypothesis of independence is obtained by pairing up tumors from different patients, which are independent by default.

Since only one gain/loss is admissible per chromosome arm it is highly recommended to apply this methodology to arrays with at most 10,000-15,000 markers. We suggest averaging blocks of consecutive probes for arrays with larger resolution, see function ave.adj.probes.

Value

If the reference is TRUE, function returns the list with 4 elements: LR, OneStepSeg, ChromClass, refLR.

LR - matrix with the within patient comparisons. Each row corresponds to a pair of samples being compared. Columns are: Sample1 - name of sample 1; Sample2 - name of sample 2; LR1 - likelihood ratio without comparisons of specific concordant gains/losses; LR2 - final likelihood ratio with individual comparisons; GGorLL - number of chromosome arms that are classified as Gains in both tumors or Losses in both tumors; NN - number of chromosome arms that are classified as Normal in both tumors; GL - number of chromosome arms that are classified as Gain in one tumors and Loss in another; GNorLN - number of chromosome arms that are classified as Gain(Loss) in one tumors and Normal in another; IndividualComparisons - list of chromosome arms that had comparisons of specific concordant gains/losses in both tumors and the corresponding likelihood ratio for them being exactly the same. p-value - quantile of the reference distribution under the null hypothesis (refLR$LR2) that the value of LR2 match.

OneStepSeg - is the output of one step segmentation of the data. It has the same structure as the output of 'segment' from DNAcopy, but only one most prominent change per arm is allowed.
ChromClass - is the matrix of chromosome classifications based on the one step segmentation. Rows correspond to chromosome arms, columns correspond to samples. Chromosome arms are classified by the middle segment if there are 3 segments, and by the most outstanding segment if there are 2 segments.

refLR - matrix with the between patient comparisons (reference distribution under the hypothesis of independence). Has the same structure as LR but the pairs of tumors are selected from different patients.

Note that calculating the reference distribution might take a long time.

If the reference is FALSE, there is no p-value column in LR and no refLR output.

Author(s)
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References

Examples

#Analysis of simulated data

set.seed(100)
chrom<-paste("chr",rep(c(1:22),each=100),"p",sep="")
chrom[nchar(chrom)==5]<-paste("chr0",substr(chrom[nchar(chrom)==5] ,4,5),sep="")
maploc<- rep(c(1:100),22)
data<-NULL
for (pt in 1:9) #first 9 patients have independent tumors
{
tumor1<-tumor2<- NULL
mean1<- rnorm(22)
mean2<- rnorm(22)
for (chr in 1:22)
{
  r<-runif(2)
  if (r[1]<0.5) tumor1<-c(tumor1,rep(0,100))
else if (r[1]>0.7) tumor1<-c(tumor1,rep(mean1[chr],100))
else { i<-sort(sample(1:100,2))
        tumor1<-c(tumor1,mean1[chr]*c(rep(0, i[1]),rep(1, i[2]-i[1]), rep(0, 100-i[2])))
    }
if (r[2]<=0.5) tumor2<-c(tumor2,rep(0,100))
else if (r[2]>0.7) tumor2<-c(tumor2,rep(mean2[chr],100))
else {i<-sort(sample(1:100,2))
        tumor2<-c(tumor2,mean2[chr]*c(rep(0, i[1]),rep(1, i[2]-i[1]), rep(0, 100-i[2])))
    }
data<-cbind(data,tumor1,tumor2)
}

#last patient has identical profiles
tumor1<- NULL
mean1<- rnorm(22)
for (chr in 1:22)
{
    r<-runif(1)
    if (r<=0.4) tumor1<-c(tumor1,rep(0,100))
    else if (r>0.6) tumor1<-c(tumor1,rep(mean1[chr],100))
    else { i<-sort(sample(1:100,2))
        tumor1<-c(tumor1,mean1[chr]*c(rep(0, i[1]),rep(1, i[2]-i[1]), rep(0, 100-i[2])))
    }
data<-cbind(data,tumor1,tumor1)
data<-data+matrix(rnorm( 44000,mean=0,sd=0.4) ,nrow=2200,ncol=20)
dataCNA<-CNA(data,chrom=chrom,maploc=maploc,sampleid=paste("pt",rep(1:10,each=2),rep(1:2,10)))
ptlist<- paste("pt",rep(1:10,each=2),sep=".")
samnms<-paste("pt",rep(1:10,each=2),rep(1:2,10),sep=".")
results<-clonality.analysis(dataCNA, ptlist, pfreq = NULL, refdata = NULL, nmad = 1, reference = TRUE, allpairs = TRUE)

#genomewide plots of pairs of tumors from the same patient
pdf("genomewideplots.pdf",height=7,width=11)
for (i in unique(ptlist))
{
    w<-which(ptlist==i)
    ns<- length(w)
    if (ns>1)
    {
        for (p1 in c(1:(ns-1)))
        for (p2 in c((p1+1):ns))
            genomewidePlots(results$OneStepSeg, results$ChromClass,ptlist , ptpair=samnms[w[p1],w[p2]],results$LR, plot.as.in.analysis = TRUE)
    }
}
dev.off()

pdf("hist.pdf",height=7,width=11)
histogramPlot(results$LR[,4], results$refLR[,4])
dev.off()
for (i in unique(ptlist))
{
    pdf(paste("Patient", i, ".pdf", sep=""), height=7, width=11)
    chromosomePlots(results$OneStepSeg, ptlist, ptname=i, nmad=1.25)
    dev.off()
}

---

**ECMtesting**

Clonality testing of $\geq$3 tumors using Extended Concordant Mutations (ECM) test based on LOH (Loss of Heterozygosity) profiles

**Description**

Function to test clonality of three and more tumors from the same patient based on their LOH profiles. This function implements Extended Concordant Mutations for all possible subsets of tumors from the same patient and minP multiplicity adjustment using simulated tumors.

**Usage**

```r
ECMtesting(LOHtable, ptlist, noloh, loh1, loh2, Nsim=100)
```

**Arguments**

- **LOHtable** Matrix of LOH calls. Each row corresponds to a marker. First column contains the names of the markers. Each other column represents a sample and contains LOH calls.
- **ptlist** Vector of the patient IDs in the order the samples appear in the data. For example, if the first three tumors (columns 2, 3, 4 of data) belong to patient A, and the following two (columns 5, 6 of data) belong to patient B, then ptlist=c(’ptA’, ’ptA’, ’ptA’, ’ptB’, ’ptB’).
- **noloh** The string or a number that denotes absence of LOH.
- **loh1** The string or a number that denotes presence of LOH.
- **loh2** The string or a number that denotes presence of LOH that is discordant from loh1.
- **Nsim** Number of simulations used to calculate minP adjusted p-values

**Details**

Extended Concordant Mutations test is done for every subset of tumors. It uses number of concordant mutations in all tumors of the subset as a test statistic, and its reference distribution is calculated assuming fixed counts of LOH per tumor and equal probability of maternal and paternal alleles being affected. Note that ECM test for 2 tumors and original CM test will give slightly different p-values since continuity correction is done in ECM test.

**Value**

The function returns a list with number of elements equal to the number of patients. Each element is a matrix with two rows: ECM p-values for all possible subsets of tumors from this patient, and minP adjusted p-values. The tumors are denoted 1,2,3,... in the order they appear in LOHtable. Any tumor subsets with minP adjusted p-value $\leq 0.05$ can be considered clonal.
References

Ostrovnaya, I. "Testing clonality of three and more tumors using their loss of heterozygosity pro-
files", Statistical Applications in Genetics and Molecular Biology, 2012

Examples

set.seed(25)
LOHtable<-cbind(1:15,matrix(sample(c(0,1,2),15*12,replace=TRUE),ncol=12))
ECMtesting(LOHtable,rep(1:3,each=4),noloh=0,loh1=1,loh2=2,Nsim=100)

```r
# See example as in clonality.analysis()
```

description

Plot contains genomewide profiles from a pair of tumors. It uses the output from the function
clonality.analysis().

Usage

genomewidePlots(data.seg1, classall, ptlist, ptpair, ptLR, plot.as.in.analysis = TRUE)

Arguments

data.seg1 Output of one-step segmentation - output OneStepSeg of clonality.analysis().
The chromosomes should be in the format "chr01p", "chr01q" etc.

classall Classifications of the chromosomes - output ChromClass of clonality.analysis()

ptlist Vector of the patient IDs in the order of the samples appearing in the data.

ptpair Two sample names for which the plot is desired

ptLR Matrix with the likelihood ratios - output LR of clonality.analysis()

plot.as.in.analysis If TRUE then the gain/loss patterns will be highlighted in accordance with the
chromosome classification. For example, if there are three segments in a chro-
mosome, then the middle one determines the chromosome status. If it is nor-
mal, no color will be plotted in the chromosome even if the 1st and 3rd seg-
ments are gains or losses. Another example: if there are 2 or 3 different seg-
ments of gains, they will be combined and only one segment will be plotted. If
plot.as.in.analysis is equal to FALSE, the original one-step CBS segmentation
will be plotted.

Details

Function produces genomewide plots of a pair of tumors. The log-ratios are plotted in grey in the
order of their genomic locations, gains are plotted in blue, and losses are plotted in red.

Examples

```r
# See example as in clonality.analysis()
```
**histogramPlot**

*Histrograms of Log-Likelihood Ratios*

**Description**

Function produces the histograms of the within-patient and between-patient log-Likelihood Ratios.

**Usage**

`histogramPlot(ptLRvec, refLRvec)`

**Arguments**

- `ptLRvec`: Vector with the within-patient likelihood ratios - output LR of clonality.analysis()
- `refLRvec`: Vector with the between-patient likelihood ratios - output refLR of clonality.analysis()

**Details**

Functions plots two overlapping histograms: within-patient log-likelihood ratios are in red and between-patient log-likelihood ratios (reference distribution under the hypothesis of independence) are in black.

**Examples**

```r
# See example as in clonality.analysis()
```

---

**LOHclonality**

*Clonality testing using LOH (Loss of Heterozygosity) profiles*

**Description**

Function to test clonality of two tumors from the same patient based on their LOH profiles. This function implements Concordant Mutations and Likelihood Ratio tests.

**Usage**

`LOHclonality(LOHtable, ptlist, refLOHtable = NULL, pfreq = NULL, noloh, loh1, loh2, method = "both")`

**Arguments**

- `LOHtable`: Matrix of LOH calls. Each row corresponds to a marker. First column contains the names of the markers. Each other column represents a sample and contains LOH calls.
- `ptlist`: Vector of the patient IDs in the order the samples appear in the data. For example, if the first three tumors (columns 3, 4, 5 of data) belong to patient A, and the following two (columns 6, 7 of data) belong to patient B, then `ptlist=c('ptA', 'ptA', 'ptA', 'ptB', 'ptB')`. 
LOHclonality

**refLOHtable** Matrix of LOH calls that should be used to calculate the LOH frequencies used in Likelihood Ration calculation. The structure is similar to LOHtable. If refLOHtable is not specified, frequencies are calculated from LOHtable.

**pfreq** Vector of LOH frequencies known from the literature. Should be in the same order as the markers in LOHtable. If pfreq is not specified, frequencies are calculated from LOHtable.

**noloh** The string or a number that denotes absence of LOH.

**loh1** The string or a number that denotes presence of LOH.

**loh2** The string or a number that denotes presence of LOH that is discordant from loh1.

**method** Takes values "CM", "LR" or "both" if only Concordant Mutations test, or only Likelihood Ratio test, or both should be performed. Default value is "both".

### Details

Function tests clonality of LOH profiles of tumors from the same patient using two tests. Concordant Mutations test has number of markers with concordant LOH as its test statistic. Its theoretical reference distribution under independence is calculated assuming that the maternal and paternal alleles are equally likely to be lost and that the frequencies of LOH are about the same across different markers.

Likelihood Ratio test uses pre-specified frequencies of LOH to compute Likelihood Ratio statistic. Its reference distribution is obtained by simulating tumors with the given LOH probabilities, and probability of maternal/paternal mutation estimated from the data. If LOH frequencies are not specified then they are estimated from the data.

### Value

The function returns a data frame where each row corresponds to the pair of samples that are compared. Columns are: Sample1 - name of sample 1; Sample2 - name of sample 2; a - number of markers with concordant LOH in both tumors (test statistic for Concordant Mutations test); e - number of markers with LOH in both tumors, concordant or discordant; f - number of markers with LOH in the first tumor and Normal in the 2nd tumor; g - number of markers with LOH in the second tumor and Normal in the first tumor; h - number of markers that are Normal in both tumors; Ntot - total number of informative markers for both tumors; CMpvalue - p-value for Concordant Mutations test; LRpvalue - p-value for Likelihood Ratio test.

### References


### Examples

```r
set.seed(25)
LOHtable<-cbind(1:20,matrix(sample(c(0,1,2),20*20,replace=TRUE),20))
LOHclonality(LOHtable,rep(1:10,each=2),pfreq=NULL,noloh=0,loh1=1,loh2=2)
```
LRtesting3or4tumors  
Clonality testing of 3 or 4 tumors using Likelihood model based on LOH (Loss of Heterozygosity) profiles

Description

Function to test clonality of 3 or 4 tumors from the same patient based on their LOH profiles.

Usage

LRtesting3or4tumors(LOHtable, ptlist, refLOHtable=NULL, pfreq=NULL, noloh, loh1, loh2, Nsim=100, m=0.5)

Arguments

- **LOHtable**: Matrix of LOH calls. Each row corresponds to a marker. First column contains the names of the markers. Each other column represents a sample and contains LOH calls.
- **ptlist**: Vector of the patient IDs in the order the samples appear in the data. For example, if the first three tumors (columns 2, 3, 4 of data) belong to patient A, and the following two (columns 5, 6 of data) belong to patient B, then ptlist=c('ptA', 'ptA', 'ptA', 'ptB', 'ptB').
- **refLOHtable**: Matrix of LOH calls that should be used to calculate the LOH frequencies used in Likelihood Ratio calculation. The structure is similar to LOHtable. If refLOHtable is not specified, frequencies are calculated from LOHtable.
- **pfreq**: Vector of LOH frequencies known from the literature. Should be in the same order as the markers in LOHtable. If pfreq is not specified, frequencies are calculated from LOHtable.
- **noloh**: The string or a number that denotes absence of LOH.
- **loh1**: The string or a number that denotes presence of LOH.
- **loh2**: The string or a number that denotes presence of LOH that is discordant from loh1.
- **Nsim**: Number of simulations used to calculate minP adjusted p-values
- **m**: Probability that a favored allele is affected given that LOH has occurred. Must be a number above 0.5 (equal probability of maternal and paternal allelic loss)

Details

Likelihood ratio test for 3 and 4 tumors. For 3 tumors there are 3 possible tumor orderings, and for 4 tumors there are 2 topologies with 3 and 12 orderings each. The test calculates maximum likelihood ratio across all possible orderings, and the p-value is calculated using simulated reference distribution.

Value

The function returns a list with number of elements equal to the number of patients. Each element is list with two elements. First contains log maximum likelihood ratio value, p-value, and estimates of parameters c, the topology and tumor ordering that have maximum likelihood ratio. If p-value is significant, then the null hypothesis that all tumors are independent can be rejected. The second element has a matrix with all possible topologies and tumor orderings and their corresponding log likelihood ratios.
splitChromosomes

References
Ostrovnaya, I. "Testing clonality of three and more tumors using their loss of heterozygosity profiles", Statistical Applications in Genetics and Molecular Biology, 2012

Examples

```r
set.seed(25)
LOHtable<-cbind(1:15,matrix(sample(c(0,1,2),15*12,replace=TRUE),ncol=12))
q<-LRtesting3or4tumors(LOHtable,rep(1:4,each=3),refLOHtable=NULL,pfreq=NULL,noloh=0,loh1=1,loh2=2,Nsim=100)
```

splitChromosomes  

**Chromosome splitting**

**Description**
Divides the chromosomes into p and q arms.

**Usage**

```r
splitChromosomes(chrom,maploc)
```

**Arguments**

- `chrom` Vector of chromosomes. They should be numeric 1 to 22.
- `maploc` Vector of genomic locations. They should be in Kilobases.

**Details**

The function returns the vector of chromosome arms labeled "chr01p", "chr01q", etc. The split into arms is accomplished using the following centers (in Kb) for chromosomes 1 through 22: (122356.96, 93189.90, 92037.54, 50854.87, 47941.40, 60438.12, 59558.27, 45458.05, 48607.50, 40434.94, 52950.78, 35445.46, 16934.00, 16570.00, 16760.00, 36043.30, 22237.13, 16082.90, 28423.62, 27150.40, 11760.00, 12830.00).

**Examples**

```r
#simulated data
set.seed(100)
chrom<rep(c(1:22),each=100)
maploc<-runif(2200)*200000
chromarm<-splitChromosomes(chrom,maploc)
```
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