Package ‘DeconRNAsEq’

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Type Package

Title Deconvolution of Heterogeneous Tissue Samples for mRNA-Seq data

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Depends R (>= 2.14.0), limSolve, pcaMethods, ggplot2, grid

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Description DeconSeq is an R package for deconvolution of heterogeneous tissues based on mRNA-Seq data. It modeled expression levels from heterogeneous cell populations in mRNA-Seq as the weighted average of expression from different constituting cell types and predicted cell type proportions of single expression profiles.

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biocViews DifferentialExpression

NeedsCompilation no

R topics documented:

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package DeconRNASeq contains function "DeconRNASeq", implementing the decomposition of RNA-Seq expression profilings of heterogeneous tissues into cell/tissue type specific expression and cell type concentration based on cell-type-specific reference measurements.

Description

Main function "DeconRNASeq" implements a nonnegative decomposition by quadratic programming as datasets = signature*A, where "datasets" are the originally measured data matrix (e.g. genes by samples), "signature" is the signature matrix (genes by cell types) and "A" the cell type concentration matrix (cell types by samples).

Details

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DeconRNASeq(datasets, signature)

Author(s)

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References


all.datasets data objects for rat liver_brain samples

Description

A data frame providing the expression profilings of GSE19830 microarray samples.

Usage

all.datasets
array.proportions

Format
A matrix with expression studies in the GSE19830 microarray samples: the first three columns are corresponding to the liver, while the last three samples are corresponding to the brain.

Author(s)
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Examples
data(rat_liver_brain)

array.proportions  proportions for rat liver and brain mixing samples

Description
array.proportions: a data frame providing the fractions for liver and brain from the microarray GSE 19830 study

Usage
array.proportions

Format
a matrix whose rows are mixing samples’ name and columns are fractions from pure live and brain tissues

Author(s)
Ting Gong <tinggong@gmail.com> Joseph D. Szustakowski <joseph.szustakowski@novartis.com>

Examples
data(rat_liver_brain)

array.signatures  data objects for rat liver and brain pure samples

Description
array.signatures: a data frame providing the expression values from rat pure liver and brain samples, each has threee replicates

Usage
array.signatures
Format

a data matrix with 30 expressions from rat pure liver and brain tissues

Author(s)

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Examples

data(rat_liver_brain)

condplot

Draw the plot of the condition numbers vs. the number of genes in the signature matrix

Description

A function is used to draw the plot of the condition number of signature matrices of all sizes, from a handful of genes in one extreme to the whole signature in the other.

Usage

condplot(step, cond)

Arguments

step an array with the number of genes used to calculate the condition numbers of signature matrices, default stepwise = 20
cond an array with the condition numbers of signature matrices

Value

a plot for the condition numbers of signature matrices

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References

library(DeconRNASeq)

### toy data example:

step <- seq(20,1000, by=20) # every 20 genes
## cell type-specific gene expression matrix:
  x.signature <- matrix(rexp(2000), ncol=2)
  sig.cond <- sapply(step, function(x) kappa(scale(x.signature[1:x,])))

## decon.bootstrap

decon.bootstrap (data.set, possible.signatures, n.sig, n.iter)
DeconRNASeq

Arguments

data.set the data object for mixing samples
possible.signatures a data frame providing the expression values from pure tissue samples
n.sig the number of genes/transcripts used for estimation of proportions from our deconvolution
n.iter the number of bootstraps for our deconvolution

Value

A three dimensional array to store means and 95% confidence interval

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References


DeconRNASeq Function for Deconvolution of Complex Samples from RNA-Seq.

Description

This function predicts proportions of constituting cell types from gene expression data generated from RNA-Seq data. Perform nonnegative quadratic programming to get per-sample based globally optimized solutions for constituting cell types.

Usage

DeconRNASeq(datasets, signatures, proportions = NULL, checksig = FALSE, known.prop = FALSE, use.scale = TRUE, fig = TRUE)

Arguments

datasets measured mixture data matrix, genes (transcripts) e.g. gene counts by samples. The user can choose the appropriate counts, RPKM, FPKM etc..

signatures signature matrix from different tissue/cell types, genes (transcripts) by cell types. For gene counts, the user can choose the appropriate counts, RPKM, FPKM etc..

proportions proportion matrix from different tissue/cell types.

checksig whether the condition number of signature matrix should be checked, default = FALSE

known.prop whether the proportions of cell types have been known in advanced for proof of concept, default = FALSE

use.scale whether the data should be centered or scaled, default = TRUE

fig whether to generate the scatter plots of the estimated cell fractions vs. the true proportions of cell types, default = TRUE
Details

Data in the originally measured mixture sample matrix: datasets and reference matrix: signatures, need to be non-negative. We recommend to deconvolute without log-scale.

Value

Function DeconRNA-Seq returns a list of results

- **out.all**: estimated cell type fraction matrix for all the mixture samples
- **out.pca**: svd calculated PCA on the mixture samples to estimate the number of pure sources according to the cumulative R2
- **out.rmse**: averaged root mean square error (RMSE)) measuring the differences between fractions predicted by our model and the truth fraction matrix for all the tissue types

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References


Examples

```r
## Please refer our demo
##source("DeconRNASeq.R")
### multi_tissue: expression profiles for 10 mixing samples from multiple tissues
#data(multi_tissue.rda)
#datasets <- x.data[,2:11]
#signatures <- x.signature.filtered.optimal[,2:6]
#proportions <- fraction

#DeconRNASeq(datasets, signatures, proportions, checksig=FALSE, known.prop = TRUE, use.scale = TRUE)
```

### Descriptive Table

| fraction | mixing fractions for multi-tissues mixing samples |

Description

A data frame providing the fractions from multiple tissues in the mixing samples

Usage

fraction

Format

A matrix whose rows are mixing samples’ name and columns are fractions from pure tissues including brain, muscle, lung, liver and heart
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Examples
data(multi_tissue)

data(liver_kidney)

Description
a list containing:
1) datasets: a data frame providing the RPKM of seven mixing samples.
2) proportions: a data frame providing the fractions for liver and kidney in the mixing samples
3) signatures: a data frame providing the expression values from pure liver and kidney samples

Usage
liver_kidney

Format
A list 1) a data frame with 31979 genes’ expression on the 7 mixing samples: reads.1, reads.2, reads.3, reads.4, reads.5, reads.6, reads.7
2) a matrix whose rows are mixing samples’ name and columns are fractions from pure liver and kidney tissues
3) a data matrix with 630 expressions from pure liver and kidney tissues

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Examples
data(liver_kidney)
multiplot

\textit{Draw the plots of proportions of cells determined from deconvolution vs. proportions of the cells actually mixed (when available) with RMSE.}

\begin{description}
\item[Description] A function is used to draw the multiple plots of proportions of cells determined from deconvolution vs. proportions of the cells actually mixed. Each plot corresponds to one tissue/cell.
\item[Usage] \texttt{multiplot(..., plotlist = NULL, cols)}
\item[Arguments]
\begin{itemize}
\item \ldots \hspace{1cm} any number of the plot objects that store the scatter plots for all the cells/tissue types
\item \texttt{plotlist} \hspace{1cm} any other plot objects
\item \texttt{cols} \hspace{1cm} columns of the plots, default = 1
\end{itemize}
\item[Value] A pdf file with the plots of proportions of cells determined from deconvolution vs. proportions of the cells actually mixed with RMSE
\item[Author(s)] Ting Gong <tinggong@gmail.com> Joseph D. Szustakowski <joseph.szustakowski@novartis.com>
\item[Examples]
\begin{verbatim}
## The function is currently defined as
function (...[, plotlist = NULL, cols]) {
  pdf("scatterplots.pdf")
  require(grid)
  plots <- c(list(...), plotlist)
  numPlots = length(plots)
  plotCols = cols
  plotRows = ceiling(numPlots/plotCols)
  grid.newpage()
  pushViewport(viewport(layout = grid.layout(plotRows, plotCols)))
  vplayout <- function(x, y) viewport(layout.pos.row = x, layout.pos.col = y)
  for (i in 1:numPlots) {
    curRow = ceiling(i/plotCols)
    curCol = (i - 1)%/%plotCols + 1
    
\end{verbatim}
\end{description}
multi_tissue

print(plots[[i]], vp = vplayout(curRow, curCol))
}
dev.off()
}

multi_tissue  

Data objects for multi-tissues mixing samples

Description

A list containing:

1) x.data: a data frame providing the RPKM of nine mixing samples.
2) x.signatures: a data frame providing the expression values from pure brain, muscle, lung, liver and heart samples.
3) x.signatures.filtered: a data frame providing the expression values from pure brain, muscle, lung, liver and heart samples after filtering.
4) x.signatures.filtered.optimal: a data frame providing the expression values from pure brain, muscle, lung, liver and heart samples used for the example in DeconRNA-Seq.
5) fraction: a data frame providing the fractions from 5 tissues in the mixing samples

Usage

multi_tissue

Format

A list 1) a matrix with all the genes’ expression in the mixing samples: the first two columns are corresponding to the RefSeq accession numbers and gene symbols
2) a matrix whose rows are gene symbols and columns are RPKM expressions from pure tissues.
3) a matrix whose rows are gene symbols and columns are RPKM expressions from pure tissues: the genes with RPKM less than 200 within any of the five tissues have been filtered.
4) a matrix whose rows are gene symbols and columns are RPKM expressions from pure tissues: based on the filtered signature matrix, the optimal number of genes have been selected for the deconvolution according to the condition numbers
5) a matrix whose rows are mixing samples’ name and columns are fractions from pure tissues including brain, muscle, lung, liver and heart

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Examples

data(multi_tissue)
**proportions**

---

**proportions**

**proportions for liver and kidney mixing samples**

---

**Description**

proportions: a data frame providing the fractions for liver and kidney in the mixing samples

**Usage**

proportions

**Format**

a matrix whose rows are mixing samples’ name and columns are fractions from pure live and kidney tissues

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**Examples**

data(liver_kidney)

---

**rmse**

*Calculate the differences between proportions predicted by deconvolution and the values actually measured*

---

**Description**

A function is used to calculate the root-mean-square error (RMSE) for the accuracy of estimated proportions.

**Usage**

rmse(x, y)

**Arguments**

- **x**: proportions from the actual measurement
- **y**: estimated proportions from our deconvolution

**Value**

A number for RMSE

**Author(s)**

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References


signatures  
data objects for liver and kidney pure samples

Description

signatures: a data frame providing the expression values from pure liver and kidney samples

Usage

signatures

Format

a data matrix with 630 expressions from pure liver and kidney tissues

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Examples

data(liver_kidney)

x.data  
data objects for multi-tissues mixing samples

Description

A data frame providing the RPKM of nine mixing samples.

Usage

x.data

Format

A matrix with all the genes’ expression in the mixing samples: the first two columns are corresponding to the RefSeq accession numbers and gene symbols

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Examples

data(multi_tissue)
**x.signature**

**data objects for multi-tissues pure samples**

**Description**
A data frame providing the expression values from pure brain, muscle, lung, liver and heart samples.

**Usage**

`x.signature`

**Format**

A martix whose rows are gene symbols and columns are RPKM expressions from pure tissues.

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**Examples**

```r
data(multi_tissue)
```

---

**x.signature.filtered**

**filtered signatures for multi-tissues pure samples**

**Description**
A data frame providing the expression values from pure brain, muscle, lung, liver and heart samples after filtering.

**Usage**

`x.signature.filtered`

**Format**

A martix whose rows are gene symbols and columns are RPKM expressions from pure tissues: the genes with RPKM less than 200 within any of the five tissues have been filtered.

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**Examples**

```r
data(multi_tissue)
```
x.signature.filtered.optimal

selected signatures from multi-tissues pure samples

Description
A data frame providing the expression values from pure brain, muscle, lung, liver and heart samples used for the example in DeconRNA-Seq.

Usage
x.signature.filtered.optimal

Format
A matrix whose rows are gene symbols and columns are RPKM expressions from pure tissues: based on the filtered signature matrix, the optimal number of genes have been selected for the deconvolution according to the condition numbers.

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Examples
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