Package ‘FEM’

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Title Identification of Functional Epigenetic Modules
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Description The FEM package performs a systems-level integrative analysis of DNA methylation and gene expression data. It seeks modules of functionally related genes which exhibit differential promoter DNA methylation and differential expression, where an inverse association between promoter DNA methylation and gene expression is assumed. For full details, see Jiao et al Bioinformatics 2014.
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The FEM package performs a systems-level integrative analysis of DNA methylation and gene expression. It seeks modules of functionally related genes which exhibit differential promoter DNA methylation and differential expression, where an inverse association between promoter DNA methylation and gene expression is assumed. For full details, see Jiao et al Bioinformatics 2014.

Details

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Author(s)

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References


DoEpiMod

Identifies differential DNA methylation network hotspots.

Description

This function aims to identify subnetworks where many members exhibit differential DNA methylation in relation to the phenotype of interest.
**Usage**

```r
DoEpiMod(intEpi.o, nseeds = 100, gamma = 0.5, nMC = 1000,
  sizeR.v = c(1, 100), minsizeOUT = 10, writeOUT = TRUE, nameSTUDY = "X", ew.v = NULL)
```

**Arguments**

- `intEpi.o`: The output of the DoIntEpi450k function.
- `nseeds`: An integer specifying the number of seeds and therefore modules to search for. By default this number is 100.
- `gamma`: A parameter of the spin-glass algorithm, which determines the average module size. Default value generally leads to modules in the desired size range (10-100 genes).
- `nMC`: Number of Monte Carlo runs for establishing statistical significance of modularity values under randomisation of the molecular profiles on the network.
- `sizeR.v`: Desired size range for modules.
- `minsizeOUT`: Minimum size of modules to report as interesting.
- `writeOUT`: A logical to indicate whether to write out tables in text format.
- `nameSTUDY`: A name for the study, to be used as label in the output files.
- `ew.v`: The edge weight vector of the integrated network. This is actually generated by the function itself, and can speed up inference significantly, if provided as argument to a 2nd instance of the function. Default value is NULL.

**Value**

A list with following entries:

- `size`: A vector of inferred module sizes for each of the ntop seeds.
- `mod`: A vector of associated modularities.
- `pv`: A vector of associated significance P-values with resolution limited by nMC runs.
- `selmod`: Index positions of significant modules of size at least minsizeOUT
- `fem`: A summary matrix of the selected modules.
- `topmod`: A list of summary matrices for each of the selected modules.
- `sgc`: A list of the spin-glass module detection algorithm for each seed.
- `ew`: The edge-weight vector of the integrated network.
- `adj`: adjacency matrix of the maximally connected integrated network (at present only maximally connected subnetwork is used). It is same to intEpi.o$adj, and will be used for FemModShow function.

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**References**

**DoExpMod**

**Identifies differential mRNA expression network hotspots.**

**Description**

This function aims to identify subnetworks where many members exhibit differential mRNA expression in relation to the phenotype of interest.

**Usage**

```r
DoExpMod(intExp.o, nseeds = 100, gamma = 0.5, nMC = 1000, sizeR.v = c(1,100),
          minsizeOUT = 10, writeOUT = TRUE, nameSTUDY = "X", ew.v = NULL)
```

**Arguments**

- `intExp.o` The output of the DoIntExp function
- `nseeds` An integer specifying the number of seeds and therefore modules to search for. By default this number is 100.
- `gamma` A parameter of the spin-glass algorithm, which determines the average module size. Default value generally leads to modules in the desired size range (10-100 genes).
- `nMC` Number of Monte Carlo runs for establishing statistical significance of modularity values under randomisation of the molecular profiles on the network.
- `sizeR.v` Desired size range for modules.
- `minsizeOUT` Minimum size of modules to report as interesting.
- `writeOUT` A logical to indicate whether to write out tables in text format.
- `nameSTUDY` A name for the study, to be used as label in the output files.
- `ew.v` The edge weight vector of the integrated network. This is actually generated by the function itself, and can speed up inference significantly, if provided as argument to a 2nd instance of the function. Default value is NULL.

**Value**

A list with following entries:

- `size` A vector of inferred module sizes for each of the ntop seeds.
- `mod` A vector of associated modularities.
DoFEMbi

Identifies interactome hotspots of differential promoter DNA methylation and differential mRNA expression.

Description

DoFEMbi identifies interactome hotspots of differential promoter methylation and differential expression in relation to a phenotype of interest, where an inverse association between methylation and gene expression is assumed.

Usage

DoFEMbi(intFEM.o, nseeds = 100, gamma = 0.5, nMC = 1000, sizeR.v = c(1,100), minsizeOUT = 10, writeOUT = TRUE, nameSTUDY = "X", ew.v = NULL)
Arguments

intFEM.o  The output of the DoFEMbi function.
nseeds  An integer specifying the number of seeds and therefore modules to search for. By default this number is 100.
gamma  A parameter of the spin-glass algorithm, which determines the average module size. Default value generally leads to modules in the desired size range (10-100 genes).
nMC  Number of Monte Carlo runs for establishing statistical significance of modularity values under randomisation of the molecular profiles on the network.
sizeR.v  Desired size range for modules.
minsizeOUT  Minimum size of modules to report as interesting.
writeOUT  A logical to indicate whether to write out tables in text format.
nameSTUDY  A name for the study, to be used as label in the output files.
ew.v  The edge weight vector of the integrated network. This is actually generated by the function itself, and can speed up inference significantly, if provided as argument to a 2nd instance of the function. Default value is NULL.

Value

A list with following entries:

size  A vector of inferred module sizes for each of the ntop seeds.
mod  A vector of associated modularities.
pv  A vector of associated significance P-values with resolution of nMC
selmod  Index positions of significant modules of size at least minsizeOUT
fem  A summary matrix of the selected modules.
topmod  A list of summary matrices for each of the selected modules.
sgc  A list of the spin-glass module detection algorithm for each seed.
ew  The edge-weight vector of the integrated network.
adj  adjacency matrix of the maximally connected integrated network (at present only maximally connected subnetwork is used). It is same to intFEM.o$adj, and will be used for FemModShow function

Author(s)

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References

DoIntEpi450k

**Examples**

```r
data(Toydata);
intFEM.o <- list(statM=Toydata$statM,statR=Toydata$statR,adj=Toydata$adj);
DoFEMbi(intFEM.o,nseeds=1,gamma=0.5,nMC=1000,sizeR.v=c(1,100),
    minsizeOUT=10,writeOUT=TRUE,nameSTUDY="TEST",ew.v=NULL);
#You can also test on the Realdata contains matched DNA methylation and RNA Expression of 17 normal and 118 cancer samples. Since running on the realdata is time-consuming, we comment it out.
#data(Realdata);
#intFEM.o <- list(statM=Realdata$statM,statR=Realdata$statR,adj=Realdata$adjacency);
#DoFEMbi(intFEM.o,nseeds=100,gamma=0.5,nMC=1000,sizeR.v=c(1,100),nameSTUDY="TEST");
```

---

**DoIntEpi450k**

Integrates statistics of differential DNA methylation for a specified contrast with a network adjacency matrix.

**Description**

Integrates statistics of differential DNA methylation for a specified contrast with a network adjacency matrix. Output of this function is required input for DoEpiMod.

**Usage**

```r
DoIntEpi450k(statM.o,adj.m,c,"dmaMode")
```

**Arguments**

- `statM.o`: The output of GenStatM, GenStatMsp, or analogous object generated by user.
- `adj.m`: The adjacency matrix defining the network of gene relations, e.g. a PPI network, annotated to NCBI entrez gene IDs.
- `c`: An integer specifying the desired contrast, i.e. column of the contrast matrix. User needs to check which column of statM.o$cont is the one of interest.
- `dmaMode`: An parameter specifying the object mode for statM.o, "avbeta" for the average beta mode object generated by GenStatM, whereas "singleProbe" for the single probe mode object generated by the GenStatMsp, default for "avbeta"

**Value**

A list with following entries:

- `statM`: matrix of DNA methylation moderated t-statistics and P-values for the genes in the integrated network
- `adj`: adjacency matrix of the maximally connected integrated network (at present only the maximally connected subnetwork is used in the subsequent inference
- `avbeta`: average DNA methylation data matrix mapped to unique Entrez IDs
- `probeID`: vector of the probe IDs that most differentially methylated for each gene

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References


DoIntExp

Integrates statistics of differential expression for a specified contrast with a network adjacency matrix.

Description

Integrates statistics of differential expression for a specified contrast with a network adjacency matrix. Output of this function is required input for DoExpMod.

Usage

DoIntExp(statR.o, adj.m, c)

Arguments

statR.o The output of GenStatR, or analogous object generated by user.
adj.m The adjacency matrix defining the network of gene relations, e.g. a PPI network, annotated to NCBI entrez gene IDs.
c An integer specifying the desired contrast, i.e. column of the contrast matrix. User needs to check which column of statR.o$cont is the one of interest.

Value

A list with following entries:

statR matrix of gene expression moderated t-statistics and P-values for the genes in the integrated network
adj adjacency matrix of the maximally connected integrated network (at present only the maximally connected subnetwork is used for subsequent inference)
avexp average expression data matrix mapped to unique Entrez IDs

Author(s)

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References

DoIntFEM450k

Integrates statistics of differential DNA methylation and differential expression for a specified contrast with a network adjacency matrix.

Description

Integrates statistics of differential DNA methylation and differential expression for a specified contrast with a network adjacency matrix. Output of this function is required input for DoFEMbi.

Usage

DoIntFEM450k(statM.o,statR.o,adj.m,cM,cR,"dmaMode")

Arguments

- **statM.o**: The output of GenStatM, or analogous object generated by user.
- **statR.o**: The output of GenStatR, or analogous object generated by user.
- **adj.m**: The adjacency matrix defining the network of gene relations, e.g. a PPI network, annotated to NCBI entrez gene IDs.
- **cR**: An integer specifying the desired contrast, i.e. column of the contrast matrix. User needs to check which column of statR.o$cont is the one of interest.
- **cM**: An integer specifying the desired contrast, i.e. column of the contrast matrix. User needs to check which column of statM.o$cont is the one of interest. Note that for this to make sense the contrast cM must correspond to the same one as specified by cR.
- **dmaMode**: An parameter specifying the object mode for statM.o, "avbeta" for the average beta mode object generated by GenStatM, whereas "singleProbe" for the single probe mode object generated by the GenStatMsp, default for "avbeta"

Value

A list with following entries:

- **statM**: matrix of DNA methylation moderated t-statistics and P-values for the genes in the integrated network
- **statR**: matrix of gene expression moderated t-statistics and P-values for the genes in the integrated network
- **adj**: adjacency matrix of the maximally connected integrated network (at present only maximally connected subnetwork is used).
- **avexp**: average expression data matrix mapped to unique Entrez IDs
- **avbeta**: average DNA methylation data matrix mapped to unique Entrez IDs
- **probeID**: vector of the probe IDs that most differentially methylated for each gene

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**FemModShow**

Generates a figure showing the inferred FEM module(s)

### Description

This function generates a network representation of an inferred FEM module.

### Usage

```r
FemModShow(mod, name, fem.o, mode="Integration")
```

### Arguments

- `mod`: the module of the FEM result object for which a figure is to be generated
- `name`: the name for the output figure file
- `fem.o`: FEM result object generated by DoFEMbi, DoEpiMod, or DoExpMod
- `mode`: There are three modes: "Integration", "Epi", "Exp". "Integration" means the module is from DoFEMbi, "Epi" means the module is from DoEpiMod, "Exp" means the module is from DoExpMod.

### Value

An igraph object representing the module. At the same time, the module graph is generated and saved as a "X.pdf" file with "X" the name given in the argument to the function.

### Author(s)

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### References


### Examples

```r
data(Realdata)
FemModShow(Realdata$fembi.o$topmod$HAND2,name="HAND2",Realdata$fembi.o)
```
**Description**

Given an Illumina 450k or EPIC (850k) data matrix and a phenotype vector, it will generate statistics of differential DNA methylation using limma, at the gene-level, for all pairwise comparisons of phenotype levels.

**Usage**

```r
GenStatM(dnaM.m, pheno.v, "chiptype")
```

**Arguments**

- `dnaM.m`: The DNA methylation beta valued data matrix with rownames annotated to Illumina 450k or EPIC(850k) probe IDs
- `pheno.v`: The phenotype vector.
- `chiptype`: A parameter specifying the input data matrix, it should be either "450k" for Illumina 450k matrix or "EPIC" for Illumina EPIC matrix. Default for "450k"

**Value**

- `top`: A list of matrices, from the limma output, ranking genes according to differential methylation between two phenotypes as specified in the contrasts matrix, i.e. `top[[i]]` will contain the ranking for the `i`'th contrast.
- `cont`: The contrasts matrix, with columns labeling the contrasts.
- `avbeta`: The beta-valued DNAm data matrix at the gene-level, following the procedure as described in Jiao et al Bioinformatics 2014.

**Author(s)**

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**References**

GenStatMsp

Generation of differential DNA methylation statistics for SingleProbe Mode

Description

Given an Illumina 450k or EPIC (850k) data matrix and a phenotype vector, similar to the function of GenStatM, it will generate statistics of differential DNA methylation using limma, at the gene-level, for all pairwise comparisons of phenotype levels. However, instead of the average beta values for the probes in each region, it will select the most significant differentially methylated CpGs among TSS200 and 1st Exon regions, if not present then select the the most significant differentially methylated CpGs among TSS1500

Usage

GenStatMsp(dnaM.m,pheno.v,"chiptype")

Arguments

dnaM.m The DNA methylation beta valued data matrix with rownames annotated to Illumina 450k or EPIC(850k) probe IDs
pheno.v The phenotype vector.
chiptype A parameter specifying the the input data matrix, it should be either "450k" for Illumina 450k matrix or "EPIC" for Illumina EPIC matrix. default for "450k"

Value

top A list of matrices, from the limma output, ranking genes according to differential methylation between two phenotypes as specified in the contrasts matrix, i.e. top[i][i] will contain the ranking for the i’th contrast.
cont The contrasts matrix, with columns labeling the contrasts.
probeID A list of vectors, with each entry a vector of probeIDs that present the most differentially methylated cpg associated with each gene, i.e. probeID[i][i] will contain the vector for the i’th contrast.

Author(s)

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References

**GenStatR**

**Generation of differential expression statistics**

**Description**

Given a gene expression data matrix annotated to entrez gene IDs and a phenotype vector, it will generate statistics of differential expression using limma, at the gene-level, for all pairwise comparisons of phenotype levels.

**Usage**

```
GenStatR(exp.m, pheno.v)
```

**Arguments**

- `exp.m`: The gene expression data matrix with rownames annotated to entrez gene IDs. If there are multiple rows with the same gene ID, function will average these.
- `pheno.v`: The phenotype vector.

**Value**

- `top`: A list of matrices, from the limma output, ranking genes according to differential expression between two phenotypes as specified in the contrasts matrix, i.e. `top[[i]]` will contain the ranking for the `i`'th contrast.
- `cont`: The contrasts matrix, with columns labeling the contrasts.
- `avexp`: The averaged expression matrix at the gene-level.

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**References**


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**probe450kfemanno**

**Processed annotation of Illumina 450k probes for use in the FEM algorithm.**

**Description**

A list of entries giving various annotation information of the 450k probes

**Usage**

```
data(probe450kfemanno)
```
**Value**

- **typeC**: a vector of 0’s and 1’s indicating whether a probe maps to a CpG (1) or not (0).
- **CGI**: a vector of 0’s and 1’s indicating whether a probe maps to a CGI (1) or not (0).
- **design**: a vector of 1’s and 2’s, specifying design type of probe (1 = type I, 2 = type II).
- **probeID**: the 450k probe IDs.
- **GeneGroup**: an integer vector specifying which gene region the probe maps to. Regions are annotated as follows: 1=TSS1500, 2=TSS200, 3=5’UTR, 4=1stExon, 5=gene body, 6=3’UTR. Probes with ambiguous mappings are assigned an NA.
- **eid**: mapping of 450k probes to Entrez Gene ID.

**Author(s)**

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**References**


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**Description**

A list of entries giving various annotation information of the EPIC probes.

**Usage**

```r
data(probeEPICfemanno)
```

**Value**

- **typeC**: a vector of 0’s and 1’s indicating whether a probe maps to a CpG (1) or not (0).
- **CGI**: a vector of 0’s and 1’s indicating whether a probe maps to a CGI (1) or not (0).
- **Design**: a vector of 1’s and 2’s, specifying Design type of probe (1 = type I, 2 = type II).
- **probeID**: the EPIC probe IDs.
- **GeneGroup**: an integer vector specifying which gene region the probe maps to. Regions are annotated as follows: 1=TSS1500, 2=TSS200, 3=5’UTR, 4=1stExon, 5=gene body, 6=3’UTR. Probes with ambiguous mappings are assigned an NA.
- **eid**: mapping of EPIC probes to Entrez Gene ID.

**Author(s)**

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Realdata

A dataset object derived from real DNA methylation and gene expression data from normal endometrial and endometrial cancer tissue

Description

Realdata is a list containing data derived from mRNA Expression and DNA methylation from an endometrial cancer study. See Jones et al PLoS Med.2013 for details about the data.

Usage

data(Realdata)

Value

statM a matrix of statistics and p-values of differential DNA Methylation between endometrial cancer and normal tissue (cancer compared to normal) with rownames annotated with entrez gene IDs.

statR a matrix of statistics and p-values of differential RNA Expression between endometrial cancer and normal tissue (cancer compared to normal) with rownames annotated with entrez gene IDs.

adjacency adjacency matrix of PPI network, with number of rows equal to the number of rows of Realdata$statM, ordered in same way and with same gene identifier. The resulting graph is connected.

fembi.o this entry represents the output of the function DoFEMbi() (see example below). This is included so as to avoid having to rerun the (lengthy) example from scratch.

References


Examples

#data(Realdata);
#intFEM.o <- list(statM=Realdata$statM,statR=Realdata$statR,adj=Realdata$adjacency);
Toydata

Artificial data set.

Description
A list object containing artificial data to test the FEM algorithm. It contains artificially created statistics of differential DNA methylation, mRNA expression, as well as an artificial adjacency matrix to illustrate the application of FEM.

Usage
data(Toydata)

Value
statM a matrix of simulated statistics and p-values of differential DNA Methylation with 84 rows (genes).
statR a matrix of simulated statistics and p-values of differential RNA Expression with 84 rows (genes).
adjacency artificial adjacency matrix of 84 rows and columns, ordered according to the rows of previous two matrices. Adjacency matrix is a random graph except for containing a clique defined by 10 nodes.
tenodes a vector of 10 integer indices, labeling the nodes/genes which form the clique in the network.

Author(s)
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References

Examples
data(Toydata);
intFEM.o <- list(statM=Toydata$statM,statR=Toydata$statR,adj=Toydata$adjacency);
DoFEMbi(intFEM.o,nseeds=1, gamma=0.5,nMC=1000, sizeR.v=c(1,100), minsizeOUT=10,
writeOUT=TRUE,nameSTUDY="TEST",ew.v=NULL);
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