Package ‘GeneMeta’

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Title MetaAnalysis for High Throughput Experiments

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Description A collection of meta-analysis tools for analysing high throughput experimental data

Maintainer Bioconductor Package Maintainer

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Depends R (>= 2.10), methods, Biobase (>= 2.5.5), genefilter

Imports methods, Biobase (>= 2.5.5)

Suggests RColorBrewer

LazyLoad yes

biocViews Sequencing, GeneExpression, Microarray

NeedsCompilation no

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CountPlot

Plots for Meta-analysis of gene expression data.

Description

Plots for meta-analysis
Usage

IDRplot(m,CombineExp=1:(length(grep("zSco_Ex",colnames(m))))),colPos="black",colNeg="red",pchPos="*",pchNeg="*",type="b",ylab="IDR",xlab="z threshold",...

CountPlot(kkk,cols,Score=c("FDR","zSco"),kindof=c("two.sided","pos","neg"),type="b",pch="*",ylab...

Arguments

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<td>m</td>
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<tr>
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<td>symbol for negative z scores</td>
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<tr>
<td>CombineExp</td>
<td>vector of integer- which experiments should be combined-default:all experiments</td>
</tr>
<tr>
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<tr>
<td>cols</td>
<td>vector of cols, one for each experiment, and one for the combination</td>
</tr>
<tr>
<td>Score</td>
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</tr>
<tr>
<td>kindof</td>
<td>&quot;pos&quot;,&quot;neg&quot; or &quot;two.sided&quot;</td>
</tr>
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Details

IDRplot produces a plot described in Choi et al.

Author(s)

M.Ruschhaupt

References

Choi et al, Combining multiple microarray studies and modeling interstudy variation. Bioinformatics, 2003, i84-i90.

dstar

Description

A small number of meta-analysis functions for comparing two gene expression experiments are provided.
Usage

dstar(d, n)
getdF(data, categ)
sigmad(d, ng1, ng2)

Arguments

d A vector of t-statistics, i.e. the output of getdF.
n The number of t-statistics.
data The data used to compute t-statistics, either a matrix or an ExpressionSet.
categ A vector of 0’s and 1’s indicating group membership.
ng1 The number of samples in group 1.
ng2 The number of samples in group 2.

Details

The functions getdF compute t-test statistics for the input data and group membership (note that group membership must be indicated by a vector of 0’s and 1’s).

The function dstar computes an unbiased estimate of the t-test. The function sigmad computes the variance estimate of dstar.

Value

The different functions have different return values, but generally they are vectors of the requested quantities.

Author(s)

L. Lusa, R. Gray and R. Gentleman

References

Choi et al, Combining multiple microarray studies and modeling interstudy variation. Bioinformatics, 2003, i84-i90.

Examples

x = matrix(rnorm(1000), ncol=10)
ds = getdF(x, rep(c(0,1), c(5,5)))
dst = dstar(ds, ncol(x))
sgd = sigmad(ds, 5, 5)
Compute Cochran’s Q statistic

Description

Compute Cochran’s Q statistic for testing whether the a fixed effects or a random effects model will be appropriate.

Usage

f.Q(dadj, varadj)

Arguments

dadj A matrix, each row is a gene, each column a study, of the estimated t-statistics.
varadj A matrix, each row is a gene, each column a study, of the estimated, adjusted variances of the t-statistics.

Details

A straightforward computation of Cochran’s Q statistic. If the null hypothesis that the data are well modeled by a fixed effects design is true then the estimate Q values will have approximately a chi-squared distribution with degrees of freedom equal to the number of studies minus one.

Value

A vector of length equal to the number of rows of dadj with the Q statistics.

Author(s)

L. Lusa and R. Gentleman

References

Choi et al, Combining multiple microarray studies and modeling interstudy variation. Bioinformatics, 2003, i84-i90.

See Also
dstar,sigmad

Examples

##none now, this requires a pretty elaborate example
Nevins

Intensity data for 46 Affymetrix slides with tissue samples of breast tumors

Description

Intensity data for 46 Affymetrix hu6800 slides with tissue samples of breast tumors. See vignette Nevins.pdf in the /scripts directory for details of the processing.

Usage

data(Nevins)

Format

Nevins is an ExpressionSet containing the data from 46 Affymetrix chips.

Source

http://data.cgt.duke.edu/west.php

References


Examples

data(Nevins)
Nevins

tau2.DL

Estimating my and tau in a REM

Description

tau2.DL is an estimation of tau in a random effects model (REM) using Cochran’s Q statistic.

Usage

tau2.DL(Q, num.studies, my.weights)
mu.tau2(my.d, my.vars.new)
var.tau2(my.vars.new)
Arguments

Q  
num.studies  
my.weights  
my.d  
my.vars.new

A vector of Cochran’s Q statistics.
The number of studies used for the meta-analysis.
A matrix with one column for each experiment containing the variances of the effects that should be combined.
A matrix, with one column for each experiment, containing the effects that should be combined.
A matrix, with one column for each experiment, containing the variances of the effects that should be combined.

Author(s)

L. Lusa and R. Gentleman

References

Choi et al, Combining multiple microarray studies and modeling interstudy variation. Bioinformatics, 2003, i84-i90.

See Also

dstar, sigmad

Examples

# please have a look at the vignette

zScores  

Tools for Meta-analysis of gene expression data.

Description

A small number of meta-analysis functions for computing zScores for FEM and REM and computing FDR.

Usage

zScores(esets, classes, useREM=TRUE, CombineExp=1:length(esets))
zScorePermuted(esets, classes, useREM=TRUE, CombineExp=1:length(esets))
zScoreFDR(esets, classes, useREM=TRUE, nperm=1000, CombineExp=1:length(esets))
multExpFDR(theScores, thePermScores, type="pos")

Arguments

esets  
classes  
useREM

A list of ExpressionSets, one expression set per experiment. All experiments must have the same variables (genes).
A list of class memberships, one per experiment. Each list can only contain 2 levels.
A logical value indicating whether or not to use a REM, TRUE, or a FEM, FALSE, for combining the z scores.
The function `zScores` implements the approach of Choi et al. for for a set of ExpressionSets. The function `zScorePermuted` applies `zScore` to a single permutation of the class labels. The function `zScoreFDR` computes a FDR for each gene, both for each single experiment and for the combined experiment. The FDR is calculated as described in Choi et al. Up to now ties in the zscores are not taken into account in the calculation. The function might produce incorrect results in that case. The function also computes zScores, both for the combines experiment and for each single experiment.

**Value**

A matrix with one row for each probe(set) and the following columns:

- `zSco_Ex_` For each single experiment the standardized mean difference, divided by the estimated standard deviation, the square root of the `EffectVar_Ex_` column.
- `MUvals` The combined standardized mean difference (using a FEM or REM).
- `MUsds` The standard deviation of the `MUvals`.
- `zSco` The z statistic - the `MUvals` divided by their standard deviations, `MUsds`.
- `Qvals` Cochran’s Q statistic for each gene.
- `df` The degree of freedom for the Chi-square distribution. This is equal to the number of combined experiments minus one.
- `Qpvalues` The probability that a Chi-square random variable, with `df` degrees of freedom) has a higher value than the value from the Q statistic.
- `Chisq` The probability that a Chi-square random variate (with 1 degree of freedom) has a higher value than the value of `zSco^2`.
- `Effect_Ex_` The standardized mean difference for each single experiment.
- `EffectVar_Ex_` The variance of the standardized mean difference for each single experiment.

Note that the three column names that end in an underscore are replicated, once for each experiment that is being analyzed.

**Author(s)**

M. Ruschhaupt

**References**

Choi et al, Combining multiple microarray studies and modeling interstudy variation. Bioinformatics, 2003, i84-i90.
Examples

data(Nevins)

##Splitting
thestatus <- Nevins$ER.status
group1 <- which(thestatus=="pos")
group2 <- which(thestatus=="neg")
rrr <- c(sample(group1, floor(length(group1)/2)),
sample(group2, ceiling(length(group2)/2)))
Split1 <- Nevins[,rrr]
Split2 <- Nevins[,-rrr]

#obtain classes
Split1.ER <- as.numeric(Split1$ER.status) - 1
Split2.ER <- as.numeric(Split2$ER.status) - 1

esets <- list(Split1,Split2)
classes <- list(Split1.ER,Split2.ER)
theScores <- zScores(esets,classes,useREM=FALSE)
theScores[1:2,]
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