# Package ‘GenoGAM’

December 21, 2016

**Type** Package  
**Title** A GAM based framework for analysis of ChIP-Seq data  
**Version** 1.2.1  
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**Description** This package allows statistical analysis of genome-wide data with smooth functions using generalized additive models based on the implementation from the R-package ‘mgcv’. It provides methods for the statistical analysis of ChIP-Seq data including inference of protein occupancy, and pointwise and region-wise differential analysis. Estimation of dispersion and smoothing parameters is performed by cross-validation. Scaling of generalized additive model fitting to whole chromosomes is achieved by parallelization over overlapping genomic intervals.

**License** GPL-2

**LazyData** true

**Depends** R (>= 3.3), Rsamtools (>= 1.18.2), SummarizedExperiment (>= 1.1.19), GenomicRanges (>= 1.23.16), methods

**Imports** BiocParallel (>= 1.5.17), data.table (>= 1.9.4), DESeq2 (>= 1.11.23), futile.logger (>= 1.4.1), GenomeInfoDb (>= 1.7.6), GenomicAlignments (>= 1.7.17), IRanges (>= 2.5.30), mgcv (>= 1.8), reshape2 (>= 1.4.1), S4Vectors (>= 0.9.34), Biostrings (>= 2.39.14)

**Suggests** BiocStyle, chipseq (>= 1.21.2), LSD (>= 3.0.0), genefilter (>= 1.54.2), ggplot2 (>= 2.1.0), testthat, knitr

**VignetteBuilder** knitr

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**biocViews** Regression, DifferentialPeakCalling, ChIPSeq, DifferentialExpression, Genetics, Epigenetics

**Collate** 'GenomicTiles-class.R' 'GenoGAMSettings-class.R' 'GenoGAM-class.R' 'GenoGAM-package.R' 'GenoGAMDataSet-class.R' 'cv.R' 'devel.R' 'diffBinding.R' 'filter.R' 'genogam.R' 'helper.R' 'peakCalling.R' 'plotting.R' 'qc.R' 'readData.R' 'sf.R'

**URL** [https://github.com/gstricker/GenoGAM](https://github.com/gstricker/GenoGAM)
BugReports  https://github.com/gstricker/GenoGAM/issues
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asDataFrame  GenomicTiles to DataFrame

Description
GenomicTiles to DataFrame

See Also
Other res: GenoGAMDataSetToDataFrame

callPeaks  Call peaks on a GenoGAM object

Description
Call narrow or broad peaks on the GenoGAM fit and computing significance, respectively

Usage
callPeaks(fit, smooth = NULL, range = NULL, peakType = c("narrow", "broad"), threshold = NULL, thresholdType = c("fdr", "pvalue"), maxgap = 500, cutoff = 0.05, minregion = 1)

Arguments

fit  A GenoGAM object
smooth  The name of the smooth, i.e. the 'by' variables in the GenoGAMDataSet design. By default the last one will be taken.
range  A GRanges object specifying a range. By default the complete fit is taken.
peakType  The type of the peak (narrow or broad). Default is narrow, see details.
threshold  The significance threshold. Keep in mind that the threshold depends on the thresholdType. By default this is 0.05 for 'pvalue' and 0.1 for 'fdr'.
thresholdType  The threshold type. Either 'fdr'(default) or 'pvalue'. If the threshold is not provided it, will be set accordingly to the thresholdType.
maxgap  For broad peaks only. The maximum gap between two broad peaks, that can be tolerated in order to identify both as part of one broad peak. All broad peaks with distances smaller or equal to the maxgap will be merged.
cutoff  A separate threshold for broad peaks. Since pointwise pvalues are available, this threshold is used to identify all significantly high positions, which then make up a broad peak.
minregion  For broad peaks only. The minimum length of a broad peak. By default 1, thus catching also narrow peaks.
changeSettings

Details

Note, that broad peaks don’t provide a specific highest location, but a region. Whereas narrow peaks provide both. However, the borders of narrow peaks are not necessarily informative. Additionally narrow peaks provide a 95% confidence interval for the position, namely 'start' and 'end', which gives a more informative uncertainty measure to the peak position. Also narrow peaks provide an occupancy estimate at the peak position, while broad peaks give the average occupancy across the region. The columns returned are:

Value

A data.table of identified peaks. The different columns loosely resemble the narrow and broad peak format (with different column order), such that it is easy to write them to a 'narrowPeak', 'broadPeak' file. See details for column description.

Author(s)

Georg Stricker <georg.stricker@in.tum.de>

Examples

load(system.file("extdata/Set1/fit.rda", package="GenoGAM"))
## calling narrow peaks
peaks <- callPeaks(fit, smooth = "genotype", threshold = 1)
peaks
## calling broad peaks
peaks <- callPeaks(fit, smooth = "genotype", threshold = 1,

changeSettings

Check data compliance with tile settings

Description

Check if the indices were build correctly, according to the specified parameters. This is the recommended way of changing tile settings, as it triggers instant recomputation of the index.

Usage

changeSettings(object, param, value)

## S4 method for signature 'GenomicTiles,character'
changeSettings(object, param, value)

Arguments

object A GenomicTiles object.
param The name of a tile settings parameter.
value An appropriate value. In most cases integer.
checkSettings

Value

A `GenomicTiles` object

Author(s)

Georg Stricker <georg.stricker@in.tum.de>

Examples

```
gt <- makeTestGenomicTiles()
gt2 <- changeSettings(gt, "chunkSize", 20)
```

---

checkSettings `Check data compliance with tile settings`

Description

Check if the indices were build correctly, according to the specified parameters

Usage

```
checkSettings(object)
```

## S4 method for signature 'GenomicTiles'

```
checkSettings(object)
```

Arguments

object `A GenomicTiles object`

Value

A logical value

Author(s)

Georg Stricker <georg.stricker@in.tum.de>

Examples

```
gt <- makeTestGenomicTiles()
checkSettings(gt)
```
computeRegionSignificance

*Compute significance for given regions*

**Description**

For a given set of regions, region-wise pvalues and FDR is computed.

**Usage**

```r
computeRegionSignificance(fit, regions, what = NULL)
```

**Arguments**

- `fit` A GenoGAM object containing the fit
- `regions` A GRanges object of regions of interest
- `what` Which fit should be used. The names should be equivalent to the column names used in the config file. Lookup with `names(colData(my_GenoGAMDataSet_object))`

**Details**

For a given set of regions, region-wise pvalues are computed by applying familywise hochberg correction and taking the minimal p-value. FDR is computed by further applying Benjamini-Hochberg correction.

**Value**

The GRanges object from the 'region' parameter extended by two columns: pvalue and FDR

**Author(s)**

Georg Stricker <georg.stricker@in.tum.de>

**Examples**

```r
gg <- makeTestGenoGAM()
gr <- GRanges("chrI", IRanges(1,100))
computeRegionSignificance(gg, gr)
```

---

computeSignificance

*Compute significance.*

**Description**

Based on the model fits this functions computes pointwise pvalues.

**Usage**

```r
computeSignificance(gg, log.p = FALSE)
```
computeSizeFactors

Arguments

  gg  A fitted GenoGAM object.
  log.p  Should pvalues be returned in log scale?

Value

A GenoGAM object which fits has been updated by the pvalue columns.

Author(s)

Georg Stricker <georg.stricker@in.tum.de>

Examples

ggd <- makeTestGenoGAM()
ggd <- computeSignificance(ggd)
head(getFits(ggd))

Description

The function computes the size factors for given factor groups based on the DESeq2 package.

Usage

computeSizeFactors(ggd, factorGroups = NULL)

Arguments

  ggd  A GenoGAMDataSet object.
  factorGroups  A list of grouped IDs (same as the colnames of the GenoGAMDataSet object). Each element of the list represents a group of samples within which size factors are computed. If NULL all samples are regarded to belong to one group. Size factors are not computed between groups.

Value

An updated GenoGAMDataSet object.

Author(s)

Georg Stricker <georg.stricker@in.tum.de>

Examples

ggd <- makeTestGenoGAMDataSet()
ggd <- computeSizeFactors(ggd)
dataRange

The GRanges of the underlying data

Description

Just like the coordinates slot but returns the genomic ranges of the underlying data.

Usage

```r
dataRange(object)
```

Arguments

- `object`: A GenomicTiles object.

Value

A GRanges object of genomic ranges of the underlying data

Author(s)

Georg Stricker <georg.stricker@in.tum.de>

Examples

```r
gt <- makeTestGenomicTiles()
dataRange(gt)
```

design,GenoGAMDataSet-method

Access the `design` slot

Description

The design slot contains the formula object which is used to fit the model.

Usage

```r
## S4 method for signature 'GenoGAMDataSet'
design(object)
```

```r
## S4 replacement method for signature 'GenoGAMDataSet,ANY'
design(object) <- value
```
filterData

Arguments

object: A GenoGAMDataSet object.
value: A formula object

Value

A formula object

Author(s)

Georg Stricker <georg.stricker@in.tum.de>

Examples

ggd <- makeTestGenoGAMDataSet()

```
filterData(ggd, windowsize = 201, mode = c("sum", "mean"))
```

Description

A function to filter the GenoGAMDataSet by the sum or mean of counts to significantly reduce the amount of models to compute

Usage

```
filterData(ggd, threshold = NULL, windowsize = 201, mode = c("sum", "mean"))
```

Arguments

ggd: A GenoGAMDataSet object
threshold: A value for the mean or sum of counts, which will be used to filter on basepair level. By default it is taken as median + 3*MAD
windowsize: The sliding window size. Should be an odd value.
mode: Should the sum or the mean of counts be used?

Value

A GenoGAMDataSet object containing the filtered regions

Author(s)

Georg Stricker <georg.stricker@in.tum.de>

Examples

ggd <- makeTestGenoGAMDataSet()

```
fdata <- filterData(ggd, windowsize = 10)
fdata
```
GenoGAM: A package providing a framework to analyse ChIP-Seq data

Description

GenoGAM: A package providing a framework to analyse ChIP-Seq data

Description

This is the fitting function for GenoGAMDataSet. It processes the data in GenoGAMDataSet, estimates the overdispersion and the penalization parameter, passes all information to mgcv::gam in parallel fashion and extracts the results. So far the model is restricted to Negativ Binomial distribution (mgcv::nb()).

Usage

```
genogam(ggd, lambda = NULL, family = mgcv::nb(), bpknots = 20, kfolds = 10, intervallSize = 20, m = 2)
```

Arguments

- `ggd`: A GenoGAMDataSet object to be fitted.
- `lambda`: The penalization parameter. Will be estimated if missing.
- `family`: A distribution family object. So far only mgcv::nb() is allowed.
- `bpknots`: Number of basepairs per one knot, that is, how dense should the knots be placed. The denser the knots, the more sensitive the fit. Note however, that computation time increases approximately cubic with every additional knot.
- `kfolds`: An integer number giving the number of k-folds to be used in cross validation, if parameters need to be estimated.
- `intervallSize`: The size of the intervalls to be used in cross validation. Short intervalls are used instead of single points to be left out due to spatial correlation. If replicates are present it is advised to make them bigger, e.g. 2*fragment size. Otherwise, depending on the density of the data, they should not exceed the size of a short read.
- `m`: The penalization order of the P-Splines.

Value

A GenoGAM object containing the fits and parameters.

Author(s)

Georg Stricker <georg.stricker@in.tum.de>
## Examples

```r
## Not run:
## simple example
config <- data.frame(ID = c("input", "IP"),
  file = c("myInput.bam", "myIP.bam"),
  paired = c(FALSE, FALSE),
  type = c(0,1), stringsAsFactors = FALSE)
bpk <- 100  ## basepairs per one knot
chunkSize <- 5000
overhang <- round(7*chunkSize/bpk)  ## overhang with 7 knots
knots <- chunkSize/bpk
## build the GenoGAMDataSet
gtiles <- GenoGAMDataSet(config = config, chunkSize = chunkSize, overhangSize = overhang,
  design = ~ s(x) + s(x, by = type))
gtiles <- computeSizeFactors(gtiles)
fits <- genogam(gtiles, bpknots = bpk)
## End(Not run)
```

---

### GenoGAM-class

**GenoGAM class**

This class is designed to represent the model object containing the estimate parameters, arguments and finals fits of the model on a basepair level.

### Slots

- **design** A mgcv-type formula object.
- **fits** A data.frame of the fits, the standard error and the first and second derivative of the fits for each experiment.
- **positions** A GPos object of the positions and seqnames corresponding to the rows in the 'fits' slot.
- **smooths** A data.frame of knot positions and base function coefficients, in order to reproduce the splines and compute derivatives.
- **vcov** A list of covariance matrices for each tile fit.
- **experimentDesign** The design matrix according to which the fitting was performed.
- **fitparams** Global parameters 'lambda', 'theta', 'Coefficient of Variation' and the 'penalty order' used to compute the model.
- **family** The distribution family.
- **cvparams** Parameters used for cross validation.
- **settings** The global and local settings that were used to compute the model.
- **tileSettings** A list of settings used to compute tiles.

### Author(s)

Georg Stricker <georg.stricker@in.tum.de>
Description
The different accessor functions for the GenoGAM object
The 'positions' slot holds the positions of the fit in GPos format
The 'design' slot holds the formula of the fit
The 'fits' slot contains the fitted values of the model
The 'experimentDesign' slot contains the experimental design of the model as specified in the config file

Usage

```r
## S4 method for signature 'GenoGAM'
rowRanges(x)

## S4 method for signature 'GenoGAM'
design(object)

getFits(x)

## S4 method for signature 'GenoGAM'
colData(x)
```

Arguments

- `x, object` A GenoGAM object.

Value

The respective slot

Author(s)

Georg Stricker <georg.stricker@in.tum.de>

Examples

```r
gg <- makeTestGenoGAM()
ranges <- rowRanges(gg)
gg <- makeTestGenoGAM()
des <- design(gg)
gg <- makeTestGenoGAM()
fits <- getFits(gg)
gg <- makeTestGenoGAM()
exdesign <- colData(gg)
```
GenoGAMDataSet

**GenoGAMDataSet constructor.**

**Description**

This is the constructor function for GenoGAMDataSet. So far a GenoGAMDataSet can be constructed from either an experiment design file or data.frame or directly from a RangedSummarizedExperiment with a GPos object being the rowRanges.

**Usage**

```
GenoGAMDataSet(experimentDesign, chunkSize, overhangSize, design, 
directory = ".", settings = NULL, ...)
```

**Arguments**

- **experimentDesign**
  Either a character object specifying the path to a delimited text file (the delimiter will be determined automatically), or a data.frame specifying the experiment design. See details for the structure of the experimentDesign.

- **chunkSize**
  An integer specifying the size of one chunk in bp.

- **overhangSize**
  An integer specifying the size of the overhang in bp. As the overhang is taken to be symmetrical, only the overhang of one side should be provided.

- **design**
  A mgcv-like formula object. See details for its structure.

- **directory**
  The directory from which to read the data. By default the current working directory is taken.

- **settings**
  A GenoGAMSettings object. This class is already present but not yet fully tested and therefore not accessible to the user. This argument exists however in order to allow some workarounds if necessary. See the vignette for a possible use.

- **...**
  Further parameters, mostly for arguments of custom processing functions or to specify a different method for fragment size estimation. See details for further information.

**Details**

The experimentDesign file/data.frame must contain at least three columns with fixed names: 'ID', 'file' and 'paired'. The field 'ID' stores a unique identifier for each alignment file. It is recommended to use short and easy to understand identifiers because they are subsequently used for labelling data and plots. The field 'file' stores the BAM file name. The field 'paired', values TRUE for paired-end sequencing data, and FALSE for single-end sequencing data. All other columns are stored in the colData slot of the GenoGAMDataSet object. Note that all columns which will be used for analysis must have at most two conditions, which are for now restricted to 0 and 1. For example, if the IP data should be corrected for input, then the input will be 0 and IP will be 1, since we are interested in the corrected IP. See examples.

Design must be a mgcv-like formula. At the moment only the following is possible: Either ~ 1 for a constant. ~ s(x) for a smooth fit over the entire data. s(x, by = "myColumn"), where 'myColumn' is a column name in the experimentDesign. This type of formula will then only fit the samples annotated with 1 in this column. Or ~ s(x) + s(x, by = "myColumn") + s(x, by = ...) + ... The last formula lets you combine any number of columns, given they are binary with 0 and 1. For example
the formula for correcting IP for input would look like this: \(~ s(x) + s(x, \text{by} = "\text{experiment}"),\) where 'experiment' is a column with 0s and 1s, with the IP samples annotated with 1 and input samples with 0. In case of single-end data it might be useful to specify a different method for fragment size estimation. The argument 'shiftMethod' can be supplied with the values 'coverage' (default), 'correlation' or 'SISSR'. See \texttt{?chipseq::estimate.mean.fraglen} for explanation.

Value

An object of class GenoGAMDataSet.

Author(s)

Georg Stricker <georg.stricker@in.tum.de>

Examples

```r
## Not run:
myConfig <- data.frame(ID = c("input","ip"),
                       file = c("myInput.bam", "myIP.bam"),
                       paired = c(FALSE, FALSE),
                       experiment = factor(c(0,1)),
                       stringsAsFactors = FALSE)
myConfig2 <- data.frame(ID = c("wildtype1","wildtype2",
                          "mutant1", "mutant2"),
                        file = c("myWT1.bam", "myWT2.bam",
                                 "myMutant1.bam", "myMutant2.bam"),
                        paired = c(FALSE, FALSE, FALSE, FALSE),
                        experiment = factor(c(0, 0, 1, 1)),
                        stringsAsFactors = FALSE)

gtiles <- GenoGAMDataSet(myConfig, chunkSize = 2000,
                          overhang = 250, design = ~s(x) + s(x, \text{by} = "experiment")

gtiles <- GenoGAMDataSet(myConfig2, chunkSize = 2000,
                          overhang = 250, design = ~s(x) + s(x, \text{by} = "experiment")

## End(Not run)
## make a test dataset
ggd <- makeTestGenoGAMDataSet()
ggd
```

---

GenoGAMDataSet-class  GenoGAMDataSet

Description

This class is designed to represent the input for the GenoGAM model. It extends the GenomicTiles class.

Details

For all other slots see SummarizedExperiment.
**GenoGAMDataSetToDataFrame**

**Description**
GenoGAMDataSet to DataFrame

**See Also**
Other res: asDataFrame

---

**GenoGAMSettings**

*The constructor function for GenoGAMSettings*

**Description**
The constructor function for GenoGAMSettings

**Usage**
GenoGAMSettings(...)

**Arguments**

... Any parameters corresponding to the slots and their possible values. See GenoGAM- Settings

**Value**
A GenoGAMSettings object.

**Author(s)**
Georg Stricker <georg.stricker@in.tum.de>
GenoGAMSettings-class  GenoGAMSettings

Description

This class is designed to store settings for the computation of the GenoGAM package

Details

Center can have three values: TRUE, FALSE, NULL. TRUE will trigger the center function, FALSE will trigger the use of the entire fragment. NULL should be used in case a custom process function is used.

Slots

center  A logical or NULL value to specify if the raw data should be centered, i.e. only the midpoint of the fragment will be used to represent its coverage. See details.

chromosomeList  A character vector of chromosomes to be used. NULL for all chromosomes.

bamParams  An object of class ScanBamParam. See ?Rsamtools::ScanBamParam.

parallel  A parallel backend of the respective class. See BiocParalell for the options

processFunction  A custom function on how to process raw data. Not used if center is TRUE/FALSE.

optimMethod  The optiomisation method to be used in cross validation.

optimControl  Settings for the optim() function.

Author(s)

Georg Stricker <georg.stricker@in.tum.de>

GenomicTiles constructor.

Description

This is the constructor function for GenomicTiles. The easiest construction is from SummarizedExperiment. However as the class operates on basepair level, the rowRanges are restricted to the GPos class.

Usage

GenomicTiles(assays, chunkSize = 10000, overhangSize = 0, ...)

Arguments

assays
One of two things. Either directly an object of type 'RangedSummarizedExperiment'. Or in case the object is created from raw data, a 'list' or 'SimpleList' of matrix-like elements, or a matrix-like object. All elements of the list must have the same dimensions, and dimension names (if present) must be consistent across elements and with the row names of 'rowRanges' and 'colData'.

chunkSize
An integer specifying the size of one chunk in bp.

overhangSize
An integer specifying the size of the overhang in bp. The overhang is regarded to be symmetric, such that only the overhang of one side should be provided.

Details

Most, but not necessary all functionalities of SummarizedExperiment are yet provided.

Value

An object of class GenomicTiles.

Author(s)

Georg Stricker <georg.stricker@in.tum.de>

Examples

## from raw data
gp <- GPos(GRanges(c("chrI", "chrII"), IRanges(c(1,1), c(5,5))))
assay <- matrix(1:10, 10, 1)
gt <- GenomicTiles(assay, chunkSize = 3, rowRanges = gp)

## from GenomicTiles
se <- SummarizedExperiment(assay, rowRanges = gp)
gt <- GenomicTiles(se, chunkSize = 3)

Description

This class is designed to represent the entire genome (or a subset of it) and any additional data associated with the samples or positions. It extends the RangedSummarizedExperiment class and adds two additional index slots to keep track of the data. The main change compared to RangedSummarizedExperiment is the use of a GPos (basepair level) instead of GRanges (ranges level) object as rowRanges and the use of two GRanges objects as indices. The GPos object allows to store raw instead of summarized data in the assays. Because of this the size of genomic data can increase tremendously. Thus the GenomicTiles class automatically divides the data in (overlapping) tiles, making any operation on this data easy executable in parallel.

Details

For all other slots see SummarizedExperiment.
Slots

index  A GRanges object that stores the tiles ranges and their index in the genome space. That is, ranges are the positions on the genome.
coordinates  A GRanges object that stores the tiles ranges and their index in the DataFrame space. That is ranges are the row positions in the DataFrame.

Author(s)

Georg Stricker <georg.stricker@in.tum.de>

---

getChromosomes  The single entries of the tile settings

Description

Returns the single elements of the tile settings

Usage

getChromosomes(object)

## S4 method for signature 'GenomicTiles'
getChromosomes(object)

getTileSize(object)

## S4 method for signature 'GenomicTiles'
getTileSize(object)

getChunkSize(object)

## S4 method for signature 'GenomicTiles'
getChunkSize(object)

getOverhangSize(object)

## S4 method for signature 'GenomicTiles'
getOverhangSize(object)

gTileNumber(object, ...)

## S4 method for signature 'GenomicTiles'
getTileNumber(object)

Arguments

object  A GenomicTiles object.
...
  Additional arguments
getChunkIndex

Value

An integer value, or in case of `getChromosomes` a `GRanges` object

Author(s)

Georg Stricker <georg.stricker@in.tum.de>

Examples

gt <- makeTestGenomicTiles()
getChromosomes(gt)
getTileSize(gt)
getChunkSize(gt)
getOverhangSize(gt)
getTileNumber(gt)

---

getChunkIndex  
*Compute the index for chunks instead tiles*

Description

The chunk index holds the Granges object that splits the entire dataset in chunk, that is non-overlapping intervals.

Usage

getChunkIndex(object, ...)

## S4 method for signature 'GenomicTiles'
getChunkIndex(object, id = NULL)

Arguments

object  
A `GenomicTiles` object.

...  
Additional arguments

id  
A vector if tile ids. By default the complete index is returned.

Value

A `GRanges` object representing the index

Author(s)

Georg Stricker <georg.stricker@in.tum.de>

Examples

gt <- makeTestGenomicTiles()
getChunkIndex(gt)
getCoordinates

Accessor to the `coordinates` slot

Description

The `coordinates` slot contains the row coordinates of each chromosome in the data. Such that taken a genomic position from a chromosome it’s easy to detect the correct row in the assay.

Usage

```r
getCoordinates(object)
```

Arguments

- `object`: A `GenomicTiles` object.

Value

A `GRanges` object of row coordinates

Author(s)

Georg Stricker <georg.stricker@in.tum.de>

Examples

```r
gt <- makeTestGenomicTiles()
getCoordinates(gt)
```

getIndex

Accessor to the `index` slot

Description

The index holds the Granges object that splits the entire dataset in tiles.

Usage

```r
getIndex(object, ...)
```

Arguments

- `object`: A `GenomicTiles` object.
- `...`: Additional arguments
- `id`: A vector if tile ids. By default the complete index is returned.
**Value**

A GRanges object representing the index

**Author(s)**

Georg Stricker <georg.stricker@in.tum.de>

**Examples**

```r
getIndexCoordinates(gt)
```

---

**getDescription**

**Compute the row coordinates for a given index**

**Description**

Given an index of genomic positions, this method computes the corresponding row positions in the assay

**Usage**

```r
getIndexCoordinates(object, ...)
```

## S4 method for signature 'GenomicTiles'

```r
getcIndexCoordinates(object, id = NULL, index = NULL)
```

**Arguments**

- `object`: A `GenomicTiles` object.
- `...`: Additional arguments Usually the original index or the chunk index.
- `id`: A vector if tile ids. By default the complete index is returned.
- `index`: A `Granges` object representing an index of genomic positions.

**Value**

A `GRanges` object of row coordinates

**Author(s)**

Georg Stricker <georg.stricker@in.tum.de>

**Examples**

```r
getIndexCoordinates(gt)
```
**makeTestGenoGAM**

**getTile**

*Tile extraction as a DataFrame*

**Description**

Extracting one or multiple tiles from a GenomicTiles object and coercing them to a DataFrameList.

**Usage**

```
getTile(object, id, ...)  
```

## S4 method for signature 'GenomicTiles'

```
getTile(object, id, size = 3e+09)
```

**Arguments**

- `object` A GenomicTiles object
- `id` A vector of tile ids
- `...` Additional arguments
- `size` The maximal number of rows that should be handled at once. If the dataset is bigger it will be processed in chunks. This is to lower memory consumption on big datasets, which in turn is slower.

**Value**

A SimpleDataFrameList

**Author(s)**

Georg Stricker <georg.stricker@in.tum.de>

**Examples**

```
gt <- makeTestGenomicTiles()
getTile(gt, 1:3)
```

**makeTestGenoGAM**

*Make an example codeGenoGAM*

**Description**

Make an example codeGenoGAM

**Usage**

```
makeTestGenoGAM()
```

**Value**

A codeGenoGAM object
makeTestGenoGAMDataSet

Examples

test <- makeTestGenoGAM()

Description

Make an example `GenoGAMDataSet`

Usage

makeTestGenoGAMDataSet()

Value

A `GenoGAMDataSet` object

Examples

test <- makeTestGenoGAMDataSet()

makeTestGenomicTiles

Description

Make an example `GenomicTile`

Usage

makeTestGenomicTiles()

Value

A `GenomicTiles` object

Examples

test <- makeTestGenomicTiles()
plot.GenoGAM  

The pot function for a GenoGAM object

Description

This function plots the fit of a given region and optionally the read counts from the GenoGAM-DataSet object.

Usage

plot.GenoGAM(x, ggd = NULL, ranges = NULL, seqnames = NULL, start = NULL, end = NULL, scale = TRUE, ...)

Arguments

x  
A GenoGAM object

ggd  
A GenoGAMDataSet object to plot raw counts

ranges  
A GRanges object specifying a particular region

seqnames  
A chromosome name. Together with start and end it is an alternative way of selecting a region

start  
The start of a region

d_end  
The end of a region

scale  
Logical, should all tracks be scaled to the same y-axis?

...  
Additional parameters that will be passed to the basic plot routine

Value

A plot of all tracks either using the ggplot2 or the base R framework

Author(s)

Georg Stricker <georg.stricker@in.tum.de>

qualityCheck  

A function to quality check the data

Description

This function checks some data attributes in the given class. Check details for more information.

Usage

qualityCheck(object, ...)

Arguments

object  
Any object for which this methods is implemented

...  
further parameters. See details.
Details

So far this method is only implemented for the class GenoGAMDataSet. In this case some general metrics are printed and some plots are stored in the folder "qc", which will be created in the working directory.

Additional parameters: factorGroups (for GenoGAMDataSet), which is used to specify factor groups for normalization plots. By default the groups will be identified automatically. See ?computeSizeFactors for parameter description.

Value

Based on the object provided, see details.

Author(s)

Georg Stricker <georg.stricker@in.tum.de>

sizeFactors,GenoGAMDataSet-method

Access the sizeFactor slot

Description

The sizeFactor slot contains the vector of normalization values for each sample

Usage

## S4 method for signature 'GenoGAMDataSet'
sizeFactors(object)

## S4 replacement method for signature 'GenoGAMDataSet,ANY'
sizeFactors(object) <- value

Arguments

object A GenoGAMDataSet object.
value A named numeric vector

Value

A named numeric vector

Author(s)

Georg Stricker <georg.stricker@in.tum.de>

Examples

```r
ggd <- makeTestGenoGAMDataSet()
sizeFactors(ggd)
sizeFactors(ggd) <- c(a = 5, b = 1/5)
```
Subset method for GenoGAM

Description
Subsetting the GenoGAM by a logical statement

Usage
```r
## S4 method for signature 'GenoGAM'
subset(x, ...)
```

Arguments
- `x` A GenoGAM object.
- `...` Further arguments. Mostly a logical statement. Note that the columnnames for chromosomes and positions are: seqnames and pos.

Value
A subsetted GenoGAM object.

Author(s)
Georg Stricker <georg.stricker@in.tum.de>

Examples
```r
gg <- makeTestGenoGAM()
subset(gg, pos <= 40)
```

Subset method for GenoGAMDataSet

Description
Subsetting the GenoGAMDataSet by a logical statement

Usage
```r
## S4 method for signature 'GenoGAMDataSet'
subset(x, ...)
```

Arguments
- `x` A GenoGAMDataSet object.
- `...` Further arguments. Mostly a logical statement. Note that the columnnames for chromosomes and positions are: seqnames and pos.
Value

A subsetted GenomicTiles object.

Author(s)

Georg Stricker <georg.stricker@in.tum.de>

Examples

```r
ggd <- makeTestGenoGAMDataSet()
res <- subset(ggd, seqnames == "chrI" & pos <= 50)
```

Description

Subsetting the GenomicTiles by a logical statement

Usage

```r
## S4 method for signature 'GenomicTiles'
subset(x, ...)
```

Arguments

- **x**: A GenomicTiles object.
- **...**: Further arguments. Mostly a logical statement. Note that the column names for chromosomes and positions are: seqnames and pos.

Value

A subsetted GenomicTiles object.

Author(s)

Georg Stricker <georg.stricker@in.tum.de>

Examples

```r
gt <- makeTestGenomicTiles()
res <- subset(gt, seqnames == "chrI" & pos <= 50)
```
subsetByOverlaps, GenoGAM, ANY-method

Subset by overlaps method for GenoGAM

Description

Subsetting the GenoGAM by a GRanges object

Usage

## S4 method for signature 'GenoGAM,ANY'
subsetByOverlaps(query, subject)

Arguments

query A GenoGAM object.
subject A GRanges object
...
Additional parameters

Value

A subsetted GenoGAM object.

Author(s)

Georg Stricker <georg.stricker@in.tum.de>

Examples

gg <- makeTestGenoGAM()
gr <- GRanges("chr1", IRanges(1,40))
subsetByOverlaps(gg, gr)

subsetByOverlaps, GenoGAMDataSet, GRanges-method

Subset by overlaps method for GenoGAMDataSet

Description

Subsetting the GenoGAMDataSet by a GRanges object

Usage

## S4 method for signature 'GenoGAMDataSet,GRanges'
subsetByOverlaps(query, subject,
maxgap = 0L, minoverlap = 1L, type = c("any", "start", "end", "within",
"equal"), ...)

...
Arguments

query A GenoGAMDataSet object.
subject A GRanges object
maxgap, minoverlap

Intervals with a separation of maxgap or less and a minimum of minoverlap
overlapping positions, allowing for maxgap, are considered to be overlapping.
maxgap should be a scalar, non-negative, integer. minoverlap should be a scalar,
positive integer.
type By default, any overlap is accepted. By specifying the type parameter, one
can select for specific types of overlap. The types correspond to operations in
Allen’s Interval Algebra (see references). If type is start or end, the intervals
are required to have matching starts or ends, respectively. While this operation
seems trivial, the naive implementation using outer would be much less effi-
cient. Specifying equal as the type returns the intersection of the start and
end matches. If type is within, the query interval must be wholly contained
within the subject interval. Note that all matches must additionally satisfy the
minoverlap constraint described above.
The maxgap parameter has special meaning with the special overlap types. For
start, end, and equal, it specifies the maximum difference in the starts, ends
or both, respectively. For within, it is the maximum amount by which the query
may be wider than the subject.

... Additional parameters

Value

A subsetted GenoGAMDataSet object.

Author(s)

Georg Stricker <georg.stricker@in.tum.de>

Examples

```r
ggd <- makeTestGenoGAMDataSet()
gr <- GRanges("chrI", IRanges(1,50))
res <- subsetByOverlaps(ggd, gr)
```

Description

Subsetting the GenomicTiles by a GRanges object

Usage

```r
## S4 method for signature 'GenomicTiles,GRanges'
subsetByOverlaps(query, subject, maxgap = 0L,
        minoverlap = 1L, type = c("any", "start", "end", "within", "equal"), ...)
```
Arguments

query A GenomicTiles object.
subject A GRanges object
maxgap, minoverlap Intervals with a separation of maxgap or less and a minimum of minoverlap overlapping positions, allowing for maxgap, are considered to be overlapping. maxgap should be a scalar, non-negative, integer. minoverlap should be a scalar, positive integer.
type By default, any overlap is accepted. By specifying the type parameter, one can select for specific types of overlap. The types correspond to operations in Allen’s Interval Algebra (see references). If type is start or end, the intervals are required to have matching starts or ends, respectively. While this operation seems trivial, the naive implementation using outer would be much less efficient. Specifying equal as the type returns the intersection of the start and end matches. If type is within, the query interval must be wholly contained within the subject interval. Note that all matches must additionally satisfy the minoverlap constraint described above.
The maxgap parameter has special meaning with the special overlap types. For start, end, and equal, it specifies the maximum difference in the starts, ends or both, respectively. For within, it is the maximum amount by which the query may be wider than the subject.

Value
A subsetted GenomicTiles object.

Author(s)
Georg Stricker <georg.stricker@in.tum.de>

Examples

gt <- makeTestGenomicTiles()
gr <- GRanges(c("chrI", "chrII"), IRanges(c(1, 120), c(40, 150)))
res <- subsetByOverlaps(gt, gr)

Summary,GenomicTiles-method

Computing metrics

Description
Computing metrics on each tile of the GenomicTiles object. So far all metrics from the Summary generics group, as well as mean, var, sd, median, mad and IQR are supported.
Summary.GenomicTiles-method

Usage

## S4 method for signature 'GenomicTiles'
Summary(x, ..., na.rm = FALSE)

## S4 method for signature 'GenomicTiles'
mean(x)

## S4 method for signature 'GenomicTiles,ANY'
var(x)

## S4 method for signature 'GenomicTiles'
sd(x)

## S4 method for signature 'GenomicTiles'
median(x)

## S4 method for signature 'GenomicTiles'
mad(x)

## S4 method for signature 'GenomicTiles'
IQR(x)

Arguments

x A GenomicTiles object
...
na.rm Should NAs be dropped. Otherwise the result is NA

Value

A list of as many elements as there are assays. Each element contains of a matrix with the specified metric computed per tile per column of the assay data.

Author(s)

Georg Stricker <georg.stricker@in.tum.de>

Examples

gt <- makeTestGenomicTiles()
sum(gt)
min(gt)
max(gt)
mean(gt)
var(gt)
sd(gt)
median(gt)
mad(gt)
IQR(gt)
tileSettings

Return tile settings

Description

Returns a list settings used to generate the tile index

Usage

tileSettings(object)

## S4 method for signature 'GenomicTiles'
tileSettings(object)

Arguments

object A GenomicTiles object.

Value

A list of tile settings

Author(s)

Georg Stricker <georg.stricker@in.tum.de>

Examples

gt <- makeTestGenomicTiles()
tileSettings(gt)

untile

Set index to chunkIndex

Description

Replace the tile index with the chunk index in GenomicTiles object

Usage

untile(object, ...)

## S4 method for signature 'GenomicTiles'
untile(object, id = NULL)

Arguments

object A GenomicTiles object.
... Additional arguments
id A vector if tile ids. By default the complete index is taken.
Value
A modified `codeGenomicTiles` object

Author(s)
Georg Stricker <georg.stricker@in.tum.de>

Examples
```
gt <- makeTestGenomicTiles()
newGT <- untile(gt)
```

Description
View the dataset

Usage
```
view(object, ...)
```  
```
## S4 method for signature 'GenomicTiles'
view(object, ranges = NULL, seqnames = NULL,
      start = NULL, end = NULL)
```

Arguments
```
object      A GenomicTiles object
...
ranges      A GRanges object. Makes it possible to select regions by GRanges. Either ranges or seqnames, start and end must be supplied
seqnames    A chromosomes name. Either ranges or seqnames, start and end must be supplied
start       A start site. Either ranges or seqnames, start and end must be supplied
end         An end site. Either ranges or seqnames, start and end must be supplied
```

Value
A data.frame of the selected data.

Author(s)
Georg Stricker <georg.stricker@in.tum.de>

Examples
```
gt <- makeTestGenomicTiles()
gr <- GRanges(c("chrI", "chrII"), IRanges(c(1, 10), c(40, 30)))
head(view(gt, ranges = gr))
head(view(gt, seqnames = "chrI", start = 1, end = 20))
```
view, GenoGAM-method  

View the dataset

Description

Cbinding the columns all together and coercing to data.frame

Usage

```r
## S4 method for signature 'GenoGAM'
view(object, ranges = NULL, seqnames = NULL,
     start = NULL, end = NULL)
```

Arguments

- `object`: A GenoGAM object
- `ranges`: A GRanges object. Makes it possible to select regions by GRanges. Either `ranges` or `seqnames`, `start` and `end` must be supplied
- `seqnames`: A chromosomes name. Either `ranges` or `seqnames`, `start` and `end` must be supplied
- `start`: A start site. Either `ranges` or `seqnames`, `start` and `end` must be supplied
- `end`: An end site. Either `ranges` or `seqnames`, `start` and `end` must be supplied

Value

A data.frame of the selected data.

Author(s)

Georg Stricker <georg.stricker@in.tum.de>

Examples

```r
gg <- makeTestGenoGAM()
gr <- GRanges("chrI", IRanges(1,40))
head(view(gg, gr))
```

writeToBEDFile  

Write peaks to BED6+3/4 format

Description

A function to write the data.table of peaks into a narrowPeaks or broadPeaks file

Usage

```r
writeToBEDFile(peaks, file = NULL)
```
Arguments

peaks A data.table or data.frame of peaks as produced by callPeaks()

file A file name without suffix. It will be determined automatically. If no file is given, it will be written to a generic `peaks_[timestamp]` file in the current working directory

Value

Nothing. A narrowPeaks or broadPeaks file written to 'file'

Author(s)

Georg Stricker <georg.stricker@in.tum.de>

Description

Providing subsetting by GRanges through the single-bracket operator

Usage

```r
## S4 method for signature 'GenoGAMDataSet,GRanges,ANY,ANY'
x[i]
```

Arguments

x A GenoGAMDataSet object

i A GRanges object

Value

A subsetted GenoGAMDataSet object

Description

Getting a specific tile

Usage

```r
## S4 method for signature 'GenomicTiles,numeric,ANY'
x[[i]]
```

```r
## S4 method for signature 'GenomicTiles,GRanges,ANY,ANY'
x[i]
```
Arguments

x  A GenomicTiles object
i  An integer (for '[i]') or a GRanges object (for 'i')

Value

A DataFrame (for '[i]') or a subsetted GenomicTiles object (for 'i')
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