## Package ‘LMGene’

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**Title** LMGene Software for Data Transformation and Identification of Differentially Expressed Genes in Gene Expression Arrays

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**Depends** R (>= 2.10.0), Biobase (>= 2.5.5), multtest, survival, affy

**Suggests** affydata

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**Description** LMGene package for analysis of microarray data using a linear model and glog data transformation

**License** LGPL

**URL** [http://dmrocke.ucdavis.edu/software.html](http://dmrocke.ucdavis.edu/software.html)

**biocViews** Microarray, DifferentialExpression, Preprocessing

**NeedsCompilation** no

### R topics documented:

<table>
<thead>
<tr>
<th>Topic</th>
<th>Page</th>
</tr>
</thead>
<tbody>
<tr>
<td>genediff</td>
<td>2</td>
</tr>
<tr>
<td>glog</td>
<td>4</td>
</tr>
<tr>
<td>LMGene</td>
<td>5</td>
</tr>
<tr>
<td>lnorm</td>
<td>6</td>
</tr>
<tr>
<td>lnormeS</td>
<td>7</td>
</tr>
<tr>
<td>neweS</td>
<td>8</td>
</tr>
<tr>
<td>norm</td>
<td>9</td>
</tr>
<tr>
<td>plotMeanSD</td>
<td>10</td>
</tr>
<tr>
<td>pmeans</td>
<td>11</td>
</tr>
<tr>
<td>pvadjust</td>
<td>12</td>
</tr>
<tr>
<td>rowaov</td>
<td>13</td>
</tr>
<tr>
<td>sample.eS</td>
<td>14</td>
</tr>
<tr>
<td>sample.ind</td>
<td>15</td>
</tr>
<tr>
<td>sample.mat</td>
<td>16</td>
</tr>
<tr>
<td>tranest</td>
<td>16</td>
</tr>
<tr>
<td>tranestAffyProbeLevel</td>
<td>19</td>
</tr>
<tr>
<td>transeS</td>
<td>21</td>
</tr>
<tr>
<td>vlist</td>
<td>23</td>
</tr>
</tbody>
</table>
**Description**

Computes two sets of p-values per gene or probe via gene-by-gene ANOVA, using both the gene-specific MSE and the posterior MSE for each term in the ANOVA. P-values are not adjusted for multiple testing.

Assumes a fixed effects model and that the correct denominator for all comparisons is the MSE.

**Usage**

```r
genediff(eS, model = NULL, method = c("MLE", "MOM", "MOMlog"), verbose = TRUE)
```

**Arguments**

- `eS`: An `ExpressionSet` object. Any transformation and normalization of `exprs(eS)` should be conducted prior to use in `genediff`.
- `model`: Model used for comparison; see details and `LMGene`.
- `method`: Method by which posterior p-values are calculated. Default "MLE".
- `verbose`: If TRUE, the prior degrees of freedom and mean reciprocal precision are printed. See details.

**Details**

The argument `eS` must be an `ExpressionSet` object from the Biobase package. If you have data in a matrix and information about experimental design factors, then you can use `neweS` to convert the data into an `ExpressionSet` object. Please see `neweS` for more detail.

The `model` argument is an optional character string, constructed like the right-hand side of a formula for `lm`. It specifies which of the variables in the `ExpressionSet` will be used in the model and whether interaction terms will be included. If `model=NULL`, it uses all variables from the `ExpressionSet` without interactions. Be careful of using interaction terms with factors; this often leads to overfitting, which will yield an error.

The method argument specifies how the adjusted MSE and degrees of freedom should be calculated for use in computation of the posterior p-values:

- "MLE": Default. Calculate adjusted MSE and degrees of freedom by maximum likelihood estimation, as described in Wright and Simon (2003).
- "MOM": Calculate adjusted MSE and degrees of freedom by method of moments, as described in Rocke (2003).
- "MOMlog": Calculate adjusted MSE and degrees of freedom by method of moments on log scale, as described in Smyth (2004). Uses functions `fitFdist` and `trigammainverse` from the package `limma`. Note that the method of Smyth (2004) is used here to calculate the posterior MSE, but not to directly calculate the posterior p-values.
All three methods assume that the gene-specific MSE’s follow a gamma distribution with mean tau. (NB: Notation and parameterization vary somewhat between each of the source papers.) The mean of the gamma distribution, tau, is modeled with an inverse gamma prior with hyperparameters alpha and beta. Empirical Bayes methods are used to estimate the prior hyperparameters, either by maximum likelihood, method of moments, or method of moments on the log scale. The "posterior MSE" is the posterior mean of the variances given the observed gene-specific MSE’s.

If verbose = TRUE, the function prints the estimated prior degrees of freedom, which equals twice the prior shape parameter alpha, and the estimated prior mean reciprocal precision, or 1/(alpha*beta).

All p-values are calculated from fixed-effects ANOVA F statistics, using either the gene-specific MSE or the posterior MSE as the denominator.

Value

A list with components:

Gene.Specific A matrix of p-values calculated using the gene-specific MSE, with one row for each gene/probe and one column for each factor

Posterior A matrix of p-values calculated using the posterior MSE, with one row for each gene/probe and one column for each factor

Author(s)

David Rocke, Geun-Cheol Lee, and Blythe Durbin-Johnson

References


http://dmrocke.ucdavis.edu

See Also

LMGene, rowaov, neweS

Examples

library(Biobase)
library(LMGene)

#data
data(sample.mat)
data(vlist)

raw.eS <- neweS(sample.mat, vlist)

# glog transform data
trans.eS <- transeS(raw.eS, lambda = 727, alpha = 56)
# calculate p-values
pvlist <- genediff(trans.eS)
pvlist$Posterior[1:5,]

---

**glog**

*Generalized log transformation function*

**Description**

This function transforms the input values by the generalized log function.

**Usage**

```r
glog(y, lambda)
```

**Arguments**

- `y`: A data matrix
- `lambda`: Transformation parameter

**Details**

The glog transformation of a variable `y` is defined as \(\log(y + \sqrt{y^2 + \lambda})\). Using \(\lambda = 0\) corresponds to the log transformation, up to a scale factor of 2. (Other, equivalent expressions exist for the glog transformation. See Durbin et al. (2002) and Huber et al. (2002) for further details.)

The input matrix `y` may be modified prior to transformation by subtracting a constant or vector ("alpha"). The parameters `lambda` and `alpha` may be estimated from `tranest`.

**Value**

- `yt`: A matrix of glog-transformed values

**Author(s)**

David Rocke and Geun-Cheol Lee

**References**

- [http://dmrocke.ucdavis.edu](http://dmrocke.ucdavis.edu)

**See Also**

- `tranest`
- `transeS`
Examples

```r
#library
library(Biobase)
library(LMGene)

#data
data(sample.mat)
sample.mat[1:5,1:4]

GloggedSmpd<-glog(sample.mat-50,500)
GloggedSmpd[1:5,1:4]
```

---

**LMGene**

**LMGene main function**

Description

LMGene calls function `genediff` to calculate the unadjusted gene-specific and posterior p-values of all genes and then calculates the FDR-adjusted p-values of all genes. Significant genes for each factor in `model` (based on either the gene-specific or posterior FDR-adjusted p-values) are output.

Usage

```r
LMGene(eS, model = NULL, level = 0.05, posterior = FALSE, method = c("MLE", "MOM", "MOMlog"))
```

Arguments

- **eS**: An ExpressionSet object. Any transformation and normalization of `exprs(eS)` should be conducted prior to use in `LMGene`.
- **model**: Specifies model to be used. Default is to use all variables from `eS` without interactions. See details.
- **level**: Significance level
- **posterior**: If `TRUE`, the posterior FDR-adjusted p-values are used in listing significant genes for each factor. Default is to use gene-specific FDR-adjusted p-values.
- **method**: Method by which the posterior p-values are calculated. Default is "MLE".

Details

If you have data in a matrix and information about experimental design factors, then you can use `neweS` to convert the data into an ExpressionSet object. Please see `neweS` for more detail.

The level argument indicates the False Discovery Rate, e.g. level=0.05 means a 5 percent FDR.

The model argument is an optional character string, constructed like the right-hand side of a formula for `lm`. It specifies which of the variables in the ExpressionSet will be used in the model and whether interaction terms will be included. If `model=NULL`, it uses all variables from the ExpressionSet without interactions. Be careful of using interaction terms with factors; this often leads to overfitting, which will yield an error.

See `genediff` for details of method.
Value

lmres A list with one component for each factor in model. Each component consists of a character vector with one element per significant gene. If no genes are significant for a given factor, the component for that factor is set to "No significant genes".

Author(s)

David Rocke and Geun-Cheol Lee

References


http://dmrocke.ucdavis.edu

See Also

genediff, neweS

Examples

library(Biobase)
library(LMGene)

#data
data(sample.mat)
data(vlist)

raw.eS <- neweS(sample.mat, vlist)

# glog transform data
trans.eS <- transeS(raw.eS, lambda = 727, alpha = 56)

# Identify significant genes, using an FDR of 1 percent
LMGene(trans.eS, level = 0.01)

---

Inorm

Lowess normalization function

Description

Lowess normalization function

Usage

lnorm(mat1, span = 0.1)

Arguments

mat1 A data matrix to be normalized
span Lowess smoother span. Larger values give more smoothness.
Details

mat1 must be a p by n matrix, where p is the number of genes and n is the number of arrays or samples.

Value

matnorm1 Normalized matrix

Author(s)

David Rocke and Geun-Cheol Lee

References

http://dmrocke.ucdavis.edu

See Also

lnormeS, norm

Examples

library(Biobase)
library(LMGene)

#data
data(sample.mat)
data(vlist)

raw.eS <- neweS(sample.mat, vlist)

# glog transform data
trans.eS <- transeS(raw.eS, lambda = 727, alpha = 56)

# normalize
normed.exprs <- lnorm(exprs(trans.eS))

lnormeS Function to apply lowess normalization to an expression set.

Description

Like lnorm, but applies to and returns an ExpressionSet or AffyBatch object instead of a matrix.

Usage

lnormeS(eS, span=0.1)

Arguments

eS An ExpressionSet or AffyBatch object
span Smoothing parameter for lowess. Larger values correspond to more smoothness.
neweS

Value

Returns an ExpressionSet with exprs(eS) normalized by lnorm.

Author(s)

John Tillinghast, Blythe Durbin-Johnson

References

http://dmrocke.ucdavis.edu

See Also

lnorm, norm

Examples

library(LMGene)
library(Biobase)
data(sample.eS)

# glog transform expression set
trsample.eS <- transeS(sample.eS, 667, 65)

# normalize expression set
normtrsample.eS <- lnormeS(trsample.eS)

---

neweS  

Coerce a matrix to class ExpressionSet

Description

This function converts a data matrix into an ExpressionSet object.

Usage

neweS(mat, vlist, vlabel = as.list(names(vlist)))

Arguments

mat  
A data matrix to be converted.

vlist  
A list, each component of which describes a factor in the experimental design.

vlabel  
A list of labels for each component of vlist.

Details

Each element of a component of vlist corresponds to a column of mat. See vlist for an example.

Value

eset  
An ExpressionSet object.
norm

Author(s)
David Rocke and Geun-Cheol Lee

References
http://dmrocke.ucdavis.edu

See Also
vlist

Examples
library(Biobase)
library(LMGene)

#data
data(sample.mat)
data(vlist)

Smpdt <- neweS(sample.mat,vlist)
data(sample.eS)
identical(exprs(sample.eS), exprs(Smpdt))
identical(pData(sample.eS), pData(Smpdt))

Description
This function normalizes a matrix by subtracting the column (sample) mean from each element and adding the grand mean.

Usage
norm(mat1)

Arguments
mat1 A matrix to be normalized

Value
matnorm Normalized matrix

Author(s)
David Rocke and Geun-Cheol Lee

References
http://dmrocke.ucdavis.edu
plotMeanSD

See Also

lnorm

Examples

library(Biobase)
library(LMGene)

#data
data(sample.mat)
data(vlist)

raw.eS <- neweS(sample.mat, vlist)

# glog transform data
trans.eS <- transeS(raw.eS, lambda = 727, alpha = 56)

# normalize
normed.exprs <- norm(exprs(trans.eS))

plotMeanSD(indata, by.rank = TRUE, line = FALSE, ymax = NULL)

Arguments

indata An object of class matrix, data.frame, ExpressionSet, or AffyBatch
by.rank If TRUE, the row standard deviations are plotted against the ranks of the row means. Otherwise, the row standard deviations are plotted against the row means themselves.
line If TRUE, a lowess smoother line is drawn on the plot.
ymax The upper limit for the plot y-axis. If missing, axis limits are generated automatically by plot.

Details

Generates a scatter plot of the row standard deviations of a matrix of expression data against the row means or ranks of the row means.

Value

NULL
Author(s)
Rachel Chen and Blythe Durbin-Johnson

Examples

```r
library(LMGene)
library(Biobase)

data(sample.eS)
# transform data
trans.eS <- transeS(sample.eS, lambda = 727, alpha = 56)

# plot SD against rank of mean
plotMeanSD(trans.eS, line = TRUE)
plotMeanSD(sample.eS, line = TRUE, ymax = 1000)
```

psmeans  Function to take means of probesets.

Description

Converts an `ExpressionSet` or `AffyBatch` object with one row of expression data per probeset into an `ExpressionSet` or `AffyBatch` object with one row per probe.

Usage

```r
psmeans(eS, ind)
```

Arguments

- `eS` An `ExpressionSet` or `AffyBatch` object
- `ind` A vector used to indicate which probes go into which probesets.

Details

Each entry of `ind` corresponds to one probe and tells the number of the probeset it belongs to. See `tranestAffyProbeLevel` and `sample.ind` for examples.

Value

Returns an `ExpressionSet` or `AffyBatch` object with the expression matrix rows corresponding to probesets instead of individual probes. Elements of the returned `ExpressionSet` or `AffyBatch` object are means over each probeset.

Author(s)

John Tillinghast

See Also

`tranestAffyProbeLevel`, `sample.ind`
pvadjust

Examples

```r
library(LMGene)
library(Biobase)

data(sample.eS)
data(sample.ind)

# glog transform data
trs.eS <- transeS(sample.eS, 667, 65)

# lowess normalize
ntrs.eS <- lnormeS(trs.eS)

# take means over probesets
genesample.eS<- psmeans(ntrs.eS, sample.ind)
```

pvadjust  

**P-value adjusting function**

Description

This function converts the given raw p-values into the FDR adjusted p-values using R package 'multtest'.

Usage

`pvadjust(pvlist)`

Arguments

pvlist  
A list containing raw p-values

Details

pvlist is the output from genediff containing p-values from gene-specific MSE's and posterior MSE's.

Value

pvlist2  
A list with the raw p-values and the newly computed FDR adjusted p-values

Author(s)

David Rocke and Geun-Cheol Lee

References

David M. Rocke (2004), Design and analysis of experiments with high throughput biological assay data, Seminars in Cell & Developmental Biology, 15, 703-713.

http://www.idav.ucdavis.edu/~dmrocke/
rowaov

**See Also**

genediff

**Examples**

```r
#library
library(Biobase)
library(LMGene)

#data
data(sample.mat)
data(vlist)
LoggedSmpd0<-neweS(lnorm(log(sample.mat)),vlist)

pvlist<-genediff(LoggedSmpd0)
pvlist$Posterior[1:5,]

apvlist<-pvadjust(pvlist)
names(apvlist)
apvlist$Posterior.FDR[1:5,]
```

---

**rowaov**

*Gene by gene ANOVA function*

**Description**

Computes the mean squares and degrees of freedom for gene-by-gene ANOVAs.

**Usage**

```r
rowaov(eS, model=NULL)
```

**Arguments**

- **eS**
  - An ExpressionSet object. Any transformation and normalization of eS should be done prior to use in rowaov.

- **model**
  - Model used for comparison. See details and LMGene.

**Details**

If you have data in a matrix and information about experimental design factors, then you can use neweS to convert the data into an ExpressionSet object. Please see neweS for more detail.

The model argument is an optional character string, constructed like the right-hand side of a formula for lm. It specifies which of the variables in the ExpressionSet will be used in the model and whether interaction terms will be included. If model=NULL, it uses all variables from the ExpressionSet without interactions. Be careful of using interaction terms with factors; this often leads to overfitting, which will yield an error.
Value

```r
resmat
```
A matrix of MSEs and degrees of freedom for all model factors and all genes. The first rows of `resmat` contain MSE’s for each effect in model, ending with the residual MSE. The remaining rows contain degrees of freedom for each effect in the model, ending with the residual d.f. Each column corresponds to a gene.

Author(s)

David Rocke and Geun-Cheol Lee

References


[http://dmrocke.ucdavis.edu](http://dmrocke.ucdavis.edu)

See Also

`genediff`, `LMGene`

Examples

```r
library(Biobase)
library(LMGene)

# data
data(sample.mat)
data(vlist)

raw.eS <- neweS(sample.mat, vlist)

# glog transform data
trans.eS <- transeS(raw.eS, lambda = 727, alpha = 56)

# Perform gene-by-gene anova
resmat <- rowaov(trans.eS)
resmat[,1:3]
```

---

**sample.eS**  
*Sample array data for LMGene*

Description

Sample ExpressionSet class data.

Usage

```r
data(sample.eS)
```

Format

Formal class ExpressionSet [package Biobase].
Details

Identical with `neweS(sample.mat, vlist)`, up to metadata

Examples

```r
library(Biobase)
library(LMGene)

#data
data(sample.mat)
data(vlist)

Smpdt <- neweS(sample.mat, vlist)

data(sample.eS)
identical(exprs(sample.eS), exprs(Smpdt))
identical(pData(sample.eS), pData(Smpdt))
```

---

**sample.ind**  
*Sample probeset index vector*

Description

Vector indicating which probeset each probe belongs to

Usage

```r
data(sample.ind)
```

Format

A vector of integers, e.g., c(1,1,1,2,2,3,3,3,4,4,...). Length is equal to the number of probes (rows) in `sample.mat`.

Examples

```r
data(sample.eS)
data(sample.ind)
trs.eS <- transeS(sample.eS, 667, 65)
ntrs.eS <- lnormeS(trs.eS)
genesample.eS <- psmeans(ntrs.eS, sample.ind)
```
sample.mat  
*Sample array data for LMGene package*

---

**Description**

A matrix of array data

**Usage**

```r
data(sample.mat)
```

**Format**

A matrix measuring 613 rows (probes) by 32 columns (samples).

**Examples**

```r
library(Biobase)
library(LMGene)

#data
data(sample.mat)
data(vlist)

Smpdt<-neweS(sample.mat,vlist)

data(sample.es)
identical(exprs(sample.es), exprs(Smpdt))
identical(pData(sample.es), pData(Smpdt))
```

---

**tranest**  
*Glog transformation parameter estimation function*

---

**Description**

Estimates parameters for the glog transformation, by maximum likelihood or by minimizing the stability score.

**Usage**

```r
tranest(es, ngenes = -1, starting = FALSE, lambda = 1000, alpha = 0, 
gradtol = 1e-3, lowessnorm = FALSE, method=1, mult=FALSE, model=NULL, 
SD = FALSE, rank = TRUE, model.based = TRUE, rep.arrays = NULL)
```
Arguments

- **eS**: An ExpressionSet object
- **ngen**es**: Number of genes to be used in parameter estimation. Default is to use all genes unless there are more than 100,000, in which case a subset of 50,000 genes is selected at random.
- **starting**: If TRUE, user-specified starting values for lambda and alpha are input to the optimization routine.
- **lambda**: Starting value for parameter lambda. Ignored unless starting = TRUE.
- **alpha**: Starting value for parameter alpha. Ignored unless starting = TRUE.
- **gradtol**: A positive scalar giving the tolerance at which the scaled gradient is considered close enough to zero to terminate the algorithm.
- **lowessnorm**: If TRUE, lowess normalization (using lnorm) is used in calculating the likelihood.
- **method**: Determines optimization method. Default is 1, which corresponds to a Newton-type method (see nlm and details.)
- **mult**: If TRUE, tranest will use a vector alpha with one (possibly different) entry per sample. Default is to use same alpha for every sample. SD and mult may not both be TRUE.
- **model**: Specifies model to be used. Default is to use all variables from eS without interactions. See details.
- **SD**: If TRUE, transformation parameters are estimated by minimizing the stability score rather than by maximum likelihood. See details.
- **rank**: If TRUE, the stability score is calculated by regressing the replicate standard deviations on the ranks of the gene/row means (rather than on the means themselves). Ignored unless SD = TRUE.
- **model.based**: If TRUE, the stability score is calculated using the standard deviations of residuals from the linear model in model. Ignored unless SD = TRUE.
- **rep.arrays**: List of sets of replicate arrays. Each element of rep.arrays should be a vector with entries corresponding to arrays (columns) in exprs(eS) conducted under the same experimental conditions, i.e., with identical rows in pData(eS). Ignored unless SD = TRUE and model.based = FALSE.

Details

If you have data in a matrix and information about experimental design factors, then you can use neweS to convert the data into an ExpressionSet object. Please see neweS for more detail.

The model argument is an optional character string, constructed like the right-hand side of a formula for lm. It specifies which of the variables in the ExpressionSet will be used in the model and whether interaction terms will be included. If model=NULL, it uses all variables from the ExpressionSet without interactions. Be careful of using interaction terms with factors; this often leads to overfitting, which will yield an error.

The default estimation method is maximum likelihood. The likelihood is derived by assuming that there exist values for lambda and alpha such that the residuals from the linear model in model, fit to glog-transformed data using those values for lambda and alpha, follow a normal distribution. See Durbin and Rocke (2003) for details.

If SD = TRUE, lambda and alpha are estimated by minimizing the stability score rather than by maximum likelihood. The stability score is defined as the absolute value of the slope coefficient from the regression of the replicate/residual standard deviation on the gene/row means, or on the...
rank of the gene/row means. If `model.based = TRUE`, the stability score is calculated using the standard deviation of residuals from the linear model in `model`. Otherwise, the stability score is calculated using the pooled standard deviation over sets of replicates in `rep.arrays`. See Wu and Rocke (2009) for details.

Optimization methods in `method` are as follows:

1 = Newton-type method, using `nlm`
2 = Nelder-Mead, using `optim`
3 = BFGS, using `optim`
4 = Conjugate gradients, using `optim`
5 = Simulated annealing, using `optim` (may only be used when `mult = TRUE`)

**Value**

A list with components:

- `lambda` Estimate of transformation parameter lambda
- `alpha` Estimate of transformation parameter alpha

**Author(s)**

David Rocke, Geun-Cheol Lee, John Tillinghast, Blythe Durbin-Johnson, and Shiquan Wu

**References**


[http://dmrocke.ucdavis.edu](http://dmrocke.ucdavis.edu)

**See Also**

`tranestAffyProbeLevel`, `lnorm`, `glog`

**Examples**

```r
library(Biobase)
library(LMGene)

#data
data(sample.eS)

tranpar <- tranest(sample.eS, 100)
tranpar
tranpar <- tranest(sample.eS, mult=TRUE)
tranpar
```
tranestAffyProbeLevel  Glog transformation parameter estimation function for probe-level Affymetrix expression data

Description

Estimates parameters for the glog transformation on probe-level Affymetrix expression data, by maximum likelihood or by minimizing the stability score.

Usage

tranestAffyProbeLevel(eS, ngenes = 5000, starting = FALSE, lambda = 1000, alpha = 0, gradtol = 0.001, lowessnorm = FALSE, method = 1, mult = FALSE, model = NULL, SD = FALSE, rank = TRUE, model.based = TRUE, rep.arrays = NULL)

Arguments

eS  An AffyBatch object
ngenes Number of randomly sampled probesets to be used in estimating the transformation parameter
starting If TRUE, user-specified starting values for lambda and alpha are input to the optimization routine
lambda Starting value for parameter lambda. Ignored unless starting = TRUE
alpha Starting value for parameter alpha. Ignored unless starting = TRUE
gradtol A positive scalar giving the tolerance at which the scaled gradient is considered close enough to zero to terminate the algorithm
lowessnorm If TRUE, lowess normalization (using lnorm) is used in calculating the likelihood.
method Determines optimization method. Default is 1, which corresponds to a Newton-type method (see nlm and details.)
mult If TRUE, tranest will use a vector alpha with one (possibly different) entry per sample. Default is to use same alpha for every sample. SD and mult may not both be TRUE.
model Specifies model to be used. Default is to use all variables from eS without interactions. See details.
SD If TRUE, transformation parameters are estimated by minimizing the stability score. See details.
rank If TRUE, the stability score is calculated by regressing the replicate standard deviation on the rank of the probe/row means (rather than on the means themselves). Ignored unless SD = TRUE
model.based If TRUE, the stability score is calculated using the standard deviation of residuals from the linear model in model. Ignored unless SD = TRUE
rep.arrays List of sets of replicate arrays. Each element of rep.arrays should be a vector with entries corresponding to arrays (columns) in exprs(eS) conducted under the same experimental conditions, i.e., with identical rows in pData(eS). Ignored unless SD = TRUE and model.based = FALSE
Details

The `model` argument is an optional character string, constructed like the right-hand side of a formula for `lm`. It specifies which of the variables in the `ExpressionSet` will be used in the model and whether interaction terms will be included. If `model=NULL`, it uses all variables from the `ExpressionSet` without interactions. Be careful of using interaction terms with factors; this often leads to overfitting, which will yield an error.

The default estimation method is maximum likelihood. The likelihood is derived by assuming that there exist values for `lambda` and `alpha` such that the residuals from the linear model in `model`, fit to glog-transformed data using those values for `lambda` and `alpha`, follow a normal distribution. See Durbin and Rocke (2003) for details.

If `SD = TRUE`, `lambda` and `alpha` are estimated by minimizing the stability score rather than by maximum likelihood. The stability score is defined as the absolute value of the slope coefficient from the regression of the replicate/residual standard deviation on the probe/row means, or on the rank of the probe/row means. If `model.based = TRUE`, the stability score is calculated using the standard deviation of residuals from the linear model in `model`. Otherwise, the stability score is calculated using the pooled standard deviation over sets of replicates in `rep.arrays`. See Wu and Rocke (2009) for details.

A random sample of probsets (of size `ngene`) is sampled from `featureNames(eS)`. Expression data from all probes in the sampled probsets is used in estimating the transformation parameters. Optimization methods in `method` are as follows:

1 = Newton-type method, using `nlm`
2 = Nelder-Mead, using `optim`
3 = BFGS, using `optim`
4 = Conjugate gradients, using `optim`
5 = Simulated annealing, using `optim` (may only be used when `mul = TRUE`)

Value

A list with components:

- `lambda` Estimate of transformation parameter lambda
- `alpha` Estimate of transformation parameter alpha

Author(s)

Lei Zhou, David Rocke, Geun-Cheol Lee, John Tillinghast, Blythe Durbin-Johnson, and Shiquan Wu

References


http://dmrocke.ucdavis.edu
See Also

tranest, lnorm, psmeans, glog

Examples

```r
library(LMGene)
library(affy)
library(Biobase)
library(affydata)

data(Dilution)

tranpar.Dilution <- tranestAffyProbeLevel(Dilution, model = "liver",
ngenex = 3000, method = 2)

# transform data
trans.Dilution <- tranS(Dilution, tranpar.Dilution$lambda,
tranpar.Dilution$alpha)

# extract transformed perfect matches
exprs(trans.Dilution) <- pm(trans.Dilution)

# lowess normalize transformed data
inorm.Dilution <- inormeS(trans.Dilution)

## Not run:
# Average over probesets
# First, create index of probes
fnames <- featureNames(Dilution)
p <- length(featureNames(Dilution))
ind <- vector()
for (i in 1:p){
nprobes <- dim(pm(Dilution,fnames[i]))[1]
ind <- c(ind, rep(i,nprobes))
}

avg.Dilution <- psmeans(lnorm.Dilution, ind)

## End(Not run)
```

**tranS**

"Function to apply the glog transform to an expression set."

**Description**

For each element in the array of expression data, this function applies the glog transform $y \rightarrow \text{glog} (y - \alpha, \lambda)$. If $\alpha$ is a vector, it must have one element for each column in exprs(eS).

**Usage**

```
tranS(eS, lambda, alpha)
```
Arguments

- **eS**: An ExpressionSet or AffyBatch object
- **lambda**: The parameter lambda to be used in the glog transform.
- **alpha**: The alpha parameter(s) for the glog transform. May be a single number used for all samples, or a vector with one entry per sample.

Details

The glog transformation of a variable y is defined as \( \log(y + \sqrt{y^2 + \lambda}) \). Using \( \lambda = 0 \) corresponds to the log transformation, up to a scale factor of 2. (Other, equivalent expressions exist for the glog transformation. See Durbin et al. (2002) and Huber et al. (2002) for further details.)

\( \text{trnseS} \) subtracts a (scalar or vector) parameter \( \alpha \) prior to application of the glog transformation, resulting in the expression \( \log(y - \alpha + \sqrt{(y - \alpha)^2 + \lambda}) \).

The parameters \( \lambda \) and \( \alpha \) may be estimated using \text{tranest}.

Value

Returns an ExpressionSet or AffyBatch object with the expression matrix glog-transformed.

Author(s)

John Tillinghast

References

- [http://dmrocke.ucdavis.edu](http://dmrocke.ucdavis.edu)

See Also

- \text{glog}, \text{tranest}

Examples

```r
library(LMGene)
library(Biobase)

data(sample.eS)
trsample.eS <- transeS(sample.eS, 667, 65)
```
Sample experimental/phenotype data for LMGene package

Description

List of experimental factors for the sample matrix array data, 'sample.mat'.

Usage

data(vlist)

Examples

library(Biobase)
library(LMGene)

#data
data(vlist)

vlist
Index

*Topic datasets
  sample.eS, 14
  sample.ind, 15
  sample.mat, 16
  vlist, 23

*Topic dplot
  plotMeanSD, 10

*Topic manip
  neweS, 8
  psmeans, 11

*Topic math
  glog, 4
  tranest, 16
  tranestAffyProbeLevel, 19
  traneS, 21

*Topic models
  genediff, 2
  pvadjust, 12
  rowaov, 13

*Topic smooth
  lnorm, 6
  lnormeS, 7
  genediff, 2, 5, 6, 13, 14
  glog, 4, 18, 21, 22

LMGene, 2, 3, 5, 13, 14
lnorm, 6, 7, 8, 10, 17–19, 21
lnormeS, 7, 7
neweS, 2, 3, 5, 6, 8, 13, 17
norm, 7, 8, 9

plotMeanSD, 10
psmeans, 11, 21
pvadjust, 12
rowaov, 3, 13

sample.eS, 14
sample.ind, 11, 15
sample.mat, 16

tranest, 4, 16, 21, 22
tranestAffyProbeLevel, 11, 18, 19