Package ‘PING’

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CoverageTrack  

**Description**

This track displays the coverage of the genomic region by the reads used as input for the segmentation step.

**Usage**

CoverageTrack(ping, reads, chr, gen="gen", FragmentLength=200, name="XSET")

**Arguments**

- **ping**: An object of class **pingList**, the output of PING.
- **reads**: An object of class **GRanges**, the reads used in the segmentation process.
- **chr**: A character string for the chromosome name.
- **gen**: An optional character string for the genome name.
- **FragmentLength**: An integer. The fixed length of the reads. If PE is set to TRUE, this argument can be ignored.
- **name**: A character string. The title of the track.

**Value**

This function returns an object of class **AnnotationTrack**.

**Author(s)**

Renan Sauteraud

**See Also**

postPING, plotSummary, AnnotationTrack, plotTracks

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NucleosomeTrack  

**Description**

This track actually creates two tracks: it displays the position of the predicted nucleosomes as well as the standard error associated with each prediction and an histogram of the score.

**Usage**

NucleosomeTrack(PS, chr, gen="gen", scoreTrack=TRUE, scoreThreshold=0.05, name="PING", ...)

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Track for the Nucleosome position
Arguments

- **PS**: A data.frame or `pingList`. The output of `postPING` or `PING`.
- **chr**: A character string for the chromosome name.
- **gen**: An optional character string for the genome name.
- **scoreTrack**: A logical. If set to FALSE, the track showing the histogram of the score is removed.
- **scoreThreshold**: A numeric. Nucleosomes predicted with a score inferior to this threshold are removed.
- **name**: A character string for the title of the track.
- **...**: A list of supplemental argument that can be provided to the `AnnotationTrack` track constructor.

Value

This function returns a list containing an object of class `AnnotationTrack` and a `DataTrack` if `scoreTrack` is set to TRUE.

Author(s)

Renan Sauteraud

See Also

- `postPING`
- `plotSummary`
- `DataTrack`
- `AnnotationTrack`
- `plotTracks`

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**Description**

Functions imported from PICS and exposed when loading PING. Refer to PICS documentation for further information.

- `?PICS::segReads`
- `?PICS::segReadsList`
- `?PICS::segReadsPE`
- `?PICS::segReadsListPE`
- `?PICS::setParaEM`
- `?PICS::setParaPrior`
- `?PICS::bam2gr`

See Also

- `makeRangedDataOutput`
- `segReads`
- `segReadsList`
- `setParaEM`
- `setParaPrior`
- `segReadsPE`
- `segReadsListPE`
ping

Estimation of binding site positions

Description
This object contains Estimation of binding site positions and has the following slots: segReadsList.

Usage
PING(segReadsList, paraEM=NULL, paraPrior=NULL, dataType="MNase", detail=0, rescale=1, nCores=1)

Arguments
segReadsList
This object contains segmentation of Genome

paraEM
A list of parameters for the EM algorithm. The default parameters should be good enough for most usages.

- minK: an integer, default=0. The minimum number of binding events per region. If the value is 0, the minimum number is automatically calculated.
- maxK: an integer, default=0. The maximum number of binding events per region. If the value is 0, the maximum number is automatically calculated.
- tol: a numeric, default=le-4. The tolerance for the EM algorithm.
- B: an integer, default=100. The maximum number of iterations to be used.
- mSelect: a character string specifying the information criteria to be used when selecting the number of binding events. Default="AIC3"
- mergePeaks: a logical stating whether overlapping binding events should be picked. Default=TRUE
- mapCorrect: a logical stating whether mappability profiles should be incorporated in the estimation, i.e: missing reads estimated. Default=TRUE

paraPrior
A list of parameters for the prior distribution. The default parameters should be good enough for most usages.

- xi: an integer, default=150. The average DNA fragment size.
- rho: an integer, default=1.2. A variance parameter for the average DNA fragment size distribution.
- alpha: an integer, default=10. First hyperparameter of the inverse Gamma distribution for sigma^2 in the PICS model
- beta: an integer, default=20000. Second hyperparameter of the inverse Gamma distribution for sigma^2 in the PING model
- lambda: an integer, default=0.000064. The precision of the prior for mu used for histone data.
- dMu: an integer, default=200. Our best guess for the distance between two neighboring nucleosomes.

dataType
A character string that can be set to use selected default parameters for the algorithm.

detail
An integer. Additional information are printed if set to a value > 0.

rescale
An integer.

nCores
An integer. The number of cores that should be used in parallel by the function.
Methods

- **code** signature(x = "ping"): return the error code for each list element (i.e. candidate region) of a PING object. If the string is empty, there were no errors.
- **plot** signature(x = "ping"): Plot all regions in the PING object. This might be long, and should only be used to plot a few regions, so subset the object before plotting.
- **sigmaSqR** signature(x = "ping"): return the variance parameter of the reverse (R) distribution for each binding event.
- **sigmaSqF** signature(x = "ping"): return the variance parameter of the forward (F) distribution for each binding event.
- **score** signature(x = "ping"): return the score for each binding event.
- **scoreF** signature(x = "ping"): return the score of the forward (F) for each binding event.
- **scoreR** signature(x = "ping"): return the score of the forward (R) for each binding event.
- **maxRange** signature(x = "ping"): return the range maximum.
- **minRange** signature(x = "ping"): return the range minimal.
- **K** signature(x = "ping"): subset PING object.
- **density** signature(x = "ping"): return the density for each binding event.

Author(s)

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Sangsoon Woo, <swoo@fhcrc.org>

See Also

- ping

ping-class  The ping class

Description

This object is used to gather all parameters from fitting PING to a single candidate region. The objet contains the following slots: 'estimates', 'infMat', 'Nmerged', 'converge', 'chr'. 'estimates' is a list containing all parameters estimates as well as standard errors. 'infMat' is the Cholesky decomposition of the information matrix, 'converge' is a logical value indicating whether the EM algorithm has converged, while 'chr' is a character string corresponding to a candidate region's chromosome. 'Nmerged' gives the number of binding events that were merged; binding events that overlap are merged (see the cited paper below for details).

Accessors

The PING package provide accessors to directly access to most of the parameters/standard errors and chromosome. In the code snippets below, 'x' is a 'ping' object.

- **`chromosome(x)`** Gets the chromosome name of the candidate region.
- **`mu(x)`** Gets the position estimates of all binding sites identified in the region.
- **`delta(x)`** Gets the average fragment lengths of all binding sites identified in the region.
‘`sigmaSqF(x)`’ Gets the F peak variances of all binding sites identified in the region.
‘`sigmaSqR(x)`’ Gets the R peak variances of all binding sites identified in the region.
‘`se(x)`’ Gets the standard errors of all binding site position estimates identified in the region.
‘`seF(x)`’ Gets the standard errors of all F peak modes identified in the region.
‘`seR(x)`’ Gets the standard errors of all R peak modes identified in the region.

`score` signature(x = "ping"): return the score for each binding event.
`scoreF` signature(x = "ping"): return the score of the forward (F) for each binding event.
`scoreR` signature(x = "ping"): return the score of the forward (R) for each binding event.

Constructor

`newPing(w, mu, delta, sigmaSqF, sigmaSqR, seMu, seMuF, seMuR, score, Nmerged, converge, infMat, chr)`
construct a new ’ping’ object with the following arguments:

`w` The mixture weights (a vector)
`mu` The binding site positions (a vector)
`delta` The DNA fragment lengths (a vector)
`sigmaSqF` The variance parameters for the forward distribution (vector)
`sigmaSqR` The variance parameters for the forward distribution (vector)
`seMu` The standard errors for mu (vector)
`seMuF` The standard errors for muF (vector)
`seMuR` The standard errors for muR (vector)
`score` The scores for each binding event (vector)
`Nmerged` The number of peaks that got merged (integer)
`converge` A logical value, TRUE, if the EM as converged
`infMat` The information matrix
`chr` The chromosome for the region

Author(s)

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References

Xuekui Zhang, Gordon Robertson, Sangsoon Woo, Brad G. Hoffman, and Raphael Gottardo, "Probabilistic Inference for Nucleosome Positioning with MNase-based or Sonicated Short-read Data” GenomeBiology, under review.

See Also

ping pingError
Examples

# Here is an example of how to construct such a region.
# Typically, you would not do this manually, you would use the ping function to return a 'pingList'
# that contains a list of 'ping' or a 'pingError' object.

w<-1
mu<-10000
delta<-150
sigmaSqF<-5000
sigmaSqR<-5000
seMu<10
seMuF<-10
seMuR<-10
score<-5
Nmerged<-0
converge<-TRUE
chr<"chr1"
range<-(1000,2000)

# Constructor
myPING<-newPing(w,mu,delta,sigmaSqF,sigmaSqR,seMu,seMuF,seMuR,score,Nmerged,as.integer(range),chr)

pingError-class

The ping class

Description

This object is used to return an error code when the PING function failed to return a valid set of estimates for a candidate regions. This could be due to non-convergence of the EM algorithm, a singular information matrix, or a number of reads below the limit specified by the user. All of these are typically due to too few reads in the region and do not affect the rest of the analysis, as such regions would most likely be labelled as false positives.

Accessors

All of the accessors defined for a 'ping' object still work for a 'pingError' object but will simply return a NULL pointer.

Constructor

newPingError(string) where 'string' is the error code.

Constructor

newPingError<-function(string)

string  The mixture weights (a vector)

Author(s)

Xuekui Zhang <xzhang@stat.ubc.ca>, Sangsoon Woo, <swoo@fhcrc.org> and Raphael Gottardo <raphael.gottardo@ircm.qc.ca>
The `pingList-class`

## Description

This object is used to gather all parameters from fitting PING to multiple candidate regions (as returned by the `segmentReads` function). The object contains the following slots: `List`, `paraPrior`, `paraEM`, `minReads`, `N`, `Nc`. `List` is a list of `ping` or `pingError` objects. `paraPrior` is a list containing the hyperparameters used for the prior, `paraEM` is a list of convergence parameters for the EM. `minReads` is a list containing the minimum number of reads used to fit a region with PING. `N` is the total number of reads in the ChIP samples while `Nc` is the total number of reads in the control sample.

## Arguments

- **object**: An object of class `ping`.

## Accessors

The PING package provides accessors to directly access to most of the parameters/standard errors and chromosomes. In the code snippets below, `x` is a `pingList` object. For all accessors, the `pingError` objects are omitted, so that the accessors only return values for the `ping` objects (i.e. all valid binding events).

- **`chromosome(x)`**: Gets the chromosome names of all candidate regions.
- **`mu(x)`**: Gets the position estimates of all binding sites identified in all candidate regions.
- **`delta(x)`**: Gets the average fragment lengths of all binding sites identified in all candidate regions.

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**References**

Xuekui Zhang, Gordon Robertson, Sangsoon Woo, Brad G. Hoffman, and Raphael Gottardo. "Probabilistic Inference for Nucleosome Positioning with MNase-based or Sonicated Short-read Data" Genome Biology, under review.

**See Also**

`ping`

**Examples**

```r
# Here is an example on how to construct such a pingError object
# Typically, you would not do this manually, you would use the ping function to return a 'pingList'
# that contains a list of 'ping' or 'pingError' object.
# Constructor
myPingError<-newPingError("Singular information matrix")
# Accessors
# Get the standard error of Mu
se(myPingError)
# Get the standard error of MuF
seF(myPingError)
# Get the scores
score(myPingError)
```
'\texttt{sigmaSqF(x)}' Gets the F peak variances of all binding sites identified in all candidate regions.

'\texttt{sigmaSqR(x)}' Gets the R peak variances of all binding sites identified in all candidate regions.

'\texttt{seF(x)}' Gets the standard errors of all binding site position estimates identified in all candidate regions.

'\texttt{seF(x)}' Gets the standard errors of all F peak modes identified in all candidate regions.

'\texttt{seR(x)}' Gets the standard errors of all R peak modes identified in all candidate regions.

'\texttt{score(x)}' Gets the scores of all binding events identified in all candidate regions.

\textbf{Constructor}

\texttt{newPingList(List, paraEM, paraPrior, minReads, N, Nc)} constructs a new \texttt{\texttt{pingList}} object with the following arguments.

\texttt{List} The mixture weights (a vector)

\texttt{paraEM} The binding site positions (a vector)

\texttt{paraPrior} The DNA fragment lengths (a vector)

\texttt{N} The variance parameters for the forward distribution (vector)

\texttt{Nc} The variance parameters for the forward distribution (vector)

\textbf{Methods}

[ \texttt{signature(x = "ping")}: subset PING object.]

\textbf{Methods}

\texttt{length \texttt{signature(x = "ping")}: subset PING object.}

\texttt{as.data.frame \texttt{signature(x = "pingList")}: Coerce a pingList to a data.frame.}

\textbf{Constructor}

\texttt{newPingList} function\texttt{(List, paraEM, paraPrior, minReads, N, Nc)} constructs a new \texttt{\texttt{pingList}} object with the following arguments.

\texttt{newPingList}

\texttt{w} The mixture weights (a vector)

\texttt{mu} The binding site positions (a vector)

\texttt{delta} The DNA fragment lengths (a vector)

\texttt{sigmaSqF} The variance parameters for the forward distribution (vector)

\texttt{sigmaSqR} The variance parameters for the reverse distribution (vector)

\texttt{seMu} The standard errors for \texttt{mu} (vector)

\texttt{seMuF} The standard errors for \texttt{muF} (vector)

\texttt{seMuR} The standard errors for \texttt{muR} (vector)

\texttt{score} The scores for each binding event (vector)

\texttt{Nmerged} The number of peaks that were merged (integer)

\texttt{converge} A logical value, \texttt{TRUE}, if the EM as converged

\texttt{infMat} The information matrix

\texttt{chr} The chromosome for the region
Author(s)

Xuekui Zhang <<xzhang@stat.ubc.ca>>, Sangsoon Woo, <swoo@fhcrc.org> and Raphael Gottardo <<raphael.gottardo@ircm.qc.ca>>

References


See Also

ping

Examples

# Here is an example of how to construct such a region
# Typically, you would not do this manually, you would use the ping function to return a 'pingList'
# that contains a list of 'ping' or 'pingError' object.

w<-1
mu<-10000
delta<-150
sigmaSqF<-5000
sigmaSqR<-5000
seMu<-10
seMuF<-10
seMuR<-10
score<-5
Nmerged<-0
converge<-TRUE
infMat<-matrix(0)
chr<="chr1"
range<-c(1000,2000)

# Constructor
#myPING1<-newPing(w,mu,delta,sigmaSqF,sigmaSqR,seMu,seMuF,seMuR,score,Nmerged,converge,infMat,as.integer(range),chr)
#myPING2<-newPing(w,mu+1000,delta,sigmaSqF,sigmaSqR,seMu,seMuF,seMuR,score,Nmerged,converge,infMat,as.integer(range),chr)

#minReads<-list(perPeak=2,perRegion=5)
#paraPrior<-list(xi=200,rho=1,alpha=20,beta=40000)
#paraEM<-list(minK=1,maxK=15,tol=10e-6,B=100)
#N<-100
#Nc<-200

#mynewPingList<-newPingList(list(myPING1,myPING2), paraEM, paraPrior, minReads, as.integer(100), as.integer(200))

# Accessors
# Get the standard error of Mu
#se(mynewPingList)
# Get the standard error of MuF
#seF(mynewPingList)
# Get the scores
#score(mynewPingList)
**plotSummary**

Plot a summary of the prediction for given ranges.

**Description**

This function uses the Gviz package to summarize the nucleosome position prediction from postPING.

**Usage**

```r
plotSummary(PS, ping, reads, chr, gen="gen", from=NULL, to=NULL, FragmentLength=200, title="", scoreThreshold=0.05)
```

**Arguments**

- **PS**: A `data.frame`. The output from `postPING`. If a list of `data.frame` is passed, multiple `NucleosomeTrack` will be plotted.
- **ping**: A `pingList`. The output of `PING`.
- **reads**: A `GRanges` object. The reads used in the segmentation process.
- **chr**: A character string for the chromosome name.
- **gen**: An optional character string for the genome name.
- **from**: An integer, the first base of the plot.
- **to**: An integer, the last base of the plot.
- **FragmentLength**: An integer, the length of XSET profile extension in the CoverageTrack. This argument will be ignored if the reads are paired-end sequencing data.
- **title**: An optional character string that can be prepended to the automatically generated title.
- **scoreThreshold**: A numeric. Removes nucleosome with a score inferior to the threshold from the plot.

**Value**

This function returns an invisible list of tracks. The tracks can be used in other Gviz plots.

**Author(s)**

Renan Sauteraud

**See Also**

- `CoverageTrack`
- `RawReadsTrack`
- `NucleosomeTrack`
- `plotTracks`
Post process Estimation of binding site positions obtained from PING

Description
Post process Estimation of binding site positions obtained from PING. Refit mixture models with stronger prior in candidate regions contain potential problems, and then convert final result into dataframe.

Usage
postPING(ping, seg, rho2=NULL, sigmaB2=NULL, alpha2=NULL, beta2=NULL, min.dist=100, paraEM=NULL, paraPrior=NULL, ... nCores=1, makePlot=FALSE, FragmentLength=100, mart=NULL, seg.boundary=NULL, DupBound=NULL, IP=NULL, datname="")

Arguments
- ping: A ‘pingList’ object containing estimation of nucleosome positions, result of ‘PING’ function.
- seg: An object of class ‘segmentReadsList’ containing the results for all regions pre-processed, ‘segmentReads’ function.
- paraEM: A list of parameters for the EM algorithm. The default parameters should be good enough for most usages.
- paraPrior: A list of parameters for the prior distribution. The default parameters should be good enough for most usages.
- minK: An integer, default=0. The minimum number of binding events per region. If the value is 0, the minimum number is automatically calculated.
- maxK: An integer, default=0. The maximum number of binding events per region. If the value is 0, the maximum number is automatically calculated.
- tol: A numeric, default=1e-4. The tolerance for the EM algorithm.
- B: An integer, default=100. The maximum number of iterations to be used.
- mSelect: A character string specifying the information criteria to be used when selecting the number of binding events. Default="AIC3"
- mergePeaks: A logical stating whether overlapping binding events should be picked. Default=TRUE
- mapCorrect: A logical stating whether mappability profiles should be incorporated in the estimation, i.e: missing reads estimated. Default=TRUE
- xi: An integer. The average DNA fragment size.
- rho: An integer. A variance parameter for the average DNA fragment size distribution.
- alpha: An integer. First hyperparameter of the inverse Gamma distribution for sigma^2 in the PICS model
- beta: An integer. Second hyperparameter of the inverse Gamma distribution for sigma^2 in the PING model
- lambda: An integer. The lambda control Gaussian Markov Random Field prior on the distance of adjacent nucleosomes, we do not recommend user change the default value.
- dMu: An integer. Our best guess for the distance between two neighboring nucleosomes.
rho2, sigmaB2, alpha2, beta2

Integer values, the parameters in the prior of mixture models to be re-fitted.

min.dist

The minimum distance of two adjacent nucleosomes predicted from different candidate regions, smaller than that will be treated as duplicated predictions for the same nucleosomes.

score

A numeric. The score threshold used when calling FilterPING.

dataType

A character string that can be set to use selected default parameters for the algorithm.

nCores

An integer. The number of cores that should be used in parallel by the function.

makePlot

A logical. Plot a summary of the output.

FragmentLength, mart, seg.boundary, DupBound, IP, datname

Plotting parameters and options.

IP: A GRanges object. The reads used in segmentation process.

FragmentLength: An integer. The length of XSET profile extension

Value

A data.frame containing the estimation of binding site positions.

Note

Based on our experiment on a few real data sets, we suggestion to use following values of parameters. For sonication data we use rho1=1.2; sigmaB2=6400; rho=15; alpha1=10; alpha2=98; beta2=200000. For MNase data we use rho1=3; sigmaB2=4900; rho=8; alpha1=20; alpha2=100; beta2=100000. The value of xi depends on specy of sample, since that affect the length of linker-DNA. For example, we use xi=160 for yeast and xi=200 for mouse.

Author(s)

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References

Xuekui Zhang, Gordon Robertson, Sangsoon Woo, Brad G. Hoffman, and Raphael Gottardo, "Probabilistic Inference for Nucleosome Positioning with MNase-based or Sonicated Short-read Data" PlosONE, under review.

See Also

PINGplotSummary
**RawReadsTrack**

*Raw reads density track*

**Description**

This track displays the start position of the forward and reverse reads used as input for the segmentation step.

**Usage**

```r
RawReadsTrack(ping, reads, chr, gen="gen", from=NULL, to=NULL, ...)
```

**Arguments**

- `ping`: A `pingList`. The output of `PING`.
- `reads`: An object of class `GRanges`. The reads used in the segmentation process.
- `chr`: A character string for the chromosome name.
- `gen`: An optional character string for the genome name.
- `from`, `to`: Two numeric. A subset of the reads can be selected in order to speed up the computation.
- `...`: Optional arguments (DisplayParameters) that can be passed on to the `DataTrack`. Refer to Gviz help and vignette for a list of arguments.

**Value**

This function returns an object of class `DataTrack`.

**Author(s)**

Renan Sauteraud

**See Also**

`segmentPING`, `plotSummary`, `DataTrack`, `plotTracks`

---

**segmentPING**

*Segment the genome into candidate regions*

**Description**

Pre-process bidirectional aligned reads data to detect candidate regions with a minimum number of forward and reverse reads. These candidate regions will then be processed by `PING`.

**Usage**

```r
segmentPING(data, dataC=NULL, map=NULL, minReads=2, minReadsInRegion=3, jitter=FALSE, maxLregion=1200, minLregion=80, step=NULL, width=NULL, islandDepth=3, min_cut=50, max_cut=1000, maxReadsWidth=500, PE=FALSE)
```
**segmentPING**

**Arguments**

- **data**: A `GRanges` object containing the IP reads. See details for more information on how to set up the data.
- **dataC**: A `GRanges` object containing the control reads. Set to NULL by default, i.e. no control.
- **map**: A ‘RangedData’ object containing the mappability profiles. Set to NULL by default, i.e. no profiles.
- **minReads**: The minimum number of F/R reads to be present in the sliding window.
- **minReadsInRegion**: The minimum number of F/R reads to be present in the region.
- **jitter**: A logical value stating whether some noise should be added to the read locations. This is recommended if the read positions have lots of duplicates.
- **step**: An integer. The step size for the sliding window.
- **width**: An integer. The size of the sliding window.
- **maxLregion**: The maximum length.
- **minLregion**: The minimum length.
- **PE**: A logical. Set to TRUE for Paired-End sequencing data.
- **islandDepth, min_cut, max_cut, maxReadsWidth**: Parameters used for paired-end sequencing data segmentation.
  - **islandDepth**: An integer. The minimum number of reads to cover a candidate region.
  - **min_cut**: An integer. The minimum length of a candidate region.
  - **max_cut**: An integer. The maximum length of a candidate region.
  - **maxReadsWidth**: An integer. Reads with width superior to this limit will be removed from the data.

**Value**

An object of class `segReadsList` containing the results for all regions pre-processed.

**Author(s)**

Xuekui Zhang, Arnaud Droit <<arnaud.droit@crchuq.ualaval.ca>> and Raphael Gottardo <<rgottard@fhcrc.org>>

**References**

Xuekui Zhang, Gordon Robertson, Sangsoon Woo, Brad G. Hoffman, and Raphael Gottardo, "Probabilistic Inference for Nucleosome Positioning with MNase-based or Sonicated Short-read Data” PlosONE.

**See Also**

`segReadsList`

**Examples**

```r
# Read data
path<-'system.file("extdata",package="PING")
dataIP<-'read.table(file.path(path,"GSM351492_R4_chr1.bed"),header=TRUE)
dataIP<-'as(dataIP,"GRanges")
seg<-segmentPING(dataIP, minReads=NULL, maxLregion=1200,minLregion=80, jitter=TRUE)
```
show

Description

This methods show the objects of PING.

Usage

```r
## S4 method for signature 'ping'
show(object)
## S4 method for signature 'pingError'
show(object)
## S4 method for signature 'pingList'
show(object)
## S4 method for signature 'segReads'
show(object)
## S4 method for signature 'segReadsList'
show(object)
```

Arguments

- `object` Object returned from `ping`.

Details

List of the slots include in the object

Author(s)

Xuekui Zhang <<xzhang@stat.ubc.ca>> Sangsoon Woo, <swoo@fhcrc.org> Raphael Gottardo <<raphael.gottardo@ircm.qc.ca>>

See Also

- `summary`

summary

Description

This methods summary the objects of PING.
Usage

## S4 method for signature 'ping'
summary(object)
## S4 method for signature 'pingList'
summary(object)
## S4 method for signature 'segReads'
summary(object)
## S4 method for signature 'segReadsList'
summary(object)

Arguments

object Object returned from ping.

Author(s)

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