Package ‘STATegRa’

January 15, 2017

**Type** Package

**Title** Classes and methods for multi-omics data integration

**Version** 1.8.0

**Date** 2016-10-10

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**Depends** R (>= 2.10)

**Imports** Biobase, gridExtra, ggplot2, methods, stats, grid, MASS, calibrate, gplots, edgeR, limma, foreach, affy

**Suggests** RUnit, BiocGenerics, knitr (>= 1.6), rmarkdown, BiocStyle (>= 1.3), roxygen2, doSNOW

**VignetteBuilder** knitr

**Encoding** UTF-8

**biocViews** Software, StatisticalMethod, Clustering, DimensionReduction, PrincipalComponent

**Description** Classes and tools for multi-omics data integration.

**License** GPL-2

**LazyLoad** yes

**Collate** ‘STATegRa_combiningMappings.R’ ‘STATegRa_fused.R’ ‘STATegRa_holistOmics.R’ ‘STATegRa_omicsCLUST_bioMap.R’ ‘STATegRa_omicsCLUST_bioDist.R’ ‘STATegRa_omicsCLUST_bioDistW.R’ ‘STATegRa_omicsNPC.R’ ‘STATegRa_omicsNPC_internal.R’ ‘STATegRa_omicsPCA_caClass.R’ ‘STATegRa_omicsPCA_methods.R’ ‘STATegRa_omicsPCA_plotting.R’ ‘STATegRa_package.R’

**RoxygenNote** 5.0.1

**NeedsCompilation** no

**R topics documented:**

bioDist .................................................. 2
bioDistClass ........................................... 5
bioDistFeature ......................................... 5
bioDist

Description

Function to compute a bioDistclass object from profile data and a mapping. For details of the process see the user’s guide, but briefly the process involves using the mapping to identify reference features appropriate to each surrogate feature (if any), aggregating the surrogate data into pseudo-data for each reference feature, and then calculating the correlation distance between the reference features according to the surrogate data.

Usage

bioDist(referenceFeatures=NULL, reference=NULL, mapping=NULL, referenceData=NULL, surrogateData=NULL, filtering=NULL, noMappingDist=NA, distance="spearman", aggregation="sum", maxitems=NULL, selectionRule="maxFC", expfac=NULL, name=NULL, ...)

Arguments

referenceFeatures
subset of features to be considered for the computation of the distances. If NULL then the features are first gathered from the features in referenceData. If referenceData is not provided then the list of features are gathered from mapping (bioMap class) and using the reference.

reference
A character indicating the variable that is being used as features to compute distance between

mapping
The mapping between feature types

referenceData
ExpressionSet object with the data from the reference features.

surrogateData
ExpressionSet object with the data from the surrogate features.

filtering
A filtering for the bioMap class. To be implemented.

noMappingDist
Distance value to be used when a reference feature do not map to any surrogate feature. If "max", maximum indirect distance among the rest of reference features is taken. If NA, distance weights are re-scaled so this surrogate association is not considered. If a number then the missing values are replaces with that value.

distance
Distance between features to be computed. Possible values are "pearson", "kendall", "spearman", "euclidean", "maximum", "manhattan", "canberra", "binary" and "minkowski". Default is "spearman".

aggregation
Action to perform when a reference feature maps to more than one surrogate feature. Options are "max", "sum", "mean" or "median" and the the values are aggregated according to the chosen statistic.

maxitems
The maximum number of surrogate features per reference feature to be used, selected according to "selectionRule" parameter. Default is 2.

selectionRule
Rule to select the surrogate features to be used (the number is determined by "maxitems"). It can be one of the following: (1) "maxcor" those presenting maximum correlation with corresponding main feature; in this case "referenceData" must be provided and the columns must overlap in at least 3 samples; (2) "maxmean": average across samples is computed and those features with higher mean are selected; case (3) is similar to (2) but considering other statistics: "maxmedian", "maxdiff", "maxFC", "sd", "ee".

expfac
Not in use yet.

name
Character that describes the nature of the bioDist class computed

... extra arguments passed to dist, eg "p=value" for the power used if calculating minkowski distance

Value
An object of class bioDistclass containing distances between the features in surrogateData.

Author(s)
David Gomez-Cabrero
Examples

data(STATegRa_S1)
data(STATegRa_S2)
require(Biobase)

# Truncate data for brevity
Block1 <- Block1[1:100,]
Block2 <- Block2[1:100,]

## Create ExpressionSets
mRNA.ds <- createOmicsExpressionSet(Data=Block1,pData=ed,pDataDescr=c("classname"))
miRNA.ds <- createOmicsExpressionSet(Data=Block2,pData=ed,pDataDescr=c("classname"))

## Create the bioMap
map.gene.miRNA<-bioMap(name = "Symbol-miRNA",
                        metadata = list(type_v1="Gene",type_v2="miRNA",
                                        source_database="targetscan.Hs.eg.db",
                                        data_extraction="July2014"),
                        map=mapdata)

# Create Gene-gene distance computed through miRNA data
bioDistmiRNA<-bioDist(referenceFeatures = rownames(Block1),
                       reference = "Var1",
                       mapping = map.gene.miRNA,
                       surrogateData = miRNA.ds, ### miRNA data
                       referenceData = mRNA.ds, ### mRNA data
                       maxitems=2,
                       selectionRule="sd",
                       expfac=NULL,
                       aggregation = "sum",
                       distance = "spearman",
                       noMappingDist = 0,
                       filtering = NULL,
                       name = "mRNAbymiRNA")

# Create Gene-gene distance through mRNA data
bioDistmRNA<-new("bioDistclass",
                 name = "mRNAbymRNA",
                 distance = cor(t(exprs(mRNA.ds)),method="spearman"),
                 map.name = "id",
                 map.metadata = list(),
                 params = list())

###### Generation of the list of Surrogated distances.

bioDistList<-list(bioDistmRNA,bioDistmiRNA)
sample.weights<-matrix(c(0,0.33,0.67,1),4,2)
sample.weights[,1]<-c(0,0.33,0.67,1)
sample.weights[,2]<-c(1,0.67,0.33,0)

###### Generation of the list of bioDistWclass objects.

bioDistWList<-bioDistW(referenceFeatures = rownames(Block1),
                       bioDistList = bioDistList,
                       weights=sample.weights)
Plot of distances.

```r
bioDistWPlot(referenceFeatures = rownames(Block1),
              listDistW = bioDistWList,
              method.cor = "spearman")
```

Computing the matrix of features/distances associated.

```r
fm <- bioDistFeature(Feature = rownames(Block1)[1],
                     listDistW = bioDistWList,
                     threshold.cor = 0.7)
bioDistFeaturePlot(data = fm)
```

---

**bioDistclass**

**Description**

Class to manage mappings between genomic features.

**Usage**

```r
bioDistclass(name, distance, map.name, map.metadata, params)
```

**Arguments**

- **name**: Name assigned to the object
- **distance**: Matrix giving the object
- **map.name**: Charactering giving the name of the bioMap object used to compute the distance
- **map.metadata**: List of parameters used to generate the mapping
- **params**: List of parameters used to generate the distance

---

**bioDistFeature**

**Description**

Function that computes for a given selected feature the closest features given a selected set of weighted distances.

**Usage**

```r
bioDistFeature(Feature, listDistW, threshold.cor)
```

**Arguments**

- **Feature**: Feature A selected as a reference.
- **listDistW**: A list of bioDistWclass objects. All the objects must contain the Feature A selected and all of them must contain the same set of features.
- **threshold.cor**: A threshold to select the features associated to Feature A
Value

Matrix with the associated features given the different weighted distances considered

Author(s)

David Gomez-Cabrero

Examples

data(STATegRa_S1)
data(STATegRa_S2)
require(Biobase)

# Truncate data for brevity
Block1 <- Block1[1:100,]
Block2 <- Block2[1:100,]

## Create ExpressionSets
mRNA.ds <- createOmicsExpressionSet(Data=Block1,pData=ed,pDataDescr=c("classname"))
miRNA.ds <- createOmicsExpressionSet(Data=Block2,pData=ed,pDataDescr=c("classname"))

## Create the bioMap
map.gene.miRNA<-bioMap(name = "Symbol-miRNA",
metadata = list(type_v1="Gene",type_v2="miRNA",
source_database="targetscan.Hs.eg.db",
data_extraction="July2014"),
map=mapdata)

# Create Gene-gene distance computed through miRNA data
bioDistmiRNA<-bioDist(referenceFeatures = rownames(Block1),
reference = "Var1",
mapping = map.gene.miRNA,
surrogateData = miRNA.ds, ### miRNA data
referenceData = mRNA.ds, ### mRNA data
maxitems=2,
selectionRule="sd",
expfac=NULL,
aggregation = "sum",
distance = "spearman",
noMappingDist = 0,
filtering = NULL,
name = "mRNAbymiRNA")

# Create Gene-gene distance through mRNA data
bioDistmRNA<-new("bioDistclass",
name = "mRNAbymRNA",
distance = cor(t(exprs(mRNA.ds)),method="spearman"),
map.name = "id",
map.metadata = list(),
params = list())

######## Generation of the list of Surrogated distances.

bioDistList<-list(bioDistmRNA,bioDistmiRNA)
sample.weights<-matrix(0,4,2)
sample.weights[,1]<-c(0,0.33,0.67,1)
sample.weights[,2]<-c(1,0.67,0.33,0)

##### Generation of the list of bioDistWclass objects.
bioDistWList<-bioDistW(referenceFeatures = rownames(Block1),
    bioDistList = bioDistList,
    weights=sample.weights)

##### Plot of distances.
bioDistWPlot(referenceFeatures = rownames(Block1),
    listDistW = bioDistWList,
    method.cor="spearman")

##### Computing the matrix of features/distances associated.
fm<-bioDistFeature(Feature = rownames(Block1)[1],
    listDistW = bioDistWList,
    threshold.cor=0.7)
bioDistFeaturePlot(data=fm)

---

**bioDistFeaturePlot**

**bioDistFeaturePlot**

---

**Description**

Function that plots the results from a bioDistFeature analysis

**Usage**

bioDistFeaturePlot(data)

**Arguments**

- data: Matrix produced by bioDistFeature

**Value**

Generates a heatmap plot

**Author(s)**

David Gomez-Cabrero

**Examples**

```r
data(STATegRa_S1)
data(STATegRa_S2)
require(Biobase)

# Truncate data for brevity
Block1 <- Block1[1:100,]
Block2 <- Block2[1:100,
```
## Create ExpressionSets
mRNA.ds <- createOmicsExpressionSet(Data=Block1,pData=ed,pDataDescr=c("classname"))
miRNA.ds <- createOmicsExpressionSet(Data=Block2,pData=ed,pDataDescr=c("classname"))

## Create the bioMap
map.gene.miRNA<-bioMap(name = "Symbol-miRNA",
                        metadata = list(type_v1="Gene",type_v2="miRNA",
                                        source_database="targetscan.Hs.eg.db",
                                        data_extraction="July2014"),
                        map=mapdata)

# Create Gene-gene distance computed through miRNA data
bioDistmiRNA<-bioDist(referenceFeatures = rownames(Block1),
                      reference = "Var1",
                      mapping = map.gene.miRNA,
                      surrogateData = miRNA.ds, ### miRNA data
                      referenceData = mRNA.ds, ### mRNA data
                      maxitems=2,
                      selectionRule="sd",
                      expfac=NULL,
                      aggregation = "sum",
                      distance = "spearman",
                      noMappingDist = 0,
                      filtering = NULL,
                      name = "mRNAbymiRNA")

# Create Gene-gene distance through mRNA data
bioDistmRNA<-new("bioDistclass",
                 name = "mRNAbymRNA",
                 distance = cor(t(exprs(mRNA.ds)),method="spearman"),
                 map.name = "id",
                 map.metadata = list(),
                 params = list())

##### Generation of the list of Surrogated distances.
bioDistList<-list(bioDistmRNA,bioDistmiRNA)
sample.weights<-matrix(0,4,2)
sample.weights[,1]<-c(0,0.33,0.67,1)
sample.weights[,2]<-c(1,0.67,0.33,0)

##### Generation of the list of bioDistWclass objects.
bioDistWList<-bioDistW(referenceFeatures = rownames(Block1),
                       bioDistList = bioDistList,
                       weights=sample.weights)

##### Plot of distances.
bioDistWPlot(referenceFeatures = rownames(Block1),
             bioDistList = bioDistWList,
             weights=sample.weights)

##### Computing the matrix of features/distances associated.
fm<-bioDistFeature(Feature = rownames(Block1)[1],
                   listDistW = bioDistWList,
                   threshold.cor=0.7)
bioDistW

bioDistFeaturePlot(data=fm)

---

**bioDistW**

**bioDistW**

**Description**

Function that computes weighted distances between a list of bioDistclass objects.

**Usage**

```r
bioDistW(referenceFeatures, bioDistList, weights)
```

**Arguments**

- `referenceFeatures`  
  The set of features that weighted distance is computed between.

- `bioDistList`  
  A list of bioDistclass objects. All the objects must contain the set of features selected.

- `weights`  
  A matrix where the number of columns equals the number of elements included in the bioDistList list.

**Value**

Returns a list of bioDistWclass objects. Each element in the list returns the weighted distance associated to each row in the "weights" matrix.

**Author(s)**

David Gomez-Cabrero

**Examples**

```r
data(STATegRa_S1)
data(STATegRa_S2)
require(Biobase)

# Truncate data for brevity
Block1 <- Block1[1:100,]
Block2 <- Block2[1:100,]

## Create ExpressionSets
mRNA.ds <- createOmicsExpressionSet(Data=Block1,pData=ed,pDataDescr=c("classname"))
miRNA.ds <- createOmicsExpressionSet(Data=Block2,pData=ed,pDataDescr=c("classname"))

## Create the bioMap
map.gene.miRNA<-bioMap(name = "Symbol-miRNA",
    metadata = list(type_v1="Gene",type_v2="miRNA",
                    source_database="targetsan.Hs.eg.db",
                    data_extraction="July2014"),
    map=mapdata)
```
# Create Gene-gene distance computed through miRNA data
bioDistmiRNA<-bioDist(referenceFeatures = rownames(Block1),
            reference = "Var1",
            mapping = map.gene.miRNA,
            surrogateData = miRNA.ds, ### miRNA data
            referenceData = mRNA.ds, ### mRNA data
            maxitems=2,
            selectionRule="sd",
            expfac=NULL,
            aggregation = "sum",
            distance = "spearman",
            noMappingDist = 0,
            filtering = NULL,
            name = "mRNAbymiRNA")

# Create Gene-gene distance through mRNA data
bioDistmRNA<-new("bioDistclass",
            name = "mRNAbymRNA",
            distance = cor(t(exprs(mRNA.ds)),method="spearman"),
            map.name = "id",
            map.metadata = list(),
            params = list())

###### Generation of the list of Surrogated distances.
bioDistList<-list(bioDistmRNA,bioDistmiRNA)
sample.weights<-matrix(0,4,2)
sample.weights[,1]<-c(0,0.33,0.67,1)
sample.weights[,2]<-c(1,0.67,0.33,0)

###### Generation of the list of bioDistWclass objects.
bioDistWList<-bioDistW(referenceFeatures = rownames(Block1),
            bioDistList = bioDistList,
            weights=sample.weights)

###### Plot of distances.
bioDistWPlot(referenceFeatures = rownames(Block1) ,
            listDistW = bioDistWList,
            method.cor="spearman")

###### Computing the matrix of features/distances associated.
fm<-bioDistFeature(Feature = rownames(Block1)[1] ,
            listDistW = bioDistWList,
            threshold.cor=0.7)
bioDistFeaturePlot(data=fm)

---

**Description**

Function that plots the "distance relation" between features computed through different surrogate features.
bioDistWPlot

Usage

bioDistWPlot(referenceFeatures, listDistW, method.cor)

Arguments

referenceFeatures

The set of features to be used.

listDistW

A list of bioDistWclass objects.

method.cor

Method to compute distances between the elements in the listDistW. The default is spearman correlation.

Value

Makes a plot with the projected distance between the listDistW objects.

Author(s)

David Gomez-Cabrero

Examples

data(STATegRa_S1)
data(STATegRa_S2)
require(Biobase)

# Truncate data for brevity
Block1 <- Block1[1:100,]
Block2 <- Block2[1:100,]

## Create ExpressionSets
mRNA.ds <- createOmicsExpressionSet(Data=Block1,pData=ed,pDataDescr=c("classname"))
miRNA.ds <- createOmicsExpressionSet(Data=Block2,pData=ed,pDataDescr=c("classname"))

## Create the bioMap
map.gene.miRNA<-bioMap(name = "Symbol-miRNA",
metadata = list(type_v1="Gene",type_v2="miRNA",
source_database="targetscan.Hs.eu.db",
data_extraction="July2014"),
map=mapdata)

# Create Gene-gene distance computed through miRNA data
bioDistmiRNA<-bioDist(referenceFeatures = rownames(Block1),
reference = "Var1",
mapping = map.gene.miRNA,
surrogateData = miRNA.ds, ### miRNA data
referenceData = mRNA.ds, ### mRNA data
maxitems=2,
selectionRule="sd",
expfac=NULL,
aggregation = "sum",
distance = "spearman",
noMappingDist = 0,
filtering = NULL,
name = "mRNAbymiRNA")
# Create Gene-gene distance through mRNA data
bioDistmRNA<-new("bioDistclass",
    name = "mRNAbymRNA",
    distance = cor(t(exprs(mRNA.ds)),method="spearman"),
    map.name = "id",
    map.metadata = list(),
    params = list())

###### Generation of the list of Surrogated distances.
bioDistList<-list(bioDistmRNA,bioDistmiRNA)
sample.weights<-matrix(0,4,2)
sample.weights[,1]<-c(0,0.33,0.67,1)
sample.weights[,2]<-c(1,0.67,0.33,0)

###### Generation of the list of bioDistWclass objects.
bioDistWList<-bioDistW(referenceFeatures = rownames(Block1),
    bioDistList = bioDistList,
    weights=sample.weights)

###### Plot of distances.
bioDistWPlot(referenceFeatures = rownames(Block1) ,
    listDistW = bioDistWList,
    method.cor="spearman")

###### Computing the matrix of features/distances associated.
fm<-bioDistFeature(Feature = rownames(Block1)[1] ,
    listDistW = bioDistWList,
    threshold.cor=0.7)
bioDistFeaturePlot(data=fm)

---

**bioMap**

**Description**

Function to generate a bioMap object.

**Usage**

bioMap(name, metadata, map)

**Arguments**

- **name**
  - Name to assign the object
- **metadata**
  - A list with information of the mapping. Elements expected in the list are: (1) "type_v1" and "type_v2", refer to the nature of the features mapped; a vocabulary we recommend is "gene", "mRNA", "miRNA", "proteins", etc. (2) "source_database", provides information on the source of the mapping; from a specific database e.g. "targetscan.Hs.eg.db" to a genomic location mapping. (3) "data_extraction" stores information on the data the mapping was generated or downloaded.
A data.frame object storing the mapping. The data.frame may include an unlimited number of columns, however the first column must be named "Var1" and refer to the elements of "type_v1" and similarly for the second column ("Var2", "type_v2").

Value

An object of class bioMap

Author(s)

David Gomez-Cabrero

Examples

data(STATegRa_S2)
map.gene.miRNA<-bioMap(name = "Symbol-miRNA",
metadata = list(type_v1="Gene",type_v2="miRNA",
source_database="targetscan.Hs.eg.db",
data_extraction="July2014"),
map=mapdata)

biplotRes

Biplot of component analysis

Description

Generate a biplot of component analysis results

Usage

biplotRes(object, type, comps, block, title=NULL, colorCol=NULL,
sizeValues=c(2, 4), shapeValues=c(17, 0), background=TRUE,
pointSize=4, labelSize=NULL, axisSize=NULL, titleSize=NULL)

Arguments

object caClass object containing component analysis results

Type

Character specifying which components to plot; "common", "individual" or "both"

Comps

Components to plot. If combined=FALSE, specifies the component indices to use as x and y for the plot. Otherwise, the component from the first block and the component from second block to plot together.

Block

Which block to plot, either "1" or "2" or the name of the block.

Title

Plot title

ColorCol

Character specifying a pData column to use to colorise the plot points

SizeValues

Vector containing sizes for scores and loadings

ShapeValues

Vector indicating the shapes for scores and loadings

Background

Logical, whether to use a grey background

PointSize

Size of plot points
biplotRes

labelSize  Size of plot labels if not NULL
axisSize    Size of axis text
titleSize   Size of title text

Value

ggplot2 object

Author(s)

Patricia Sebastian-Leon

Examples

data("STATegRa_S3")
B1 <- createOmicsExpressionSet(Data=Block1.PCA,pData=ed.PCA,
pDataDescr=c("classname"))
B2 <- createOmicsExpressionSet(Data=Block2.PCA,
pData=ed.PCA,pDataDescr=c("classname"))

# Omics components analysis
discoRes <- omicsCompAnalysis(Input=list(B1,B2),Names=c("expr","mirna"),
method="DISCOSCA",Rcommon=2,Rspecific=c(2,2),
center=TRUE,scale=TRUE,weight=TRUE)
jiveRes <- omicsCompAnalysis(Input=list(B1,B2),Names=c("expr","mirna"),
method="JIVE",Rcommon=2,Rspecific=c(2,2),
center=TRUE,scale=TRUE,weight=TRUE)
o2plsRes <- omicsCompAnalysis(Input=list(B1,B2),Names=c("expr","mirna"),
method="O2PLS",Rcommon=2,Rspecific=c(2,2),
center=TRUE,scale=TRUE,weight=TRUE)

# Biplot common part. DISCO-SCA
biplotRes(object=discoRes,type="common",comps=c(1,2),block="",
title=NULL,colorCol="classname",sizeValues=c(2,4),
shapeValues=c(17,0),background=TRUE,pointSize=4,labelSize=NULL,axisSize=NULL,titleSize=NULL)

# Biplot common part. O2PLS
p1 <- biplotRes(object=o2plsRes,type="common",comps=c(1,2),
block="expr",title=NULL,colorCol="classname",
sizeValues=c(2,4),shapeValues=c(17,0),background=TRUE,pointSize=4,labelSize=NULL,
axisSize=NULL,titleSize=NULL)
p2 <- biplotRes(object=o2plsRes,type="common",comps=c(1,2),
block="mirna",title=NULL,colorCol="classname",
sizeValues=c(2,4),shapeValues=c(17,0),background=TRUE,pointSize=4,labelSize=NULL,
axisSize=NULL,titleSize=NULL)

# Biplot distinctive part. O2PLS
p1 <- biplotRes(object=discoRes,type="individual",comps=c(1,2),
block="expr",title=NULL,colorCol="classname",
sizeValues=c(2,4),shapeValues=c(17,0),
caClass-class

Stores the results of any of the omicsPCA analyses.

Slots

- **InitialData**: List of ExpressionSets, one for each set of omics data
- **Names**: Character vector giving names for the input data
- **preprocessing**: Character vector describing the preprocessing applied to the data
- **preproData**: List of matrices containing data after preprocessing
- **caMethod**: Character giving the component analysis method name
- **commonComps**: Numeric giving the number of common components
- **distComps**: Numeric vector giving the number of distinctive components for each block
- **scores**: List of matrices of common and distinctive scores
- **loadings**: List of matrices of common and distinctive loadings
- **VAF**: List of matrices indicating VAF (Variability Explained For) for each component in each block of data
- **others**: List containing other miscellaneous information specific to different SCA methods

Author(s)

Patricia Sebastian Leon

combiningMappings

combiningMappings, combining several mappings for use in the omicsNPC function

Description

This function combines several annotation so that measurements across different datasets are mapped to the same reference elements (e.g., genes). The annotations should all be either data frame / matrices, named vectors/lists, or bioMap objects. See the examples for further details

Usage

combiningMappings(mappings, reference = NULL, retainAll = FALSE)
createOmicsExpressionSet

Arguments
mappings List of annotations.
reference If the annotations are data frame, matrices or bioMap objects, the name of the column containing the reference elements
retainAll Logical, if set to TRUE measurements that have no counterparts in other datasets are retained

Value
A data frame encoding the mapping across several dataset

Author(s)
Vincenzo Lagani

References
Nestoras Karathanasis, Ioannis Tsamardinos and Vincenzo Lagani. omicsNPC: applying the Non-Parametric Combination methodology to the integrative analysis of heterogeneous omics data. Submitted to PlosONE.

Examples
#Example 1
#Mapping with data frames
mRNA <- data.frame(gene = rep(c('G1', 'G2', 'G3'), each = 2), probeset = paste('p', 1:6, sep = ''));
methylation <- data.frame(gene = c(rep('G1', 3), rep('G2', 4)),
                         methy = paste('metyh', 1:7, sep = ''));
miRNA <- data.frame(gene = c(rep('G1', 2), rep('G2', 1), rep('G3', 2)),
                    miR = c('miR1', 'miR2', 'miR1', 'miR1', 'miR2'));
mappings <- list(mRNA = mRNA, methylation = methylation, miRNA = miRNA);
combiningMappings(mappings = mappings, retainAll = TRUE)

#Example 2
#Mapping with character vectors
mRNA <- rep(c('G1', 'G2', 'G3'), each = 2);
names(mRNA) = paste('p', 1:6, sep = '');
methylation <- c(rep('G1', 3), rep('G2', 4));
names(methylation) = paste('metyh', 1:7, sep = '');
miRNA <- c(rep('G1', 2), rep('G2', 1), rep('G3', 2));
names(miRNA) = c('miR1', 'miR2', 'miR1', 'miR1', 'miR2');
mappings <- list(mRNA = mRNA, methylation = methylation, miRNA = miRNA);
combiningMappings(mappings = mappings, retainAll = TRUE)
getInitialData

Usage

createOmicsExpressionSet(Data, pData = NULL, pDataDescr = NULL, feaData = NULL, feaDataDescr = NULL)

Arguments

Data | Omics data
pData | Data associated with the samples/phenotype
pDataDescr | Description of the phenotypic data
feaData | Data associated with the variables/features annotation
feaDataDescr | Description of the feature annotation

Details

In Data matrix, samples has to be in columns and variables has to be in rows

Value

ExpressionSet with the data provided

Author(s)

Patricia Sebastian-Leon

Examples

data(STATegRa_S3)
B1 <- createOmicsExpressionSet(Data=Block1.PCA, pData=ed.PCA, pDataDescr=c("classname"))
B2 <- createOmicsExpressionSet(Data=Block2.PCA, pData=ed.PCA, pDataDescr=c("classname"))

getInitialData | Retrieve initial data from caClass objects

Description

Generic function to retrieve the initial data used for by omicsCompAnalysis from a caClass-class object

Usage

getInitialData(x, block=NULL)

Arguments

x | caClass-class object.
block | Character indicating the block of data to be returned. It can be specified by the position of the block ("1" or "2") or the name assigned in the caClass-class object. If it is NULL both blocks are displayed.
Value

The requested data block or blocks

Author(s)

Patricia Sebastian-Leon

See Also

omicsCompAnalysis, caClass-class

Examples

data("STATegRa_S3")
B1 <- createOmicExpressionSet(Data=Block1.PCA, pData=ed.PCA,
pDataDescr=c("classname"))
B2 <- createOmicExpressionSet(Data=Block2.PCA, pData=ed.PCA, pDataDescr=c("classname"))
# Omics components analysis
res <- omicsCompAnalysis(Input=list(B1, B2), Names=c("expr", "mirna"),
method="DISCOSCA", Rcommon=2, Rspecific=c(2, 2),
center=TRUE, scale=TRUE, weight=TRUE)
getInitialData(res)
getInitialData(res, block="expr")

getLoadings

Retrieve component analysis loadings

Description

Generic function to retrieve loadings (common and distinctive) found by omicsCompAnalysis on a caClass-class object.

Usage

getLoadings(x, part=NULL, block=NULL)

Arguments

x caClass-class object.
part Character indicating whether "common" or "distinctive" loadings should be displayed
block Character indicating the block of data for which the loadings will be given. It can be specified by the position of the block ("1" or "2") or the name assigned in the caClass-class object. If it is NULL both blocks are displayed.

Value

A list containing the requested information.

Author(s)

Patricia Sebastian-Leon
getMethodInfo

Retrieve information about component analysis method

Description

Generic function to retrieve information about the method used by omicsCompAnalysis on a caClass-class object.

Usage

getMethodInfo(x, method=FALSE, comps=NULL, block=NULL)

Arguments

x caClass-class object.
method Logical indicating whether to return the method name.
comps Character indicating which component number to return ("common", "distinctive" or "all")
block Character indicating the block of data for which the component count will be given. It can be specified by the position of the block ("1" or "2") or the name assigned in the caClass-class object. If it is NULL both blocks are displayed.

Value

A list containing the requested information.

Author(s)

Patricia Sebastian-Leon

See Also

omicsCompAnalysis, caClass-class
**getPreprocessing**

Retrieve information about preprocessing

**Description**

Generic function to retrieve information about the preprocessing done by `omicsCompAnalysis` on a `caClass-class` object.

**Usage**

```r
getPreprocessing(x, process=FALSE, preproData=FALSE, block=NULL)
```

**Arguments**

- `x`  
  `caClass-class` object.
- `process`  
  Logical indicating whether to return information about the processing done.
- `preproData`  
  Logical indicating whether to return the pre-processed data matrices.
- `block`  
  Character indicating the block of data to be returned. It can be specified by the position of the block ("1" or "2") or the name assigned in the `caClass-class` object. If it is NULL both blocks are displayed.

**Value**

If both `process` and `preproData` are specified, a list containing (otherwise the individual item):

- `process` Character indicating the processing done
- `preproData` Matrix (or list of matrices, depending on `block`) containing pre-processed data

**Author(s)**

Patricia Sebastian-Leon

**See Also**

`omicsCompAnalysis`, `caClass-class`
getScores

Examples

data("STATegRa_S3")
B1 <- createOmicsExpressionSet(Data=Block1.PCA, pData=ed.PCA,
    pDataDescr=c("classname"))
B2 <- createOmicsExpressionSet(Data=Block2.PCA,
    pData=ed.PCA, pDataDescr=c("classname"))

# Omics components analysis
res <- omicsCompAnalysis(Input=list(B1, B2), Names=c("expr", "mirna"),
    method="DISCOSCA", Rcommon=2, Rspecific=c(2, 2),
    center=TRUE, scale=TRUE, weight=TRUE)
getPreprocessing(res, process=TRUE)
getPreprocessing(res, preproData=TRUE, block="1")

getScores

Retrieve component analysis scores

description

Generic function to retrieve scores (common and distinctive) found by omicsCompAnalysis on a caClass-class object.

Usage

gscores(x, part=NULL, block=NULL)

Arguments

x caClass-class object.
part Character indicating whether "common" or "distinctive" scores should be displayed
block Character indicating the block of data for which the scores will be given. It can be specified by the position of the block ("1" or "2") or the name assigned in the caClass-class object. If it is NULL both blocks are displayed.

Value

A list containing the requested information.

Author(s)

Patricia Sebastian-Leon

See Also

omicsCompAnalysis, caClass-class
getVAF

Examples

data("STATegRa_S3")
B1 <- createOmicsExpressionSet(Data=Block1.PCA, pData=ed.PCA,
pDataDescr=c("classname"))
B2 <- createOmicsExpressionSet(Data=Block2.PCA,
pData=ed.PCA, pDataDescr=c("classname"))
# Omics components analysis
res <- omicsCompAnalysis(Input=list(B1, B2), Names=c("expr", "mirna"),
  method="DISCOSCA", Rcommon=2, Rspecific=c(2, 2),
  center=TRUE, scale=TRUE, weight=TRUE)
getScores(res)
getScores(res, part="common")
getScores(res, part="distinctive", block="expr")

getVAF  Retrieve information about VAF

Description

Generic function to retrieve VAF found by omicsCompAnalysis on a caClass-class object.

Usage

getVAF(x, part=NULL, block=NULL)

Arguments

x  caClass-class object.
part Character indicating whether "common" or "distinctive" VAF should be displayed
block Character indicating the block of data for which the VAF will be given. It can be specified by the position of the block ("1" or "2") or the name assigned in the caClass-class object. If it is NULL both blocks are displayed.

Value

A list containing the requested information.

Author(s)

Patricia Sebastian-Leon

See Also

omicsCompAnalysis, caClass-class
Examples

```r
data("STATegRa_S3")
B1 <- createOmicsExpressionSet(Data=Block1.PCA, pData=ed.PCA,
pDataDescr=c("classname"))
B2 <- createOmicsExpressionSet(Data=Block2.PCA,
pData=ed.PCA, pDataDescr=c("classname"))

# Omics components analysis
res <- omicsCompAnalysis(Input=list(B1, B2), Names=c("expr", "mirna"),
method="DISCOSCA", Rcommon=2, Rspecific=c(2, 2),
center=TRUE, scale=TRUE, weight=TRUE)

getVAF(res)
getVAF(res, part="common")
getVAF(res, part="distinctive", block="expr")
```

holistOmics

HolistOmics an application of NPC on omics datasets

Description

This function is now deprecated. Use omicsNPC instead.

Usage

```r
holistOmics(dataInput, dataTypes, comb.method = c("Fisher", "Liptak", "Tippett"),
numPerm = 1000, numCores = 1, verbose = FALSE)
```

Arguments

- `dataInput`: List of ExpressionSet objects, one for each data modality.
- `dataTypes`: Character vector with possible values: 'RNA-seq', 'microarray'
- `comb.method`: Character vector with possible values: 'Fisher', 'Liptak', 'Tippett', if more than one is specified, all will be used.
- `numPerm`: Number of permutations
- `numCores`: Number of CPU cores to use
- `verbose`: Logical, if set to TRUE holistOmics prints out the step that it performs

Value

A data.frame

Author(s)

Nestoras Karathanasis

References

modelSelection

Examples

# Load the data
data("TCGA_BRCA_Batch_93")
# Setting dataTypes, the first two ExpressionSets include RNAseq data,
# the third ExpressionSet includes Microarray data.
dataTypes <- c("RNAseq", "RNAseq", "Microarray")
# Setting methods to combine pvalues
comb.method = c("Fisher", "Liptak", "Tippett")
# Setting number of permutations
numPerm = 1000
# Setting number of cores
numCores = 1
# Setting holistOmics to print out the steps that it performs.
verbose = TRUE
# Run holistOmics analysis.
# The output is a data.frame of p-values.
# Each row corresponds to a gene name. Each column corresponds to a method
# used in the analysis.
## Not run: out <- holistOmics(dataInput = TCGA_BRCA_Data, dataTypes = dataTypes,
##                             comb.method = comb.method, numPerm = numPerm,
##                             numCores = numCores, verbose = verbose)
## End(Not run)

modelSelection Find optimal common and distinctive components

Description

Uses selectCommonComps and PCA.selection to estimate the optimal number of common and distinctive components according to given selection criteria.

Usage

modelSelection(Input, Rmax, fac.sel, varthreshold=NULL, nvar=NULL, PCnum=NULL)

Arguments

Input List of two ExpressionSet objects
Rmax Maximum common components (see selectCommonComps)
fac.sel PCA criteria (see PCA.selection)
varthreshold Cumulative variance criteria (see PCA.selection)
nvar Relative variance criteria (see PCA.selection)
PCnum Fixed component number (see PCA.selection)

Value

List containing:

common Number of common components
dist Number of distinct components per input block
omicsCompAnalysis

Author(s)

Patricia Sebastian-Leon

See Also

selectCommonComps, PCA.selection, omicsCompAnalysis

Examples

data(STATegRa_S3)
B1 <- createOmicsExpressionSet(Data=Block1.PCA,pData=ed.PCA,pDataDescr=c("classname"))
B2 <- createOmicsExpressionSet(Data=Block2.PCA,pData=ed.PCA,pDataDescr=c("classname"))
ms <- modelSelection(Input=list(B1, B2), Rmax=4, fac.sel="single\%", varthreshold=0.03)
ms

omicsCompAnalysis  Components analysis for multiple objects

Description

This function performs a components analysis of object wise omics data to understand the mechanisms that underlay all the data blocks under study (common mechanisms) and the mechanisms underlying each of the data block independently (distinctive mechanisms). This analysis include both, the preprocessing of data and the components analysis by using three different methodologies.

Usage

omicsCompAnalysis(Input, Names, method, Rcommon, Rspecific,
  convThres=1e-10, maxIter=600, center=FALSE,
  scale=FALSE, weight=FALSE)

Arguments

Input  List of ExpressionSet objects, one for each block of data.
Names  Character vector giving names for each Input object.
method  Method to use for analysis (either "DISCOSCA", "JIVE", or "O2PLS").
Rcommon  Number of common components between all blocks
Rspecific  Vector giving number of unique components for each input block
convThres  Stop criteria for convergence
maxIter  Maximum number of iterations
center  Character (or FALSE) specifying which (if any) centering will be applied before analysis. Choices are "PERBLOCKS" (each block separately) or "ALL-BLOCKS" (all data together).
scale  Character (or FALSE) specifying which (if any) scaling will be applied before analysis. Choices are "PERBLOCKS" (each block separately) or "ALL-BLOCKS" (all data together).
weight  Logical indicating whether weighting is to be done.
Value

An object of class `caClass-class`.

Author(s)

Patricia Sebastian Leon

Examples

```r
data("STATegRa_S3")
B1 <- createOmicsExpressionSet(Data=Block1.PCA,pData=ed.PCA,
pDataDescr=c("classname"))
B2 <- createOmicsExpressionSet(Data=Block2.PCA,
pData=ed.PCA,pDataDescr=c("classname"))

# Omics components analysis
discoRes <- omicsCompAnalysis(Input=list(B1,B2),Names=c("expr","mirna"),
method="DISCOSCA",Rcommon=2,Rspecific=c(2,2),
center=TRUE,scale=TRUE,weight=TRUE)
jiveRes <- omicsCompAnalysis(Input=list(B1,B2),Names=c("expr","mirna"),
method="JIVE",Rcommon=2,Rspecific=c(2,2),
center=TRUE,scale=TRUE,weight=TRUE)
o2plsRes <- omicsCompAnalysis(Input=list(B1,B2),Names=c("expr","mirna"),
method="O2PLS",Rcommon=2,Rspecific=c(2,2),
center=TRUE,scale=TRUE,weight=TRUE)
```

Description

This function applies the NonParametric Combination methodology on the integrative analysis of different omics data modalities. It retrieves genes associated to a given outcome, taking into account all omics data. First, each datatype is analyzed independently using the appropriate method. omicsNPC analyses continuous data (microarray) using limma, while count data (e.g., RNAseq) are first preprocessed with using the "voom" function. The user can also specify their own function for computing deregulation / association. The p-values from the single dataset analysis are then combined employing Fisher, Liptak and Tippett combining functions. The Tippett function returns findings which are supported by at least one omics modality. The Liptak function returns findings which are supported by most modalities. The Fisher function has an intermediate behavior between those of Tippett and Liptak.

Usage

```r
omicsNPC(dataInput, dataMapping, dataTypes = rep('continuous', length(dataInput)),
combMethods = c("Fisher", "Liptak", "Tippett"), numPerms = 1000,
numCores = 1, verbose = FALSE, functionGeneratingIndex = NULL,
outcomeName, allCombinations = FALSE,
dataWeights = rep(1, length(dataInput)) / length(dataInput), ...)
```
Arguments

dataInput List of ExpressionSet objects, one for each data modality.
dataMapping A data frame describing how to map measurements across datasets. See details for more information.
dataTypes Character vector with possible values: ‘continuous’, ‘count’. Alternatively, a list of functions for assessing deregulation / association with an outcome
combMethods Character vector with possible values: ‘Fisher’, ‘Liptak’, ‘Tippett’. If more than one is specified, all will be used.
numPerms Number of permutations
numCores Number of CPU cores to use
verbose Logical, if set to TRUE omicsNPC prints out the step that it performs
functionGeneratingIndex Function generating the indices for randomly permuting the samples
outcomeName Name of the outcome of interest / experimental factor, as reported in the design matrices. If NULL, the last column of the design matrices is assumed to be the outcome of interest.
allCombinations Logical, if TRUE all combinations of omics datasets are considered
dataWeights A vector specifying the weight to give to each dataset. Note that sum(dataWeights) should be 1.
... Additional arguments to be passed to the user-defined functions

Value

A list containing: stats0 Partial deregulation / association statistics pvalues0 The partial p-values computed on each dataset pvaluesNPC The p-values computed through NPC.

Author(s)

Nestoras Karathanasis, Vincenzo Lagani

References


Examples

# Load the data
data("TCGA_BRCA_Batch_93")
# Setting DataTypes, the first two ExpressionSets include RNAseq data,
# the third ExpressionSet includes Microarray data.
dataTypes <- c("count", "count", "continuous")
# Setting methods to combine pvalues
combMethods = c("Fisher", "Liptak", "Tippett")
# Setting number of permutations
numPerms = 1000
# Setting number of cores
numCores = 1
# Setting omicsNPC to print out the steps that it performs.
verbose = TRUE
# Run omicsNPC analysis.
# The output contains a data.frame of p-values, where each row corresponds to a gene,
# and each column corresponds to a method used in the analysis.

## Not run: out <- omicsNPC(dataInput = TCGA_BRC_A_Data, dataTypes = dataTypes,
combMethods = combMethods, numPerms = numPerms,
numCores = numCores, verbose = verbose)
## End(Not run)

PCA.selection

Select an optimal number of components using PCA

Description

Selects the optimal number of components from data using PCA. There are four different criteria available: accumulated variance explained, individual explained variance of each component, absolute value of variability or fixed number of components.

Usage

PCA.selection(Data, fac.sel, varthreshold=NULL, nvar=NULL, PCnum=NULL)

Arguments

- **Data**: Data matrix (with samples in columns and features in rows)
- **fac.sel**: Selection criteria ("%accum", "single]%", "rel.abs", "fixed.num")
- **varthreshold**: Threshold for ">% accum" or "single]%" criteria
- **nvar**: Threshold for "rel.abs"
- **PCnum**: Fixed number of components for "fixed.num"

Value

List containing:

- **PCAres**: List containing results of PCA, with fields "eigen", "var.exp", "scores" and "loadings"
- **numComps**: Number of components selected

Author(s)

Patricia Sebastian Leon

Examples

data(STATegRa_S3)
ps <- PCA.selection(Data=Block2.PCA, fac.sel="single%", varthreshold=0.03)
ps$numComps
plotRes

Plot component analysis results

Description
Plot scatterplots of scores or loadings, for common and distinctive parts as well as combined plots.

Usage
plotRes(object, comps=c(1, 2), what, type, combined, block, color=NULL, shape=NULL, labels=NULL, background=TRUE, palette=NULL, pointSize=4, labelSize=NULL, axisSize=NULL, titleSize=NULL)

Arguments

- **object**: caClass object containing component analysis results
- **comps**: If combined=FALSE, it indicates the x and y components of the type and block chosen. If combined=TRUE, it indicates the component to plot for the first block of information and the component for the second block of information to plot together. By default the components are set to c(1,2) if combined=FALSE and to c(1,1) if combined=TRUE.
- **what**: Either "scores" or "loadings"
- **type**: Either "common", "individual" or "both"
- **combined**: Logical indicating whether to make a simple plot of two components from one block, or components from different blocks
- **block**: Which block to plot, either "1" or "2" or the name of the block.
- **color**: Character specifying a pData column from the original data to use to color points
- **shape**: Character specifying a pData column to select point shape
- **labels**: Character specifying a pData column from which to take point labels
- **background**: Logical specifying whether to make a grey background
- **palette**: Vector giving the color palette for the plot
- **pointSize**: Size of plot points
- **labelSize**: Size of point labels if not NULL
- **axisSize**: Size of axis text
- **titleSize**: Size of title text

Value
ggplot object

Author(s)
Patricia Sebastian-Leon
Examples

data("STATegRa_S3")

B1 <- createOmicsExpressionSet(Data=Block1.PCA,pData=ed.PCA,
pDataDescr=c("classname"))

B2 <- createOmicsExpressionSet(Data=Block2.PCA,
pData=ed.PCA,pDataDescr=c("classname"))

# Omics components analysis
discoRes <- omicsCompAnalysis(Input=list(B1,B2),Names=c("expr","mirna"),
method="DISCOSCA",Rcommon=2,Rspecific=c(2,2),
center=TRUE,scale=TRUE,weight=TRUE)
jiveRes <- omicsCompAnalysis(Input=list(B1,B2),Names=c("expr","mirna"),
method="JIVE",Rcommon=2,Rspecific=c(2,2),
center=TRUE,scale=TRUE,weight=TRUE)
o2plsRes <- omicsCompAnalysis(Input=list(B1,B2),Names=c("expr","mirna"),
method="O2PLS",Rcommon=2,Rspecific=c(2,2),
center=TRUE,scale=TRUE,weight=TRUE)

# Scatterplot of scores variables associated to common components
# DISCO-SCA
plotRes(object=discoRes,comps=c(1,2),what="scores",type="common",
combined=FALSE,block="",color="classname",shape=NULL,labels=NULL,
background=TRUE,palette=NULL,pointSize=4,labelSize=NULL,
axisSize=NULL,titleSize=NULL)

# JIVE
plotRes(object=jiveRes,comps=c(1,2),what="scores",type="common",
combined=FALSE,block="",color="classname",shape=NULL,labels=NULL,
background=TRUE,palette=NULL,pointSize=4,labelSize=NULL,
axisSize=NULL,titleSize=NULL)

# O2PLS
# Scatterplot of scores variables associated to common components
# Associated to first block
p1 <- plotRes(object=o2plsRes,comps=c(1,2),what="scores",type="common",
combined=FALSE,block="expr",color="classname",shape=NULL,
labels=NULL,background=TRUE,palette=NULL,pointSize=4,labelSize=NULL,
axisSize=NULL,titleSize=NULL)

# Associated to second block
p2 <- plotRes(object=o2plsRes,comps=c(1,2),what="scores",type="common",
combined=FALSE,block="mirna",color="classname",shape=NULL,
labels=NULL,background=TRUE,palette=NULL,pointSize=4,labelSize=NULL,
axisSize=NULL,titleSize=NULL)

# Combined plot of scores variables associated to common components
plotRes(object=o2plsRes,comps=c(1,1),what="scores",type="common",
combined=TRUE,block="",color="classname",shape=NULL,
labels=NULL,background=TRUE,palette=NULL,pointSize=4,labelSize=NULL,
axisSize=NULL,titleSize=NULL)

# Loadings plot for individual components
# Separately for each block
p1 <- plotRes(object=discoRes,comps=c(1,2),what="loadings",type="individual",
combined=FALSE,block="expr",color="classname",shape=NULL,
labels=NULL,background=TRUE,palette=NULL,pointSize=4,labelSize=NULL,
axisSize=NULL,titleSize=NULL)
plotVAF

p2 <- plotRes(object=discoRes,comps=c(1,2),what="loadings",type="individual", 
combined=FALSE,block="mirna",color="classname",shape=NULL, 
labels=NULL,background=TRUE,palette=NULL,pointSize=4, 
labelSize=NULL,axisSize=NULL,titleSize=NULL)

# Combined plot
plotRes(object=discoRes,comps=c(1,1),what="loadings",type="individual", 
combined=TRUE,block="",color="classname",shape=NULL, 
labels=NULL,background=TRUE,palette=NULL,pointSize=4, 
labelSize=NULL,axisSize=NULL,titleSize=NULL)

---

plotVAF

Plot VAF (Variance Explained For) from Component Analysis

Description

This function visualises the VAF results from component analysis. The input is a `caClass-class` object from `omicsCompAnalysis`. VAF cannot be calculated if mode "O2PLS" was used. The plots for modes "DISCOSCA" and "JIVE" are different since DISCO-SCA distinctive components have some VAF in the other block. This VAF can be interpreted as an error in the rotation.

Usage

plotVAF(object, mainTitle="")

Arguments

object caClass-class object containing component analysis results
mainTitle Plot title

Value

ggplot object

Author(s)

Patricia Sebastian-Leon

Examples

data("STATegRa_S3")
require(ggplot2)
B1 <- createOmicsExpressionSet(Data=Block1.PCA,pData=ed.PCA, 
pDataDescr=c("classname"))
B2 <- createOmicsExpressionSet(Data=Block2.PCA, 
pData=ed.PCA,pDataDescr=c("classname"))
# Omics components analysis
discoRes <- omicsCompAnalysis(Input=list(B1,B2),Names=c("expr","mirna"), 
method="DISCOSCA",Rcommon=2,Rspecific=c(2,2), 
center=TRUE,scale=TRUE,weight=TRUE)
jiveRes <- omicsCompAnalysis(Input=list(B1,B2),Names=c("expr","mirna"), 
method="JIVE",Rcommon=2,Rspecific=c(2,2), 
center=TRUE,scale=TRUE,weight=TRUE)
# DISCO-SCA plotVAF
plotVAF(discoRes)

# JIVE plotVAF
plotVAF(jiveRes)

selectCommonComps

Select common components in two data blocks

Description
This function applies a Simultaneous Component Analysis (SCA). The idea is that the scores for both blocks should have a similar behaviour if the components are in the common mode. Evaluation is by the ratios between the explained variances (SSQ) of each block and the estimator. The highest component count with $0.8 < \text{ratio} < 1.5$ is selected.

Usage
selectCommonComps(X, Y, Rmax)

Arguments

X  Matrix of omics data
Y  Matrix of omics data
Rmax  Maximum number of common components to find

Value
A list with components:

- **common**  Optimal number of common components
- **ssqs**  Matrix of SSQ for each block and estimator
- **pptsq**  ggplot object showing SSQ for each block and estimator
- **pratios**  ggplot object showing SSQ ratios between each block and estimator

Author(s)
Patricia Sebastian-Leon

Examples

data(STATegRa_S3)
c <- selectCommonComps(X=Block1.PCA, Y=Block2.PCA, Rmax=3)
c$common
c$c$ssqs
c$c$pratios
STATegRa

Description

STATegRa is a package for the integrative analysis of multi-omic data-sets.
For full information, see the user’s guide.

See Also

STATegRaUsersGuide

STATegRa-deprecated

Description

These functions have been deprecated in STATegRa

Details

- holistOmics: omicsNPC

STATegRaUsersGuide

Description

Finds the location of the STATegRa User’s Guide and optionally opens it.

Usage

STATegRaUsersGuide(view = TRUE)

Arguments

view Whether to open a browser

Value

The path to the documentation

Author(s)

David Gomez-Cabrero

Examples

STATegRaUsersGuide(view=FALSE)
Description

mRNA data (Block1), miRNA data (Block2) and the design matrix (ed), from STATegRa_S1, provides selected data downloaded from https://tcga-data.nci.nih.gov/docs/publications/gbm_exp/. The mapping between miRNA and mRNA (mapdata, available in STATegRa_S2) contains, as a processed matrix, selected information available from TargetScan; we selected the set of miRNA target predictions for humans for those miRNA-mRNA pairs where both miRNA and mRNA were in Block1 and Block2 respectively.

The PCA version of the data (Block1_PCA, Block2_PCA, ed_PCA; available in STATegRa_S3), provides a similar data-set to Block1, Block2 and ed data; however in this case the data has been processed in order to provide a pedagogic example of OmicsPCA. Results obtained from OmicsPCA (omicsCompAnalysis) with the existing data should not be taken as clinically valid.

Format

Two matrices with mRNA and miRNA expression data, a design matrix that describes both and a mapping between miRNA and genes.

Author(s)

David Gomez-Cabrero, Patricia Sebastian-Leon, Gordon Ball

Source

(a) See https://tcga-data.nci.nih.gov/docs/publications/gbm_exp/. (b) Gabor Csardi, targetscan.Hs.eg.db: TargetScan miRNA target predictions for human. R package version 0.6.1

Examples

data(STATegRa_S1)
data(STATegRa_S2)
data(STATegRa_S3)

Description

Data were downloaded from TCGA data portal, https://tcga-data.nci.nih.gov/tcga/. We downloaded sixteen tumour samples and the sixteen matching normal, for Breast invasive carcinoma, BRCA, batch 93. Herein, three types of data modalities are included, RNAseq (TCGA_BRCA_Data$RNAseq), RNAseqV2 (TCGA_BRCA_Data$RNAseqV2) and Expression-Genes (TCGA_BRCA_Data$Microarray). The Data Level was set to Level 3. For each data type, we pooled all data to one matrix, where rows corresponded to genes and columns to samples. Only the first 100 genes are included.
STATegRa_data_TCGA_BRCA

Format

One list, which contains three ExpressionSet objects.

Author(s)

Nestoras Karathanasis, Vincenzo Lagani

Source

See https://tcga-data.nci.nih.gov/tcga/.

Examples

# load data
data(TCGA_BRCA_Batch_93)
Index

*Topic datagen
  - createOmicsExpressionSet, 16
  - getPreprocessing, caClass-method (getPreprocessing), 20
  - getScores, 21
  - getScores, caClass-method (getScores), 21
  - getVAF, 21
  - getVAF, caClass-method (getVAF), 22
  - ggplot, 32

bioDist, 2
  - character, character, bioMap, ExpressionSet-expression
  - (bioDist), 2
bioDistclass, 5
bioDistFeature, 5
bioDistFeature, character, list, numeric-method (bioDistFeature), 5
bioDistFeaturePlot, 7
bioDistW, 9
bioDistW, character, list, matrix-method (bioDistW), 9
bioDistWPlot, 10
bioDistWPlot, character, list, character-method (bioDistWPlot), 10
bioMap, 12
biplotRes, 13
biplotRes, caClass, character, numeric, character
  - (biplotRes), 13
  - Block1 (STATegRa_data), 34
  - Block2 (STATegRa_data), 34
caClass-class, 15
combiningMappings, 15
createOmicsExpressionSet, 16
createOmicsExpressionSet, matrix-method (createOmicsExpressionSet), 16
dist, 3
ed (STATegRa_data), 34
getInitialData, 17
getInitialData, caClass-method (getInitialData), 17
getLoadings, 18
getLoadings, caClass-method (getLoadings), 18
getMethodInfo, 19
getMethodInfo, caClass-method (getMethodInfo), 19
getPreprocessing, 20
getPreprocessing, caClass-method (getPreprocessing), 20
getScores, 21
getScores, caClass-method (getScores), 21
getVAF, 21
getVAF, caClass-method (getVAF), 22
ggplot, 32
holistOmics, 23
holistOmics, list, character-method (holistOmics), 23
mapdata (STATegRa_data), 34
modelSelection, 24
modelSelection, list, numeric, character-method (modelSelection), 24
omicsCompAnalysis, 17-22, 25, 28, 31, 34
omicsCompAnalysis, list, character, character, numeric, numeric-method (omicsCompAnalysis), 25
omicsNPC, 26, 33
omicsNPC, list, data.frame-method (omicsNPC), 26
omicsNPC, list, missing-method (omicsNPC), 26
PCA.selection, 24, 25, 28
PCA.selection, matrix, character-method (PCA.selection), 28
plotRes, 29
plotRes, caClass, numeric, character, character, logical, character
  - (plotRes), 29
plotVAF, 31
plotVAF, caClass-method (plotVAF), 31
selectCommonComps, 24, 25, 32
selectCommonComps, matrix, matrix, numeric-method (selectCommonComps), 32
STATegRa, 33
STATegRa-deprecated, 33
STATegRa-package (STATegRa), 33
STATegRa_data, 34
STATegRa_data_TCGA_BRCA, 34
STATegRaUsersGuide, 33, 33
TCGA_BRCA_Data
(STATegRa_data_TCGA_BRCA), 34