Package ‘biovizBase’

November 21, 2016

Version 1.22.0

Title Basic graphic utilities for visualization of genomic data.

Description The biovizBase package is designed to provide a set of utilities, color schemes and conventions for genomic data. It serves as the base for various high-level packages for biological data visualization. This saves development effort and encourages consistency.

Depends R (>= 2.10), methods

Imports grDevices, stats, scales, Hmisc, RColorBrewer, dichromat, BiocGenerics, S4Vectors (>= 0.9.25), IRanges (>= 1.99.28), GenomeInfoDb (>= 1.5.14), GenomicRanges (>= 1.23.21), SummarizedExperiment, Biostrings (>= 2.33.11), Rsamtools (>= 1.17.28), GenomicAlignments (>= 1.1.16), GenomicFeatures (>= 1.21.19), AnnotationDbi, VariantAnnotation (>= 1.11.4), ensembledb (>= 1.3.8)

Suggests BSgenome.Hsapiens.UCSC.hg19, TxDb.Hsapiens.UCSC.hg19.knownGene, BSgenome, rtracklayer, EnsDb.Hsapiens.v75, RUnit

License Artistic-2.0

LazyLoad Yes


biocViews Infrastructure, Visualization, Preprocessing

NeedsCompilation yes

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R topics documented:

biovizBase-package
biovizBase-package

biovizBase is a package which provides utilities and color scheme...

Description

biovizBase is a package which provides utilities and color scheme for higher level graphic package which aim to visualize biological data especially genetic data.
addStepping-method

Details

This package provides default color scheme for nucleotide, strand, amino acid, try to pass colorblind checking as possible as we can. And also provide giemsa stain result color scheme used to show cytoband. This package also provides utilities to manipulate and summarize raw data to get them ready to be visualized.

Description

Adding disjoint levels to a GenomicRanges object

Usage

```r
## S4 method for signature 'GenomicRanges'
addStepping(obj, group.name, extend.size = 0,
             fix = "center",
             group.selfish = TRUE)
```

Arguments

- `obj` A GenomicRanges object
- `group.name` Column name in the elementMetadata which specify the grouping information of all the entries. If provided, this will make sure all intervals belong to the same group will try to be on the same level and nothing falls in between.
- `extend.size` Adding invisible buffered region to the GenomicRanges object, if it's 10, then adding 5 at both end. This make the close neighbors assigned to the different levels and make your eyes easy to identify.
- `fix` "start", "end", or "center" (default) passed to `resize`, denoting what to use as an anchor for each element of `obj`, and add `extend.size` to it's current width.
- `group.selfish` Passed to `addStepping`, control whether to show each group as unique level or not. If set to FALSE, if two groups are not overlapped with each other, they will probably be layout in the same level to save space.

Details

This is a tricky question, for example, pair-end RNA-seq data could be represented as two set of GenomicRanges object, one indicates the read, one indicates the junction. At the same time, we need to make sure pair-ended read are shown on the same level, and nothing falls in between. For better visualization of the data, we may hope to add invisible extended buffer to the reads, so closely neighbored reads will be on the different levels.

Value

A modified GenmicRanges object with stepping as one column.

Author(s)

Tengfei Yin
Examples

library(GenomicRanges)
set.seed(1)
N <- 500
## sample GRanges
gr <- GRanges(seqnames =
  sample(c("chr1", "chr2", "chr3", "chrX", "chrY"),
         size = N, replace = TRUE),
IRanges(
  start = sample(1:300, size = N, replace = TRUE),
  width = sample(70:75, size = N, replace = TRUE)),
strand = sample(c("+", "-", "*"), size = N,
                replace = TRUE),
value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
group = sample(c("Normal", "Tumor"),
               size = N, replace = TRUE),
pair = sample(letters, size = N,
              replace = TRUE))

## grouping and extending
head(addStepping(gr))
head(addStepping(gr, group.name = "pair"))
gr.close <- GRanges(c("chr1", "chr1"), IRanges(c(10, 20), width = 9))
addStepping(gr.close)
addStepping(gr.close, extend.size = 5)

colorBlindSafePal  

Description

This function help users create a function based on specified color blind safe palette. And the returned function could be used for color generation.

Usage

genDichromatPalInfo()
genBrewerBlindPalInfo()
genBlindPalInfo()
colorBlindSafePal(palette)
blind.pal.info
brewer.pal.blind.info
dichromat.pal.blind.info

Arguments

palette  
A index numeric value or character. Please see blind.pal.info, the palette could be "pal_id" or names the row in which users could specify the palette you want to use.
Details

We get those color-blind safe palette based on http://colorbrewer2.org/ and http://geography.uoregon.edu/datagraphics/color_scales.htm those color are implemented in two packages, RColorBrewer and dichromat. But RColorBrewer doesn’t provide subset of color-blind safe palette info. And dichromat doesn’t group color based on “quality”, “sequential” and “divergent”, so we pick those color manually and create a combined palette, blind.pal.info.

colorBlindSafePal will return a function, this function will take two arguments, n and repeatable. If n is smaller than 3(n >= 3 is required by RColorBrewer), we use 3 instead and return a color vector. If n is over the maxcolors column in blind.pal.info, we use maxcolors instead when repeatable set to FALSE, if repeatable set to TRUE we repeat the color of all the allowed colors(length equals maxcolors) in the same order. This has special case in certain graphics which is always displayed side by side and don’t worry about the repeated colors being neighbors.

genBrewerBlindPalInfo return brewer.pal.blind.info data frame containing all color-blind safe palettes from brewer.pal.info defined in RColorBrewer, but it’s not only just subset of it, it also changes some maxcolors info.

genDichromatPalInfo return dichromat.pal.blind.info data frame.

genBlindPalInfo return blind.pal.info data frame.

Value

A color generating function with arguments n and repeatable. n specifying how many different discrete colors you want to get from them palette, and if repeatable turned on and set to TRUE, you can specify n even larger than maximum color. The color will be repeated following the same order.

Author(s)

Tengfei Yin

Examples

## Not run:
library(scales)
## brewer subse of only color blind palette
brewer.pal.blind.info
genBrewerBlindPalInfo()
## dichromat info
dichromat.pal.blind.info
genDichromatPalInfo()
## all color blind palette, adding id/pkg.
blind.pal.info
## with no parameters, just return blind.pal.info
colorBlindSafePal()
mypal <- colorBlindSafePal(20)
## or pass character name
mypal <- colorBlindSafePal("Set2")
mypal12 <- colorBlindSafePal(22)
show_col(mypal(12, repeatable = FALSE)) # warning
show_col(mypal(11, repeatable = TRUE)) # no warning, and repeat
show_col(mypal12(12))

## End(Not run)
**Description**

Test if a string contain any letters

**Usage**

`containLetters(obj, all=FALSE)`

**Arguments**

- `obj` String
- `all` If set to FALSE, return TRUE when any letters appears; if all is set to TRUE, return TRUE only when the string is composed of just letters.

**Details**

Useful when processing/sorting seqnames

**Value**

Logical value

**Author(s)**

tengfei

**Examples**

```r
containLetters("XYZ123")
containLetters("XYZ123", TRUE)
```

---

**Description**

Genomic sequencing of colorectal adenocarcinomas identifies a recurrent VTI1A-TCF7L2 fusion sample data set

**Usage**

`data(CRC)`

**Details**

The data used in this study is from this a paper http://www.nature.com/ng/journal/v43/n10/full/ng.936.html.

**Examples**

`data(CRC)`
Description
CRC-1 mutation and structural rearrangement

Usage
data(crc1.GeRL)

description
GenomicRangesList contains somatic mutation, rearrangement etc for a tumor sample, please check the reference for details.

References
Genomic sequencing of colorectal adenocarcinomas identifies a recurrent VTI1A-TCF7L2 fusion

Examples
data(crc1.GeRL)
crc1.GeRL

crunch Fetching GRanges from various data source

Description
Fetching Granges from various data source, currently supported objects are TxDb, EnsDb, GAlignments and BamFile.

Usage
## S4 method for signature 'TxDb'
crunch(obj, which, columns = c("tx_id", "tx_name","gene_id"),
       type = c("all", "reduce"), truncate.gaps = FALSE,
       truncate.fun = NULL, ratio = 0.0025)
## S4 method for signature 'EnsDb'
crunch(obj, which, columns = c("tx_id", "gene_name","gene_id"),
       type = c("all", "reduce"), truncate.gaps = FALSE,
       truncate.fun = NULL, ratio = 0.0025)
## S4 method for signature 'GAlignments'
crunch(obj, which, truncate.gaps = FALSE,
       truncate.fun = NULL, ratio = 0.0025)
## S4 method for signature 'BamFile'
crunch(obj, which, ..., type = c("gapped.pair", "raw", "all"),
       truncate.gaps = FALSE, truncate.fun = NULL, ratio = 0.0025)
Arguments

- **obj**: supported objects are TxDb, GAlignments and BamFile.
- **which**: GRanges object. For TxDb object, could also be a list. For EnsDb, it can also be a single filter object extending `BasicFilter-class` or a list of such objects.
- **columns**: columns to include in the output.
- **type**: default 'all' is to show the full model, 'reduce' is to show a single model.
- **truncate.gaps**: logical value, default FALSE. Whether to truncate gaps or not.
- **truncate.fun**: shrinkage function.
- **ratio**: numeric value, shrinking ratio.
- **...**: arguments passed to function `readGAlignments`.

Value

GRanges object.

Author(s)

Tengfei Yin

Examples

```r
library(biovizBase)
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
data(genesymbol, package = "biovizBase")
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
obj <- txdb
temp <- crunch(txdb, which = genesymbol["BRCA1"], type = "all")
temp <- crunch(txdb, which = genesymbol["BRCA1"], type = "reduce")

## Fetching data from a EnsDb object.
library(EnsDb.Hsapiens.v75)
edb <- EnsDb.Hsapiens.v75
gr <- genesymbol["BRCA1"]
seqlevels(gr) <- sub(seqlevels(gr), pattern="chr", replacement="")
temp <- crunch(edb, which=gr)

## Alternatively, use the GenenameFilter from the ensembldb package:
temp <- crunch(edb, which=GenenameFilter("BRCA1"))
```

---

darned_hg19_subset500  Subset of RNA editing sites in hg19...

Description

Subset of RNA editing sites in hg19

Usage

data(darned_hg19_subset500)
This data set provides a subset (500 sites only) of hg19 RNA editing sites, and originally from DARNED http://darned.ucc.ie/ for the hg19 assembly.

Examples

data(darned_hg19_subset500)
darned_hg19_subset500

estimateCoverage

Estimation of Coverage

## S4 method for signature 'BamFile'
estimateCoverage(x, maxBinSize = 2^14)

Arguments

x A BamFile object.
maxBinSize Max bin size.

Value

A GRanges object.

Author(s)

Michael Lawrence

flatGrl

Transform GRangesList to GRanges

Description

Transform GRangesList to GRanges.

Usage

flatGrl(object, indName = "grl_name")

Arguments

object a GRangesList object.
indName column named by 'indName' that groups the records by their original element in 'object'.
Details
This method is different from default stack, it integrate elementMetadata of GRangesList to the final coerced GRanges.

Value
A GRanges object.

Author(s)
Tengfei Yin

Examples
library(GenomicRanges)
g1 <- GRanges("chr1", IRanges(1:10, width = 5))
g2 <- GRanges("chr2", IRanges(1:10, width = 5))
obj <- GRangesList(g1, g2)
values(obj) <- data.frame(a = 1:2, b = letters[1:2])
stack(obj)
flatGrl(obj)

---

GCcontent

GC content computation for BSgenome

Description
Compute GC content in a certain region of a BSgenome object

Usage
GCcontent(obj, ..., view.width, as.prob = TRUE)

Arguments

obj
BSgenome object

...  
Arguments passed to getSeq method for BSgenome package.

view.width
Passed to letterFrequencyInSlidingView, the constant (e.g. 35, 48, 1000) size of the "window" to slide along obj. The specified letters are tabulated in each window of length view.width. The rows of the result (see value) correspond to the various windows.

as.prob
If TRUE return percentage of GC content, otherwise return counts.

Details
GC content is an interesting variable may be related to various biological questions. So we need a way to compute GC content in a certain region of a reference genome. GCcontent function is a wrapper around getSeq function in BSgenome package and letterFrequency, letterFrequencyInSlidingView in Biostrings package.

if the view.width is specified, the GC content will be computed in the sliding view
Value
Numeric value indicate count or percentage

Author(s)
Tengfei Yin

Examples
library(BSgenome.Hsapiens.UCSC.hg19)
GCcontent(Hsapiens, GRanges("chr1", IRanges(1e6, 1e6 + 1000)))

genesymbol

Description
Gene symbols with position

Usage
data(genesymbol)

details
This data set provides gene symbols in human with position and strand information, stored as GRanges object.

Examples
data(genesymbol)
head(genesymbol)
genesymbol["RBM17"]

getBioColor

Description
Color scheme getter for biological data

Usage
getBioColor(type = c("DNA_BASES_N", "DNA_BASES", "DNA_ALPHABET",
"RNA_BASES_N", "RNA_BASES", "RNA_ALPHABET",
"IUPAC_CODE_MAP", "AMINO_ACID_CODE", "AA_ALPHABET",
"STRAND", "CYTOBAND"),
source = c("option", "default"))
Arguments

type Color set based on which you want to get.
source "option" tries to get color scheme from Options. This allow user to edit the color globally. "default" gets fixed color scheme.

Details

Most data set specified in the type argument are defined in Biostrings package, such as "DNA_BASES", "DNA_ALPHABET", "RNA_BASES", "RNA_ALPHABET", "IUPAC_CODE_MAP", "AMINO_ACID_CODE", "AA_ALPHABET", please check the manual for more details.

"DNA_BASES_N" is just "DNA_BASES" with extra "N" used in certain cases, like the result from applyPileup in Rsamtools package. We start with the five most used nucleotides, A, T, C, G, N, In genetics, GC-content usually has special biological significance because GC pair is bound by three hydrogen bonds instead of two like AT pairs. So it has higher thermostability which could result in different significance, like higher annealing temperature in PCR. So we hope to choose warm color for G, C and cold color for A, T, and a color in between to represent N. They are chosen from diverging color set of color brewer. So we should be able to easily tell the GC enriched region. This set of color also passed vischeck for colorblind people.

In GRanges object, we have strand which contains three levels, +, -, *. We are using a qualitative color set from Color Brewer. This color set pass the colorblind test. It should be a safe color set to use to color strand.

For most default color scheme if they are under 18, we are trying to use package dichromat to set color for color blind people. But for data set that contains more than 18 objects, it’s not possible to assign colorblind-safe color to them anymore. So we need to repeat some color. It should not matter too much, because even normal people cannot tell the difference anymore.

Here are the definition for the data sets.

DNA_BASES Contains A, C, T, G.
DNA_ALPHABET This alphabet contains all letters from the IUPAC Extended Genetic Alphabet (see "?IUPAC_CODE_MAP") + the gap ("-") and the hard masking ("+") letters.
DNA_BASES_N Contains A, C, T, G, N
RNA_BASES_N Contains A, C, U, G, N
RNA_BASES Contains A, C, T, G
RNA_ALPHABET This alphabet contains all letters from the IUPAC Extended Genetic Alphabet (see ?IUPAC_CODE_MAP) where "T" is replaced by "U" + the gap ("-")) and the hard masking ("+") letters.
IUPAC_CODE_MAP The "IUPAC_CODE_MAP" named character vector contains the mapping from the IUPAC nucleotide ambiguity codes to their meaning.
AMINO_ACID_CODE Single-Letter Amino Acid Code (see "?AMINO_ACID_CODE").
AA_ALPHABET This alphabet contains all letters from the Single-Letter Amino Acid Code (see "?AMINO_ACID_CODE") + the stop ("*") the gap ("-")) and the hard masking ("+") letters
STRAND Contains "+", "+" and "-".
CYTOBAND Contains giemsa stain results: gneg, gpos25, gpos50, gpos75, gpos100, gvar, stalk, acen. Color defined in package geneplotter.

Value

A character of vector contains color(rgb), and the names of the vector is originally from the name of different data set. e.g. for DNA_BASES, it’s just A, C, T, G. This allow users to get color for a vector of specified names. Please see the examples below.
getFormalNames

Author(s)

Tengfei Yin

Examples

```r
opts <- getOption("biovizBase")
opts$DNABasesNColor[1] <- "red"
options(biovizBase = opts)
## get from option(default)
getBioColor("DNA_BASES_N")
## get default fixed color
getBioColor("DNA_BASES_N", source = "default")
## get colors for a sequence.
getBioColor("DNA_BASES_N")[seqs]
```

getFormalNames  Get formals from functions

Description

Get formals from functions, used for dispatching arguments inside.

Usage

```r
getFormalNames(..., remove.dots = TRUE)
```

Arguments

- `...` functions.
- `remove.dots` logical value, indicate remove dots in formals or not, default is TRUE.

Value

A character vector for formal arguments.

Author(s)

Tengfei Yin

Examples

```r
getFormalNames(plot, sapply)
```
getGaps

get gaps for a stepping transformed GRanges object

Description
Get gaps for a stepping transformed GRanges object, for visualization purpose. So an extra "stepping" column is required. Please see details below for motivation.

Usage
## S4 method for signature 'GRanges'
getGaps(obj, group.name = NULL, facets = NULL)

Arguments
- obj: A GRanges object.
- group.name: group name, such as transcript ID, this is the group method within each panel of facets and gaps will be computed for each group of intervals.
- facets: formula used for creating graphics, all variables must be present in the data.

Details
Since faceting is a subset and group process in visualization stage, some statistical computation need to be taken place after that. This leaves some computation like computing gaps hard based on solely GRanges object. Extra information like facets formula and group method would help to generate gaps which make sure they are aligned on the same level and within the same panel for grouped intervals. facets variables will be added to gaps GRanges along with group.name.

Value
A GRanges object representing gaps and for each row, the "stepping" column help later visualization and make sure gaps and intervals they are generated from are showed on the expected place.

Author(s)
Tengfei Yin

Examples
set.seed(1)
N <- 100
library(GenomicRanges)
gr <- GRanges(seqnames = sample(c("chr1", "chr2", "chr3"),
                                  size = N, replace = TRUE),
              IRanges(
                      start = sample(1:300, size = N, replace = TRUE),
                      width = sample(70:75, size = N, replace = TRUE),
                      strand = sample(c("+", "+", "-"), size = N,
                                      replace = TRUE),
                      value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
                      sample = sample(c("Normal", "Tumor"),
                                      size = N)),
              strand = sample(c("+", "+", "-"), size = N,
                               replace = TRUE),
              value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
              sample = sample(c("Normal", "Tumor"),
                               size = N))
getIdeoGR

getIdeoGR

Description

This function tries to parse ideogram information from seqlengths of a GRanges object. This information is usually used to plot chromosome background for karyogram or estimate proper lengths of chromosomes from data space for showing overview.

Usage

getIdeoGR(gr)

Arguments

gr

A GRanges object with or without lengths information.

Value

A ideogram GRanges object, each row indicate one single chromosome, and start with 1 and end with real chromosome length or estimated lengths.

Author(s)

Tengfei Yin

Examples

library(GenomicRanges)
data("hg19IdeogramCyto", package = "biovizBase")
hg19IdeogramCyto  
## without seqlengths, simply reduce  
getIdeoGR(hg19IdeogramCyto)  
## with seqlengths  
gr <- GRanges("chr1", IRanges(1,3))  
seqlevels(gr) <- c("chr1", "chr2", "chr3")  
nms <- c(100, 200, 300)  
names(nms) <- c("chr1", "chr2", "chr3")  
seqlengths(gr) <- nms  
gr  
getIdeoGR(gr)
getIdeogram  
Get ideogram.

Description
Get ideogram w/o cytoband for certain genome

Usage
getIdeogram(genome, subchr, cytobands=TRUE)

Arguments
<table>
<thead>
<tr>
<th>genome</th>
<th>Single specie names, which must be one of the result from ucscGenomes()$db. If missing, will invoke a menu for users to choose from.</th>
</tr>
</thead>
<tbody>
<tr>
<td>subchr</td>
<td>A character vector used to subset the result.</td>
</tr>
<tr>
<td>cytobands</td>
<td>If TRUE, return ideogram with gieStain and name column. If FALSE, simply return the genome information for each chromosome.</td>
</tr>
</tbody>
</table>

Details
This function require a network connection, it will parse the data on the fly, function is a wrapper of some functionality from rtracklayer package to get certain table like cytoBand, a full table schema could be found http://genome.ucsc.edu/cgi-bin/hgTables in UCSC genome browser. This is useful for visualization of the whole genome or single chromosome, you can see some examples in ggbio package.

Value
A GRanges object.

Author(s)
Tengfei Yin

Examples
## Not run: hg19IdeogramCyto <- getIdeogram("hg19", cytobands = TRUE)

getScale  
Get scale information from a GRanges

Description
Trying to get scale information from a GRanges object, used for circular view for geom "scale".

Usage
getScale(gr, unit = NULL, n = 100, type = c("M", "B", "sci"))
**getXScale**

**Arguments**

- **gr**: a GRanges object.
- **unit**: A numeric value for scale unit. Default NULL use argument n to estimate the unit.
- **n**: Integer value to indicate how many scale ticks to make.
- **type**: unit types to shown.

**Value**

A GRanges object, with extra column: "type" indicate it's longer major ticks or shorter minor ticks. "scale.y" indicates y height for major and minor ticks. Default ratio is 3:1.

**Author(s)**

Tengfei Yin

---

**Description**

get x scale breaks and labels for GRanges with different coordinates (currently only "truncate_gaps" and "genome" supported).

**Usage**

```r
## S4 method for signature 'GRanges'
getXScale(obj, type = c("default", "all", "left", "right"))
```

**Arguments**

- **obj**: a GRanges object. "coord" in metadata shows proper coordinates transformation for this object.
- **type**: types of labels for transformed data.

**Value**

list of breaks and labels.

**Author(s)**

Tengfei Yin
**Examples**

```r
library(GenomicRanges)
gr1 <- GRanges("chr1", IRanges(start = c(100, 300, 600),
                              end = c(200, 400, 800)))
shrink.fun1 <- shrinkageFun(gaps(gr1), max.gap = maxGap(gaps(gr1), 0.15))
shrink.fun2 <- shrinkageFun(gaps(gr1), max.gap = 0)
s1 <- shrink.fun1(gr1)
getXScale(s1)
# coord: genome
set.seed(1)
gr1 <- GRanges("chr1", IRanges(start = as.integer(runif(20, 1, 100)),
                              width = 5))
gr2 <- GRanges("chr2", IRanges(start = as.integer(runif(20, 1, 100)),
                              width = 5))
gr <- c(gr1, gr2)
fr.t <- transformToGenome(gr, space.skip = 1)
getXScale(fr.t)
```

---

**getYLab-method**

```r
parse x and y label information from a specific object
```

### Description

parse y label information, object specific.

### Usage

```r
## S4 method for signature 'TxDb'
getYLab(obj)
## S4 method for signature 'GRanges'
getXLab(obj)
## S4 method for signature 'GRangesList'
getXLab(obj)
## S4 method for signature 'GAlignments'
getXLab(obj)
```

### Arguments

- **obj**
  - A TxDb object.

### Value

- a string.

### Author(s)

Tengfei Yin
**hg19Ideogram**

| hg19Ideogram | *Hg19 ideogram without cytoband information* |

**Description**

Hg19 ideogram without cytoband information

**Usage**

data(hg19Ideogram)

**Details**

This data set provides hg19 genome information without cytoband information.

**Examples**

data(hg19Ideogram)
hg19Ideogram

---

**hg19IdeogramCyto**

| hg19IdeogramCyto | *Hg19 ideogram with cytoband information* |

**Description**

Hg19 ideogram with cytoband information

**Usage**

data(hg19IdeogramCyto)

**Details**

This data set provides hg19 genome information with cytoband information.

**Examples**

data(hg19IdeogramCyto)
hg19IdeogramCyto
ideogram without cytoband information

**Description**
ideogram without cytoband information

**Usage**
data(ideo)

**Details**
This data set provides hg19, hg18, mm10, mm9 genome information without cytoband information as a lit.

**Examples**
data(ideo)
ideo

ideogram with cytoband information

**Description**
ideogram with cytoband information

**Usage**
data(ideoCyto)

**Details**
This data set provides hg19, hg18, mm10, mm9 genome information with cytoband information as a lit.

**Examples**
data(ideoCyto)
ideoCyto
isIdeogram  

**Ideogram checking**

**Description**

Check if an object is ideogram or not

**Usage**

```r
isIdeogram(obj)
```

**Arguments**

- `obj` object

**Details**

Simply test if it’s the result coming from `getIdeogram` function or not, make sure it’s a GRanges and with extra column

**Value**

A logical value to indicate it’s a ideogram or not.

**Author(s)**

Tengfei Yin

**Examples**

```r
data(hg19IdeogramCyto)
data(hg19Ideogram)
isIdeogram(hg19IdeogramCyto)
isIdeogram(hg19Ideogram)
```

isMatchedWithModel  

**Utils for Splicing Summary**

**Description**

Utilities used for summarizing isoforms

**Usage**

```r
isJunctionRead(cigar)
isMatchedWithModel(model, gr)
```

**Arguments**

- `cigar` A CIGAR string vector.
- `model` A GRanges object.
- `gr` A GRanges object.
isSimpleIdeogram

Details

isJunctionRead simply parsing the CIGAR string to see if there is "N" in between and return a logical vector of the same length as cigar parameters, indicate it's junction read or not.

isMatchedWithModel mapping gr to model, and counting overlapped cases for each row of model. If gr contains all the read, this will return a logical vector of the same length as gr, and indicate if each read is the support for this model. NOTICE: we only assume it's a full model, so each model here is simply one isoform. So we only treat the gaped reads which only overlapped with two consecutive exons in model as one support for it.

Value

Logical vectors.

Author(s)

Tengfei Yin

Examples

library(GenomicAlignments)
bamfile <- system.file("extdata", "SRR027894subRBM17.bam", package="biovizBase")
## get index of junction read
which(isJunctionRead(cigar(readGAlignments(bamfile))))
##
model <- GRanges("chr1", IRanges(c(10, 20, 30, 40), width = 5))
gr <- GRanges("chr1", IRanges(c(10, 10, 12, 22, 33), c(31, 40, 22, 32, 44)))
isMatchedWithModel(model, gr)

isSimpleIdeogram

Simple ideogram checking

Description

Check if an object is a simple ideogram or not

Usage

isSimpleIdeogram(obj)

Arguments

obj object

Details

test if it's GRanges or not, doesn't require cytoband information. But it double check to see if there is only one entry per chromosome

Value

A logical value to indicate it's a simple ideogram or not.
**maxGap-method**

**Author(s)**

tengfei

**Examples**

```r
data(hg19IdeogramCyto)
data(hg19Ideogram)
isSimpleIdeogram(hg19IdeogramCyto)
isSimpleIdeogram(hg19Ideogram)
```

**Description**

Compute an estimated max gap information, which could be used as max.gap argument in `shrinkageFun`.

**Usage**

```r
## S4 method for signature 'GenomicRanges'
maxGap(obj, ratio = 0.0025)
```

**Arguments**

- `obj` GenomicRanges object
- `ratio` Multiple by the range of the provided gaps as the max gap.

**Details**

This function tries to estimate an appropriate max gap to be used for creating a shrinkage function.

**Value**

A numeric value

**Author(s)**

Tengfei Yin

**See Also**

`shrinkageFun`
Examples

```r
require(GenomicRanges)
gr1 <- GRanges("chr1", IRanges(start = c(100, 300, 600),
end = c(200, 400, 800)))
gr2 <- GRanges("chr1", IRanges(start = c(100, 350, 550),
end = c(220, 500, 900)))
gaps.gr <- intersect(gaps(gr1, start = min(start(gr1)));
gaps(gr2, start = min(start(gr2))))
shrink.fun <- shrinkageFun(gaps.gr, max.gap = maxGap(gaps.gr))
shrink.fun(gr1)
shrink.fun(gr2)
```

*mold*

*mold data into data.frame*

Description

*mold data into data.frame* usued for visualization, so it’s sometimes not as simple as coercion, for example, we add midpoint varialbe for mapping.

Usage

```r
## S4 method for signature 'IRanges'
mold(data)
## S4 method for signature 'GRanges'
mold(data)
## S4 method for signature 'GRangesList'
mold(data, indName = "grl_name")
## S4 method for signature 'Seqinfo'
mold(data)
## S4 method for signature 'matrix'
mold(data)
## S4 method for signature 'eSet'
mold(data)
## S4 method for signature 'ExpressionSet'
mold(data)
## S4 method for signature 'RangedSummarizedExperiment'
mold(data, assay.id = 1)
## S4 method for signature 'Views'
mold(data)
## S4 method for signature 'Rle'
mold(data)
## S4 method for signature 'RleList'
mold(data)
## S4 method for signature 'VRanges'
mold(data)
```

Arguments

data  data to be mold into data.frame with additional column that helped mapping. For example we add ‘midpoint’ variable to a ranged converted data.frame.
When collapsing a `GRangesList`, list names are aggregated into an extra column named by this parameter.

Define the assay id you want to convert into a `data.frame`.

**Value**

A `data.frame` object.

**Author(s)**

Tengfei Yin

---

**Description**

Utilities for parsing (un)evaluated arguments

**Usage**

```r
parseArgsForAes(args)
parseArgsForNonAes(args)
```

**Arguments**

- `args` arguments list.

**Value**

For `parseArgsForAes` return an unevaluated arguments.

For `parseArgsForNonAes` return a evaluated/quoted arguments.

**Author(s)**

Tengfei Yin

**Examples**

```r
args <- alist(a = color, b = "b")
# parseArgsForAes(args)
```
**Description**

This function summarize reads from bam files for nucleotides on single base unit in a given region, this allows the downstream mismatch summary analysis.

**Usage**

```r
pileupAsGRanges(bams, regions, DNABases=c("A", "C", "G", "T", "N"), ...)
```

**Arguments**

- `bams` A character which specify the bam file path.
- `regions` A GRanges object specifying the region to be summarized. This passed to which arguments in `ApplyPileupsParam`.
- `DNABases` Nucleotide type you want to summarize in the result and in specified order. It must be one or more of A,C,G,T,N.
- `...` Extra parameters passed to `ApplyPileupsParam`.

**Details**

It’s a wrapper around `applyPileup` function in Rsamtools package. more detailed control could be found under manual of `ApplyPileupsParam` function in Rsamtools. `pileupAsGRanges` function return a GRanges object which including summary of nucleotides, depth, bam file path. This object could be read directly into `pileupGRangesAsVariantTable` function for mismatch summary.

**Value**

A GRanges object, each row is one single base unit. and elementMetadata contains summary about this position about all nucleotides specified by DNABases. and depth for total reads, bam for file path.

**Author(s)**

Michael Lawrence, Tengfei Yin

**Examples**

```r
## Not run:
library(Rsamtools)
data(genesymbol)
library(BSgenome.Hsapiens.UCSC.hg19)
bamfile <- system.file("extdata", "SRR027894subRBM17.bam", package="biovizBase")
test <- pileupAsGRanges(bamfile, region = genesymbol["RBM17"])
test.match <- pileupGRangesAsVariantTable(test, Hsapiens)
head(test[, -7])
head(test.match[, -5])
## End(Not run)
```
pileupGRangesAsVariantTable

Mismatch summary

Description

Compare to reference genome and compute mismatch summary for certain region of reads.

Usage

```
pileupGRangesAsVariantTable(gr, genome, DNABases=c("A", "C", "G", "T", "N"))
```

Arguments

- `gr`: A GRanges object, with nucleotides summary, each base take one column in elementMetadata or user can simply passed the returned result from `pileupAsGRanges` function to this function.
- `genome`: BSgenome object, need to be the reference genome.
- `DNABases`: Nucleotide types contained in passed GRanges object. Default is A/C/G/T/N, it tries to match the column names in elementMetadata to those default nucleotides. And treat the matched column as base names.

Details

User need to make sure to pass the right reference genome to this function to get the right summary. The function drop the position has no reads and only keep the region with coverage in the summary. The result could be used to show stacked barchart for mismatch summary.

Value

A GRanges object. Containing the following elementMetadata

- `refNucleotide` in reference genome.
- `readNucleotide` contained in the reads at particular position, if multiple nucleotide, either matched or unmatched are found, they will be summarized in different rows.
- `countCount` for read column.
- `matchLogical` value, whether matched to reference genome or not
- `bamCharacter` indicate bam file path.

Author(s)

Michael Lawrence, Tengfei Yin

Examples

```r
## Not run:
library(Rsamtools)
data(genesymbol)
library(BSgenome)
bamfile <- system.file("extdata", "SRR027894subRBM17.bam", package="biovizBase")
test <- pileupAsGRanges(bamfile, region = genesymbol["RBM17"])
```
plotColorLegend <- pileupGRangesAsVariantTable(test, Hsapiens)
head(test[,7])
head(test.match[,5])
## End(Not run)

---

plotColorLegend

Show colors

Description

Plot color legend, simple way to check your default color scheme

Usage

plotColorLegend(colors, labels, title)

Arguments

colors       A character vector of colors
labels       Labels to put aside colors, if missing, use names of the colors character vector.
title        Title for the color legend.

Details

Show color scheme as a legend style, labels

Value

A graphic device showing color legend.

Author(s)

Tengfei Yin

Examples

cols <- getOption("biovizBase")$baseColor
plotColorLegend(cols, title = "strand legend")
**showColor**  
*Show colors*

**Description**

Show colors with color string or names of color vectors.

**Usage**

```r
showColor(colors, label = c("color", "name"))
```

**Arguments**

- `colors`: A color character vector.
- `label`: "color" label color with simple color string, and "name" label color with names of the vectors.

**Value**

A plot.

**Author(s)**

Tengfei Yin

**Examples**

```r
## Not run:
showColor(getBioColor("CYTOBAND"))
## End(Not run)
```

---

**shrinkageFun-method**  
*Shrinkage function*

**Description**

Create a shrinkage function based on specified gaps and shrinkage rate.

**Usage**

```r
## For IRanges
## S4 method for signature 'IRanges'
shrinkageFun(obj, max.gap = 1L)

## For GenomicRanges
## S4 method for signature 'GenomicRanges'
shrinkageFun(obj, max.gap = 1L)

is_coord_truncate_gaps(obj)
```
**splitByFacets**

**Description**

Split a GRanges by formula into GRangesList. Parse variables in formula and form a interaction factors then split the GRanges by the factors.

**Arguments**

- **obj**: GenomicRanges object which represent gaps
- **max.gap**: Gaps to be kept, it’s a fixed value, if this value is bigger than certain gap interval, then that gap is not going to be shrunk.

**Details**

shrinkageFun function will read in a GenomicRanges object which represent the gaps, and return a function which works for another GenomicRanges object, to shrink that object based on previously specified gaps shrinking information. You could use this function to treat multiple tracks(e.g. GRanges) to make sure they shrunk based on the common gaps and the same ratio.

is_coord_truncate_gaps is used to check if a GRanges object is in "truncate_gaps" coordinates or not.

**Value**

A shrinkage function which could shrink a GenomicRanges object, this function will add coord "truncate_gaps" and max.gap to metadata, ".ori" for orignal data as extra column

**Author(s)**

Michael Lawrence, Tengfei Yin

**Examples**

```r
library(GenomicRanges)
g1 <- GRanges("chr1", IRanges(start = c(100, 300, 600),
                    end = c(200, 400, 800)))
shrink.fun1 <- shrinkageFun(gaps(g1), max.gap = maxGap(gaps(g1), 0.1))
shrink.fun2 <- shrinkageFun(gaps(g1), start(g1), end(g1)), max.gap = maxGap(gaps(g1), 0.1)
shrink.fun3 <- shrinkageFun(gaps(g1), max.gap = 0)
s1 <- shrink.fun1(g1)
s2 <- shrink.fun2(g1)
s3 <- shrink.fun3(g1)
metadata(s1)$coord
values(s1)$ori
is_coord_truncate_gaps(s1)
```

---

**splitByFacets**

**split a GRanges by formula**
### Usage

```r
## S4 method for signature 'GRanges,formula'
splitByFacets(object, facets)
## S4 method for signature 'GRanges,GRanges'
splitByFacets(object, facets)
## S4 method for signature 'GRanges,NULL'
splitByFacets(object, facets)
## S4 method for signature 'GRanges,missing'
splitByFacets(object, facets)
```

### Arguments

- **object**: GRanges object.
- **facets**: formula object, such as `~ seqnames`. Or GRanges object, or NULL.

### Details

- If facets is formula, factors are created based on interaction of variables in formula, then split it with this factor. If facets is GRanges object, it first subset the original data by facets GRanges, then split by each region in the facets. If facets is NULL, split just by seqnames. This function is used to perform computation in conjunction with facets arguments in higher level graphic function.

### Value

A GRangesList.

### Examples

```r
library(GenomicRanges)
N <- 1000
gr <- GRanges(seqnames =
sample(c("chr1", "chr2", "chr3"),
size = N, replace = TRUE),
IRanges(
    start = sample(1:300, size = N, replace = TRUE),
    width = sample(70:75, size = N, replace = TRUE)),
strand = sample(c("+", "-", "*"), size = N,
    replace = TRUE),
value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
sample = sample(c("Normal", "Tumor"),
    size = N, replace = TRUE),
pair = sample(letters, size = N,
    replace = TRUE))

facets <- sample ~ seqnames
splitByFacets(gr, facets)
splitByFacets(gr)
gr.sub <- GRanges("chr1", IRanges(c(1, 200, 250), width = c(50, 10, 30)))
splitByFacets(gr, gr.sub)
```
### strip_formula_dots

*strip dots around a formula variables*

**Description**

Strip dots around variables in a formula.

**Usage**

```r
strip_formula_dots(formula)
```

**Arguments**

- `formula` A formula, such as `coverage ~ seqnames`, or `..coverage.. ~ seqnames`.

**Value**

A formula without ".." around for all variables.

**Author(s)**

Tengfei Yin

**Examples**

```r
obj <- ..coverage.. ~ seqnames
strip_formula_dots(obj)
```

---

### subsetArgsByFormals

*Subset list of arguments by functions*

**Description**

Find arguments matched by formals of passed functions.

**Usage**

```r
subsetArgsByFormals(args, ..., remove.dots = TRUE)
```

**Arguments**

- `args` list of arguments with names indicate the formals.
- `...` functions used to parse formals.
- `remove.dots` logical value indicate whether to include dots in formals or not.

**Value**

Arguments that matched with passed function.
transformGRangesForEvenSpace

Author(s)

Tengfei Yin

Examples

args <- list(x = 1:3, simplify = TRUE, b = "b")
subsetArgsByFormals(args, plot, sapply)

transformGRangesForEvenSpace

Transform GRanges with New Coordinates

Description

For graphics, like linked plot, e.g. generated by qplotRangesLinkedToData function in package ggbio. we need to generate a new set of coordinates which is used for even spaced statistics track.

Usage

transformGRangesForEvenSpace(gr)

Arguments

gr  
A GRanges object.

Details

Most used internally for special graphics, like qplotRangesLinkedToData function in package ggbio.

Value

A GRanges object as passed in, with new column x.new which indicate the static track coordinates, in this way, we could map the new coordinates with the old one.

Author(s)

Tengfei Yin

Examples

library(GenomicRanges)
gr <- GRanges("chr1", IRanges(seq(1,100, length.out = 10), width = 5))
library(biovizBase)
transformGRangesForEvenSpace(gr)
transformToGenome

Transform GRanges to different coordinates and layout

Description

Used for coordinate genome transformation, other transformation in circular view.

Usage

```r
## S4 method for signature 'GRanges'
transformToGenome(data, space.skip = 0.1, chr.weight = NULL)
## S4 method for signature 'GRangesList'
transformToGenome(data, space.skip = 0.1, chr.weight = NULL)
## S4 method for signature 'GRanges'
transformToArch(data, width = 1)
transformToCircle(data, x = NULL, y = NULL, ylim = NULL,
                   radius = 10, trackWidth = 10,
                   direction = c("clockwise", "anticlockwise"),
                   mul = 0.05)

transformToRectInCircle(data, y = NULL, space.skip = 0.1, trackWidth = 10, radius = 10,
                         direction = c("clockwise", "anticlockwise"),
                         n = 100, mul = 0.05, chr.weight = NULL)

transformToBarInCircle(data, y = NULL, space.skip = 0.1, trackWidth = 10, radius = 10,
                        direction = c("clockwise", "anticlockwise"),
                        n = 100, mul = 0.05, chr.weight = NULL)

transformToSegInCircle(data, y = NULL, space.skip = 0.1, trackWidth = 10, radius = 10,
                        direction = c("clockwise", "anticlockwise"), n =
                        100, chr.weight = NULL)

transformToLinkInCircle(data, linked.to, space.skip = 0.1, trackWidth = 10, radius = 10,
                         link.fun = function(x, y, n = 100) bezier(x, y, evaluation = n),
                         direction = c("clockwise", "anticlockwise"), chr.weight = NULL)

transformDfToGr(data, seqnames = NULL, start = NULL, end = NULL,
                 width = NULL, strand = NULL,
                 to.seqnames = NULL, to.start = NULL, to.end = NULL,
                 to.width = NULL, to.strand = NULL, linked.to = to.gr)

## S4 method for signature 'GRanges'
transformToDf(data)
```
is_coord_genome(data)

**Arguments**

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>data</td>
<td>a GRanges object. For function <code>transformDfToGr</code> it's data frame.</td>
</tr>
<tr>
<td>x</td>
<td>character for variable as x axis used for transformation.</td>
</tr>
<tr>
<td>y</td>
<td>character for variable as y axis used for transformation.</td>
</tr>
<tr>
<td>ylim</td>
<td>numeric range to control the y limits on tracks when 'y' information is involved.</td>
</tr>
<tr>
<td>space.skip</td>
<td>numeric values indicates skipped ratio of whole space, not skipped space is identical between each space.</td>
</tr>
<tr>
<td>radius</td>
<td>numeric value, indicates radius when transform to a circle.</td>
</tr>
<tr>
<td>trackWidth</td>
<td>numeric value, for track width.</td>
</tr>
<tr>
<td>direction</td>
<td>&quot;clockwise&quot; or &quot;counterclockwise&quot;, for layout or transform direction to circle.</td>
</tr>
<tr>
<td>mul</td>
<td>numeric value, passed to <code>expand_range</code> function, to control margin of y in the track.</td>
</tr>
<tr>
<td>n</td>
<td>integer value, control interpolated points numbers.</td>
</tr>
<tr>
<td>linked.to</td>
<td>a column name of GRanges object, indicate the linked line’s end point which represented as a GRanges too.</td>
</tr>
<tr>
<td>link.fun</td>
<td>function used to generate linking lines.</td>
</tr>
<tr>
<td>seqnames</td>
<td>character or integer values for column name or index indicate variable mapped to seqnames, default NULL use &quot;seqnames&quot;.</td>
</tr>
<tr>
<td>start</td>
<td>character or integer values for column name or index indicate variable mapped to start, default NULL use &quot;start&quot;.</td>
</tr>
<tr>
<td>end</td>
<td>character or integer values for column name or index indicate variable mapped to end, default NULL use &quot;end&quot;.</td>
</tr>
<tr>
<td>width</td>
<td>character or integer values for column name or index indicate variable mapped to width, default NULL use &quot;width&quot;.</td>
</tr>
<tr>
<td>strand</td>
<td>character or integer values for column name or index indicate variable mapped to strand, default NULL use &quot;strand&quot;.</td>
</tr>
<tr>
<td>to.seqnames</td>
<td>character or integer values for column name or index indicate variable mapped to linked seqnames, default NULL, create GRanges without new GRanges attached as column. If this variable is not NULL, this mean you try to parse linked GRanges object.</td>
</tr>
<tr>
<td>to.start</td>
<td>character or integer values for column name or index indicate variable mapped to start of linked GRanges, default NULL use &quot;to.start&quot;.</td>
</tr>
<tr>
<td>to.end</td>
<td>character or integer values for column name or index indicate variable mapped to end of linked GRanges, default NULL use &quot;to.end&quot;.</td>
</tr>
<tr>
<td>to.width</td>
<td>character or integer values for column name or index indicate variable mapped to width of linked GRanges, default NULL use &quot;to.width&quot;.</td>
</tr>
<tr>
<td>to.strand</td>
<td>character or integer values for column name or index indicate variable mapped to strand, default NULL use &quot;to.strand&quot; or just use *.</td>
</tr>
<tr>
<td>chr.weight</td>
<td>numeric vectors which sum to &lt;1, the names of vectors has to be matched with seqnames in seqinfo, and you can only specify part of the seqnames, other lengths of chromosomes will be assined proportionally to their seqlengths, for example, you could specify chr1 to be 0.5, so the chr1 will take half of the space and other chromosomes squeezed to take left of the space.</td>
</tr>
</tbody>
</table>
Value

A GRanges object, with calculated new variables, including ".circle.x" for transformed x position, ".circle.y" for transformed y position, ".circle.angle" for transformed angle.

Author(s)

Tengfei Yin

Examples

```r
library(biovizBase)
library(GenomicRanges)
set.seed(1)
gr1 <- GRanges("chr1", IRanges(start = as.integer(runif(20, 1, 2e9)),
width = 5))
gr2 <- GRanges("chr2", IRanges(start = as.integer(runif(20, 1, 2e9)),
width = 5))
gr <- c(gr1, gr2)
gr.t <- transformToGenome(gr, space.skip = 0.1)
is_coord_genome(gr.t)
transformToCircle(gr.t)
transformToRectInCircle(gr)
transformToSegInCircle(gr)
values(gr1)$to.gr <- gr2
transformToLinkInCircle(gr1, linked.to = "to.gr")
```
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