Package ‘doppelgangR’

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Maintainer  Levi Waldron <lwaldron.research@gmail.com>
Depends    R (>= 3.3), Biobase, BiocParallel
Suggests   BiocStyle, knitr, rmarkdown, curatedOvarianData, ROCR, pROC, RUnit, simulatorZ, proxy
Imports    sva, impute, digest, mnormt, methods, grDevices, graphics, stats, utils
Author     Levi Waldron, Markus Riester, Marcel Ramos
Version    1.4.0
Date       2017-04-12
License    GPL (>=2.0)
Title      Identify likely duplicate samples from genomic or meta-data
Description The main function is doppelgangR(), which takes as minimal input a list of ExpressionSet object, and searches all list pairs for duplicated samples. The search is based on the genomic data (exprs(eset)), phenotype/clinical data (pData(eset)), and "smoking guns" - supposedly unique identifiers found in pData(eset).
URL        https://github.com/lwaldron/doppelgangR
BugReports https://github.com/lwaldron/doppelgangR/issues
VignetteBuilder knitr
biocViews   RNASeq, Microarray, GeneExpression, QualityControl
NeedsCompilation no

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Details

The DESCRIPTION file:

```
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Maintainer: Levi Waldron <lwaldron.research@gmail.com>
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```

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- doppelgangR: doppelgangR
- doppelgangR-package: Identify likely duplicate samples from genomic or meta-data
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vectorHammingDist Calculate Hamming Distance between two vectors, using pairwise complete observations.
vectorWeightedDist Calculate a weighted distance between two vectors, using pairwise complete observations.

Author(s)

Levi Waldron, Markus Riester, Marcel Ramos

Maintainer: Levi Waldron <lwaldron.research@gmail.com>

corFinder Calculate pair-wise correlations between samples using the expr() slots of a list of two ExpressionSets.

Description

This function acts as a wrapper around ComBat (sva package) and cor(), to calculate pairwise correlations within one or between two ExpressionSets.

Usage

corFinder(eset.pair, separator = ":", use.ComBat = TRUE, ...)

Arguments

eset.pair a list of ExpressionSets, with two elements. If the two elements are identical, return the correlation matrix for pairs of samples in the first element. If not identical, return pairs between the two elements.
separator Separator between dataset name and sample name. Dataset names are added to sample names to keep track of dataset of origin.
use.ComBat Use the sva::ComBat function for batch correction of the expr() data between the two datasets.
... Extra arguments passed to the cor() function.
Value
Returns a matrix of sample-wise Pearson Correlations.

Author(s)
Levi Waldron, Markus Riester, Marcel Ramos

Examples
```r
eexample("phenoFinder")
corFinder(esets2)
```

DoppelGang-class

Description
S4 class containing results of doppelgangR() function.

Objects from the Class
Objects can be created by calls of the form new(DoppelGang ...)

Slots
- `fullresults`: Object of class "list"
- `summaryresults`: Object of class "data.frame"
- `inputargs`: "list" object containing input arguments to doppelgangR() function.

Methods
- `plot` signature(x = "DoppelGang"): See ?plot,DoppelGang-method
- `summary` signature(object = "DoppelGang"): produces a data.frame of potential doppel-gangers, with similarity measures and available phenoData for both of each potential doppel-ganger pair. Can be saved as a csv table and inspected carefully in a spreadsheet.
- `print` signature(x = "DoppelGang"): ...
- `show` signature(object = "DoppelGang"): ...

Author(s)
Levi Waldron and Markus Riester

See Also
plot,DoppelGang-method
Description

Identify samples with suspiciously high correlations and phenotype similarities

Usage
doppelgangR(esets, separator = ":", corFinder.args = list(separator = separator,
use.ComBat = TRUE, method = "pearson"), phenoFinder.args = list(separator = separator,
vectorDistFun = vectorWeightedDist), outlierFinder.expr.args = list(bonf.prob = 0.5,
transFun = atanh, tail = "upper"), outlierFinder.pheno.args = list(normal.upper.thresh = 0.99,
bonf.prob = NULL, tail = "upper"), smokingGunFinder.args = list(transFun = I),
impute.knn.args = list(k = 10, rowmax = 0.5, colmax = 0.8,
maxp = 1500, rng.seed = 362436069), manual.smokingguns = NULL,
automatic.smokingguns = FALSE, within.datasets.only = FALSE,
intermediate.pruning = FALSE, cache.dir = "cache", BPPARAM = bpparam(),
verbose = TRUE)

Arguments

esets a list of ExpressionSets, containing the numeric and phenotypic data to be analyzed.
separator a delimiter to use between dataset names and sample names
corFinder.args a list of arguments to be passed to the corFinder function.
phenoFinder.args a list of arguments to be passed to the phenoFinder function. If NULL, samples
with similar phenotypes will not be searched for.
outlierFinder.expr.args a list of arguments to be passed to outlierFinder when called for expression data
outlierFinder.pheno.args a list of arguments to be passed to outlierFinder when called for phenotype data
smokingGunFinder.args a list of arguments to be passed to smokingGunFinder
impute.knn.args a list of arguments to be passed to impute::impute.knn. Set to NULL to do no
knn imputation.
manual.smokingguns a character vector of phenoData columns that, if identical, will be considered
evidence of duplication
automatic.smokingguns automatically look for "smoking guns." If TRUE, look for phenotype variables
that are unique to each patient in dataset 1, also unique to each patient in dataset
2, but contain exact matches between datasets 1 and 2.
within.datasets.only If TRUE, only search within each dataset for doppelgangers.
The default setting FALSE will result in output with no missing values, but uses extra memory because all results from the expression, phenotype, and smoking gun doppelganger searches must be saved until the end. Setting this to TRUE will save memory for very large searches, but distance metrics will only be available if that value was identified as a doppelganger (for example, phenotype doppelgangers will have missing values for the expression and smoking gun similarity).

The name of a directory in which to cache or look up results to save re-calculating correlations. Set to NULL for no caching.

Argument for `BiocParallel::bplapply()`, by default will use all cores of a multi-core machine.

Print progress information

Returns an object of S4-class "DoppelGang". See ?DoppelGang-class.

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?BiocParallel::'BiocParallelParam-class'

Example

```r
example("phenoFinder")
results2 <- doppelgangR(esets2, cache.dir = NULL)
results2
plot(results2)
summary(results2)
## Set phenoFinder.args=NULL to ignore similar phenotypes, and
## turn off ComBat batch correction:
## results2 <- doppelgangR(testesets, corfinder.args=list(use.ComBat=FALSE), phenoFinder.args=NULL, cache.dir = NULL)
## summary(results2)
```

### dst

**Skew-t Distribution**

Density function, distribution function and random number generation for the skew-t (ST) distribution. Functions copied from sn CRAN library v0.4.18 for argument name compatibility with `st.mle` function from the same version.

#### Usage

```r
dst(x, location = 0, scale = 1, shape = 0, df = Inf, dp = NULL, log = FALSE)
pst(x, location = 0, scale = 1, shape = 0, df = Inf, dp = NULL, ...) 
qst(p, location = 0, scale = 1, shape = 0, df = Inf, tol = 1e-06, dp = NULL, ...) 
rst(n = 1, location = 0, scale = 1, shape = 0, df = Inf, dp = NULL)
```
**Arguments**

- **x** vector of quantiles. Missing values (NAs) are allowed.
- **p** vector of probabilities
- **location** vector of location parameters.
- **scale** vector of (positive) scale parameters.
- **shape** vector of shape parameters. With `pst` and `qst`, it must be of length 1.
- **df** degrees of freedom (scalar); default is `df=Inf` which corresponds to the skew-normal distribution.
- **dp** a vector of length 4, whose elements represent location, scale (positive), shape and `df`, respectively. If `dp` is specified, the individual parameters cannot be set.
- **n** sample size.
- **log** logical; if TRUE, densities are given as log-densities.
- **tol** a scalar value which regulates the accuracy of the result of `qsn`.
- **...** additional parameters passed to `integrate`.

**Value**

Density (`dst`), probability (`pst`), quantiles (`qst`) and random sample (`rst`) from the skew-t distribution with given location, scale, shape and `df` parameters.

**Details**

Typical usages are

```r
dst(x, location=0, scale=1, shape=0, df=Inf, log=FALSE)
dst(x, dp=, log=FALSE)
pst(x, location=0, scale=1, shape=0, df=Inf, ...)
pst(x, dp=, log=FALSE)
qst(p, location=0, scale=1, shape=0, df=Inf, tol=1e-8, ...)
qst(x, dp=, log=FALSE)
rst(n=1, location=0, scale=1, shape=0, df=Inf)
rst(x, dp=, log=FALSE)
```

**Background**

The family of skew-t distributions is an extension of the Student’s t family, via the introduction of a shape parameter which regulates skewness; when `shape=0`, the skew-t distribution reduces to the usual Student’s t distribution. When `df=Inf`, it reduces to the skew-normal distribution. A multivariate version of the distribution exists. See the reference below for additional information.

**References**


**See Also**

`st.mle`
mst.mle

Maximum likelihood estimation for a (multivariate) skew-t distribution

Description

Fits a skew-t (ST) or multivariate skew-t (MST) distribution to data, or fits a linear regression model with (multivariate) skew-t errors, using maximum likelihood estimation. Functions copied from an CRAN library v0.4.18 because they were later deprecated in that library.

Usage

mst.mle(X, y, freq, start, fixed.df=NA, trace=FALSE, algorithm = c("nlminb","Nelder-Mead", "BFGS", "CG", "SANN"), control=list())
st.mle(X, y, freq, start, fixed.df=NA, trace=FALSE, algorithm = c("nlminb","Nelder-Mead", "BFGS", "CG", "SANN"), control=list())

Arguments

y
a matrix (for mst.mle) or a vector (for st.mle). If y is a matrix, rows refer to observations, and columns to components of the multivariate distribution.

X
a matrix of covariate values. If missing, a one-column matrix of 1’s is created; otherwise, it must have the same number of rows of y. If X is supplied, then it must include a column of 1’s.

freq
a vector of weights. If missing, a vector of 1’s is created; otherwise it must have length equal to the number of rows of y.

start
for mst.mle, a list containing the components beta, Omega, alpha, df of the type described below; for st.mle, a vector whose components contain analogous ingredients as before, with the exception that the scale parameter is the square root of Omega. In both cases, the dp component of the returned list from a previous call has the required format and it can be used as a new start. If the start parameter is missing, initial values are selected by the function.

fixed.df
a scalar value containing the degrees of freedom (df), if these must be taken as fixed, or NA (default value) if df is a parameter to be estimated.

trace
logical value which controls printing of the algorithm convergence. If trace=TRUE, details are printed. Default value is FALSE.

algorithm
a character string which selects the numerical optimization procedure used to maximize the loglikelihood function. If this string is set equal to "nlminb", then this function is called; in all other cases, optim is called, with method set equal to the given string. Default value is "nlminb".

control
this parameter is passed to the chose optimizer, either nlminb or optim; see the documentation of this function for its usage.

Examples

pdf <- dst(seq(-4,4,by=0.1), shape=3, df=5)
rnd <- rst(100, 5, 2, -5, 8)
q <- qst(c(0.25,0.5,0.75), shape=3, df=5)
stopifnot(identical(all.equal(pst(q, shape=3, df=5), c(0.25,0.5,0.75)), TRUE))
Details

If \( y \) is a vector and it is supplied to \texttt{mst.mle}, then it is converted to a one-column matrix, and a scalar skew-t distribution is fitted. This is also the mechanism used by \texttt{st.mle} which is simply an interface to \texttt{mst.mle}.

The parameter \texttt{freq} is intended for use with grouped data, setting the values of \( y \) equal to the central values of the cells; in this case the resulting estimate is an approximation to the exact maximum likelihood estimate. If \texttt{freq} is not set, exact maximum likelihood estimation is performed.

Numerical search of the maximum likelihood estimates is performed in a suitable re-parameterization of the original parameters with aid of the selected optimizer (\texttt{nlminb} or \texttt{optim}) which is supplied with the derivatives of the log-likelihood function. Notice that, in case the optimizer is \texttt{optim}, the gradient may or may not be used, depending on which specific method has been selected. On exit from the optimizer, an inverse transformation of the parameters is performed. For a specific description on the re-parametrization adopted, see Section 5.1 and Appendix B of Azzalini \\& Capitanio (2003).

Value

A list containing the following components:

- **call**: a string containing the calling statement.
- **dp**
  - for \texttt{mst.mle}, this is a list containing the direct parameters \( \beta \), \( \Omega \), \( \alpha \). Here, \( \beta \) is a matrix of regression coefficients with \( \text{dim}(\beta)=c(ncol(X), ncol(y)) \), \( \Omega \) is a covariance matrix of order \( ncol(y) \), \( \alpha \) is a vector of shape parameters of length \( ncol(y) \). For \texttt{st.mle}, \( dp \) is a vector of length \( ncol(X)+3 \), containing \( c(\beta, \omega, \alpha, df) \), where \( \omega \) is the square root of \( \Omega \).
- **se**: a list containing the components \( \beta \), \( \alpha \), \( \text{info} \). Here, \( \beta \) and \( \alpha \) are the standard errors for the corresponding point estimates; \( \text{info} \) is the observed information matrix for the working parameter, as explained below.
- **algorithm**: the list returned by the chose optimizer, either \texttt{nlminb} or \texttt{optim}, plus an item with the name of the selected algorithm; see the documentation of either \texttt{nlminb} or \texttt{optim} for explanation of the other components.

Background

The family of multivariate skew-t distributions is an extension of the multivariate Student’s t family, via the introduction of a shape parameter which regulates skewness; when \( \text{shape}=0 \), the skew-t distribution reduces to the usual t distribution. When \( df=\text{Inf} \) the distribution reduces to the multivariate skew-normal one; see \texttt{dmsn}. See the reference below for additional information.

References


See Also

\texttt{dst}
outlierFinder

Identifies outliers in a similarity matrix.

Description

By default uses the Fisher z-transform for Pearson correlation (atanh), and identifies outliers as those above the quantile of a skew-t distribution with mean and standard deviation estimated from the z-transformed matrix. The quantile is calculated from the Bonferroni-corrected cumulative probability of the upper tail.

Usage

outlierFinder(similarity.mat, bonf.prob = 0.05, transFun = atanh,
normal.upper.thresh = NULL, tail = "upper")

Arguments

similarity.mat  A matrix of similarities - larger values mean more similar.
bonf.prob  Bonferroni-corrected probability. A raw.prob is calculated by dividing this by the number of non-missing values in similarity.mat, and the rejection threshold is qnorm(1-raw.prob, mean, sd) where mean and sd are estimated from the transFun-transformed similarity.mat.
transFun  A function applied to the numeric values of similarity.mat, that should result in normally-distributed values.
normal.upper.thresh  Instead of specifying bonf.prob and transFun, an upper similarity threshold can be set, and values above this will be considered likely duplicates. If specified, this over-rides bonf.prob.
tail  "upper" to look for samples with very high similarity values, "lower" to look for very low values, or "both" to look for both.

Value

Returns either NULL or a dataframe with three columns: sample1, sample2, and similarity.

Author(s)

Levi Waldron, Markus Riester, Marcel Ramos

Examples

library(curatedOvarianData)
data(GSE32063_eset)
cormat <- cor(exprs(GSE32063_eset))
outlierFinder(cormat, bonf.prob = 0.05)
### phenoDist

**Calculate distance between two vectors, rows of one matrix/dataframe, or rows of two matrices/dataframes.**

#### Description

This function does some simple looping to allow x and y to be various combinations of vectors and matrices/dataframes.

#### Usage

```r
phenoDist(x, y = NULL, bins = 10, vectorDistFun = vectorWeightedDist, ...)
```

#### Arguments

- **x**
  - A vector, matrix or dataframe
- **y**
  - NULL, a vector, matrix, or dataframe. If x is a vector, y must also be specified.
- **bins**
  - discretize continuous fields in the specified number of bins
- **vectorDistFun**
  - A function of two vectors that returns the distance between those vectors.
- **...**
  - Extra arguments passed on to vectorDistFun

#### Value

A matrix of distances between pairs of rows of x (if y is unspecified), or between all pairs of rows between x and y (if both are provided).

#### Author(s)

Levi Waldron, Markus Riester, Marcel Ramos

#### Examples

```r
example("phenoFinder")
pdat1 <- pData(esets2[[1]])
pdat2 <- pData(esets2[[2]])

## Use phenoDist() to calculate a weighted distance matrix
distmat <- phenoDist(as.matrix(pdat1), as.matrix(pdat2))
## Note outliers with identical clinical data, these are probably the same patients:
graphics::boxplot(distmat)
```
Calculate pairwise similarities of phenoData between samples for a list containing two ExpressionSets

Description

This function acts as a wrapper to phenoDist to handle cases of one ExpressionSet, a list of two identical ExpressionSets, or a list of two different ExpressionSets.

Usage

```
phenoFinder(eset.pair, separator = "\n", ...)  
```

Arguments

- `eset.pair`: input: a list of ExpressionSets with two elements, or an ExpressionSet. If the two elements are identical, return the correlation matrix for pairs of samples in the first element. If not identical, return pairs between the two elements.
- `separator`: a separator between dataset name (taken from the list names) and sample name (taken from sampleNames(eset), to keep track of which samples come from which dataset).
- `...`: Extra arguments passed on to phenoDist

Value

A matrix of similarities between the phenotypes of pairs of samples.

Author(s)

Levi Waldron, Markus Riester, Marcel Ramos

Examples

```
library(curatedOvarianData)
data(GSE32063_eset)
data(GSE17260_eset)
esets2 <- list(JapaneseB=GSE32063_eset, Yoshihara2010=GSE17260_eset)

## standardize the sample ids to improve matching based on clinical annotation
esets2 <- lapply(esets2, function(X){
    X$alt_sample_name <- paste(X$sample_type, gsub("[^0-9]", "", X$alt_sample_name), sep="_")
}

## Removal of columns that cannot possibly match also helps duplicated patients to stand out
pData(X) <- pData(X)[, !grepl("uncurated_author_metadata", colnames(pData(X)))]
X <- X[, 1:20]  ##speed computations
return(X) })

## See first six samples in both rows and columns
phenoFinder(esets2)[1:6, 1:6]
```
Histograms of all pairwise sample correlations, showing identified doppelgangers.

Description
Identified doppelgangers are shown with a red vertical line overlaid on a histogram of pairwise sample correlations. One plot is made per pair of datasets.

Arguments
- **x**: An object of class DoppelGang
- **skip.no.doppels**: (default FALSE) If TRUE, do not plot histograms where no doppelgangers were identified.
- **plot.pair**: An optional character vector of length two, providing the names of two datasets. If provided, only the comparison of these two datasets will be plotted.
- **...**: Additional arguments passed on to hist.

Value
None

Methods
- signature(x = "DoppelGang") Histograms of all pairwise sample correlations, showing identified doppelgangers.

Author(s)
Levi Waldron

Examples
```r
library(curatedOvarianData)
data(TCGA_eset)
data(GSE26712_eset)
## Remove some TCGA samples to speed computation:
"TCGA.24.2289", "TCGA.29.2414", "TCGA.59.2352", "TCGA.36.2532",
"TCGA.36.2529", "TCGA.36.2551", "TCGA.42.2590", "TCGA.13.2071",
"TCGA.29.2422", "TCGA.36.2537", "TCGA.36.2547", "TCGA.4.1369",
"TCGA.42.2591", "TCGA.23.2641", "TCGA.29.2434", "TCGA.36.2538",
"TCGA.36.2548", "TCGA.5.1516", "TCGA.42.2593", "TCGA.36.2549",
"TCGA.4.1644", "TCGA.3.2057", "TCGA.23.2647", "TCGA.36.2530",
"TCGA.36.2552", "TCGA.42.2587", "TCGA.13.2061", "TCGA.42.2588";
```
"TCGA.36.2544", "TCGA.42.2589", "TCGA.13.2066", "TCGA.61.2613",
"TCGA.61.2614", "TCGA.24.1852", "TCGA.29.1704", "TCGA.13.1819"
)
keep.tcga <- unique(c(keep.tcga, sampleNames(TCGA_eset)[1:200]))
testesets <- list(Bonome08=GSE26712_eset, TCGA=TCGA_eset[, keep.tcga])
results1 <- doppelgangR(testesets,
        corFinder.args=list(use.ComBat=FALSE), phenoFinder.args=NULL, cache.dir=NULL)
plot(results1)

print-methods

Print a DoppelGang object

Description

Prints the @summaryresults slot of a DoppelGang object.

Methods

signature(x = "DoppelGang")  Prints the @summaryresults slot of a DoppelGang object.

show-methods

Show a DoppelGang object

Description

Formats a summary of the DoppelGang object.

Methods

signature(object = "DoppelGang")  Summarizes the @summaryresults slot of a DoppelGang object

smokingGunFinder

Find doppelgangers based on "smoking gun" phenotypes - those that should be unique to each patient.

Description

Checks all pairwise combinations of samples for values of the "smoking" gun phenotypes that are identical.

Usage

smokingGunFinder(eset.pair, smokingguns, transFun = I, separator = ":")
Arguments

eset.pair a list of ExpressionSets, with two elements. If the two elements are identical, the function will check for duplicate IDs within one element. If not identical, it will check for duplicate IDs between elements.

smokingguns phenoData column names found in multiple elements of eset.pair that may contain "smoking guns" such as identifiers that should be unique to each sample.

transFun a function to apply to IDs before comparing. By default apply no transformation.

separator Separator between dataset name and sample name. Dataset names are added to sample names to keep track of dataset of origin.

Value

Returns an adjacency matrix for samples where matches have value 1, non-matches have value zero. Value for a sample against itself is NA.

Author(s)

Levi Waldron, Markus Riester, Marcel Ramos

Examples

example("phenoFinder")

smokingGunFinder(esets2, "days_to_death")

summary-methods Summarizes a DoppelGang object

Description

Summarizes the @summaryresults slot of a DoppelGang object.

Methods

signature(object = "DoppelGang") Summarizes the @summaryresults slot of a DoppelGang object.

vectorHammingDist Calculate Hamming Distance between two vectors, using pairwise complete observations.

Description

Simple function to count the fraction of different elements (in the same position) between two vectors of the same length, after removing elements from both vectors corresponding to positions that are NA in either vector.
vectorWeightedDist

Calculate a weighted distance between two vectors, using pairwise complete observations.

Description

Simple function to count the fraction of different elements (in the same position) between two vectors of the same length, after removing elements from both vectors corresponding to positions that are NA in either vector. Distance is the probability for observing the matches and mismatches in two random patients.

Usage

vectorWeightedDist(x, y, k, l)

Arguments

x  
a matrix
y  
a matrix with the same number of columns as x
k  
row in x to test for differences
l  
row in y to test for differences
**Value**

Returns a numeric value, the log of the probability of observing the matches in x and y

**Author(s)**

Levi Waldron, Markus Riester, Marcel Ramos

**Examples**

```r
mymat1 <- matrix(rnorm(20), ncol = 5)
mymat1[1, 4] <- NA
mymat2 <- matrix(rnorm(20), ncol = 5)
vectorWeightedDist(mymat1, mymat2, 1, 2)
```
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