Package ‘flagme’

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**Description**

Reads ASCII ELU-format files (output from AMDIS) and attaches them to an already created `peaksDataset` object.

**Usage**

```r
addAMDISPeaks(object,fns=dir(.)[^lu],verbose=TRUE,...)
```

**Arguments**

- `object` a `peaksDataset` object.
- `fns` character vector of same length as `object@rawdata` (user ensures the order matches)
- `verbose` whether to give verbose output, default `TRUE`
- `...` arguments passed on to `parseELU`

**Details**

Repeated calls to `parseELU` to add peak detection results to the original `peaksDataset` object.

**Value**

A `peaksDataset` object

**Author(s)**

Mark Robinson

**References**

addChromaTOFPeaks

See Also

parseELU, peaksDataset

Examples

# need access to CDF (raw data) and ELU files
require(gcspikelite)
gcmsPath<-paste(find.package("gcspikelite"),"data",sep="/"")

# full paths to file names
cdfFiles<-dir(gcmsPath,"CDF",full=TRUE)
eluFiles<-dir(gcmsPath,"ELU",full=TRUE)

# create a 'peaksDataset' object and add AMDIS peaks to it
pd<-peaksDataset(cdfFiles[1],mz=seq(50,550),rtrange=c(7.5,8.5))
pd<-addAMDISPeaks(pd,eluFiles[1])

addChromaTOFPeaks  Add ChromaTOF peak detection results

Description

Reads ASCII tab-delimited format files (output from ChromaTOF) and attaches them to an already created peaksDataset object

Usage

addChromaTOFPeaks(object,fns=dir(,"[Tt][Xx][Tt]"),rtDivide=60,verbose=TRUE,...)

Arguments

  object    a peaksDataset object.
  fns       character vector of same length as object@rawdata (user ensures the order matches)
  rtDivide  number giving the amount to divide the retention times by.
  verbose   whether to give verbose output, default TRUE
  ...       arguments passed on to parseChromaTOF

Details

Repeated calls to parseChromaTOF to add peak detection results to the original peaksDataset object.

Value

peaksDataset object

Author(s)

Mark Robinson
addXCMSPeaks

Add xcms/CAMERA peak detection results

Description
Reads the raw data using xcms, group each extracted ion according to their retention time using CAMERA and attaches them to an already created peaksDataset object

Usage
addXCMSPeaks(files, object, peakPicking=c('cwt', 'mF'), ...)

Arguments
files character vector of same length as object@rawdata (user ensures the order matches)
object a peaksDataset object.
peakPicking Methods to use for peak detection. See details.
... arguments passed on to xcmsSet and annotate

Details
Repeated calls to xcmsSet and annotate to perform peak-picking and deconvolution. The peak detection results are added to the original peaksDataset object. Two peak detection algorithms are available: continuous wavelet transform (peakPicking=c('cwt')) and the matched filter approach (peakPicking=c('mF')) described by Smith et al (2006). For further information consult the xcms package manual.

References

See Also
parseChromaTOF, peaksDataset

Examples
# need access to CDF (raw data) and ChromaTOF files
require(gcspikelite)
gcmsPath<-paste(find.package("gcspikelite"),"data",sep="/"

# full paths to file names
cdfFiles<-dir(gcmsPath,"CDF",full=TRUE)
# [not run] cToFFiles<-dir(gcmsPath,"txt",full=TRUE)

# create a 'peaksDataset' object and add ChromaTOF peaks to it
pd<-peaksDataset(cdfFiles[1],mz=seq(50,550),rtrange=c(7.5,8.5))
# [not run] pd<-addChromTOFPeaks(pd,...)
Value

peaksDataset object

Author(s)

Riccardo Romoli <riccardo.romoli@unifi.it>

See Also

peaksDataset findPeaks.matchedFilter findPeaks.centWave xcmsRaw-class

Examples

# need access to CDF (raw data)
require(gcspikelite)
gcmsPath <- paste(find.package("gcspikelite"), "data", sep="/"

# full paths to file names
cdfFiles <- dir(gcmsPath, "CDF", full=TRUE)

# create a 'peaksDataset' object and add XCMS peaks to it
pd <- peaksDataset(cdfFiles[1], mz=seq(50,550), rtrange=c(7.5,8.5))
pd <- addXCMSPeaks(cdfFiles[1], pd, peakPicking=c("mF"),
                   snthresh=3, fwhm=4, step=1, steps=2, mzdiff=0.5)

# need access to CDF (raw data)
require(gcspikelite)
gcmsPath <- paste(find.package("gcspikelite"), "data", sep="/"

# full paths to file names
cdfFiles <- dir(gcmsPath, "CDF", full=TRUE)

# create a 'peaksDataset' object and add XCMS peaks to it
pd <- peaksDataset(cdfFiles[1], mz=seq(50,550), rtrange=c(7.5,8.5))
pd <- addXCMSPeaks(cdfFiles[1], pd, peakPicking=c("mF"),
                   snthresh=3, fwhm=4, step=1, steps=2, mzdiff=0.5)

# need access to CDF (raw data)
require(gcspikelite)
gcmsPath <- paste(find.package("gcspikelite"), "data", sep="/"

# full paths to file names
cdfFiles <- dir(gcmsPath, "CDF", full=TRUE)

# create a 'peaksDataset' object and add XCMS peaks to it
pd <- peaksDataset(cdfFiles[1], mz=seq(50,550), rtrange=c(7.5,8.5))
pd <- addXCMSPeaks(cdfFiles[1], pd, peakPicking=c("mF"),
                   snthresh=3, fwhm=4, step=1, steps=2, mzdiff=0.5)

betweenAlignment

Data Structure for "between" alignment of many GCMS samples

Description

This function creates a "between" alignment (i.e. comparing merged peaks)

Usage

betweenAlignment(pD, cAList, pAList, impList, filterMin=3, gap=0.7, D=10, usePeaks=TRUE, df=30, verbose=TRUE)

Arguments

- **pD** a peaksDataset object
- **cAList** list of clusterAlignment objects, one for each experimental group
- **pAList** list of progressiveAlignment objects, one for each experimental group
- **impList** list of imputation lists
- **filterMin** minimum number of peaks within a merged peak to be kept in the analysis
- **gap** gap parameter
- **D** retention time penalty parameter
- **usePeaks** logical, whether to use peaks (if TRUE) or the full 2D profile alignment (if FALSE)
- **df** distance from diagonal to calculate similarity
- **verbose** logical, whether to print information
calcTimeDiffs

Details

betweenAlignment objects gives the data structure which stores the result of an alignment across several "pseudo" datasets. These pseudo datasets are constructed by merging the "within" alignments.

Value

betweenAlignment object

Author(s)

Mark Robinson

References


See Also

multipleAlignment

Examples

```
require(gcspikelite)
# see 'multipleAlignment'
```

## Description

This function takes the set of all pairwise profile alignments and use these to estimate retention time shifts between each pair of samples. These will then be used to normalize the retention time penalty of the signal peak alignment.

## Usage

```
calcTimeDiffs(pd,ca.full,verbose=TRUE)
```

## Arguments

- `pd` a peaksDataset object
- `ca.full` a clusterAlignment object, fit with
- `verbose` logical, whether to print out information

## Details

Using the set of profile alignments,
Value

A list of same length as `ca.full@alignments` with the matrices giving the retention time penalties.

Author(s)

Mark Robinson

References


See Also

`peaksAlignment`, `clusterAlignment`

Examples

```r
require(gcspikelite)

# paths and files
gcmsPath <- paste(find.package("gcspikelite"),"data",sep="/")
cdfFiles <- dir(gcmsPath,"CDF",full=TRUE)
eluFiles <- dir(gcmsPath,"ELU",full=TRUE)

# read data, peak detection results
pd <- peaksDataset(cdfFiles[1:2],mz=seq(50,550),rtrange=c(7.5,8.5))
pd <- addAMDISPeaks(pd,eluFiles[1:2])

# pairwise alignment using all scans
fullca <- clusterAlignment(pd, usePeaks=FALSE, df=100)

# calculate retention time shifts
timedf <- calcTimeDiffs(pd, fullca)
```

Description

Store the raw data and optionally, information regarding signal peaks for a number of GCMS runs

Usage

```r
clusterAlignment(pD,runs=1:length(pD@rawdata),timedf=NULL,usePeaks=TRUE,verbose=TRUE,...)
```
clusterAlignment

Arguments

- **pD**: a peaksDataset object.
- **runs**: vector of integers giving the samples to calculate set of pairwise alignments over.
- **timedf**: list (length = the number of pairwise alignments) of matrices giving the expected time differences expected at each pair of peaks (used with `usePeaks=TRUE`, passed to peaksAlignment).
- **usePeaks**: logical, TRUE uses peakdata list, FALSE uses rawdata list for computing similarity.
- **verbose**: logical, whether to print out info.
- **...**: other arguments passed to peaksAlignment.

Details

clusterAlignment computes the set of pairwise alignments.

Value

clusterAlignment object

Author(s)

Mark Robinson

References


See Also

peaksDataset, peaksAlignment

Examples

```r
require(gcspikelite)

# paths and files
gcmsPath<-paste(find.package("gcspikelite"),"data",sep="/")
cddfFiles<dir(gcmsPath,"CDF",full=TRUE)
eluFiles<dir(gcmsPath,"ELU",full=TRUE)

# read data, peak detection results
pd<-peaksDataset(cddfFiles[1:2],mz=seq(50,550),rtrange=c(7.5,8.5))
pd<-addAMDISPeaks(pd,eluFiles[1:2])

cac<-clusterAlignment(pd, gap = .5,D=.05,df=30)
```
Description

Many of the peaks are not similar. So, the set of pairwise similarity matrices can be compressed.

Usage

compress(object, verbose=TRUE, ...)
decompress(object, verbose=TRUE, ...)

Arguments

object a peaksAlignment, peaksAlignment or peaksAlignment object to be compressed
verbose logical, whether to print out information
... further arguments

Details

Using sparse matrix representations, a significant compression can be achieved. Here, we use the matrix.csc class of the SpareM package.

Value

an object of the same type as the input object

Author(s)

Mark Robinson

References


See Also

peaksAlignment, clusterAlignment, progressiveAlignment

Examples

require(gcspikelite)

# paths and files
gcmsPath<-paste(find.package("gcspikelite"),"data",sep="/"

cdfFiles<dir(gcmsPath,"CDF",full=TRUE)
eluFiles<dir(gcmsPath,"ELU",full=TRUE)

# read data, peak detection results
pd<-peaksDataset(cdfFiles[1:2],mz=seq(50,550),rtrange=c(7.5,8.5))
correlationAlignment

Function for correlation-based alignment strategy of multiple GCMS samples

Description

Performs a correlation-based peak alignment of multiple GCMS peak lists; this function is able to align multiple samples, by a center-star strategy.

Usage

correlationAlignment(object, thr=0.85, D=20, penalty=0.2, normalize=TRUE, minFilter=1)

Arguments

object a peaksDataset object
thr correlation threshold from 0 (min) to 1 (max)
D retention time window in seconds
penalty the penalty inflicted to a match between two peaks when the retention time difference exceed the parameter D
normalize logical, whether to use normalized-to-100 peaks intensity or as such
minFilter if a feature is matched in a number of samples less than minFilter, this feature is trashed. The value of minFilter must be smaller than the number of samples

Details

The correlation-based peak alignment for multiple GCMS peak lists uses a center-star technique to the alignment of the peaks. The combination of the D and penalty parameters allow the users to force the algorithm to match the peaks close to the reference. The thr parameter controls the matching factor.

Value

correlationAlignment object

Author(s)

Riccardo Romoli <riccardo.romoli@unifi.it>

See Also

peaksDataset, addXCMSPeaks, correlationAlignment-class
Examples

```r
require(gcspikelite)

# paths and files
gcmsPath <- paste(find.package("gcspikelite"), "data", sep="/"

cdfFiles <- dir(gcmsPath,"CDF",full=TRUE)

# read data, peak detection results
pd <- peaksDataset(cdfFiles[1:2], mz=seq(50,550), rtrange=c(7.5,8.5))
pd <- addXCMSPeaks(files=cdfFiles[1:2], object=pd, peakPicking=c('mF'),
                   snthresh=3, fwhm=4, step=1, steps=2, mzdiff=0.5)
mp <- correlationAlignment(object=pd, thr=0.85, D=20, penalty=0.2,
                           normalize=TRUE, minFilter=1)
```

Description

A class containing the index of the aligned chromatographic peaks.

Objects from the Class

Objects can be created by calls of the form new("correlationAlignment", ...). The object created contains both the results of the alignment procedure and the file used and as center-star.

Slots

- **Alignment**: Object of class "align" contain the matrix of the aligned features. The rows represent the different peaks while the columns represent the files. The values of the matrix refers to peaksind slot of the peaksDataSet object.
- **Center**: Object of class "character" contain the file used as a center-star.

Methods

- **show** signature(object = "correlationAlignment")

Author(s)

Riccardo Romoli <riccardo.romoli@unifi.it>

See Also

- `correlationAlignment` for further information about the alignment function.

Examples

```r
showClass("correlationAlignment")
Dynamic programming algorithm, given a similarity matrix

Description
This function calls C code for a bare-bones dynamic programming algorithm, finding the best cost path through a similarity matrix.

Usage
dp(M,gap=.5,big=10000000000,verbose=FALSE)

Arguments
- `M`: similarity matrix
- `gap`: penalty for gaps
- `big`: large value used for matrix margins
- `verbose`: logical, whether to print out information

Details
This is a pretty standard implementation of a bare-bones dynamic programming algorithm, with a single gap parameter and allowing only simple jumps through the matrix (up, right or diagonal).

Value
list with element `match` with the set of pairwise matches.

Author(s)
Mark Robinson

References

See Also
- `normDotProduct`

Examples
```r
require(gcspikelite)

# paths and files
gcmsPath<-paste(find.package("gcspikelite"),"data",sep="/"

cdfFiles<-dir(gcmsPath,"CDF",full=TRUE)
eluFiles<-dir(gcmsPath,"ELU",full=TRUE)

# read data, peak detection results
pd<-peaksDataset(cdfFiles[1:2],mz=seq(50,550),rtrange=c(7.5,8.5))
```
### eitherMatrix-class

**Description**

A container to store either matrix or matrix.csc objects

**Author(s)**

Mark Robinson

**References**


**See Also**

- `peaksAlignment`

---

### exportSpectra

**Description**

Write the deconvoluted mass spectra to an external file

**Usage**

```r
exportSpectra(object, sample, spectraID, normalize = TRUE)
```

**Arguments**

- **object**: an object of class "peaksDataset" where to keep the mass spectra; both abundance (y) than m/z (x)
- **sample**: character, the sample from were to plot the mass spectra
- **spectraID**: numerical, a vector containing the index of the spectra to be plotted.
- **normalize**: logical, if TRUE normalize the intensity of the mass peak to 100, the most abundant is 100 other peaks are scaled consequetially
gatherInfo

Details
Write a .msp file of the deconvoluted mass spectra. Useful to try to identify the unknown spectra using NIST Search.

Value
a .msp file ready to be read using NIST search

Author(s)
riccardo.romoli@unifi.it

gatherInfo Gathers abundance informations from an alignment

Description
Given an alignment table (indices of matched peaks across several samples) such as that within a progressiveAlignment or multipleAlignment object, this routine goes through the raw data and collects the abundance of each fragment peak, as well as the retention times across the samples.

Usage
gatherInfo(pD, obj, newind = NULL, method = c("apex"), findmzind = TRUE, useTIC = FALSE, top = NULL, intensity.cut = 0.05)

Arguments
pD a peaksDataset object, to get the abundance data from
obj either a multipleAlignment or progressiveAlignment object
newind list giving the
method method used to gather abundance information, only apex implemented currently.
findmzind logical, whether to take a subset of all m/z indices
useTIC logical, whether to use total ion current for abundance summaries
top only use the top top peaks
intensity.cut percentage of the maximum intensity

Details
This procedure loops through the table of matched peaks and gathers the

Value
Returns a list (of lists) for each row in the alignment table. Each list has 3 elements:
mz a numerical vector of the m/z fragments used
rt a numerical vector for the exact retention time of each peak across all samples
data matrix of fragment intensities. If useTIC=TRUE, this matrix will have a single row
**imputePeaks**

**Author(s)**
Mark Robinson

**References**

**See Also**
imputePeaks

**Examples**

```r
require(gcspikelite)

# paths and files
gcmsPath<-paste(find.package("gcspikelite"),"data",sep="/")
cdfFiles<-dir(gcmsPath,"CDF",full=TRUE)
eluFiles<-dir(gcmsPath,"ELU",full=TRUE)

# read data, peak detection results
pd<-peaksDataset(cdfFiles[1:2],mz=seq(50,550),rtrange=c(7.5,8.5))

# multiple alignment
ma<-multipleAlignment(pd,c(1,1),wn.gap=0.5,wn.D=.05,bw.gap=0.6,bw.D=.2,usePeaks=TRUE,filterMin=1,df=50,verbose=TRUE)

d<-gatherInfo(pd,ma)

# table of retention times
nm<-list(paste("MP",1:length(d),sep=""),c("S1","S2"))

# Imputatin of locations of peaks that were undetected

imputePeaks

**Description**

Using the information within the peaks that are matched across several runs, we can impute the location of the peaks that are undetected in a subset of runs.

**Usage**

imputePeaks(pD, obj, type = 1, obj2 = NULL, filterMin = 3, verbose = TRUE)

**Arguments**

- **pD** a peaksDataset object
- **obj** the alignment object, either multipleAlignment or progressiveAlignment, that is used to infer the unmatched peak locations
imputePeaks

type  type of imputation to do, 1 for simple linear interpolation (default), 2 only works if obj2 is a clusterAlignment object
obj2  a clusterAlignment object
filterMin minimum number of peaks within a merged peak to impute
verbose logical, whether to print out information

Details

If you are aligning several samples and for a (small) subset of the samples in question, a peak is undetected, there is information within the alignment that can be useful in determining where the undetected peak is, based on the surrounding matched peaks. Instead of moving forward with missing values into the data matrices, this procedures goes back to the raw data and imputes the location of the apex (as well as the start and end), so that we do not need to bother with post-hoc imputation or removing data because of missing components.

We realize that imputation is prone to error and prone to attributing intensity from neighbouring peaks to the unmatched peak. We argue that this is still better than having to deal with these in statistical models after that fact. This may be an area of future improvement.

Value

list with 3 elements apex, start and end, each masked matrices giving the scan numbers of the imputed peaks.

Author(s)

Mark Robinson

References


See Also

multipleAlignment, progressiveAlignment, peaksDataset

Examples

require(gcspikelite)

# paths and files
gcmsPath<-paste(find.package("gcspikelite"),"data",sep="/")
cddfFiles<-dir(gcmsPath,"CDF",full=TRUE)
eluFiles<-dir(gcmsPath,"ELU",full=TRUE)

# read data, peak detection results
pd<-peaksDataset(cddfFiles[1:3],mz=seq(50,550),rtrange=c(7.5,8.5))
        pd<-addAMDISPeaks(pd,eluFiles[1:3])

# alignments
ca<-clusterAlignment(pd, gap = .5,D=.05,df=30)
           pa<-progressiveAlignment(pd, ca, gap = .6, D=.1,df=30)

v<-imputePeaks(pd,pa,filterMin=1)
Data Structure for multiple alignment of many GCMS samples

Description
Store the raw data and optionally, information regarding signal peaks for a number of GCMS runs

Usage
multipleAlignment(pd, group, bw.gap=0.8, wn.gap=0.6, bw.D=.20, wn.D=.05, filterMin=3, lite=FALSE, usePeaks=TRUE, df=50, verbose=TRUE, timeAdjust=FALSE, doImpute=FALSE)

Arguments
- **pd**: a peaksDataset object
- **group**: factor variable of experiment groups, used to guide the alignment algorithm
- **bw.gap**: gap parameter for "between" alignments
- **wn.gap**: gap parameter for "within" alignments
- **bw.D**: distance penalty for "between" alignments
- **wn.D**: distance penalty for "within" alignments
- **filterMin**: minimum number of peaks within a merged peak to be kept in the analysis
- **lite**: logical, whether to keep "between" alignment details (default, FALSE)
- **usePeaks**: logical, whether to use peaks (if TRUE) or the full 2D profile alignment (if FALSE)
- **df**: distance from diagonal to calculate similarity
- **verbose**: logical, whether to print information
- **timeAdjust**: logical, whether to use the full 2D profile data to estimate retention time drifts (Note: time required)
- **doImpute**: logical, whether to impute the location of unmatched peaks

Details
multipleAlignment is the data structure giving the result of an alignment across several GCMS runs. Multiple alignments are done progressively. First, all samples with the same group label with be aligned (denoted a "within" alignment). Second, each group will be summarized into a pseudo-data set, essentially a spectrum and retention time for each matched peak of the within-alignment. Third, these "merged peaks" are aligned in the same progressive manner, here called a "between" alignment.

Value
multipleAlignment object

Author(s)
Mark Robinson
References


See Also

peaksDataset, betweenAlignment, progressiveAlignment

Examples

```
require(gcspikelite)
# paths and files
gcmsPath<-paste(find.package("gcspikelite"),"data",sep="/")
cdfFiles<-dir(gcmsPath,"CDF",full=TRUE)
eluFiles<-dir(gcmsPath,"ELU",full=TRUE)

# read data, peak detection results
pd<-peaksDataset(cdfFiles[1:2],mz=seq(50,550),rtrange=c(7.5,8.5))
pd<-addAMDISPeaks(pd,eluFiles[1:2])

# multiple alignment
ma<-multipleAlignment(pd,c(1,1),wn.gap=0.5,wn.D=.05,bw.gap=0.6,bw.D=.2,usePeaks=TRUE,filterMin=1,df=50,verbose=TRUE)
```

```
# normalized dot product

```
Value
matrix of similarities

Author(s)
Mark Robinson

References

See Also
dp, peaksAlignment

Examples
require(gcspikelite)

# paths and files
gcmsPath<-paste(find.package("gcspikelite"),"data",sep="/")
cdfFiles<-dir(gcmsPath,"CDF",full=TRUE)
eluFiles<-dir(gcmsPath,"ELU",full=TRUE)

# read data, peak detection results
pd<-peaksDataset(cdfFiles[1:2],mz=seq(50,550),rtrange=c(7.5,8.5))
pd<-addAMDISPeaks(pd,eluFiles[1:2])

r<-normDotProduct(pd@peaksdata[[1]],pd@peaksdata[[2]])

parseChromaTOF  Parser for ChromaTOF files

Description
Reads ASCII ChromaTOF-format files from AMDIS (Automated Mass Spectral Deconvolution and Identification System)

Usage
parseChromaTOF(fn,min.pc=.01,mz=seq(85,500),rt.cut=.008,rtrange=NULL,skip=1,rtDivide=60)

Arguments

fn  ChromaTOF filename to read.
min.pc  minimum percent of maximum intensity.
mz  vector of mass-to-charge bins of raw data table.
rt.cut  the difference in retention time, below which peaks are merged together.
rtrange  retention time range to parse peaks from, can speed up parsing if only interested in a small region (must be numeric vector of length 2)
skip  number of rows to skip at beginning of the ChromaTOF
rtDivide  multiplier to divide the retention times by (default: 60)
Details

`parseChromaTOF` will typically be called by `addChromaTOFPeaks`, not called directly.

Peaks that are detected within `rt.cut` are merged together. This avoids peaks which are essentially overlapping.

Fragments that are less than `min.pc` of the maximum intensity fragment are discarded.

Value

A list with components `peaks` (table of spectra – rows are mass-to-charge and columns are the different detected peaks) and `tab` (table of features for each detection), according to what is stored in the ChromaTOF file.

Author(s)

Mark Robinson

References


See Also

`addAMDISPeaks`

Examples

```r
require(gcspikelite)

# paths and files
gcmsPath<-paste(find.package("gcspikelite"),"data",sep="/")
tofFiles<-dir(gcmsPath,"tof",full=TRUE)

# parse ChromaTOF file
cTofList<-parseChromaTOF(tofFiles[1])
```

Description

Parser for ELU files

Usage

```r
parseELU(f,min.pc=.01,mz=seq(50,550),rt.cut=.008,rtrange=NULL)
```
parseELU

Arguments

f  ELU filename to read.
min.pc  minimum percent of maximum intensity.
mz  vector of mass-to-charge bins of raw data table.
rt.cut  the difference in retention time, below which peaks are merged together.
rtrange  retention time range to parse peaks from, can speed up parsing if only interested in a small region (must be numeric vector of length 2)

Details

parseELU will typically be called by addAMDISPeaks, not called directly.
Peaks that are detected within rt.cut are merged together. This avoids peaks which are essentially overlapping.
Fragments that are less than min.pc of the maximum intensity fragment are discarded.

Value

list with components peaks (table of spectra – rows are mass-to-charge and columns are the different detected peaks) and tab (table of features for each detection), according to what is stored in the ELU file.

Author(s)

Mark Robinson

References


See Also

addAMDISPeaks

Examples

require(gcaspikelite)

# paths and files
gcspPath<-paste(find.package("gcaSpikelite"),"data",sep="/")
eluFiles<-dir(gcspPath,"ELU",full=TRUE)

# parse ELU file
eluList<-parseELU(eluFiles[1])
peaksAlignment-class

Data Structure for pairwise alignment of 2 GCMS samples

Description

Store the raw data and optionally, information regarding signal peaks for a number of GCMS runs

Usage

peaksAlignment(d1, d2, t1, t2, gap=.5, D=1000, timedf=NULL, df=30, verbose=TRUE, usePeaks=TRUE, compress=TRUE)

Arguments

- **d1**: matrix of MS intensities for 1st sample (if doing a peak alignment, this contains peak apexes/areas; if doing a profile alignment, this contains scan intensities. Rows are m/z bins, columns are peaks/scans.
- **d2**: matrix of MS intensities for 2nd sample
- **t1**: vector of retention times for 1st sample
- **t2**: vector of retention times for 2nd sample
- **gap**: gap penalty for dynamic programming algorithm
- **D**: time penalty (on same scale as retention time differences, t1 and t2)
- **timedf**: list (length = the number of pairwise alignments) of matrices giving the expected time differences expected at each pair of peaks (used with usePeaks=TRUE.
- **df**: integer, how far from the diagonal to go to calculate the similarity of peaks. Smaller value should run faster, but be careful not to choose too low.
- **verbose**: logical, whether to print out info.
- **usePeaks**: logical, TRUE uses peakdata list, FALSE uses rawdata list for computing similarity.
- **compress**: logical, whether to compress the similarity matrix into a sparse format.

Details

peaksAlignment is a hold-all data structure of the raw and peak detection data.

Value

peaksAlignment object

Author(s)

Mark Robinson

References


See Also

peaksDataset, clusterAlignment
**peaksDataset**

**Examples**

```r
# see clusterAlignment, it calls peaksAlignment
```

---

**peaksDataset**  
*Data Structure for raw GCMS data and peak detection results*

**Description**

Store the raw data and optionally, information regarding signal peaks for a number of GCMS runs.

**Usage**

```r
peaksDataset(fns=dir("[Cc][Dd][Ff]"),verbose=TRUE,mz=seq(50,550),rtDivide=60,rtrange=NULL)
```

**Arguments**

- `fns` character vector, filenames of raw data in CDF format.
- `verbose` logical, if TRUE then iteration progress information is output.
- `mz` vector giving bins of raw data table.
- `rtDivide` number giving the amount to divide the retention times by.
- `rtrange` retention time range to limit data to (must be numeric vector of length 2)

**Details**

peaksDataset is a hold-all data structure of the raw and peak detection data.

**Value**

peaksDataset object

**Author(s)**

Mark Robinson

**References**


**Examples**

```r
require(gcspikelite)

# paths and files
gcmsPath<-paste(find.package("gcspikelite"),"data",sep="/"

cdfFiles<-dir(gcmsPath,"CDF",full=TRUE)
eluFiles<-dir(gcmsPath,"ELU",full=TRUE)

# read data
pd<-peaksDataset(cdfFiles[1:2],mz=seq(50,550),rtrange=c(7.5,8.5))
show(pd)
```
plot.peaksDataset  
Plotting functions for GCMS data objects

Description

Store the raw data and optionally, information regarding signal peaks for a number of GCMS runs

Usage

.plotpD(object, runs=1:length(object@rawdata), mzind=1:nrow(object@rawdata[[1]]),
    mind=NULL, plotSampleLabels=TRUE, calcGlobalMax=FALSE, peakCex = 0.8, plotPeaks=TRUE,
    plotPeakBoundaries=FALSE, plotPeakLabels=FALSE, plotMergedPeakLabels=TRUE, mlwd=3,
    usePeaks=TRUE, plotAcrossRuns=FALSE, overlap=F, rrange=NULL, cols=NULL, thin=1,
    max.near=median(object@rawrt[[1]]), how.near=50, scale.up=1,...)

.plotpA(object, xlab="Peaks - run 1", ylab="Peaks - run 2", plotMatches=TRUE, matchPch=19, matchLwd=3,
    matchCex=.5, matchCol="black", col=colorpanel(50,"black", "blue", "white"),
    breaks=seq(0,1,length=51),...)

.plotcA(object, alignment=1,...)

Arguments

object  
a peaksDataset, peaksAlignment or clusterAlignment object.

runs  
for peaksDataset only: set of run indices to plot

mzind  
for peaksDataset only: set of mass-to-charge indices to sum over (default, all)

mind  
for peaksDataset only: matrix of aligned indices

plotSampleLabels  
for peaksDataset only: logical, whether to display sample labels

calcGlobalMax  
for peaksDataset only: logical, whether to calculate an overall maximum for scaling

peakCex  
character expansion factor for peak labels

plotPeaks  
for peaksDataset only: logical, whether to plot hashes for each peak

plotPeakBoundaries  
for peaksDataset only: logical, whether to display peak boundaries

plotPeakLabels  
for peaksDataset only: logical, whether to display peak labels

plotMergedPeakLabels  
for peaksDataset only: logical, whether to display 'merged' peak labels

mlwd  
for peaksDataset only: line width of lines indicating the alignment

usePeaks  
for peaksDataset only: logical, whether to plot alignment of peaks (otherwise, scans)

plotAcrossRuns  
for peaksDataset only: logical, whether to plot across peaks when unmatched peak is given

overlap  
for peaksDataset only: logical, whether to plot TIC/XICs overlapping

rrange  
for peaksDataset only: vector of length 2 giving start and end of the X-axis

cols  
for peaksDataset only: vector of colours (same length as the length of runs)
For peakDataset objects, each TIC is scale to the maximum value (as specified by the how.near and max.near values). The many parameters gives considerable flexibility of how the TICs can be visualized.

For peakAlignment objects, the similarity matrix is plotted and optionally, the set of matching peaks. clusterAlignment objects are just a collection of all pairwise peakAlignment objects.

Author(s)

Mark Robinson

References


See Also

plotImage, peaksDataset

Examples

require(gcspikelite)

# paths and files
gcmsPath<-paste(find.package("gcspikelite"),"data",sep="/")
cdfFiles<-dir(gcmsPath,"CDF",full=TRUE)
eluFiles<-dir(gcmsPath,"ELU",full=TRUE)

# read data
pd<-peaksDataset(cdfFiles[1:3],mz=seq(50,550),rtrange=c(7.5,8.5))
plotImage

Plot of images of GCMS data

Description
Image plots (i.e. 2D heatmaps) of raw GCMS profile data

Usage
plotImage(object, run=1, rtrange=c(11,13), main=NULL, mzrange=c(50,200), SCALE=log2,...)

Arguments
- object: a peaksDataset object
- run: index of the run to plot an image for
- rtrange: vector of length 2 giving start and end of the X-axis (retention time)
- main: main title (auto-constructed if not specified)
- mzrange: vector of length 2 giving start and end of the Y-axis (mass-to-charge ratio)
- SCALE: function called to scale the data (default: log2)
- ... further arguments passed to the image command

Details
For peakDataset objects, each TIC is scaled to the maximum value (as specified by the how.near and max.near values). The many parameters give considerable flexibility of how the TICs can be visualized.

For peakAlignment objects, the similarity matrix is plotted and optionally, the set of matching peaks. clusterAlignment objects are just a collection of all pairwise peakAlignment objects.

Author(s)
Mark Robinson

References

See Also
plot, peaksDataset
plotSpectra

Examples

```r
require(gcspikelite)
# paths and files
gcmsPath<-paste(find.package("gcspikelite"),"data",sep="/"

cdfFiles<-dir(gcmsPath,"CDF",full=TRUE)
eluFiles<-dir(gcmsPath,"ELU",full=TRUE)

# read data
pd<-peaksDataset(cdfFiles[1],mz=seq(50,550),rtrange=c(7.5,8.5))

# image plot
plotImage(pd,run=1,rtrange=c(7.5,8.5),main="")
```

Description

Plot the mass spectra from the profile matrix

Usage

```r
plotSpectra(object, sample, spectraID, normalize = TRUE, ...)
```

Arguments

- `object`: an object of class "peaksDataset" where to keep the mass spectra; both abundance (y) than m/z (x)
- `sample`: character, the sample from were to plot the mass spectra
- `spectraID`: numerical, a vector containing the index of the spectra to be plotted.
- `normalize`: logical, if TRUE normalize the intensity of the mass peak to 100, the most abundant is 100 other peaks are scaled consequetially
- `...`: other parameter passed to the plot() function

Details

Plot the deconvoluted mass spectra from the profile matrix

Author(s)

riccardo.romoli@unifi.it

Examples

```r
## need access to CDF (raw data)
require(gcspikelite)
gcmsPath <- paste(find.package("gcspikelite"), "data", sep="/"

## full paths to file names
cdfFiles <- dir(gcmsPath, "CDF", full=TRUE)
```
## create a `peaksDataset` object and add XCMS peaks to it

```r
pd <- peaksDataset(cdfFiles[1:3], mz=seq(50,550), rtrange=c(7.5,8.5))
pd <- addXCMSPeaks(cdfFiles[1:3], pd, peakPicking=c('mF'), snthresh=5, fwhm=10, step=1, steps=2, mzdif=0.5, sleep=0)
```

## align the chromatograms

```r
mp <- correlationAlignment(object=pd, thr=0.8, D=20, penalty=0.2, normalize=TRUE, minFilter=2)
```

## view the alignment results

```r
mp@Alignment
```

## plot the mass spectra

```r
par(mfrow=c(3,1))
plotSpectra(object=pd, sample=cdfFiles[1], spectraID=2)
plotSpectra(object=pd, sample=cdfFiles[2], spectraID=3)
plotSpectra(object=pd, sample=cdfFiles[3], spectraID=4)
```

---

### progressiveAlignment-class

*Data Structure for progressive alignment of many GCMS samples*

**Description**

Performs a progressive peak alignment (clustalw style) of multiple GCMS peak lists

**Usage**

```r
progressiveAlignment(pD, cA, D=1000, gap=.5, verbose=TRUE, usePeaks=TRUE, df=30, compress=TRUE)
```

**Arguments**

- `pD`: a `peaksDataset` object
- `cA`: a `clusterAlignment` object
- `D`: retention time penalty
- `gap`: gap parameter
- `verbose`: logical, whether to print information
- `usePeaks`: logical, whether to use peaks (if `TRUE`) or the full 2D profile alignment (if `FALSE`)
- `df`: distance from diagonal to calculate similarity
- `compress`: logical, whether to store the similarity matrices in sparse form

**Details**

The progressive peak alignment we implemented here for multiple GCMS peak lists is analogous to how `clustalw` takes a set of pairwise sequence alignments and progressively builds a multiple alignment. More details can be found in the reference below.

**Value**

`progressiveAlignment` object
Author(s)
Mark Robinson

References

See Also
peaksDataset, multipleAlignment

Examples

```r
require(gcspikelite)

# paths and files
gcmsPath<-paste(find.package("gcspikelite"),"data",sep="/")
cdfFiles<-dir(gcmsPath,"CDF",full=TRUE)
eluFiles<-dir(gcmsPath,"ELU",full=TRUE)

# read data, peak detection results
dd<-peaksDataset(cdfFiles[1:2],mz=seq(50,550),rtrange=c(7.5,8.5))
dd<-addAMDISPeaks(dd,eluFiles[1:2])

cac<-clusterAlignment(dd, gap = .5,D=.05,df=30)
pa<-progressiveAlignment(dd, cac, gap = .6, D=.1,df=30)
```

Description
Build a fat data matrix

Usage

```r
retFatMatrix(data)
```

Arguments

- `data` the list obtained from gatherInfo()

Details
This function allows to extract the data from an object created using gatherInfo and build a data matrix using the area of the deconvoluted and aligned peaks. The row are the samples while the column represent the different peaks.

Value
A fat data matrix containing the area of the deconvoluted and aligned peaks. The row are the samples while the column represent the different peaks.
rmaFitUnit

Fits a robust linear model (RLM) for one metabolite

Description
Using rlm from MASS, this procedure fits a linear model using all the fragments

Usage
rmaFitUnit(u,maxit=5,mzEffect=TRUE,cls=NULL,fitSample=TRUE,fitOrCoef=c("coef","fit"),TRANSFORM=log2)

Arguments
u a metabolite unit (list object with vectors mz and rt for m/z and retention times, respectively and a data element giving the fragmentxsample intensitiy matrix)
maxit maximum number of iterations (default: 5)
mzEffect logical, whether to fit m/z effect (default: TRUE)
cls class variable
fitSample whether to fit individual samples (alternative is fit by group)
fitOrCoef whether to return a vector of coefficients (default: "coef"), or an rlm object ("fit")
TRANSFORM function to transform the raw data to before fitting (default: log2)

Details
Fits a robust linear model.
Value
list giving elements of fragment and sample coefficients (if fitOrCoef="coef") or a list of elements from the fitting process (if fitOrCoef="fit")

Author(s)
Mark Robinson

References

See Also
peaksAlignment, clusterAlignment

Examples
require(gcspikelite)

# paths and files
gcmsPath<-paste(find.package("gcspikelite"),"data",sep="/")
cdfFiles<dir(gcmsPath,"CDF",full=TRUE)
eluFiles<dir(gcmsPath,"ELU",full=TRUE)

# read data, peak detection results
pd<-peaksDataset(cdfFiles[1:2],mz=seq(50,550),rtrange=c(7.5,8.5))
pd<-addAMDISPeaks(pd,eluFiles[1:2])

# pairwise alignment using all scans
fullca<-clusterAlignment(pd, usePeaks = FALSE, df = 100)

# calculate retention time shifts
timedf<-calcTimeDiffs(pd, fullca)
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