Package ‘genoset’

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Type Package

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Description GenoSet provides an extension of the RangedSummarizedExperiment class with additional API features. This class provides convenient and fast methods for working with segmented genomic data. Additionally, GenoSet provides the class RleDataFrame which stores runs of data along the genome for multiple samples and provides very fast summaries of arbitrary row sets (regions of the genome).

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LazyLoad yes

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### R topics documented:

- genoset-package
- baf2mbaf
- boundingIndices
- boundingIndicesByChr
- bounds2Rle
- calcGC
- calcGC2
- chr
- chrIndices
- chrInfo
- chrNames
- chrOrder
- chrPartitioning
- cn2lr
- fixSegNAs
- gcCorrect
- genome
- genomeAxis
- genoPlot
- genoPos
- GenoSet
- GenoSet-class
- genoset-datasets
- isGenomeOrder
- lr2cn
- modeCenter
- nrow,GenomicRanges-method
- numCallable
- pos,GenoSetOrGenomicRanges-method
- rangeSampleMeans
- rangeSegMeanLength
- rbindDataframe
- readGenoSet
- RleDataFrame-class
- RleDataFrame-views
- runCBS
- segPairTable
- segs2Granges
- segs2Rle
- segs2RleDataFrame
- segTable
- toGenomeOrder
- [,GenoSet,ANY-method

---

**Index**

3
3
4
5
6
6
6
7
7
8
9
9
10
11
11
12
12
13
13
14
15
16
17
18
18
19
20
20
21
21
22
22
23
24
24
25
27
28
29
31
31
32
33
34
35
36
**GenoSet: An eSet for data with genome locations**

**Description**
Load, manipulate, and plot copynumber and BAF data.

**See Also**
genoset-datasets

---

**baf2mbaf**
Calculate mBAF from BAF

**Description**
Calculate Mirrored B-Allele Frequency (mBAF) from B-Allele Frequency (BAF) as in Staaf et al., Genome Biology, 2008. BAF is converted to mBAF by folding around 0.5 so that is then between 0.5 and 1. HOM value are then made NA to leave only HET values that can be easily segmented. Values > hom.cutoff are made NA. Then, if genotypes (usually from a matched normal) are provided as the matrix ‘calls’ additional HOMs can be set to NA. The argument ‘call.pairs’ is used to match columns in ‘calls’ to columns in ‘baf’.

**Usage**
baf2mbaf(baf, hom.cutoff = 0.95, calls = NULL, call.pairs = NULL)

**Arguments**
- **baf**: numeric matrix of BAF values
- **hom.cutoff**: numeric, values above this cutoff to be made NA (considered HOM)
- **calls**: matrix of NA, CT, AG, etc. genotypes to select HETs (in normals). Dimnames must match baf matrix.
- **call.pairs**: list, names represent target samples for HOMs to set to NA. Values represent columns in "calls" matrix.

**Value**
numeric matrix of mBAF values

**Examples**
```r
data(genoset.package="genoset")
baf = baf2mbaf( genoset.ds[, , "baf"], hom.cutoff=0.9 )
calls = matrix(sample(c("AT","AA","CG","GC","AT","GG"),(nrow(genoset.ds) * 2),replace=TRUE),ncol=2,dimnames=list(rownames(genoset.ds),c("K","L")))
mbaf = baf2mbaf( genoset.ds[, , "baf"], hom.cutoff=0.9, calls = calls, call.pairs = list(K="L",L="L") ) # Sample L is matched normal for tumor sample K, M only uses hom.cutoff
mbaf = baf2mbaf( genoset.ds[, , "baf"], hom.cutoff=0.9 ) # Put mbaf back into the BAFSet object
```
boundingIndices

Find indices of features bounding a set of chromosome ranges/genes

Description

This function is similar to findOverlaps but it guarantees at least two features will be covered. This is useful in the case of finding features corresponding to a set of genes. Some genes will fall entirely between two features and thus would not return any ranges with findOverlaps. Specifically, this function will find the indices of the features (first and last) bounding the ends of a range/gene (start and stop) such that first <= start < stop <= last. Equality is necessary so that multiple conversions between indices and genomic positions will not expand with each conversion. Ranges/genes that are outside the range of feature positions will be given the indices of the corresponding first or last index rather than 0 or n + 1 so that genes can always be connected to some data.

Usage

boundingIndices(starts, stops, positions, all.indices = FALSE)

Arguments

starts integer vector of first base position of each query range
stops integer vector of last base position of each query range
positions Base positions in which to search
all.indices logical, return a list containing full sequence of indices for each query

Details

This function uses some tricks from findIntervals, where is for k queries and n features it is O(k * log(n)) generally and ~O(k) for sorted queries. Therefore will be dramatically faster for sets of query genes that are sorted by start position within each chromosome. The index of the stop position for each gene is found using the left bound from the start of the gene reducing the search space for the stop position somewhat. boundingIndices does not check for NAs or unsorted data in the subject positions. These assumptions are safe for position info coming from a GenoSet or GRanges.

Value

integer matrix of 2 columns for start and stop index of range in data or a list of full sequences of indices for each query (see all.indices argument)

See Also

Other "range summaries": boundingIndicesByChr, rangeSampleMeans

Examples

starts = seq(10,100,10)
boundingIndices( starts=starts, stops=starts+5, positions = 1:100 )
boundingIndicesByChr

Find indices of features bounding a set of chromosome ranges/genes, across chromosomes

Description

Finds subject ranges corresponding to a set of genes (query ranges), taking chromosome into account. Specifically, this function will find the indices of the features (first and last) bounding the ends of a range/gene (start and stop) such that first <= start < stop <= last. Equality is necessary so that multiple conversions between indices and genomic positions will not expand with each conversion. Ranges/genes that are outside the range of feature positions will be given the indices of the corresponding first or last index on that chromosome, rather than 0 or n + 1 so that genes can always be connected to some data. Checking the left and right bound for equality will tell you when a query is off the end of a chromosome.

Usage

boundingIndicesByChr(query, subject)

Arguments

query GRanges or something coercible to GRanges
subject GenomicRanges

Details

This function uses some tricks from findIntervals, where is for k queries and n features it is O(k * log(n)) generally and ~O(k) for sorted queries. Therefore will be dramatically faster for sets of query genes that are sorted by start position within each chromosome. The index of the stop position for each gene is found using the left bound from the start of the gene reducing the search space for the stop position somewhat.

This function differs from boundingIndices in that 1. it uses both start and end positions for the subject, and 2. query and subject start and end positions are processed in blocks corresponding to chromosomes.

Both query and subject must be in at least weak genome order (sorted by start within chromosome blocks).

Value

integer matrix with two columns corresponding to indices on left and right bound of queries in subject

See Also

Other "range summaries": boundingIndices, rangeSampleMeans
bounds2Rle  

Convert bounding indices into a Rle

Description

Given a matrix of first/last indices, like from boundingIndicesByChr, and values for each range, convert to a Rle. This function takes the expected length of the Rle, n, so that any portion of the full length not covered by a first/last range will be a run with the value NA. This is typical in the case where data is segmented with CBS and some of the data to be segmented is NA.

Usage

bounds2Rle(bounds, values, n)

Arguments

- bounds: matrix, two columns, with first and last index, like from boundingIndicesByChr
- values: ANY, some value to be associated with each range, like segmented copy number.
- n: integer, the expected length of the Rle, i.e. the number of features in the genome/target ranges processed by boundingIndicesByChr.

Value

Rle

See Also

Other "segmented data": rangeSegMeanLength, runCBS, segPairTable, segTable, segs2Granges, segs2RleDataFrame, segs2Rle

---

calcGC  

Calculate GC Percentage in windows

Description

Local GC content can be used to remove GC artifacts from copynumber data (see Diskin et al, Nucleic Acids Research, 2008, PMID: 18784189). This function will calculate GC content fraction in expanded windows around a set of ranges following example in http://www.bioconductor.org/help/course-materials/2012/useR2012/Bioconductor-tutorial.pdf. Currently all ranges are tabulated, later I may do letterFrequencyInSlidingWindow for big windows and then match to the nearest.

Usage

calcGC(object, bsgenome, expand = 1e+06, bases = c("G", "C"))

Arguments

- object: GenomicRanges or GenoSet
- bsgenome: BSgenome, like Hsapiens from BSgenome.Hsapiens.UCSC.hg19 or DNAStringSet.
- expand: scalar integer, amount to expand each range before calculating gc
- bases: character, alphabet to count, usually c("G", "C"), but "N" is useful too
**calcGC2**

**Value**
numeric vector, fraction of nucleotides that are G or C in expanded ranges of object

**Examples**
```r
## Not run: library(BSgenome.Hsapiens.UCSC.hg19)
## Not run: gc = calcGC(genoset.ds, Hsapiens)
```

---

**Description**
Local GC content can be used to remove GC artifacts from copynumber data (see Diskin et al, Nucleic Acids Research, 2008, PMID: 18784189). This function will calculate GC content fraction in expanded windows around a set of ranges following example in http://www.bioconductor.org/help/course-materials/2012/useR2012/Bioconductor-tutorial.pdf. Values are as.integer( 1e4 * fraction ) for space reasons.

**Usage**
```r
calcGC2(dna)
```

**Arguments**
- `dna`: BSgenome or DNAStringSet

**Value**
SimpleRleList, integer 1e4 * GC fraction, chromosomes 1:22, X and Y

**Examples**
```r
## Not run: library(BSgenome.Hsapiens.UCSC.hg19)
## Not run: gc = calcGC2(Hsapiens)
```

---

**chr**

**Chromosome name for each feature**

**Description**
Get chromosome name for each feature. Returns character.

**Usage**
```r
chr(object)
```

**Examples**
```r
## S4 method for signature 'GenoSet'
chr(object)
## S4 method for signature 'GenomicRanges'
chr(object)
```
chrIndices

Arguments

  object   GRanges GenoSet

Value

  character vector of chromosome positions for each feature

Examples

```r
data(genoset, package="genoset")
chr(genoset.ds) # c("chr1","chr1","chr1","chr3","chr3","chrX","chrX","chrX")
chr(rowRanges(genoset.ds)) # The same
```

chrIndices

*Get a matrix of first and last index of features in each chromosome*

Description

Sometimes it is handy to know the first and last index for each chr. This is like chrInfo but for feature indices rather than chromosome locations. If chr is specified, the function will return a sequence of integers representing the row indices of features on that chromosome.

Usage

```r
chrIndices(object, chr = NULL)
```

## S4 method for signature 'GenoSetOrGenomicRanges'

```r
chrIndices(object, chr = NULL)
```

Arguments

  object   GenoSet or GRanges
  chr       character, specific chromosome name

Value

  data.frame with "first" and "last" columns

Examples

```r
data(genoset, package="genoset")
chrIndices(genoset.ds)
chrIndices(rowRanges(genoset.ds)) # The same
chrInfo

Get chromosome start and stop positions

Description
Provides a matrix of start, stop and offset, in base numbers for each chromosome.

Usage
chrInfo(object)

## S4 method for signature 'GenoSetOrGenomicRanges'
chrInfo(object)

Arguments

object A GenoSet object or similar

Value
list with start and stop position, by ordered chr

Examples
data(genoset, package="genoset")
chrInfo(genoset.ds)
chrInfo(rowRanges(genoset.ds)) # The same

chrNames

Get list of unique chromosome names

Description
Get list of unique chromosome names

Usage
chrNames(object)

## S4 method for signature 'GenoSet'
chrNames(object)

## S4 method for signature 'GenomicRanges'
chrNames(object)

chrNames(object) <- value

## S4 replacement method for signature 'GenoSet'
chrNames(object) <- value

## S4 replacement method for signature 'GenomicRanges'
chrNames(object) <- value
chrOrder

Arguments

- **object**: GenomicRanges or GenoSet
- **value**: return value of chrNames

Value

character vector with names of chromosomes

Examples

```r
data(genoset, package="genoset")
chrNames(genoset.ds) # c("chr1","chr3","chrX")
chrNames(rowRanges(genoset.ds)) # The same
chrNames(genoset.ds) = sub("^chr","",chrNames(genoset.ds))
```

chrOrder

Order chromosome names in proper genome order

Description

Chromosomes make the most sense ordered by number, then by letter.

Usage

```r
chrOrder(chr.names)
```

Arguments

- **chr.names**: character, vector of unique chromosome names

Value

character vector of chromosome names in proper order

See Also

Other "genome ordering": isGenomeOrder, toGenomeOrder

Examples

```r
chrOrder(c("chr5","chrX","chr3","chr7","chrY")) # c("chr3","chr5","chr7","chrX","chrY")
```
chrPartitioning

Partitioning by Chromosome

Description
Get indices of first and last element in each chromosome.

Usage
chrPartitioning(object)

Arguments
object GenoSet or GenomicRanges

Value
PartitioningByEnd

cn2lr
Take vector or matrix of copynumber values, convert to log2ratios

Description
Utility function for converting copynumber units (2 is normal) to log2ratio units (two is normal). If ploidy is provided lr is log2(cn/ploidy), otherwise log2(cn/2).

Usage
cn2lr(x, ploidy)

## S4 method for signature 'numeric'
cn2lr(x, ploidy)

## S4 method for signature 'matrix'
cn2lr(x, ploidy)

## S4 method for signature 'DataFrame'
cn2lr(x, ploidy)

Arguments
x numeric vector or matrix, or DataFrame with numeric-like columns (Rle typically). Assumed to be in copynumber units.
ploidy numeric, of length ncol(x). Ploidy of each sample.

Value
data of same type as "x" transformed into log2ratio units
See Also

lr2cn

Fix NA runs in a Rle

Description

Fix NA runs in a Rle when the adjacent runs have equal values

Usage

fixSegNAs(x, max.na.run = 3)

Arguments

x
Rle to be fixed
max.na.run
integer, longest run of NAs that will be fixed

Value

Rle

gcCorrect

Correct copy number for GC content

Description

Copy number estimates from various platforms show "Genomic Waves" (Diskin et al., Nucleic Acids Research, 2008, PMID: 18784189) where copy number trends with local GC content. This function regresses copy number on GC percentage and removes the effect (returns residuals). GC content should be smoothed along the genome in wide windows >= 100kb.

Usage

gcCorrect(ds, gc, retain.mean = TRUE)

Arguments

ds
numeric matrix of copynumber or log2ratio values, samples in columns

gc
numeric vector, GC percentage for each row of ds, must not have NAs
retain.mean
logical, center on zero or keep same mean?

Value

numeric matrix, residuals of ds regressed on gc

Examples

gc = runif(n=100, min=1, max=100)
ds = rnorm(100) + (0.1 * gc)
gcCorrect(ds, gc)
genome

Get and set the genome universe annotation.

Description
Genome version

Arguments
x GenoSet

Details
The genome positions of the features in locData. The UCSC notation (e.g. hg18, hg19, etc.) should be used.

Value
character, e.g. hg19

Examples
data(genoset)
genome(genoset.ds)
genome(genoset.ds) = "hg19"

---

geneomeAxis

Label axis with base pair units

Description
Label an axis with base positions

Usage
genomeAxis(locs = NULL, side = 1, log = FALSE, do.other.side = TRUE)

Arguments
locs GenomicRanges to be used to draw chromosome boundaries, if necessary. Usually rowRanges slot from a GenoSet.
side integer side of plot to put axis
log logical Is axis logged?
do.other.side logical, label non-genome side with data values at tick marks?

Details
Label a plot with Mb, kb, bp as appropriate, using tick locations from axTicks
Value
nothing

See Also
Other "genome plots": \texttt{genoPlot}

Examples

\begin{verbatim}
data(genoset, package="genoset")
genoPlot(genoPos(genoset.ds), genoset.ds[,1, "baf"])
genomeAxis( locs=rowRanges(genoset.ds) )  # Add chromosome names and boundaries to a plot assuming genome along x-axis
genomeAxis( locs=rowRanges(genoset.ds), do.other.side=FALSE )  # As above, but do not label y-axis with data values at tickmarks
genomeAxis()  # Add nucleotide position in sensible units assuming genome along x-axis
\end{verbatim}

\texttt{genoPlot}

Plot data along the genome

Description

Plot location data and chromosome boundaries from a GenoSet or GRanges object against data from a numeric or Rle. Specifying a chromosome name and optionally a \texttt{"xlim"} will zoom into one chromosome region. If more than one chromosome is present, the chromosome boundaries will be marked. Alternatively, for a numeric \texttt{x} and a numeric or Rle \texttt{y}, data in \texttt{y} can be plotted at genome positions \texttt{x}. In this case, chromosome boundaries can be taken from the argument \texttt{locs}. If data for \texttt{y}-axis comes from a Rle lines are plotted representing segments. X-axis tickmarks will be labeled with genome positions in the most appropriate units.

Usage

\begin{verbatim}
\texttt{genoPlot(x, y, \ldots)}
\end{verbatim}

Arguments

\begin{itemize}
\item \texttt{x} GenoSet (or descendant) or GRanges
\item \texttt{y} numeric or Rle
\item \texttt{\ldots} Additional plotting args
\item \texttt{add} Add plot to existing plot
\end{itemize}
genoPos

xlab character, label for x-axis of plot
ylab character, label for y-axis of plot
col character, color to plot lines or points
locs GRanges, like rowRanges slot of GenoSet
lwd numeric, line width for segment plots from an Rle
xlim integer, length two, bounds for genome positions. Used in conjunction with "chr" to subset data for plotting.
chr Chromosome to plot, NULL by default for full genome
pch character or numeric, printing character, see points

Value
TRUE

Methods
signature(x = "GenoSetOrGenomicRanges", y = "ANY") Plot feature locations and data from one sample.
signature(x = "numeric", y = "numeric") Plot numeric location and a vector of numeric data.
signature(x = "numeric", y = "Rle") Plot numeric location and a vector of Rle data. Uses lines for Rle runs.

See Also
Other "genome plots": genomeAxis

Examples

data(genoset,package="genoset")
genoPlot( x=genoset.ds,y=genoset.ds[,1,"lrr"] )
genoPlot( genoPos(genoset.ds), genoset.ds[,1,"lrr"], locs=rowRanges(genoset.ds) ) # The same
genoPlot( 1:10, Rle(c(rep(0,5),rep(3,4),rep(1,1))) )

---

genoPos Get base positions of features in genome-scale units

Description
Get base positions of array features in bases counting from the start of the genome. Chromosomes are ordered numerically, when possible, then lexically.

Usage
genoPos(object)

## S4 method for signature 'GenoSetOrGenomicRanges'
genoPos(object)
GenoSet

Arguments

object A GenoSet object or a GenomicRanges object

Value

numeric position of each feature in whole genome units, in original order

Examples

data(genoset, package="genoset")
head(genoPos(genoset.ds))
head(genoPos(rowRanges(genoset.ds))) # The same

GenoSet Create a GenoSet object

Description

This function is the preferred method for creating a new GenoSet object. Currently, a GenoSet is simply a RangedSummarizedExperiment with some API changes and extra methods. Therefore, a GenoSet must always have a rowRanges.

Usage

GenoSet(rowRanges, assays, colData, metadata = list())

## S4 method for signature 'GenoSet'
lengths(x)
locData(object, ...) <- value
locData(object)

Arguments

rowRanges GenomicRanges, not a GenomicRangesList
assays list, SimpleList or matrix-like object
colData a data.frame or DataFrame of sample metadata with rownames matching the colnames of the matrices in assays
metadata a list of any other data you want to attach to the GenoSet object
x A GenoSet

Details

locations. Rownames are required to match featureNames.

Value

A GenoSet object
**GenoSet-class**

**Examples**

```r
test.sample.names = LETTERS[11:13]
probe.names = letters[1:10]
assays = list(matrix(31:60,nrow=10,ncol=3,dimnames=list(probe.names,test.sample.names)))
rowRanges = GRanges(ranges=IRanges(start=1:10,width=1,names=probe.names),seqnames=c(rep("chr1",4),rep("chr3",2),rep("chrX",4)))
colData = data.frame(matrix(LETTERS[1:15],nrow=3,ncol=5,dimnames=list(test.sample.names,letters[1:5])))
rse = SummarizedExperiment(rowRanges=rowRanges,assays=assays,colData=colData,metadata=metadata)
gs = GenoSet(rowRanges, assays, colData)
```

**Description**

`GenoSet` extends `RangedSummarizedExperiment` by adding some additional methods to the API. Examples include subsetting rows with a GenomicRanges and combining this with access to assays like `genoset[i,j,assay]`.

**Extends**

Class `RangedSummarizedExperiment`, directly.

**Methods**

- `[` signature(x = "GenoSet", i = "ANY", j = "ANY", drop = "ANY"): ...
- `[` signature(x = "GenoSet", i = "character", j = "ANY", drop = "ANY"): ...
- `[-` signature(x = "GenoSet", i = "ANY", j = "ANY", value = "ANY"): ...
- `chr` signature(object = "GenoSet"): ...
- `chrNames` signature(object = "GenoSet"): ...
- `dim` signature(object = "GenoSet"): ...
- `genoPlot` signature(x = "GenoSet", y = "ANY"): ...
- `rowRanges` signature(object = "GenoSet"): ...
- `names` signature(x = "GenoSet"): ...
- `ranges` signature(x = "GenoSet"): ...
- `chrInfo` signature(x = "GenoSet"): ...
- `chrIndices` signature(x = "GenoSet"): ...
- `show` signature(object = "GenoSet"): ...
- `toGenomeOrder` signature(ds = "GenoSet"): ...
- `isGenomeOrder` signature(ds = "GenoSet"): ...
- `assays` signature(x = "GenoSet"): ...
- `assay` signature(x = "GenoSet", i="ANY"): ...
- `assay<-` signature(x = "GenoSet", i="ANY",value="ANY"): ...
- `assayNames` signature(x = "GenoSet"): ...
- `colData` signature(x = "GenoSet"): ...
- `locData` signature(x = "GenoSet"): ...
- `locData<-` signature(x = "GenoSet",value="GenomicRanges"): ...
See Also

GenoSet

Examples

```r
test.sample.names = LETTERS[11:13]
probe.names = letters[1:10]
assays=list(matrix(31:60,nrow=10,ncol=3,dimnames=list(probe.names,test.sample.names)))
rowRanges=GRanges(ranges=IRanges(start=1:10,width=1,names=probe.names),seqnames=c(rep("chr1",4),rep("chr3",2),rep("chrX",4)))
colData=data.frame(matrix(LETTERS[1:15],nrow=3,ncol=5,dimnames=list(test.sample.names,letters[1:5])))
rse=SummarizedExperiment(rowRanges=rowRanges,assays=assays,colData=colData,metadata=metadata)

gs = GenoSet(rowRanges, assays, colData)
```

Description

A GenoSet object the 'baf' (B-Allele Frequency) and 'lrr' (Log-R Ratio) assay matrices. The 'lrr' assay matrix contains DNA copy number on the scale of tumor/ploidy and the 'baf' assay matrix contains data in the range 0 to 1 where 0 indicates the AA genotype, 0.5 indicates the AB genotype and 1 indicates the BB genotype.

Source

Simulated data

isGenomeOrder

Check if a GRanges or GenoSet is in genome order

Description

Checks that rows in each chr are ordered by start. If strict=TRUE, then chromosomes must be in order specified by chrOrder. isGenomeOrder for GRanges differs from order in that it orders by chromosome and start position only, rather than chromosome, strand, start, and width.

Usage

```r
isGenomeOrder(ds, strict = TRUE)
## S4 method for signature 'GenoSet'
isGenomeOrder(ds, strict = TRUE)
## S4 method for signature 'GRanges'
isGenomeOrder(ds, strict = TRUE)
```

Arguments

- `ds` : GenoSet or GRanges
- `strict` : logical, should space/chromosome order be identical to that from chrOrder?
Value

logical

See Also

Other "genome ordering": `chrOrder`, `toGenomeOrder`

Examples

data(genoset, package = "genoset")
isGenomeOrder( rowRanges(genoset.ds) )

isGenomeOrder
Check if a GRanges or GenoSet is in genome order

Description

Checks that rows in each chr are ordered by start. If strict=TRUE, then chromosomes must be in order specified by chrOrder. isGenomeOrder for GRanges differs from order in that it orders by chromosome and start position only, rather than chromosome, strand, start, and width.

Usage

isGenomeOrder(ds, strict = TRUE)

Arguments

ds GenoSet or GRanges
strict logical, should space/chromosome order be identical to that from chrOrder?

Value

logical

See Also

Other "genome ordering": `chrOrder`, `toGenomeOrder`

Examples

data(genoset, package = "genoset")
isGenomeOrder( rowRanges(genoset.ds) )
1r2cn

*Take vector or matrix of log2 ratios, convert to copynumber*

---

### Description

Utility function for converting log2ratio units (zero is normal) to copynumber units (two is normal)

### Usage

```r
1r2cn(x)
```

### Arguments

- `x`: numeric data in log2ratio values

### Value

data of same type as "x" transformed into copynumber units

### See Also

cn2lr

---

### modeCenter

*Center continuous data on mode*

---

### Description

Copynumber data distributions are generally multi-modal. It is often assumed that the tallest peak represents "normal" and should therefore be centered on a log2ratio of zero. This function uses the density function to find the mode of the dominant peak and subtracts that value from the input data.

### Usage

```r
modeCenter(ds)
```

### Arguments

- `ds`: numeric matrix

### Value

numeric matrix

### Examples

```r
modeCenter( matrix( rnorm(150, mean=0), ncol=3 ))
```
**nrow, GenomicRanges-method**

*GenomicRanges API Additions*

**Description**

I have extended the API for GenomicRanges a bit so that genoset and GenomicRanges can have the same API, at least as far as genome location based features go.

**Usage**

```r
## S4 method for signature 'GenomicRanges'
nrow(x)
```

**Arguments**

- `x` A GenomicRanges

---

**numCallable**

*Count Rle positions >= min*

**Description**

For Rle coverage vector, count number of positions where value >= min, think callable bases.

**Usage**

```r
numCallable(rle, bounds, min)
```

**Arguments**

- `rle` integer Rle, no NAs
- `bounds` IRanges or matrix, positions in Rle to consider. If bounds is a matrix, the first two columns are used as start and end.
- `min` scalar integer, count Rle positions >= this value.

**Value**

integer vector of length nrow(bounds)
pos, GenoSetOrGenomicRanges-method

Chromosome position of features

Description

Get chromosome position of features/ranges. Defined as floor of mean of start and end.

Usage

## S4 method for signature 'GenoSetOrGenomicRanges'
pos(x)

Arguments

x

GRanges GenoSet

Value

numeric vector of feature positions within a chromosome

Examples

data(genoset, package = "genoset")
pos(genoset. ds) # 1:10
pos(rowRanges(genoset. ds)) # The same

rangeSampleMeans

Average features in ranges per sample

Description

This function takes per-feature genomic data and returns averages for each of a set of genomic ranges. The most obvious application is determining the copy number of a set of genes. The features corresponding to each gene are determined with boundingIndices such that all features with the bounds of a gene (overlaps). The features on either side of the gene unless those positions exactly match the first or last base covered by the gene. Therefore, genes falling between two features will at least cover two features. Range bounding is performed by the boundingIndices function.

Usage

rangeSampleMeans(query, subject, assay. element, na.rm = FALSE)

Arguments

query

GRanges object representing genomic regions (genes) to be averaged.

subject

A GenoSet object or derivative

assay. element

character, name of element in assayData to use to extract data

na.rm

scalar logical, ignore NAs?
rangeSegMeanLength

Value
numeric matrix of features in each range averaged by sample

See Also
Other "range summaries": boundingIndicesByChr, boundingIndices

Examples
data(genoset)
my.genes = GRanges( ranges=IRanges(start=c(35e6,128e6),end=c(37e6,129e6),names=c("HER2","CMYC")), seqnames=c("chr17","chr8") )
rangeSampleMeans( my.genes, genoset.ds, "lrr" )

rangeSegMeanLength

Get segment widths

Description
The width of a genomic segment helps inform us about the importance of a copy number value. Focal amplifications are more interesting than broad gains, for example. Given a range of interesting regions (i.e. genes) this function determines all genomics segments covered by each gene and returns the average length of the segments covered by each gene in each sample. Often only a single segment covers a given gene in a given sample.

Usage
rangeSegMeanLength(range.gr, segs)

## S4 method for signature 'GRanges,list'
rangeSegMeanLength(range.gr, segs)

## S4 method for signature 'GRanges,data.frame'
rangeSegMeanLength(range.gr, segs)

Arguments

range.gr     GRanges, genome regions of interest, usually genes
segs         data.frame of segments, like from segTable, or a list of these

Value
named vector of lengths, one per item in range.gr, or a range x length(segs) of these if segs is also list-like.

See Also
Other "segmented data": bounds2Rle, runCBS, segPairTable, segTable, segs2Granges, segs2RleDataFrame, segs2Rle
**rbindDataframe**  
*A fast method for concatenating data.frames*

**Description**  
Performs the same action as `do.call(rbind, list_of_dataframes)`, but dramatically faster. Part of the speed comes from assuming that all of the data.frames have the same column names and types. If desired an additional factor column can be added that specifies the original list element associated with each row. The argument ‘element.colname’ is used to name this column.

**Usage**  
rbindDataframe(dflist, element.colname)

**Arguments**

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>dflist</td>
<td>list of data.frames</td>
</tr>
<tr>
<td>element.colname</td>
<td>scalar character, name for additional factor column giving the name of the element of ‘dflist’ corresponding to each row. ‘dflist’ must be named to use this feature.</td>
</tr>
</tbody>
</table>

**Details**

For a list of 1000 data.frames with 884 rows and 12 columns ‘rbindDataframe’ takes 0.553s and ‘do.call(rbind,x)’ takes 327.304s, a 600X speedup. This pure-R solution is made possible by the lovely shallow copy features Michael Lawrence has added to base R.

**Value**

data.frame

---

**readGenoSet**  
*Load a GenoSet from a RData file*

**Description**

Given a rds file or a rda file with one GenoSet, load it, and return. Objects that pre-date the switch to a RangedSummarizedExperiment internal representation (V 1.29.0) are automatically switched to the new format.

**Usage**

readGenoSet(path)

**Arguments**

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>path</td>
<td>character, path to rds or rda file</td>
</tr>
</tbody>
</table>
Value
GenoSet or related object (only object in RData file)

Examples
## Not run: ds = readGenoSet("/path/to/genoset.RData")
## Not run: ds = readGenoSet("/path/to/genoset.rda")
## Not run: ds = readGenoSet("/path/to/genoset.rds")

Description
The RleDataFrame class serves to hold a collection of Run Length Encoded vectors (Rle objects) of the same length. For example, it could be used to hold information along the genome for a number of samples, such as sequencing coverage, DNA copy number, or GC content. This class inherits from both DataFrame and SimpleRleList (one of the AtomicVector types). This means that all of the usual subsetting and applying functions will work. Also, the AtomicList functions, like mean and sum, that automatically apply over the list elements will work. The scalar mathematical AtomicList methods can make this class behave much like a matrix (see Examples).

New objects can be created with the RleDataFrame constructor: RleDataFrame(..., row.names=NULL), where ... can be a list of Rle objects, or one or more individual Rle objects.

Use in Biobase eSet objects
The genoset class defines an annotatedDataFrameFrom method for DataFrame, which makes it possible to include DataFrames as assayData elements. The column names for DataFrame cannot be NULL, which makes it impossible to use them as assays in SummarizedExperiment at this time.

Row and Column Summaries
These objects will sometimes be in place of a matrix, as in the eSet example above. It is convenient to have some of the summarization methods for matrices. Each of these methods takes an RleDataFrame and returns a single Rle. The time required is similar to that required for a matrix. For an RleDataFrame x,
rowSums: Sum across 'rows'.
rowMeans: Means across 'rows'.
colSums: Sum each Rle. This is just the sum method for SimpleRleList.
colSums: Mean of each Rle. This is just the mean method for SimpleRleList.

Slots
rownames: Object of class "character" OR NULL" Names to describe each row of the DataFrame.
These may end up taking more space than your collection of Rle objects, so consider leaving this NULL.
nrows: Object of class "integer" Number of rows.
elementType: Object of class "character" Notes that elements of the internal list are Rle objects.
elementMetadata: Object of class "DataTable" OR NULL" Metadata on the elements, see DataFrame.
metadata: Object of class "list" Metadata on the whole object, see DataFrame.
listData: Object of class "list" Base list containing the Rle objects.
Extends

Class "SimpleRleList", directly. Class "DataFrame", directly.

Methods

as.matrix signature(x = "RleDataFrame"): Convert to matrix.
coerce signature(x = "RleDataFrame"): Convert to other classes.
colMeans signature(x = "RleDataFrame"): Mean of each column.
colSums signature(x = "RleDataFrame"): Sum of each column.
rowMeans signature(x = "RleDataFrame"): Mean of each 'row'.
rowSums signature(x = "RleDataFrame"): Sum of each 'row'.
show signature(object = "RleDataFrame"): Short and pretty description of an object of this type.

Author(s)

Peter M. Haverty, design suggestion from Michael Lawrence.

See Also

DataFrame AtomicList Rle RleList rowMeans colMeans rowSums colSums view-summarization-methods

Examples

showClass("RleDataFrame")

## Constructors
df = new("RleDataFrame", listData=list(A=Rle(c(NA, 2:3, NA, 5), rep(2, 5)), B=Rle(c(6:7, NA, 8:10),c(3,2,1,2,1,1))), nrows=10L)

df2 = RleDataFrame(list(A=Rle(c(NA, 2:3, NA, 5), rep(2, 5)),
B=Rle(c(6:7, NA, 8:10),c(3,2,1,2,1,1))))

df3 = RleDataFrame(A=Rle(c(NA, 2:3, NA, 5), rep(2, 5)), B=Rle(c(6:7,
NA, 8:10),c(3,2,1,2,1,1))))

## AtomicList Methods
runValue(df)
runLength(df)
ranges(df)
mean(df)
sum(df)
 df + 5
log2(df) - 1

## Row and Column Summaries
rowSums(df)
colSums(df)
rowMeans(df)
colMeans(df)

## Coercion
as(df, "matrix")
as(df, "list")
as(df, "RleList")
as(df, "DataFrame")
as(df, "data.frame")

---

**RleDataFrame-views**  
*Calculate summary statistics on views of an RleDataFrame*

---

**Description**

These methods mirror the viewMeans type functions from IRanges for SimpleRleList. They differ in that they work on an RleDataFrame and an IRanges directly and also have a simplify argument. This works out to be faster (compute-wise) and also convenient.

Still, an RleDataFrame inherits from SimpleRleList, so all of the views functions will work.

**Usage**

```r
rangeSums(x, bounds, na.rm=FALSE, simplify=TRUE)
rangeMeans(x, bounds, na.rm=FALSE, simplify=TRUE, ...)
rangeMins(x, bounds, na.rm=FALSE, simplify=TRUE)
rangeMaxs(x, bounds, na.rm=FALSE, simplify=TRUE)
rangeWhichMins(x, bounds, na.rm=FALSE, simplify=TRUE)
rangeWhichMaxs(x, bounds, na.rm=FALSE, simplify=TRUE)
```

**Arguments**

- `x`  
  RleDataFrame

- `bounds`  
  Matrix with two columns or IRanges representing ranges of rows of `x` to process. If `bounds` is a matrix, an IRanges is constructed assuming the first two columns represent the start and end of the ranges. The names for the IRanges is taken from the rownames of the matrix. Such a matrix can constructed with `boundingIndicesByChr` and is the preferred input.

- `na.rm`  
  Scalar logical. Ignore NAs in calculations?

- `simplify`  
  Scalar logical. Simplify result? If TRUE, the return value will be a vector or matrix. For a single view, a vector will be returned. Otherwise a matrix with one row per view and one column per column of `x` will be returned. If FALSE, the return value will be a list of length `nrow(x)` of vectors of length `nrow(bounds)`.

- `...`  
  Additional arguments for other methods.

**Details**

The "range" name prefixes here serve to differentiate these functions from the "view" functions. This may change. I will be asking the IRanges team to add "..." and "simplify" to the "view" methods so that I can just make additional methods for RleDataFrame.

**Value**

With `simplify == TRUE`, a vector for single view or a matrix otherwise. When `simplify == FALSE`, a list of vectors length `nrow(x)` where each element is of length `nrow(bounds)`.
See Also

RleDataFrame boundingIndicesByChr

Examples

df = RleDataFrame(list(a=Rle(1:5, rep(2, 5)), b=Rle(1:5, rep(2, 5)),
row.names=LETTERS[1:10])
mat = matrix(c(1,4,3,5),ncol=2,dimnames=list(c("Gene1","Gene2"),c("start","end"))
bounds = IRanges(start=c(1, 4), end=c(3, 5), names=c("Gene1","Gene2"))

rangeMeans(df,bounds,simplify=FALSE)
rangeMeans(df,bounds,simplify=TRUE)
rangeMeans(df,mat,simplify=TRUE)

rangeMeans(df,bounds)
rangeSums(df,bounds)
rangeMins(df,bounds)
rangeMaxs(df,bounds)
rangeWhichMins(df,bounds)
rangeWhichMaxs(df,bounds)

# RleDataFrame isa SimpleRleList, so all the IRanges view* methods work too:
v = RleViewsList( lapply( df, Views, start=bounds ) )
viewMeans(v)

runCBS

Run CBS Segmentation

Description

Utility function to run CBS's three functions on one or more samples

Usage

runCBS(data, locs, return.segs = FALSE, n.cores = 1, smooth.region = 2,
outlier.SD.scale = 4, smooth.SD.scale = 2, trim = 0.025,
alpha = 0.001)

Arguments

data numeric matrix with continuous data in one or more columns
locs GenomicRanges, like rowRanges slot of GenoSet
return.segs logical, if true list of segment data.frames return, otherwise a DataFrame of Rle vectors. One Rle per sample.
n.cores numeric, number of cores to ask mclapply to use
smooth.region number of positions to left and right of individual positions to consider when smoothing single point outliers
outlier.SD.scale number of SD single points must exceed smooth.region to be considered an outlier
segPairTable

smooth.SD.scale
  floor used to reset single point outliers
trim
  fraction of sample to smooth
alpha
  pvalue cutoff for calling a breakpoint

Details
  Takes care of running CBS segmentation on one or more samples. Makes appropriate input, smooths outliers, and segment

Value
  data frame of segments from CBS

See Also
  Other "segmented data": bounds2Rle, rangeSegMeanLength, segPairTable, segTable, segs2Granges, segs2RleDataFrame, segs2Rle

Examples
  sample.names = paste("a",1:2,sep="")
  probe.names = paste("p",1:30,sep="")
  ds = matrix(c(rep(5,20),rep(3,10)),c(rep(2,10),rep(7,10),rep(9,10))),ncol=2,dimnames=list(probe.names,sample.names))
  locs = GRanges(ranges=IRanges(start=c(1:20,1:10),width=1,names=probe.names),seqnames=paste("chr",c(rep(1,20),rep(2,10)),sep=""))
  seg.rle.result = RleDataFrame( a1 = Rle(c(rep(5,20),rep(3,10))), a2 = Rle(c(rep(2,10),rep(7,10),rep(9,10))))
  seg.list.result = list( a1 = data.frame( ID=rep("a1",2), chrom=factor(c("chr1","chr2")), loc.start=c(1,1), loc.end=c(20,10), num.mark=rep(10,2)),
                           a2 = data.frame( ID=rep("a2",3), chrom=factor(c("chr1","chr1","chr2")), loc.start=c(1,11,1), loc.end=c(20,10), num.mark=rep(10,3))
  runCBS(ds,locs) # Should give seg.rle.result
  runCBS(ds,locs,return.segs=TRUE) # Should give seg.list.result

segPairTable
  Convert Rle objects to tables of segments

Description
  Like segTable, but for two Rle objects. Takes a pair of Rle or DataFrames with Rle columns and makes one or more data.frames with bounds of each new segment. Rle objects are broken up so that each resulting segment has one value from each Rle. For a DataFrame, the argument stack combines all of the individual data.frames into one large data.frame and adds a "Sample" column of sample ids.

Usage
  segPairTable(x, y, ...)

  ## S4 method for signature 'Rle,Rle'
  segPairTable(x, y, locs = NULL, chr.ind = NULL,
### Arguments

- **x**: Rle or list/DataFrame of Rle vectors
- **y**: Rle or list/DataFrame of Rle vectors
- **...**: in generic, extra arguments for methods
- **locs**: GenomicRanges with rows corresponding to rows of df
- **chr.ind**: matrix, like from chrIndices method
- **start**: integer, vector of feature start positions
- **end**: integer, vector of feature end positions
- **factor.chr**: scalar logical, make 'chrom' column a factor?
- **stack**: logical, rbind list of segment tables for each sample and add "Sample" column?

### Details

For a Rle, the user can provide `locs` or `chr.ind`, `start` and `stop`. The latter is surprisingly much faster and this is used in the DataFrame version.

### Value

one or a list of data.frames with columns chrom, loc.start, loc.end, num.mark, seg.mean

### See Also

Other "segmented data": `bounds2Rle`, `rangeSegMeanLength`, `runCBS`, `segTable`, `segs2Granges`, `segs2RleDataFrame`, `segs2Rle`

### Examples

```r
cn = Rle(c(3,4,5,6),rep(3,4))
loh = Rle(c(2,4,6,8,10,12),rep(2,6))
start = c(9:11,4:9,15:17)
end = start
locs = GRanges(IRanges(start=start,end=end),seqnames=c(rep("chr1",3),rep("chr2",6),rep("chr3",3)))
segPairTable(cn,loh,locs)
```
segs2Granges

GRanges from segment table

Description

GenoSet contains a number of functions that work on segments. Many work on a data.frame of segments, like segTable and runCBS. This function converts one of these tables in a GRanges. The three columns specifying the ranges become the GRanges and all other columns go into the ‘mcols’ portion of the GRanges object.

Usage

segs2Granges(segs)

Arguments

segs   data.frame with loc.start, loc.end, and chrom columns, like from segTable or runCBS

Value

GRanges

See Also

Other "segmented data": bounds2Rle, rangeSegMeanLength, runCBS, segPairTable, segTable, segs2RleDataFrame, segs2Rle

segs2Rle

Make Rle from segments for one sample

Description

Take output of CBS, make Rle representing all features in ‘locs’ ranges. CBS output contains run length and run values for genomic segments, which could very directly be converted into a Rle. However, as NA values are often removed, especially for mBAF data, these run lengths do not necessarily cover all features in every sample. Using the start and top positions of each segment and the location of each feature, we can make a Rle that represents all features.

Usage

segs2Rle(segs, locs)

Arguments

segs   data.frame of segments, formatted as output of segment function from DNAcopy package
locs   GenomicRanges, like rowRanges slot of a GenoSet
Value

Rle with run lengths and run values covering all features in the data set.

See Also

Other "segmented data": `bounds2Rle`, `rangeSegMeanLength`, `runCBS`, `segPairTable`, `segTable`, `segs2Granges`, `segs2RleDataFrame`

Examples

data(genoset, package="genoset")
segs = runCBS( genoset.ds[, , "lrr"], rowRanges(genoset.ds), return.segs=TRUE )
segs2Rle( segs[[1]], rowRanges(genoset.ds) ) # Take a data.frame of segments, say from DNAcopy's segment function

segs2RleDataFrame

CBS segments to probe matrix

Description

Given segments, make an RleDataFrame of Rle objects for each sample

Usage

`segs2RleDataFrame(seg.list, locs)`

Arguments

- `seg.list`: list, list of data frames, one per sample, each is result from CBS
- `locs`: rowRanges from a GenoSet object

Details

Take table of segments from CBS, convert DataTable of Rle objects for each sample.

Value

RleDataFrame with nrows same as locs and one column for each sample

See Also

Other "segmented data": `bounds2Rle`, `rangeSegMeanLength`, `runCBS`, `segPairTable`, `segTable`, `segs2Granges`, `segs2Rle`

Examples

data(genoset, package="genoset")
seg.list = runCBS( genoset.ds[, , "lrr"], rowRanges(genoset.ds), return.segs=TRUE )
segs2RleDataFrame( seg.list, rowRanges(genoset.ds) ) # Loop segs2Rle on list of data.frames in seg.list
segTable

Convert Rle objects to tables of segments

Description
Like the inverse of segs2Rle and segs2RleDataFrame. Takes a Rle or a RleDataFrame and the rowRanges both from a GenoSet object and makes a list of data.frames each like the result of CBS’s segment. Note the loc.start and loc.stop will correspond exactly to probe locations in rowRanges and the input to segs2RleDataFrame are not necessarily so. For a DataFrame, the argument stack combines all of the individual data.frames into one large data.frame and adds a "Sample" column of sample ids.

Usage
segTable(object, ...)

## S4 method for signature 'Rle'
segTable(object, locs = NULL, chr.ind = NULL,
         start = NULL, end = NULL, factor.chr = TRUE)

## S4 method for signature 'DataFrame'
segTable(object, locs, factor.chr = TRUE,
         stack = FALSE)

Arguments

object Rle or RleDataFrame
...
in generic, for extra args in methods
locs GenomicRanges with rows corresponding to rows of df
chr.ind matrix, like from chrIndices method
start integer, vector of feature start positions
end integer, vector of feature end positions
factor.chr scalar logical, make 'chrom' column a factor?
stack logical, rbind list of segment tables for each sample and add "Sample" column?

Details
For a Rle, the user can provide locs or chr.ind, start and stop. The latter is surprisingly much faster and this is used in the DataFrame version.

Value
one or a list of data.frames with columns chrom, loc.start, loc.end, num.mark, seg.mean

See Also
Other "segmented data": bounds2Rle, rangeSegMeanLength, runCBS, segPairTable, segs2Granges, segs2RleDataFrame, segs2Rle
Examples

```r
data(genoset, package="genoset")
seg.list = runCBS( genoset.ds[, , "lrr"], rowRanges(genoset.ds), return.segs=TRUE )
df = segs2RleDataFrame( seg.list, rowRanges(genoset.ds) )  # Loop segs2Rle on list of data.frames in seg.list
genoset.ds[, , "lrr.segs"] = df
segTable( df, rowRanges(genoset.ds) )
segTable( genoset.ds[, , "lrr.segs"], rowRanges(genoset.ds) )
segTable( genoset.ds[, 1, "lrr.segs"], rowRanges(genoset.ds), colnames(genoset.ds)[1] )
```

toGenomeOrder

Set a GRanges or GenoSet to genome order

Description

Returns a re-ordered object sorted by chromosome and start position. If strict=TRUE, then chromosomes must be in order specified by chrOrder. If ds is already ordered, no re-ordering is done. Therefore, checking order with isGenomeOrder, is unnecessary if order will be corrected if isGenomeOrder is FALSE.

Usage

toGenomeOrder(ds, strict = TRUE)

Arguments

ds GenoSet or GRanges
strict logical, should chromosomes be in order specified by chrOrder?

Details

toGenomeOrder for GRanges differs from sort in that it orders by chromosome and start position only, rather than chromosome, strand, start, and width.

Value

re-ordered ds

See Also

Other "genome ordering": chrOrder, isGenomeOrder

Examples

```r
data(genoset, package="genoset")
toGenomeOrder( genoset.ds, strict=TRUE )
toGenomeOrder( genoset.ds, strict=FALSE )
toGenomeOrder( rowRanges(genoset.ds) )
```
[.,GenoSet,ANY-method

Description

Subset a GenoSet

Usage

## S4 method for signature 'GenoSet,ANY'

x[i, j, k, ...., withDimnames = TRUE,
drop = FALSE]

## S4 replacement method for signature 'GenoSet,ANY,ANY,ANY'

x[i, j, k] <- value

Arguments

x          GenoSet
i           character, GRanges, logical, integer
j           character, logical, integer
k           character or integer
...         additional subsetting args
withDimnames scalar logical, put dimnames on returned assay?
drop        logical drop levels of space factor?
value       incoming data for assay "k", rows "i" and cols "j"

Examples

data(genoset,package="genoset")
genoset.ds[1:5,2:3]  # first five probes and samples 2 and 3
genoset.ds[, "K"]  # Sample called K
g = GRanges(ranges=IRanges(start=seq(from=15e6,by=1e6,length=7),width=1,names=letters[8:14]),seqnames=rep("chr17",7))
genoset.ds[ gr, "K" ]  # sample K and probes overlapping those in rd, which overlap specified ranges on chr17
Index

*Topic classes
- GenoSet-class, 17
- RleDataFrame-class, 25
*Topic datasets
- genoset-datasets, 18
*Topic methods
- RleDataFrame-class, 25
- RleDataFrame-views, 27
- [,GenoSet,ANY-method, 35
- [,GenoSet,ANY,ANY,ANY-method ([,GenoSet,ANY-method), 35
- as.matrix,RleDataFrame-method (RleDataFrame-class), 25
- AtomicList, 26
- baf2mbaf, 3
- boundingIndices, 4, 5, 23
- boundingIndicesByChr, 4, 5, 23, 28
- bounds2Rle, 6, 23, 29–33
- calcGC, 6
- calcGC2, 7
- chr, 7
- chr,GenomicRanges-method (chr), 7
- chr,GenoSet-method (chr), 7
- chrIndices, 8
- chrIndices,GenoSetOrGenomicRanges-method (chrIndices), 8
- chrInfo, 9
- chrInfo,GenoSetOrGenomicRanges-method (chrInfo), 9
- chrNames, 9
- chrNames,GenomicRanges-method (chrNames), 9
- chrNames,GenoSet-method (chrNames), 9
- chrNames<- (chrNames), 9
- chrNames<-,GenomicRanges-method (chrNames), 9
- chrNames<-,GenoSet-method (chrNames), 9
- chrOrder, 10, 19, 34
- chrPartitioning, 11
- cn2lr, 11
- cn2lr,DataFrame-method (cn2lr), 11
- cn2lr,matrix-method (cn2lr), 11
- cn2lr,numeric-method (cn2lr), 11
- coerce,RleDataFrame,matrix-method (RleDataFrame-class), 25
- colMeans, 26
- colMeans,DataFrame-method (RleDataFrame-class), 25
- colMeans,RleDataFrame-method (RleDataFrame-class), 25
- colSums, 26
- colSums,RleDataFrame-method (RleDataFrame-class), 25
- DataFrame, 26
- fixSegNAs, 12
- gcCorrect, 12
- genome, 13
- genomeAxis, 13, 15
- genoPlot, 14, 14
- genoPlot,GenoSetOrGenomicRanges,ANY-method (genoPlot), 14
- genoPlot,numeric,numeric-method (genoPlot), 14
- genoPlot,numeric,Rle-method (genoPlot), 14
- genoPos, 15
- genoPos,GenoSetOrGenomicRanges-method (genoPos), 15
- GenoSet, 16, 18
- genoset (genoset-package), 3
- GenoSet-class, 17
- genoset-datasets, 18
- genoset-package, 3
- genoset.ds (genoset-datasets), 18
- GenoSetOrGenomicRanges-class (GenoSet-class), 17
- isGenomeOrder, 10, 18, 19, 34
- isGenomeOrder,GenoSet-method (isGenomeOrder), 18
- isGenomeOrder,GRanges-method (isGenomeOrder), 18