Package ‘graphite’

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Author  Gabriele Sales <gabriele.sales@unipd.it>, Enrica Calura
         <enrica.calura@gmail.com>, Chiara Romualdi
         <chiara.romualdi@unipd.it>
Maintainer Gabriele Sales <gabriele.sales@unipd.it>
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R topics documented:

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as.list.PathwayList

Conversion of PathwayLists into lists.

Description
Converting a PathwayList object into a list of Pathways.

Usage
```r
## S3 method for class 'PathwayList'
as.list(x, ...)
```

Arguments
- `x`: a PathwayList object
- `...`: extra arguments to as.list

Value
A list of pathways.

Author(s)
Gabriele Sales

See Also
PathwayList

Examples
```r
as.list(pathways("hsapiens", "kegg"))
```
buildPathway  

Build a Pathway object.

Description
This function creates a new object of type Pathway given a data frame describing its edges.

Usage
buildPathway(id, title, species, database, proteinEdges, metaboliteEdges = NULL, mixedEdges = NULL, timestamp = NULL)

Arguments
- id: the pathway identifier.
- title: the title of the pathway.
- species: the species the pathway belongs to.
- database: the name of the database the pathway derives from.
- proteinEdges: a data.frame of edges between proteins (or genes).
  Must have the following columns: src_type, src, dest_type, dest, direction and type.
  Direction must be one of the two strings: "directed" or "undirected".
- metaboliteEdges: interactions between metabolites.
  Can be NULL. Otherwise, it must have the same structure as proteinEdges.
- mixedEdges: interactions between metabolites and proteins.
  Can be NULL. Otherwise, it must have the same structure as proteinEdges.
- timestamp: when the pathway was annotated, by default the time buildPathway is called.

See Also
Pathway-class

Examples
```r
dat <- data.frame(src_type = "ENTREZID", src="672",
dest_type = "ENTREZID", dest="7157",
direction="undirected", type="binding")
pathway <- buildPathway("#1", "example", "hsapiens", "database", dat)
# Example with metabolites:
dat <- data.frame(src_type = "CHEBI", src="77750",
dest_type = "ENTREZID", dest="7157",
direction="undirected", type="binding")
mixed <- data.frame(src_type = "CHEBI", src="77750",
dest_type = "ENTREZID", dest="7157",
direction="undirected", type="binding")
pathway <- buildPathway("#1", "example", "hsapiens", "database",
dat, mixed)
```

**convertIdentifiers**  
*Convert the node identifiers of a pathway.*

**Description**

Converts the node identifiers of pathways.

If the option `Ncpus` is set to a value larger than 1 and the package `parallel` is installed, the conversion procedure will automatically use multiple cores.

**Usage**

```r
convertIdentifiers(x, to)
```

**Arguments**

- `x` can be a list of pathways or a single pathway
- `to` a string describing the type of the identifier. Can assume the values "entrez", "symbol" or the name of one of the columns provided by an Annotation package (for example, "UNIPROT").

**Value**

A Pathway object.

**See Also**

*Pathway*

**Examples**

```r
r <- pathways("hsapiens", "reactome")
convertIdentifiers(r$'mTOR signalling', "symbol")
```

---

**cytoscapePlot**  
*Plot a pathway graph in Cytoscape*

**Description**

Renders the topology of a pathway as a Cytoscape graph.

**Usage**

```r
cytoscapePlot(pathway, ..., cy.ver = 3)
```

**Arguments**

- `pathway` a Pathway object.
- `...` optional arguments forwarded to `pathwayGraph`.
- `cy.ver` select a Cytoscape version. Only version 3 is supported in this release.
Details

Requires the RCy3 package.

Value

An invisible list with two items:

- `graph`: the `graphNEL` object sent to Cytoscape.
- `suid`: the RCy3 network SUID.

See Also

- `Pathway`
- `pathwayGraph`

Examples

```r
## Not run:
r <- pathways()
cytoscapePlot(convertIdentifiers(reactome$"Unwinding of DNA", "symbol"))
## End(Not run)
```

Pathway-class

Class "Pathway"

Description

A biological pathway.

Variants

A Pathway instance actually stores multiple variants of the same biological data. This is the list of included variants:

- `proteins`: includes only interactions among proteins;
- `metabolites`: includes only interactions among metabolites;
- `mixed`: includes all available interactions.

Methods

- `pathwayId(p)`: Returns the native ID of the pathway.
- `pathwayTitle(p)`: Returns the title of the pathway.
- `pathwayDatabase(p)`: Returns the name of the database the pathway was derived from.
- `pathwaySpecies(p)`: Returns the name of the species in which the pathway was annotated.
- `pathwayTimestamp(p)`: Returns the date of pathway data retrieval.
- `pathwayURL(p)`: Returns the URL of the pathway in its original database, if available.
- `convertIdentifiers(p, to)`: Returns a new pathway using a different type of node identifiers.
edges(p, which = c("proteins", "metabolites", "mixed"), stringsAsFactors = TRUE):
  Returns a data.frame describing the edges of this pathway.
  The option which selects the desired pathway variant (see section "Variants" above).
  If stringsAsFactors is TRUE, strings are converted to factors.

nodes(p, which = c("proteins", "metabolites", "mixed")): Returns the names of the nodes
  belonging to this pathway.
  The option which selects the desired pathway variant (see section "Variants" above).

plot(p): Shows the pathway topology in Cytoscape.

runClipper(p, expr, classes, method, ...): Runs a clipper analysis over the pathway.
runTopologyGSA(p, test, exp1, exp2, alpha, ...): Runs a topologyGSA analysis over the pathway.

Author(s)
Gabriele Sales

See Also
pathways

pathwayDatabases

List the available pathway databases.

Description
Obtains the list of pathway databases available through graphite.

Usage
pathwayDatabases()

Value
Returns a data.frame with two columns: species and database.

Author(s)
Gabriele Sales

See Also
pathways

Examples
pathwayDatabases()
pathwayGraph

Graph representing the topology of a pathway

Description

Builds a graphNEL object representing the topology of a pathway.

Usage

pathwayGraph(pathway, which = "proteins", edge.types = NULL)

Arguments

pathway a Pathway object.
which the pathway variant you want.
See Pathway documentation for a list of the supported variants.
edge.types keep only the edges matching the type names in this vector.

Value

A graphNEL object.

See Also

Pathway

Examples

r <- pathways("hsapiens", "reactome")
pathwayGraph(r$"mTOR signalling", edge.types="Binding")

PathwayList-class

Class "PathwayList"

Description

A collection of pathways from a single database.

Extends

Class "Pathways", directly.
Methods

`l[i]`: Returns a selection of the pathways contained in the pathway list.
`l[[i]]`: Access one of the pathways contained in the pathway list.
`l$'title'`: Access one of the pathways by its title.
`convertIdentifiers(l, to)` Returns a new list of pathways using a different type of node identifiers.
`length(l)` Returns the number of pathways contained in the list.
`names(l)` Returns the titles of the pathways contained in the list.
`prepareSPIA(l, pathwaySetName, print.names=FALSE)` Prepares the pathways for a SPIA analysis.
`runClipper(l, expr, classes, method, maxNodes=150, ...)` Runs a clipper analysis over all the pathways in the list.
`runTopologyGSA(l, test, exp1, exp2, alpha, maxNodes=150, ...)` Runs a topologyGSA analysis over all the pathways in the list.

Author(s)

Gabriele Sales

See Also

`pathways`

---

pathways

Retrieve a list of pathways.

Description

Retrieve a list of pathways from a database for a given species.

graphite currently supports the following databases:

- BioCarta
- HumanCyc
- KEGG
- NCI-Nature Pathway Interaction Database
- PANTHER
- PharmGKB
- Reactome
- SMPDB

Call the `pathwayDatabase` function for more details.

Usage

`pathways(species, database)`
Arguments

species one of the supported species
database the name of the pathway database

Value

A PathwayList object.

See Also

PathwayList, pathwayDatabases

Examples

pathways("hsapiens", "reactome")
prepareSPIA

Prepare pathway dataset needed by runSPIA.

Description

Prepare pathway dataset needed by runSPIA. See runSPIA and spia for more details.

Usage

prepareSPIA(db, pathwaySetName, print.names = FALSE)

Arguments

db a PathwayList object or a list of Pathways.
pathwaySetName name of the output pathway set.
print.names print pathway names as the conversion advances.

References


See Also

runSPIA
spia
PathwayList

runClipper

Run a topological analysis on an expression dataset using clipper.

Description

clipper is a package for topological gene set analysis. It implements a two-step empirical approach based on the exploitation of graph decomposition into a junction tree to reconstruct the most relevant signal path. In the first step clipper selects significant pathways according to statistical tests on the means and the concentration matrices of the graphs derived from pathway topologies. Then, it "clips" the whole pathway identifying the signal paths having the greatest association with a specific phenotype.

If the option Ncpus is set to a value larger than 1 and the package parallel is installed, the conversion procedure will automatically use multiple cores.
Usage

runClipper(x, expr, classes, method, which = "proteins", seed = NULL, ...)

Arguments

- **x**: a `PathwayList`, a list of `Pathways` or a single `Pathway` object.
- **expr**: a matrix (size: number of genes x number of samples) of gene expression.
- **classes**: a vector (length: n) of class assignments.
- **method**: the kind of test to perform on the cliques. It could be either "mean" or "variance".
- **which**: the pathway variant you want. See `Pathway` documentation for a list of the supported variants.
- **seed**: if not NULL, set the seed for the random number generator used by clipper.
- **...**: additional options: see for details `easyClip`.

When invoked on a `PathwayList`, you can use the named option `maxNodes` to limit the analysis to those pathways with at most a given number of nodes.

Details

The expression data and the pathway have to be annotated in the same set of identifiers.

References


See Also

clipper

easyClip

easyClipper

easyClipper

Examples

```r
if (require(clipper) & require(ALL) & require(a4Preproc)) {
  data(ALL)
  pheno <- as(phenoData(ALL), "data.frame")
  samples <- unlist(lapply(c("NEG", "BCR/ABL"), function(t) {
    which(grepl("B\d\d", pheno$BT) & (pheno$mol.biol == t))[1:10]
  })))
  classes <- c(rep(1,10), rep(2,10))
  expr <- exprs(ALL)[,samples]
  rownames(expr) <- paste("ENTREZID", featureData(addGeneInfo(ALL))$ENTREZID,
    sep = ".")
  k <- as.list(pathways("hsapiens", "kegg"))
  selected <- k[c("Bladder cancer", "Hippo signaling pathway - multiple species")]
  runClipper(selected, expr, classes, "mean", pathThr = 0.1)
}
```
runSPIA

Description

Run a topological analysis on an expression dataset using SPIA.

Usage

```r
runSPIA(de, all, pathwaySetName, ...)
```

Arguments

- `de`: A named vector containing log2 fold-changes of the differentially expressed genes. The names of this numeric vector are Entrez gene IDs.
- `all`: A vector with the Entrez IDs in the reference set. If the data was obtained from a microarray experiment, this set will contain all genes present on the specific array used for the experiment. This vector should contain all names of the `de` argument.
- `pathwaySetName`: The name of a pathway set created with `prepareSPIA`.
- `...`: Additional options to pass to `spia`.

Details

The `spia` option "organism" is internally used. It is an error use it in the additional options.

Value

The same of `spia`, without KEGG links. A data frame containing the ranked pathways and various statistics: `pSize` is the number of genes on the pathway; `NDE` is the number of DE genes per pathway; `tA` is the observed total perturbation accumulation in the pathway; `pNDE` is the probability to observe at least `NDE` genes on the pathway using a hypergeometric model; `pPERT` is the probability to observe a total accumulation more extreme than `tA` only by chance; `pG` is the p-value obtained by combining `pNDE` and `pPERT`; `pGfdr` and `pGfwer` are the False Discovery Rate and respectively Bonferroni adjusted global p-values; and the `Status` gives the direction in which the pathway is perturbed (activated or inhibited).

References


See Also

`spia`
Examples

```r
if (require(SPIA) && require(hgu133plus2.db)) {
  data(colorectalCancer)

  top$ENTREZ <- mapIds(hgu133plus2.db, top$ID, "ENTREZID", "PROBEID", multiVals = "first")
  top <- top[!is.na(top$ENTREZ) & !duplicated(top$ENTREZ),]
  top$ENTREZ <- paste("ENTREZID", top$ENTREZ, sep = ":")
  tg1 <- top[top$adj.P.Val < 0.05,]
  DE_Colorectal = tg1$logFC
  names(DE_Colorectal) <- tg1$ENTREZ
  ALL_Colorectal <- top$ENTREZ

  biocarta <- pathways("hsapiens", "biocarta")[1:20]
  biocarta <- convertIdentifiers(biocarta, "ENTREZID")
  prepareSPIA(biocarta, "biocartaEx")
  runSPIA(de = DE_Colorectal, all = ALL_Colorectal, "biocartaEx")
}
```

---

runTopologyGSA

**Run a topological analysis on an expression dataset using topologyGSA.**

**Description**

Use graphical models to test the pathway components highlighting those involved in its deregulation.

If the option Ncpus is set to a value larger than 1 and the package parallel is installed, the conversion procedure will automatically use multiple cores.

**Usage**

`runTopologyGSA(x, test, exp1, exp2, alpha, ...)`

**Arguments**

- `x` a `PathwayList`, a list of `Pathways` or a single `Pathway` object.
- `test` Either "var" and "mean". Determine the type of test used by topologyGSA.
- `exp1` Experiment matrix of the first class, genes in columns.
- `exp2` Experiment matrix of the second class, genes in columns.
- `alpha` Significance level of the test.
- `...` Additional parameters forwarded to topologyGSA.

When invoked on a `PathwayList`, can use the named option "maxNodes" to limit the analysis to those pathways having up to this given number of nodes.

**Details**

This function produces a warning and returns NULL when the number of genes in common between the expression matrices and the pathway is less than 3.
References

Massa MS, Chiogna M, Romualdi C. Gene set analysis exploiting the topology of a pathway. BMC System Biol. 2010 Sep 1;4:121.

See Also

`pathway.var.test` `pathway.mean.test`

Examples

```r
if (require(topologyGSA)) {
  data(examples)
  colnames(y1) <- paste("SYMBOL", colnames(y1), sep = ";")
  colnames(y2) <- paste("SYMBOL", colnames(y2), sep = ";")

  k <- pathways("hsapiens", "kegg")
  p <- convertIdentifiers(k[["Fc epsilon RI signaling pathway"]], "SYMBOL")
  runTopologyGSA(p, "var", y1, y2, 0.05)
}
```
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