Package ‘mAPKL’

April 14, 2017

Type Package

Title A Hybrid Feature Selection method for gene expression data

Version 1.6.0

Date 2015-07-21

Author Argiris Sakellariou

Maintainer Argiris Sakellariou <argisake@gmail.com>

Depends R (>= 3.2.0), Biobase

Imports multtest, clusterSim, apcluster, limma, AnnotationDbi, methods, parmigene.igraph.reactome.db

Suggests BiocStyle, knitr, mAPKLData, hgu133plus2.db, RUnit, BiocGenerics

VignetteBuilder knitr

Description We propose a hybrid FS method (mAP-KL), which combines multiple hypothesis testing and affinity propagation (AP)-clustering algorithm along with the Krzanowski & Lai cluster quality index, to select a small yet informative subset of genes.

License GPL (>= 2)

biocViews FeatureExtraction, DifferentialExpression, Microarray, GeneExpression

NeedsCompilation no

R topics documented:

mAPKL-package ........................................... 2
Annot-class ............................................ 2
annotate ............................................... 3
classification ......................................... 4
Classify-class ......................................... 5
DataLD-class .......................................... 6
loadFiles ............................................. 6
mAPKL .................................................. 7
mAPKLRes-class ....................................... 8
metrics ............................................... 9
NetAttr-class ......................................... 10
netwAttr ............................................. 10
preprocess ........................................... 11
Description

A hybrid FS method (mAP-KL), which combines multiple hypothesis testing and affinity propagation (AP) clustering algorithm along with the Krzanowski & Lai cluster quality index, to select an informative subset of genes.

Details

Package: mAPKL
Type: Package
Version: 0.99.01
Date: 2014-05-07
License: GPL (>= 2)

Author(s)

Argiris Sakellariou <a.sakellariou@gonkhosp.gr>

References


Annot-class

Description

S4 class for storing Annot analysis results

Slots

The following slots are defined for Annot objects:

- **results**: The accumulated annotation results
- **probe**: The probe id
- **symbol**: The official gene symbol
**Entrez gene identifier**

- **entrezId**: The Entrez gene Identifier
- **ensemblId**: The ensembl ID as indicated by ensembl
- **map**: The cytoband locations of the gene

**Author(s)**

Argiris Sakellariou <a.sakellariou@gonkhosp.gr>

---

**annotate**

*Genome annotation of the "exemplars".*

**Description**

The user may extract several genome specific information for the "exemplars" as described in the microarray annotation file.

**Usage**

```r
annotate(exemplars,chip)
```

**Arguments**

- **exemplars**: The "exemplars" of the mAPKL class.
- **chip**: The platforms’s name of the microarray chip used (e.g. "hgu133plus2.db")

**Details**

This function uses as key the probe id and returns the matching information as described in the gene chip annotation file. The returned information are usually multiple to the number of probe ids (one to many relationship).

**Value**

- **results**: The accumulated annotation results.
- **probe**: The probe id.
- **symbol**: The official gene symbol.
- **entrezId**: The Entrez gene Identifier.
- **ensemblId**: The ensembl ID as indicated by ensembl.
- **map**: The cytoband locations of the gene.

**Author(s)**

Argiris Sakellariou
Examples

```r
## We use the "exemplars" from the mAPKL.Rd example

exemplrs <- c(24, 26, 42, 45, 63, 81, 95, 99, 102, 113, 134, 135, 145, 152, 168)
names(exemplrs)<- c("215717_s_at", "1561358_a_at", "222752_s_at", "233922_at", "218871_x_at", "33323_r_at", "244311_at", "228932_at", "205088_at", "209566_at", "215180_at", "1560638_a_at", "201852_x_at", "229947_at", "221731_x_at")

gene.info <- annotate(exemplrs,"hgu133plus2.db")
```

Classification

Classify samples according to the SVM algorithm

Description

This function performs classification through the Support Vector Machines (SVM) algorithm. The algorithm applies on the "exemplars" dataset. It produces a classification result either on the training set or on a validation set. This function estimates how well the selected "genes" from mAP-KL method discriminate between two phenotypes. The default SVM settings are: "linear" kernel and 5-folds cross-validation. Regarding the parameters for the "linear" kernel, cost parameter, and for the "radial" kernel, cost and gamma parameters, are estimated automatically through the tune.svm function as described in e1071 r-package.

Usage

```r
classification(trExemplObj,classLabels,valExemplObj=NULL,kf=5,kernel="linear")
```

Arguments

- `trExemplObj`: The exemplars train eSet object.
- `classLabels`: The varLabels name in the eSet object where the class labels are stored e.g. "type".
- `valExemplObj`: The exemplars validation eSet object (if not NULL).
- `kf`: The k-folds value of the cross-validation parameter. The default value is 5-folds. By setting "Loo" or "LOO" a Leave-One-Out Cross Validation is performed.
- `kernel`: The type of kernel used for the classification analysis. The default kernel is "linear".

Value

- `classL`: The labels of the train set
- `valClassL`: The labels of the validation set if not NULL
- `predLbls`: The predicted labels according to the classification analysis

Author

Argiris Sakellariou
Classify-class

Examples

```r
library(mAPKLData)
data(mAPKLData)
breast <- sampling(Data=mAPKLData, valPercent=40, classLabels="type", seed=135)
normTrainData <- preprocess(breast$trainData)
normTestData <- preprocess(breast$testData)

exprs(breast$trainData)<-normTrainData$clL2.normdata
exprs(breast$testData)<-normTestData$clL2.normdata

out.clL2 <- mAPKL(trObj=breast$trainData, classLabels="type", valObj=breast$testData,dataType=7)
clasPred <- classification(trExemplObj=out.clL2@exemplTrain, classLabels="type", valExemplObj=out.clL2@exemplTest)
```

---

Classify-class

Class "Classify"

Description

S4 class for storing Classify analysis results

Slots

The following slots are defined for Classify objects:

- `classL`: Number of samples in the data set
- `valClassL`: Subset of samples used for leveraged clustering
- `predLbls`: Vector containing indices of exemplars
- `AUC`: The Area Under the ROC curve as a degree of samples discrimination
- `Accuracy`: The classification accuracy
- `MCC`: The MCC classification measure
- `Specificity`: The degree of true negative’s identification
- `Sensitivity`: The degree of true positive’s identification

Author(s)

Argiris Sakellariou <a.sakellariou@gonkhosp.gr>
DataLD-class

Description
S4 class for storing DataLD analysis results

Slots
The following slots are defined for DataLD objects:

  trainObj: An eSet class object of a training set
  valObj: An eSet class object of a validation set

Author(s)
Argiris Sakellariou <a.sakellariou@gonkhosp.gr>

loadFiles

Description
This function loads the train set, the class labels files as well as the test or validation file if any. Then we may perform normalization and (or) log2 transformation.

Usage
loadFiles(filesPath, trainFile, labelsFile, validationFile=NULL, validationLabels=NULL)

Arguments
filesPath The path where the files are stored
trainFile The genes and the relevant intensity values for feature selection analysis. The file should be of tab-delimited format
labelsFile The class labels of the samples
validationFile A further file with genes and intensity values used for validation purposes
validationLabels The class labels of the validation samples

Value
An object of Class DataLD

Author(s)
Argiris Sakellariou
The \textit{mAP-KL} algorithm

\textbf{Description}

We first employ the \texttt{mt.maxT} function from the "multtest" package to rank the genes of the training set and then we reserve the top \(N\) genes e.g. (\(N = 200\)) for further exploitation. Prior to clustering analysis with Affinity Propagation we apply the index of Krzanowski and Lai as included in the "ClusterSim" package to determine the number of clusters solely on the disease samples of the training test set. The final step involves the cluster analysis with the AP clustering method as in the "apcluster" package, which detects \(n\) (\(n = k\), the Krzanowski and Lai index) clusters among the top \(N\) genes and provides us with a list of the most representative genes of each cluster, the so called exemplars.

\textbf{Usage}

\texttt{mAPKL(trObj, classLabels, valObj=NULL, dataType=6, statTest="t", permutations=1000, features=200, minClusters=2, maxClusters=50, FC="limma", bimaxit=50, r=2)}

\textbf{Arguments}

\begin{itemize}
  \item \texttt{trObj} The train eSet object.
  \item \texttt{classLabels} The \texttt{varLabels} name in the eSet object where the class labels are stored e.g "type".
  \item \texttt{valObj} The validation eSet object (if not NULL).
  \item \texttt{dataType} The type of the data e.g 6-ratio data without normalization and 7-interval or mixed (ratio & interval) data without normalization as described in "clusterSim" package.
  \item \texttt{statTest} The statistical test applied to the geneIntensities. The available tests described in \texttt{mt.maxT} documentation in "multtest" package.
  \item \texttt{permutations} The number of permutations.
  \item \texttt{features} The top \(N\) genes to be kept.
  \item \texttt{minClusters} The minimum number of clusters that can be identified.
  \item \texttt{maxClusters} The maximum number of clusters that can be identified.
  \item \texttt{FC} The Fold Change of the exemplars according to "Limma" (default). Alternatively the "SAM" approach may be computed.
  \item \texttt{bimaxit} The maximum number of bisection steps performed by the AP algorithm. The (default) value is "50".
  \item \texttt{r} The argument \(r\) is used to transform the resulting distances by computing the \(r\)-th power. To obtain negative squared distances as in Frey’s and Dueck’s (use \(r=2\) as default).
\end{itemize}

\textbf{Value}

\begin{itemize}
  \item \texttt{rankedIntens} The top \(N\) ranked genes with their intensity values
  \item \texttt{exemplTrain} The intensity values of the exemplars in the training set
  \item \texttt{exemplTest} The intensity values of the exemplars in the validation set if not NULL
\end{itemize}
mAPKLRes-class

A list with the overall results of the "mt.maxT" analysis

The adjusted p-values according to the statistical analysis

The raw p-values according to the statistical analysis

The Fold Change of the exemplars

The selected "significant" probe ids/genes

The probe ids/genes per cluster

Author(s)

Argiris Sakellariou

References


Examples

## Using separate train-test samples
## Load the necessary files based on Breast cancer data as included in the
## package mAPKLData

library(mAPKLData)
data(mAPKLData)
breast <- sampling(Data=mAPKLData, valPercent=40, classLabels="type", seed=135)
normTrainData <- preprocess(breast$trainData)
normTestData <- preprocess(breast$testData)

exprs(breast$trainData) <- normTrainData$clL2.normdata
exprs(breast$testData) <- normTestData$clL2.normdata

out.clL2 <- mAPKL(trObj=breast$trainData, classLabels="type",
valObj=breast$testData, dataType=7)

mAPKLRes-class

Class “mAPKLRes”

Description

S4 class for storing mAPKL analysis results

Slots

The following slots are defined for mAPKLRes objects:

- rankedIntens: The top N ranked genes along with their intensity values
- exemplTrain: An eSet class object formed with the exemplars of the training set
- exemplTest: An eSet class object formed with the exemplars of the validation set if not NULL
metrics

statistic: A list with the overall results of the "mt.maxT" analysis
adjp: The adjusted p-values according to the statistical analysis
pVal: The raw p-values according to the statistical analysis
fc: The Fold Change of the exemplars
exemplars: The selected "significant" probe ids/genes
clusters: The probe ids/genes per cluster

Author(s)
Argiris Sakellariou <a.sakellariou@gonkhosp.gr>

Description
This function calculates several classification related metrics. It uses the original and the predicted samples' labels to quantify the quality of the classification process. Those measures give us a direct outlook of the selected "genes" and how well discriminate between two phenotypes.

Usage
metrics(classLbls, predLbls)

Arguments

  classLbls The initial class labels.
  predLbls The predicted class labels.

Value

  AUC The Area Under the ROC curve as a degree of samples discrimination
  Accuracy The classification accuracy
  MCC The MCC classification measure
  Specificity The degree of true negative's identification
  Sensitivity The degree of true positive's identification

Author(s)
Argiris Sakellariou

Examples

  ## Suppose 'val' represent the correct validation set labels
  ## and 'predictions' the predicted labels according to an SVM model

  val <- c(rep(0,8),rep(1,4))
predictions <- c(rep(0,6),1,1,rep(1,3),0)
  perfMetrics <- metrics(classLbls=val, predLbls=predictions)
NetAttr-class  

Class "NetAttr"

Description

S4 class for storing some network characteristics of the top ranked genes

Slots

The following slots are defined for NetAttr objects:

- edgelist: The Node1 & Node2 & Weight edgelist matrix
- degree: The Local, Global and their Weighted values of the "degree" characteristic
- closeness: The Weighted values of the Local and Global "closeness" characteristic
- betweenness: The Weighted values of the Local and Global "betweenness" characteristic
- transitivity: The Weighted values of the Local and Global "transitivity" characteristic

Author(s)

Argiris Sakellariou <a.sakellariou@gonkhosp.gr>

netwAttr  

Calculates network characteristics

Description

Calculate some basic network characteristics of the top ranked genes

Usage

netwAttr(mAPKLObj, net="clr")

Arguments

- mAPKLObj: An object of mAPKL class.
- net: The network reconstruction method to be employed. The user may select between "clr" (default), "aracne.a" and "aracne.m".

Details

It calculates some basic network characteristics. Those include the "degree", the "closeness", the "betweenness", and finally the "transitivity" or else clustering coefficient. We calculate the weighted values for both local and global scores.

The three available network reconstruction options are:
- clr: Context Likelihood or Relatedness Network
- aracne.a: Algorithm for the Reconstruction of Accurate Cellular Networks (additive model)
- aracne.m: Algorithm for the Reconstruction of Accurate Cellular Networks (multiplicative model)
**preprocess**

**Value**

Upon successful completion, the function returns an **NetAttr** object.

**Author(s)**

Argiris Sakellariou

**Examples**

```r
library(mAPKLDdata)
data(mAPKLDdata)
breast <- sampling(Data=mAPKLDdata, valPercent=40, classLabels="type", seed=135)
normTrainData <- preprocess(breast$trainData)
normTestData <- preprocess(breast$testData)

exprs(breast$trainData) <- normTrainData$clL2.normdata
eexprs(breast$testData) <- normTestData$clL2.normdata

out.clL2 <- mAPKL(trObj=breast$trainData, classLabels="type", valObj=breast$testData, dataType=7)

net.attr <- netwAttr(mAPKLObj=out.clL2)
```

---

**preprocess**  
*Performs normalization and/or log2 transformation*

**Description**

This function performs normalization and/or log2 transformation on gene expression data.

**Usage**

```r
preprocess(exprsObj, log2=TRUE, norm="ALL", destname=NULL)
```

**Arguments**

- `exprsObj`  
  An eSet object where its assay data will be normalized
- `log2`  
  Performs logarithmic transformation of base 2 prior to any normalization. The default value is TRUE
- `norm`  
  The user may define a specific normalization method rather than "ALL" which is the default case. The available abbreviations are described in the details section
- `destname`  
  Here we define the destination path and the name of the jpeg file with the density plots. The default path is the working directory
Details

The available normalization methods are:
- Mean-centering normalization "mc"
- z-score normalization "z"
- Quantile normalization "q"
- Cyclic loess normalization "cl"
- Mean-centering normalization and log2 transformation "mcL2"
- z-score normalization and log2 transformation "zL2"
- Quantile normalization and log2 transformation "qL2"
- Cyclic loess normalization and log2 transformation "clL2"

Value

<table>
<thead>
<tr>
<th>Raw data</th>
<th>The initial gene expression values</th>
</tr>
</thead>
<tbody>
<tr>
<td>mc.normdata</td>
<td>The values after 'mean-centering' normalization</td>
</tr>
<tr>
<td>z.normdata</td>
<td>The values after 'z-score' normalization</td>
</tr>
<tr>
<td>q.normdata</td>
<td>The values after 'quantile' normalization</td>
</tr>
<tr>
<td>cl.normdata</td>
<td>The values after 'cyclic loess' normalization</td>
</tr>
<tr>
<td>mcL2.normdata</td>
<td>The values after 'mean-centering' normalization and log2</td>
</tr>
<tr>
<td>zL2.normdata</td>
<td>The values after 'z-score' normalization and log2</td>
</tr>
<tr>
<td>qL2.normdata</td>
<td>The values after 'quantile' normalization and log2</td>
</tr>
<tr>
<td>clL2.normdata</td>
<td>The values after 'cyclic loess' normalization and log2</td>
</tr>
</tbody>
</table>

Author(s)

Argiris Sakellariou

Examples

```r
library(mAPKLData)
data(mAPKLData)
varLabels(mAPKLData)
breast <- sampling(Data=mAPKLData, valPercent=40, classLabels="type", seed=135)
normTrainData <- preprocess(exprsObj=breast$trainData)
```

---

probes2pathways

**Extract pathways from "exemplars"**

Description

The user may extract the pathways where the "exemplars" are involved with the aid of the reactome database.

Usage

```r
probes2pathways(annotObj)
```
**Arguments**

annotObj  
The "Annot" class object.

**Details**

This function utilizes the "Annot" class object as returned by the "annotate" function to extract the pathways where the "exemplars" are involved with the aid of the "reactome" database. We employ the probe Ids for the matching.

**Author(s)**

Argiris Sakellariou

**Examples**

```r
## We use the "exemplars" from the mAPKL.Rd example
exemplrs <- c(24, 26, 42, 45, 63, 81, 95, 99, 102, 113, 134, 135, 145, 152, 168)
names(exemplrs)<- c("215717_s_at", "1561358_at", "222752_s_at", "233922_at",
"218871_x_at", "33323_r_at", "244311_at", "220932_at", "205508_at", "209596_at",
"215180_at", "1560638_s_at", "201852_x_at", "229947_at", "221731_x_at")
gene.info <- annotate(exemplrs,"hgu133plus2.db")
## We now use the "gene.info" to identify the relevant pathways
probes2pathways(gene.info)
```

**Description**

This function gathers the results of several analysis sessions (feature selection, classification, annotation and network) and produces a report in HTML format.

**Usage**

```r
report(mAPKLObj, ClassifyObj, AnnotObj=NULL, netObj=NULL, file)
```

**Arguments**

mAPKLObj  
An object of mAPKL class.

ClassifyObj  
An object of mAPKL class.

AnnotObj  
An object of Annot class.

netObj  
An object of NetAttr class.

file  
The full path and the name of the produced report
It presents the data samples with their phenotype labels, the exemplars with their statistical scores (adj.p-value, p-value and fc), and network characteristics (like weighted local degree, closeness, betweenness, transitivity) if such a network analysis has been performed. In addition, the report presents the classification performance achieved by those exemplars (either cross-validation or hold-out), and finally several annotation information (symbol, entrez id, ensemble id, and their chromosomal location) if an annotation analysis has been carried out.

Value

Upon successful completion an HTML report is saved in the working directory.

Author(s)

Argiris Sakellariou

Examples

```r
## When a network attributes object is present
## Not run: report(out,class.pred,class.metrics,netObj=net.attr, "C:.../.../mAPKreport.html")#Define a local path to store your report
## End(Not run)

## When an annotation object is present
## Not run: report(out,class.pred,class.metrics,gene.info, "C:.../.../mAPKreport.html")#Define a local path to store your report
## End(Not run)

## Without annotation and network attributes objects
## Not run: report(out,class.pred,class.metrics, file="C:.../.../mAPKreport.html")#Define a local path to store your report
## End(Not run)
```

Description

This function takes as input a dataset and splits it into a train and a test set based to a user defined percentage.

Usage

```r
sampling(Data,valPercent,classLabels,seed)
```

Arguments

- **Data**: The input dataset to be split as an eSet object.
- **valPercent**: The percentage of the input dataset used for validation purposes e.g. 40.
- **classLabels**: The varLabels name in the eSet object where the class labels are stored e.g "type".
- **seed**: Setting the seed number for reproducible sampling. The default value is 1.
Value

- **trainData** The data used for training as an eSet object
- **testData** The data used for validation as an eSet object

Author(s)

Argiris Sakellariou

Examples

```r
library(mAPKLData)
data(mAPKLData)
breast <- sampling(Data=mAPKLData, valPercent=40, classLabels="type", seed=135)
```
Index

*Topic **IO**
  loadFiles, 6
*Topic **classes**
  Annot-class, 2
  Classify-class, 5
  DataLD-class, 6
  mAPKLRRes-class, 8
  NetAttr-class, 10
*Topic **classif**
  classification, 4
*Topic **datagen**
  sampling, 14
*Topic **htest**
  mAPKL, 7
  mAPKL-package, 2
*Topic **methods**
  annotate, 3
  metrics, 9
  netwAttr, 10
  preprocess, 11
  probes2pathways, 12
  report, 13

Annot, 2
Annot (Annot-class), 2
Annot-class, 2
annotate, 3

classification, 4
Classify, 5
Classify (Classify-class), 5
Classify-class, 5

DataLD, 6
DataLD (DataLD-class), 6
DataLD-class, 6

loadFiles, 6
mAPKL, 7
mAPKL-package, 2
mAPKLRRes, 8
mAPKLRRes (mAPKLRRes-class), 8
mAPKLRRes-class, 8
metrics, 9