Package ‘metaMS’

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**Description** MS-based metabolomics data processing and compound annotation pipeline.

**License** GPL (>= 2)

**biocViews** MassSpectrometry, Metabolomics

**Author** Ron Wehrens [aut, cre] (author of GC-MS part), Pietro Franceschi [aut] (author of LC-MS part), Nir Shahaf [ctb], Matthias Scholz [ctb], Georg Weingart [ctb] (development of GC-MS approach), Elisabete Carvalho [ctb] (testing and feedback of GC-MS pipeline)

**Maintainer** Ron Wehrens <ron.wehrens@gmail.com>

**NeedsCompilation** no

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metaMS-package

Analysis pipeline for MS-based metabolomics data

Description

Analysis pipeline for MS-based metabolomics data: basic peak picking and grouping is done using functions from packages xcms and CAMERA. The main output is a table of feature intensities in all samples, which can immediately be analysed with multivariate methods. The package supports the creation of in-house databases of mass spectra (including retention information) of pure chemical compounds. Such databases can then be used for annotation purposes.

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addRI

Description

Given an msp object and the retention times and indices of a series of reference compounds, the function adds an RI field to every entry of the msp object. This will only be done if there is not already an RI field: existing information will not be overwritten.

Usage

addRI(mspobj, RIstandards, isMSP = TRUE)
alignmentLC

Arguments

mspobj An msp object.
RIstandards A two-column matrix containing for the standards defining the RI scale both retention times and retention indices.
isMSP Logical: if TRUE, then the spectra are stored in slot pspectrum; otherwise the spectra are simply the list elements of DB - matrices with three columns.

Value

An msp object, now also containing an RI slot.

Note

If the retention time of a compound is outside the range of the RI standards, NA will be used as RI value.

Author(s)

Ron Wehrens

Examples

if (require(metaMSdata)) {
  manual.fname <- list.files(system.file("extdata", package = "metaMSdata"),
                              pattern = "msp", full.names = TRUE)
  manual <- read.msp(manual.fname)
  RIstandards <- cbind("rt" = c(1.54, 1.68, 1.99, 2.7, 4.36, 6.81, 9.43,
                            11.88, 14.17, 16.34, 18.39, 20.33, 22.18,
                            23.93, 25.5, 27.18, 28.72, 30.26, 31.75,
                            33.19, 34.58, 35.95),
                "RI" = (6:27)*100)
  manualRI <- addRI(manual, RIstandards)
}

alignmentLC

LC alignment

Description

Performs grouping and retention time correction via xcms. The settings are specified as a list with a form similar to the one discussed in the help of FEMsettings. The sequence of actions depends on the type of retention time correction, which is specified inside the Retcor list inside the Alignment slot (see details). Integration of areas of missing peaks is performed depending on the fillPeaks setting.

Usage

alignmentLC(xset, settings)
Arguments

xset The xcmsSet object resulting from peak detection
settings The subset of settings contained into the "Alignment" element of the XCMSsettings list.

Details

The sequence of actions depends on the algorithm used by xcms for retention time correction. For the density-based approach the sequence is: 1) across sample grouping, 2) retention time correction, 3) second across sample grouping, 4) optional fill-peaks. For "obiwarp", instead, the sequence of actions is: 1) retention time correction, 2) across sample grouping, 3) optional fill-peaks. For across-sample grouping in xcms, the minsamp parameter can be specified in the settings either as a minimum number of samples (min.class.size) or as the fraction of samples per class (min.class.fraction). If both are given the smallest number is used. If the retention time correction is performed by the density-based approach, the settings allow to express the xcms parameters missing and extra as fractions. When "obiwarp" is used for retention time correction, the sample with the bigger number of features is automatically selected as the "reference" sample. The xcms parameters for the retcor xcms function can be specified in the Retcor list included in the Alignment slot of FEMsettings.

Value

A grouped and rt-aligned xcmsSet object.

Author(s)

Ron Wehrens and Pietro Franceschi

See Also

FEMsettings

AnnotateTable Annotate a Peaktable

Description

Functions which annotate a peaktable on the bases af a database of standards. Not meant to be called directly by the user.

Usage

## Annotate one feature
AnnotateFeature(input, DB, settings, errf)

## Annotate a full table of features
AnnotateTable(peaktable, errf, DB, settings)
Arguments

input A vector with three elements in the form \((mz, rt, I)\).

peaktable A peaktable (matrix) with three columns corresponding to \(mz, rt\) and \(I\) values for a series of features.

errf The file containing the error function used to predict the tolerance on the \(m/z\) value used for the matching against the DB.

DB A dataframe used for the annotation. See the help of LCDBtest for a description of the DB.

settings The subset of settings contained into the "match2DB" element of the XCMSsettings list.

Details

The annotation of each feature is performed by comparing its \(m/z\) value and its retention time to a database provided by the user. To account for shifts in retention time and mass occurring during data acquisition, the matching of a specific feature against the DB is done with a specific tolerance in mass and in retention time.

**Retention time tolerance**
The retention time tolerance is specified (in minutes) in the settings list (field \(rttol\)). This value is instrument- and chromatography-dependent.

**\(m/z\) tolerance**
The tolerance on the mass scale mainly depends on the characteristics of the spectrometer used for the acquisition. For Q-TOF instruments it has been recently shown (see references) that the optimal mass tolerance can be expressed as a function of the \(m/z\) value and of the logarithm of the ion intensity \(\log_{10}(I)\). As a trend, the mass drift will be bigger for smaller ions and for low intensity signals.

In the present implementation the tolerance in mass can be either fixed over the complete mass range or calculated as a function of the \(mz\) and \(I\) values of each feature. In the simplest case, the fixed mass tolerance is provided in the \(mzwindow\) (in Dalton!) element of the list of settings. Alternatively, one can provide (supplying the \(errf\) argument) a function used to calculate the \(mz\) tolerance (in ppm!) as a function of the fields of the input vector \((mz, rt, I)\).

As discussed in the publication, for a Waters Synapt Q-TOF the function is a linear model taking as inputs \(M = input["mz"]\), \(\log I = \log_{10}(input["I"]\). This error function can be calculated by analyzing the results of the injections of the chemical standards. To avoid unreasonable small errors where data for \(mz\) and \(I\) are not available, the minimum value for the mass tolerance is explicitly set in the settings (ppm). This value should match the technical characteristics of the spectrometer.

To reduce the number of false positives and make the annotation more reliable, a match is retained only if more than one feature associated to a specific compound is found in the list of features. How many "validation" features are required is defined in the list of settings in the \(minfeat\) element. At this validation level, another retention time tolerance is introduced: two or more features validate one specific annotation if their retention time are not very much different. This \(rt\) tolerance is also defined in the settings (the \(rtval\) field). As a general suggestion, \(rtval\) should be kept smaller than \(rttol\). The latter, indeed, refers to the matching of a peaktable with a database which has been created from the injections of the chemical standards during different instrumental runs (maybe also with different columns). On the other hand, \(rtval\) accounts for smaller retention time shifts, occurring within the same LC run.

For the description of the structure of the DB, refer to the help of the LCDBtest dataset.
Value

A list with the following elements

- **annotation.table**
  - A data.frame with the results of the annotation and the reference to the DB
- **compounds**
  - The names of the annotated compounds
- **IDs**
  - The IDs of the annotated compounds
- **multiple.annotations**
  - The features with multiple annotations
- **ann.features**
  - The features with annotation

Author(s)

Pietro Franceschi

References


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**annotations2tab**  
*Conversion of a list of annotation results into a table*

**Description**

Function `annotations2tab` converts the output of `matchSamples2DB` into a table, in the case of multiple DB hits for one pseudospectrum distinguishing the “best” hit from the “alternative” hits. Function `makeAnnotation` prepares a data.frame object with the correct number of rows. Both functions are not meant to be called directly by the user.

**Usage**

```r
annotations2tab(annlist, matches)
makeAnnotation(n)
```

**Arguments**

- **annlist**  
  - Annotation list, output of `match Samples2DB`.
- **matches**  
  - Object containing all match factors - needed to distinguish the best match from the rest in the case of a double annotation.
- **n**  
  - Number of rows in the annotation data.frame (possibly zero).

**Value**

A three-column data.frame, containing the numbers of the pseudospectrum in the experimental data, the numbers of the best hits in the DB, and finally the alternative hits in the DB

Author(s)

Ron Wehrens
**constructExpPseudoSpectra**

Create a list of all pseudospectra found in a GC-MS experiment of several samples.

**Description**

Function `constructExpPseudoSpectra` creates an msp object containing all the patterns referenced to in the annotation. The first argument is the output of function `matchSamples2Samples` and contains the full annotation matrix and the pseudospectra of the known unknowns; the second is the msp object containing the standards that are actually found. Not meant to be called directly by the user.

**Usage**

`constructExpPseudoSpectra(allMatches, standardsDB)`

**Arguments**

- `allMatches`: Result of a call to `matchSamples2DB` or `matchSamples2Samples`: a list containing the element `annotation`.
- `standardsDB`: Database of standard spectra, in msp format.

**Value**

A list of spectra. All spectra from the database that are referenced to in the annotation slot will be present, and will be followed by all unknown spectra. In order to be able to see which patterns in the experimental data are pointing to which spectra, an extra slot `DB.id` is added, containing the position of the reference spectrum in the original database (which is what the numbers of the annotation object are referring to). Renumbering this is impractical since some patterns may be seen as alternative annotations, but may have no hits of their own - including them would lead to rows containing only zeros.

**Author(s)**

Ron Wehrens

**See Also**

`matchSamples2DB`, `matchSamples2Samples`

---

**createSTDdbGC**

Create an in-house database for GC-MS annotation

**Description**

For creating an in-house instrument-specific annotation database, injections of pure standards need to be processed. All patterns in the vicinity of the retention time of the standard (to be provided by the user) will be compared to an external database - in case of a sufficient match, they will be retained in the database. The `generateStdDBGC` is not meant to be called directly by the user.
**createSTDdbGC**

**Usage**
```
createSTDdbGC(stdInfo, settings, extDB = NULL, manualDB = NULL, RIstandards = NULL, nSlaves = 0)
generateStdDBGC(totalXset, settings, extDB = NULL, manualDB = NULL, RIstandards = NULL)
```

**Arguments**
- **stdInfo**: Information of the standards, given in the form of a data.frame. Minimal information: `stdFile`, `Name`, `CAS`, monoisotopic mass (`monoMW`), and retention time (`rt`). The filenames in slot `stdFile` should include path information. If this argument is NULL, this function can be used to process a manually curated DB. Arguments `stdInfo` and `manualDB` cannot be both NULL.
- **settings**: A list of settings, to be used in peak picking and pattern comparison.
- **extDB**: The external database containing spectra, with which to compare the patterns found in the standards files.
- **manualDB**: A database of manually curated spectra, that will be incorporated in the final DB without any further checks.
- **totalXset**: A list of xset objects, as generated by `peakDetection`.
- **RIstandards**: A two-column matrix containing for the standards defining the RI scale both retention times and retention indices. If not given, no RI values will be calculated and retention times will be used instead.
- **nSlaves**: Number of cores to be used in peak picking.

**Details**
Function `createSTDdbGC` creates a database object containing validated pseudospectra for a number of compounds. The injections of the standards, described in the input object `stdInfo`, are processed using function `processStandards`; comparison with the external database, inclusion of manual compounds and final formatting are done in function `generateStdDBGC`. Several situations can be envisaged:

A: a series of injections of standards needs to be compared with a standard library, such as the NIST. In this case, both `stdInfo` and `extDB` need to be non-null, and the result will be a database in which the entries have a sufficient match with the external DB. If `manualDB` is also non-null, these entries will be added too (without checking).

B: for a series of injections no standard library information is available (`extDB` is NULL, and `stdInfo` is not), and the function simply returns all patterns eluting around the indicated retention time. This allows for subsequent manual validation and pruning. If `manualDB` is non-null, these entries will be added, but since this is a somewhat unusual thing to do, a warning will be given.

C: a manual database needs to be processed to be useable as a real database. This basically entails renaming the `rt` and `rt.sd` fields into `std.rt` and `std.rt.sd`, and a similar action for any RI field.

**Value**
The output of `createSTDdbGC` (and `generateStdDBGC`, which is the last function called in `createSTDdbGC`) is a list, where every entry describes one compound/spectrum combination. For use in annotation, the following fields are mandatory: `Name`, `std.rt`, `pspectrum` and `monoMW`. 
createSTDdbLC

Create an in-house database for LC-MS annotation

Description

For creating an in-house instrument-specific annotation database, injections of pure standards need to be processed. For each standard the analyst provides a validated reference value for retention time and m/z, generally corresponding to the major ionic signal for this compound. On the bases of this data, the database is constructed by automatically extracting the features identified in the vicinity of the retention time of the standard.

The function `generateSTDdbLC` is not meant to be called directly - use `createSTDdbLC` instead.
**createSTDdbLC**

**Usage**

```r
createSTDdbLC(stdInfo, settings, polarity, Ithr = 10, nSlaves = 0)
generateStdDBLC(stdxsets, settings, Ithr)
```

**Arguments**

- `stdInfo` Information of the standards, given in the form of a data.frame. Minimal information: `stdFile`, `compound`, `ChemSpiderID`, `observed m/z (mz.observed)`, and retention time in minutes (`RTman`). The filenames in slot `stdFile` should include path information.
- `settings` A list of settings, to be used in peak picking and pattern comparison (see details).
- `polarity` The polarity of the injection: "positive" or "negative"
- `Ithr` The intensity threshold used to decide whether or not a feature should be included in the DB. Typically acting on the `maxo` value.
- `stdxsets` A list of CAMERA objects resulting from the analysis (performed by `processStandards`) of the standard injections listed in the `stdInfo` table.
- `nSlaves` Number of cores to be used in peak picking.

**Details**

The DB is created with the following workflow. Peak picking is performed on each standard file by using the settings specified in the settings list. CAMERA is used to group together the different features by considering their retention time and the correlation among the extracted ion traces. The list of features is searched looking for the values for `mz` and `Rt` included in the `stdInfo` table (see the help of `exptable` for more details), with the mass and retention time tolerances specified in the "DBconstruction" element of the settings list. In presence of positive match for the feature `f`, a spectral fingerprint is constructed by using all the features with an intensity bigger than `Ithr` which are in the same pcgroup of `f`. A match is retained only if the spectral fingerprint is composed of more than `minfeat` elements. This parameter is also included in the list of settings.

**Value**

A list with three elements.

- `RefTable` the original table used for the creation of the DB.
- `Info` a list with the settings used for the DB generation and the date.
- `DB` the DB which can be used in `runLC` for annotation.

**Author(s)**

Pietro Franceschi

**Examples**

```r
if (require(metaMSdata)) {
  ## load the manually curated table for the standards
  data(exptable)
  ## add location of cdf file from which the standards DB is going to be
  ## built - this depends on your platform and requires the metaMSdata package
  cdfpath <- system.file("extdata", package = "metaMSdata")
  ## files
```
```r
files <- list.files(cdfpath, ".RP\", full.names=TRUE)

## get the complete names for the files
exptable$stdFile <-
  sapply(exptable$stdFile,
         function(x)
         files[grep(x,files)])

## Not run:
## load the settings for the analysis
data(FEMsettings)

## set the minimum number of features to 2
metaSetting(Synapt.RP, "DBconstruction")$minfeat <- 2

## create the DB
LCDBtest <- createSTDdbLC(stdInfo=exptable,
                          settings = Synapt.RP,
                          polarity = "positive",
                          Ithr = 20)

## End(Not run)
## saved in "LCDBtest.RData" in the data directory of the metaMS
## package
```

---

**errf**

*Mass error surface for Waters Synapt Q-TOF spectrometers*

**Description**

This function can be used to calculate the optimal mz tolerance (in ppm) for annotation. The surface has been developed for a Waters Synapt QTOF spectrometer.

**Usage**

```r
data(errf)
```

**Format**

`errf` is a linear model of the form $M + \log I + M \times \log I$:

- **M**: The mz of the ion
- **logI**: Logarithm of the intensity of the ion.

**Details**

The function is a linear approximation of the complete error function (see Reference). The latter has been calculated by comparing the measured and nominal mass of several hundreds of standards. The experiments were performed by using a Waters Synapt Q-TOF spectrometer so this specific surface is valid only for this model of spectrometers.
Author(s)

Pietro Franceschi

References


Examples

```r
## <-------------- direct use of the error function -------------- >
## load the Synapt-QTOF error function
data(errf)

## predict the mass error in ppm
newdata <- data.frame(M = c(105, 131, 157), logI = c(1, .5, 1.4))
predict(errf, newdata) ## mass error in ppm

## <-------------- create a dummy error function -------------- >
## dataset to evaluate it:
## "M" is the mz,
## "logI" is the log of the intensity
## "err" is the mass error in ppm. The error is the difference between the
## actual m/z of a known ion, and the one measured with the spectrometer
MErr.data <- data.frame("M" = seq(1,500,2),
                        "logI" = rnorm(250, mean = 5, sd = 1),
                        "err" = rnorm(250, mean = 40, sd = 5))

dummy.model <- lm(err~M+logI, data = MErr.data)

data(LCresults)
data(FEMsettings)

## Run the analysis with an adaptive mass tolerance
result.adaptive.dummy <- runLC(xset = LCresults$xset,
                               settings = Synapt.RP,
                               DB = LCDBtest$DB,
                               errf = dummy.model)

## <-------------- Use the Synapt Q-TOF error function -------------- >
## load the Synapt-QTOF error function
data(errf)

result.adaptive <- runLC(xset = LCresults$xset,
                          settings = Synapt.RP,
                          DB = LCDBtest$DB,
                          errf = errf)
```
Sample table for the generation of a database of standards (LCMS)

### Description

An example of a table used to construct a database of standards. This particular table is used to obtain object LCDBtest.

### Usage

```r
data(exptable)
```

### Format

exptable is a data.frame summarizing information on the chemical standards used for creating a database.

Column description:

- **ChemSpiderID** The Chemspider ID of a specific compound.
- **compound** A string containing the (human) readable name of the compound.
- **formula** The formula of the compound.
- **M.ref** The theoretical value for the observed m/z reference ion. This can be, for example, the protonated (de-protonated) molecular ion, a known adduct or a characteristic fragment.
- **mz.observed** The manually validated m/z value of the reference ion.
- **RTman** The manually validated retention time value for the standard.
- **stdFile** The name (including the path) of the raw data file of the standard.

### Details

Mandatory fields are: ChemSpiderID, mz.observed, RTman, and stdFile.

In the current implementation the M.ref value is not used in the creation of the DB. The difference between M.ref and m.observed, however, could be used to construct the mass error surface used during feature annotation.

The sample table is also an element (Reftable) of the LCDB list generated by `createSTDdbLC`.

### Author(s)

Pietro Franceschi

### See Also

`createSTDdbLC`
FEMsettings

Example settings for metaMS

Description

Examples of settings needed for functions runLC and runGC: Synapt.RP, Synapt.NP, TSQXLS.GC and Orbitrap.RP. These four particular settings are fine-tuned for the analysis of LC-MS runs, both normal-phase and reverse-phase chromatography (Waters Synapt G1-Thermo Orbitrap) and GC-MS experiments (ThermoXLS TQQ).

Usage

data(FEMsettings)

Format

Four objects of class metaMSsettings.

Author(s)

Ron Wehrens and Pietro Franceschi

See Also

findPeaks, annotate

Examples

## Not run:
## The three sets of settings are created as follows:
Synapt.NP <- metaMSsettings(protocolName = "Synapt.QTOF.NP",
  chrom = "LC",
  PeakPicking = list(
    method = "matchedFilter",
    step = 0.05,
    fwhm = 20,
    snthresh = 4,
    max = 50),
  Alignment = list(
    min.class.fraction = .3,
    min.class.size = 3,
    mzwid = 0.1,
    bws = c(130, 10),
    missingratio = 0.2,
    extraratio = 0.1,
    Retcor = list(
      method = "linear",
      family = "symmetric"),
    fillPeaks = TRUE),
  CAMERA = list(
    perfwhm = 0.6,
    cor_eic.th = 0.7,
    ppem = 5))
metaSetting(Synapt.NP, "match2DB") <- list(
FEMsettings

rtdiff = 1.5,
rtval = .1,
mzdiff = 0.005,
ppm = 5,
minfeat = 2)
metaSetting(Synapt.NP, "DBconstruction") <- list(
  minfeat = 3,
  rttol = .3,
  mztol = .01)

## For reverse-phase LC, settings are very similar: the only difference
## is in the alignment settings
Synapt.RP <- Synapt.NP
metaSetting(Synapt.RP, "protocolName") <- "Synapt.QTOF.RP"
metaSetting(Synapt.RP, "Alignment") <- list(
  min.class.fraction = .3,
  min.class.size = 3,
  mzwid = 0.1,
  bws = c(30, 10),
  missingratio = 0.2,
  extraratio = 0.1,
  Retcor = list(
    method = "linear",
    family = "symmetric"),
  fillPeaks = TRUE)

## For the orbitrap.RP
Orbitrap.RP <- metaMSsettings(protocolName = "Orbitrap",
  chrom = "LC",
  PeakPicking = list(
    method = "centWave",
    ppm = 5,
    prefilter = c(3,10000),
    peakwidth = c(15,40)),
  Alignment = list(
    bws = 30,
    min.class.fraction = 0.3,
    min.class.size = 3,
    mzwid = 0.01,
    Retcor = list(
      method = "obiwarp",
      profStep = 0.2),
    fillPeaks = TRUE),
  CAMERA = list(
    perfwhm = 0.6,
    cor_eic_th = 0.7,
    ppm = 5))
metaSetting(Orbitrap.RP, "match2DB") <- list(
  rtdiff = 1.5,
  rtval = .1,
  mzdiff = 0.005,
  ppm = 5,
  minfeat = 2)
metaSetting(Orbitrap.RP, "DBconstruction") <- list(
  minfeat = 3,
  rttol = .3,
GCresults

mztol = .01)

## For the thermo TQ

TSQXLS.GC <- metaMSsettings("protocolName" = "TSQXLS.QQQ.GC",
  "chrom" = "GC",
  PeakPicking = list(
    method = "matchedFilter",
    step = 0.5,
    steps = 2,
    mzdiff = .5,
    fwhm = 5,
    snthresh = 2,
    max = 500),
    CAMERA = list(perfwhm = 1))

metaSetting(TSQXLS.GC, "DBconstruction") <- list(
  minintens = 0.0,
  rttol = .1,
  intensityMeasure = "maxo",
  DBthreshold = .80,
  minfeat = 5)

metaSetting(TSQXLS.GC, "match2DB") <- list(
  simthresh = 0.80,
  timeComparison = "rt",
  rtdiff = .5,
  RIdiff = 5,
  minfeat = 2)

metaSetting(TSQXLS.GC, "matchIrrelevants") <- list(
  irrelevantClasses = c("Bleeding", "Plasticizers"),
  timeComparison = "rt",
  RIdiff = 2,
  rtdiff = .05,
  simthresh = 0.70)

metaSetting(TSQXLS.GC, "betweenSamples") <- list(
  min.class.fraction = .5,
  min.class.size = 5,
  timeComparison = "rt",
  rtdiff = .05,
  RIdiff = 2,
  simthresh = .95)

## End(Not run)

---

**GCresults**

*Results of metaMS for a small GC-MS data set*

**Description**

This data file contains the results of applying the metaMS pipeline to a small GC-MS data set consisting of repeated injections of a set of chemical standards. Its aim is to demonstrate the structure of the (intermediate) outcomes without having to do the slightly time-consuming calculations.

**Usage**

```r
data(GCresults)
```
getAnnotationLC

Details

Data object GCresults is the result of a complete execution of runGC, using the example database for annotation purposes.

See Also

runGC

getAnnotationLC            Get LC annotation

Description

Main function for the annotation of an xcmsSet or CAMERA object. This function is not meant to be called directly. Use runLC instead.

Usage

getAnnotationLC(xs, settings, DB, errf)

Arguments

xs            The xcmsSet (or CAMERA) object to be annotated.
settings      The subset of settings contained into the match2DB elements of the settings list. See the help of FEMsettings for details.
DB            The database used within AnnotateTable for the annotation of the peaklist. See the help of FLCDBtest for details.
errf          The file containing the error function used to predict the m/z tolerance. See the help of AnnotateTable for details.

Details

The function extracts from the xs object a Peaktable with the intensities of the features across all the samples. Since this Peaktable is meant to be used only for annotation (and not for subsequent statistical analysis), the intensities are expressed as maxo - the absolute maximum of the signal over the detected chromatographic peak (see the documentation of xcms for more details). Within the function the peaktable is converted into a matrix in the form (mz,rt,I) used by AnnotateTable. If xs contains more than one sample, the intensity is the maximum intensity of each feature across all the samples.

Value

A list with two elements. raw is the complete output of AnnotateTable. for_table is a data.frame which summarizes the outputs of the annotation (see AnnotateTable) and it is included in the output of the runLC main function.

Author(s)

Pietro Franceschi
**References**


**See Also**

AnnotateTable, LCDBtest, FEMsettings

---

**getAnnotationMat**

**Obtain relative quantitative annotation results for GC-MS**

**Description**

Function `getAnnotationMat` obtains relative intensities for features in individual samples. A robust linear regression is performed when the number of common features is five or larger; for 2-4 peaks a weighted linear regression is used, and if only one peak is in common the intensity ratio for this peak is used. Reference patterns are the patterns in the database of standards or the patterns in the “unknowns” element of the `allMatches` object. Not meant to be called directly by the user.

**Usage**

```
getAnnotationMat(exp.msp, pspectra, allMatches)
relInt(pat, refpat)
```

**Arguments**

- `exp.msp`: List of experimental pseudospectra.
- `pspectra`: Spectra from the in-house database.
- `allMatches`: Match information in the form of a list.
- `pat, refpat`: Both pseudospectra.

**Value**

Function `getAnnotationMat` returns a matrix containing all patterns (standards as well as unknowns) in the rows, and numeric values signifying relative intensities in all samples in the columns. These relative intensities are the quantities calculated by `relInt`, which simply returns one number.

**Author(s)**

Ron Wehrens
getFeatureInfo

Construct an object containing all meta-information of the annotated pseudospectra (GC-MS).

Description

Function getFeatureInfo yields a data.frame with meta-information on all features detected in the samples. Features are rows; information is in the columns. Not meant to be called directly by the user.

Usage

getFeatureInfo(stdDB, allMatches, sampleList)

Arguments

stdDB In-house database of standards.
allMatches Object containing annotation information, generated by functions like matchSamples2DB and matchSamples2Samples.
sampleList Object containing pseudospectra of samples.

Value

A data.frame summarizing all meta-information of the pseudospectra found in the samples. Exactly what columns are included depends on the information contained in the in-house database. The last column is always "rt".

Author(s)

Ron Wehrens

getPeakTable

Extract a peak table from an xcms or CAMERA object

Description

Extracts the peak table (a data.frame) from an xcms (or CAMERA) object (without compound annotation). The peak table contains the mass and retention time for all the features and their intensities across the samples. This function is not meant to be called directly, but it is internally used by runLC, getAnnotationLC, createSTDdbLC.

Usage

getPeakTable(xs, intval = "into")

Arguments

xs The xcmsSet (or CAMERA) object
intval The intensity measure extracted form xs.
Details

The function process an xs object and returns for it a PeakTable which associates intensities to features and samples. The default measure for the intensity is into (the chromatographic peak area for a feature), but in the case of annotation, maxo (value for the intensity of the ion over the chromatographic peak) is used to measure the intensity. For a more detailed description of the possible intensity measures refer to the documentation of xcms.

Value

A data frame with the intensity for each feature (rows) in all the samples (columns). Each feature is identified by its m/z value and retention time (in minutes). If the xs object is of class CMAERA, the results of the camera annotation (isotope, adduct, pcgroup) are included in the table.

Author(s)

Pietro Franceschi

---

LCDBtest

Sample DB for LC-MS annotation

Description

This database has been generated using the exptable dataset to process the data included in the metaMSdata package.

Usage

data(LCDBtest)

Format

LCDBtest is a list with three elements:

- The initial table used to generate the DB.
- RefTable Info Some info on the DB: date of creation, and settings used.
- DB A data frame containing the actual information. See the details section.

Details

The DB data frame contains the following fields:

ChemSpiderID The Chem spider identifier for the compound.
compound A human-readable name for the compound.
adduct The output of the CAMERA annotation of the standard file.
isotopes The output of the CAMERA annotation of the standard file.
mz, rt the mz and rt values.
maxo the maxo intensity value for a given feature in the injection.
validated a string which defines the origin of a specific feature.
Author(s)

Pietro Franceschi

See Also

createSTDdbLC, exptable, FEMsettings

LCresults Result metaMS for a small LC-MS data set

Description

This data file contains the result of applying the metaMS pipeline to a small LC-MS data set consisting of repeated injections of a set of chemical standards. Its aim is to demonstrate the structure of the outcomes without having to do the slightly time-consuming calculations.

Usage

data(LCresults)

Details

Data object LCresult contains the output list of runLC.

match2ExtDB Match GC-MS spectra to an external reference DB

Description

When building an in-house database, it is imperative that pseudospectra are validated. This function provides the possibility of automatic validation by comparing the spectra to an external reference database, such as the NIST. Not meant to be called directly by the user.

Usage

match2ExtDB(xsetList, extDB, settings)

Arguments

xsetList A list of xcmsSet objects.

extDB External database in msp format.

settings Settings for the comparison, including as the most important the minimal number of features required in a valid pseudospectrum (minfeat), and the minimal agreement to speak of a match (DBthreshold).

Value

An msp object containing validated pseudospectra.

Author(s)

Ron Wehrens
Function `matchExpSpec` calculates match factors for a pseudospectrum with all entries in the database. A plot of the best match can be provided. Function `mzmatch` is an auxiliary function, not meant to be called directly, that provides the match factor, given two appropriately scaled patterns.

**Usage**

```r
matchExpSpec(pspec, DB, DB.treated, plotIt = FALSE,
            scale.p = c("sqrt", "none"), mass.weight = TRUE, ...)
mzmatch(spec1, spec2)
```

**Arguments**

- `pspec` The pseudospectrum, a two- or three-column matrix where the third column (the retention time) will be ignored.
- `DB` Database of standards.
- `DB.treated` Logical, indicating whether the database has already been scaled (TRUE) or not.
- `plotIt` Logical: show best match?
- `scale.p` indicates whether "sqrt" scaling or no scaling ("none") is to be applied. This should usually be "sqrt".
- `mass.weight` Logical, indicating whether heavier masses receive higher weight. Should usually be TRUE.
- `...` Further arguments for the pseudospectrum plot (if shown).
- `spec1, spec2` Preprocessed pseudospectra, here two-column matrices where the first column is the (integer) m/z value, and the second column the (scaled) intensity.

**Value**

A vector of match factors.

**Author(s)**

Ron Wehrens

**Examples**

```r
data(threeStdsNIST) ## gives smallDB, containing 78 patterns
data(threeStdsDB) ## gives DB, containing 3 patterns :-D

matchExpSpec(DB[[1]]$pspectrum, smallDB, DB.treated = FALSE, plotIt = TRUE)
```
**matchSamples2DB**  
*Match pseudospectra from several samples to an in-house DB (GC-MS)*

**Description**

Compare experimental results to a database of pseudospectra. The result is a nested list, containing for every pseudospectrum in every sample either an index of a corresponding pattern in the DB (if a hit is found) or nothing (if no hit is found). Not meant to be called directly by the user.

**Usage**

```r
matchSamples2DB(xset.msp, DB, settings, quick)
```

**Arguments**

- `xset.msp`: An object containing a list of pseudospectra.
- `DB`: The in-house database.
- `settings`: The settings, in the form of a list.
- `quick`: Logical. If TRUE, scaling of the pseudospectra (which is necessary for comparing to the database) will be done once using all masses in the pseudospectrum. This mode is routinely applied when comparing with a database of artefacts such as bleeding patterns or plasticizers. When comparing with a database of chemical standards, however, only peaks smaller than the molecular weight (+ 4, to allow for isotopes) should be taken into account in the scaling, and hence the scaling needs to be re-done for every comparison. This is _not_ so quick...

**Value**

A list object, with one element for each experimental sample. Every element again is a list with an entry for each pseudospectrum from that sample: if the element is empty, there is no match with the DB - a number points to the DB entry that gives a hit. Elements can contain more than one number; these will be split into one “best” annotation and “alternative” annotations in function `annotations2tab`.

**Author(s)**

Ron Wehrens

---

**matchSamples2Samples**  
*Compare pseudospectra across samples (GC-MS)*

**Description**

Function `matchSamples2Samples` matches pseudospectra across all samples - if a pseudospectrum is present at more or less the same retention time in several samples, it can get the status of “unknown”. Exactly how much difference there may be between the pseudospectra and retention times, and how often it should be present, is determined by the settings. The auxiliary function `match.unannot.patterns` compares two msp objects, representing experimental samples. Both are not meant to be called directly by the user.
Usage

matchSamples2Samples(xset.msp.scaled, xset.msp, annotations, settings)
match.unannot.patterns(msp1, msp2, settings)

Arguments

xset.msp.scaled  
Scaled version of all pseudospectra in the experimental patterns - a nested list, with one entry for each sample, and within every entry an element for each pseudospectrum.

xset.msp  
Unscaled version of the first argument: both arguments are provided for efficiency reasons.

annotations  
Annotations of all pseudospectra: only patterns without annotations will be considered.

settings  
Settings determining what a valid “unknown” is. For an example, see the man page of FEMsettings, field matchIrrelevants.

msp1, msp2  
lists of pseudospectra

Value

Function matchSamples2Samples returns an updated annotation object such as the one returned by matchSamples2DB, but now with an additional unknowns element, containing the pseudospectra that are recognized as “unknowns”.

Function match.unannot.patterns returns a list of combinations of pseudospectra IDs, retention times (or retention indices) and match factors (only for those combinations that satisfy the criteria on retention time (index) and match factor).

Author(s)

Ron Wehrens

Objects from the Class

Objects can be created by calls of the form metaMSsettings(...). See the example below.
Slots

Note: all slots describing retention times or retention time differences use minutes and not seconds. If a slot is only relevant for either GCMS or LCMS, this is indicated explicitly.

protocolName: Object of class "character": the name of the instrumental protocol, a unique identifier.

chrom: Object of class "character": chromatography. Either "LC" or "GC".

PeakPicking: Object of class "list": The parameters used for xcms peakpicking. See the arguments of findPeaks.

Alignment: Object of class "list": The parameters used for grouping and alignment. min.class.fraction and min.class.size are used to calculate the minsample xcms parameter. bws is a vector of the two bandwidths used for grouping before and after retention time alignent. missingratio and extraratio are used to set the values for missing and extra as a function of the number of samples. LC only.

CAMERA: Object of class "list": The parameters for CAMERA.

match2DB.rtdiff: Object of class "numeric": the maximal difference in retention time to match each feature with the entry in the DB.

match2DB.minfeat: Object of class "numeric": for LC, the minimal number of matching features within a retention time interval of width rtval before we speak of a hit. For GC, the minimal number of common masses for calculating a match factor.

match2DB.rtval: Object of class "numeric": the tolerance in retention time for features used in annotation. LC only.

match2DB.mzdiff: Object of class "numeric": the mass accuracy which is used if no error surface is provided. LC only.

match2DB.ppm: Object of class "numeric": the minimum mass tolerance allowed when the error surface is used. LC only.

match2DB.simthresh: Object of class "numeric": the minimal match factor to speak of a hit. GC only.

match2DB.timeComparison: Object of class "character": either "rt" or "RI". GC only.

match2DB.RIdiff: Object of class "numeric": maximal retention index difference with DB entry (GC only).

DBconstruction.minfeat: Object of class "numeric": the minimum number of features necessary to include a compound in the DB.

DBconstruction.rttol: Object of class "numeric": the tolerance in retention time to match experimental features with the reference table.

DBconstruction.mztol: Object of class "numeric": the tolerance in m/z (in dalton) to match experimental features with the reference table. LC only.

DBconstruction.minintens: Object of class "numeric": the minimum intensity for a feature to be included in the list. GC only.

DBconstruction.intensityMeasure: Object of class "character": either "into" or "maxo". GC only.

DBconstruction.DBthreshold: Object of class "numeric": minimal match factor with an external DB for a pseudospectrum to be included in the DB of standards. GC only.

matchIrrelevants.irrelevantClasses: Object of class "character": classes of compounds are considered as irrelevant (a vector of string constants, which should exactly match the Class element in the DB entries). GC only.
metaMSsettings-class

matchIrrelevants.simthresh: Object of class "numeric": the minimal match factor to speak of a hit. GC only.
matchIrrelevants.timeComparison: Object of class "character": either "rt" or "RI". GC only.
matchIrrelevants.rtdiff: Object of class "numeric": maximal retention time difference between two unknowns - this can be set to a very high value if a pattern is to be removed whatever the retention time. GC only.
matchIrrelevants.RIdiff: Object of class "numeric": maximal retention index difference between two unknowns - this can be set to a very high value if a pattern is to be removed whatever the Retention Index. GC only.
betweenSamples.min.class.fraction: Object of class "numeric": fraction of samples in which a pseudospectrum is present before it is regarded as an unknown. GC only.
betweenSamples.min.class.size: Object of class "numeric": absolute number of samples in which a pseudospectrum is present before it is regarded as an unknown. GC only.
betweenSamples.timeComparison: Object of class "character": either "rt" or "RI". GC only.
betweenSamples.rtdiff: Object of class "numeric": max retention time difference between pseudospectra in different samples. GC only.
betweenSamples.RIdiff: Object of class "numeric": max retention index difference between pseudospectra in different samples. GC only.
betweenSamples.simthresh: Object of class "numeric": similarity threshold fo comparing pseudospectra in different samples. GC only.

Methods

metaSetting<- signature(object = "metaMSsettings"): change values in a metaMSsettings object.
metaSetting signature(object = "metaMSsettings"): get values from a metaMSsettings object.
show signature(object = "metaMSsettings"): show a metaMSsettings object.

Author(s)

Ron Wehrens

See Also

FEMsettings

Examples

showClass("metaMSsettings")

## Not run:
## The three sets of settings are created as follows:
Synapt.NP <- metaMSsettings(protocolName = "Synapt.QTOF.NP",
                         chrom = "LC",
                         PeakPicking = list(
                           method = "matchedFilter",
                           step = 0.05,
                           fwhm = 20,
                           snthresh = 4,
                           max = 50),
                         ...)
Alignment = list(
  min.class.fraction = .3,
  min.class.size = 3,
  mzwid = 0.1,
  bws = c(130, 10),
  missingratio = 0.2,
  extraratio = 0.1,
  retcormethod = "linear",
  retcorfamily = "symmetric",
  fillPeaks = TRUE),
CAMERA = list(
  perfwhm = 0.6,
  cor_eic.th = 0.7,
  ppm = 5))

metaSetting(Synapt.NP, "match2DB") <- list(
  rtdiff = 1.5,
  rtval = .1,
  mzdiff = 0.005,
  ppm = 5,
  minfeat = 2)
metaSetting(Synapt.NP, "DBconstruction") <- list(
  minfeat = 3,
  rttol = .3,
  mztol = .01)

## End(Not run)

metaSetting-methods  Get or set values in metaMSsettings objects

Description
Accessor function for metaMSsettings objects, allowing to get or set values from individual slots.

Methods
signature(object = "metaMSsettings")  Get or set values from individual slots in a metaMSsettings objects.

See Also
FEMsettings

Examples
## Not run:
## The three sets of settings are created as follows:
Synapt.NP <- metaMSsettings(protocolName = "Synapt.QTOF.NP",
  chrom = "LC",
  PeakPicking = list(
    method = "matchedFilter",
    step = 0.05,
    fwhm = 20,
    snthresh = 4,
max = 50),
Alignment = list(
  min.class.fraction = .3,
  min.class.size = 3,
  mzwid = 0.1,
  bws = c(130, 10),
  missingratio = 0.2,
  extraratio = 0.1,
  retcormethod = "linear",
  retcorfamily = "symmetric",
  fillPeaks = TRUE),
CAMERA = list(
  perfwhm = 0.6,
  cor_eic_th = 0.7,
  ppm = 5)

metaSetting(Synapt.NP, "match2DB") <- list(
  rtdiff = 1.5,
  rtval = .1,
  mzdiff = 0.005,
  ppm = 5,
  minfeat = 2)

metaSetting(Synapt.NP, "DBconstruction") <- list(
  minfeat = 3,
  rttol = .3,
  mztol = .01)

## End(Not run)

### msp

*Functions to handle msp-type objects (GC-MS)*

#### Description

Functions to construct, read, write and filter so-called msp objects, collections of spectra with additional information. This other information is of free format, but typically contains fields like “Name”, “CAS”, “ChemspiderID”, and “rt”.

#### Usage

- `construct.msp(spectra, extra.info)`
- `read.msp(file, only.org = FALSE, org.set = c("C", "H", "D", "N", "O", "P", "S"), noNumbers = NULL)`
- `filter.msp(msp, rtrange = NULL, mzrange = NULL, minintens = 0, minPeaks = 0)`
- `write.msp(msp, file, newFile = TRUE)`
- `xset2msp(xsetList, settings)`
- `to.msp(object, file = NULL, settings = NULL, ndigit = 0, minfeat, minintens, intensity = c("maxo", "into"), secs2mins = TRUE)`

#### Arguments

- `spectra` a list of two-column or three-column numerical matrices. Two-column matrices only contain mz and intensity information, three-column matrices have a third column that gives retention times for the individual peaks.
extra.info  a nested list containing all other slots that should be included in the msp object. The length of this list should be equal to the number of spectra in the first argument.

file  name of input or output file containing data in msp format. If NULL in function to.msp, the input object is returned as an msp object. If non-null, the information will be written to the file with this name.

only.org  logical: if TRUE this will skip all entries that have non-organical elements in the “Formula” field.

org.set  List of elements to be considered as organic.

noNumbers  Optional argument, indicating which fields are not to be converted into numeric entries. If not given, this currently defaults to the following fields: Name, CAS, stdFile, date, validated, ChemspiderID, SMILES, InChI, Class, comment, csLinks.

msp  msp object.

rtrange, mzrange, minfeat, minintens, minPeaks  restrictions to what constitutes genuine pseudospectra in terms of minimal numbers of features, minimum feature intensity, and the intensity measure used (peak area or peak height).

intensity  the measure to use for the intensity of a peak: either the peak height (“maxo”) or area (“into”).

ndigit  number of digits for the mz values - use 0 for nominal mass data.

newFile  logical: if TRUE starts a new file, otherwise appends.

xsetList  a list of xcmsSet objects.

settings  list containing settings.

object  either an object of class "xsAnnotate" or a peaktable containing mz, rt and I information. Function to.msp will convert this object into a list that can be written as an msp file.

secs2mins  logical: if TRUE converts retention times to minutes (otherwise seconds).

Details

Even though the msp format handled by these functions is quite flexible, there are a couple of requirements that are not always satisfied by msp files generated by other software, the most important one being that one line may only contain one keyword. If more than one keyword is present, the second will likely not be read. Furthermore, the current implementation assumes that peaks in pseudospectra are represented in the form mz1, I1; mz2, I2; etcetera - in other cases these are in brackets. To solve such issues, the most easy fix for the moment is to edit the msp file and change things globally.

Other issues may arise with the keywords. While read.msp will be able to read msp files with non-standard keywords, the metaMS package expects at least the following list to be present (case-sensitive): “Name”, “validated”, “CAS”, “rt”, “monoMW”, and “Class”. The final keyword is always “Num Peaks”, and should be followed by the list of mz-I combinations.

Author(s)

Ron Wehrens
Examples

data("threeStdsDB")
## Not run:
write.msp(DB, file = "huhn.msp")
DB2 <- read.msp("huhn.msp")
## End(Not run)

Description

XCMS peak detection using settings, defined for individual laboratories and depending on the chromatographic and mass-spectrometric characteristics of the instruments at hand.

Usage

peakDetection(files, settings, rtrange = NULL, mzrange = NULL,
convert2list = FALSE, nSlaves = 0)

Arguments

files input files (including path names) that will be processed by xcms.
settings a list of settings that will be passed on to the xcmsSet function. See the help of FEMsettings for a detailed description of the fields in this list.
rtrange If non-NULL, a vector to subset the region of the chromatography retained for further analysis (given in minutes).
mzrange If non-NULL, a vector indicating the subset of the mass spectrum retained for the analysis.
convert2list logical. If TRUE, the xcmsSet object will be split into a list of one-sample objects which will be treated separately. This is useful not only in GC-MS data, where peak alignment is avoided, but also in setting up databases of standards, where no common peaks between injections are expected to occur.
nSlaves Number of cores to be used in the peak picking.

Value

Either an xcmsSet object, or a list of one-sample xcmsSet objects.

Author(s)

Ron Wehrens and Pietro Franceschi
plotPseudoSpectrum  

Plot a pseudospectrum.

Description

Auxiliary function for plotting a particular pseudospectrum. M/z values are in the first column of the matrix, and an intensity measure (either maxo, into or something else) in the second. The third column is disregarded, usually contains retention time information.

Usage

plotPseudoSpectrum(psspc, ...)

Arguments

psspc  Pseudospectrum, consisting of a two- or three-column matrix. The first column contains the m/z values, the second the intensities. A third column containing retention time information may be present, but is not used in this function.

...  Additional graphical parameters.

Details

A stick spectrum is shown on the graphical device.

Author(s)

Ron Wehrens

Examples

data("threeStdsDB")
plotPseudoSpectrum(DB[[1]]$pspectrum)

printString  

Functions for metaMS-formatted text output

Description

Functions to present progress output, warnings or information, in a consistent way in the console window. Not meant to be called by the user.

Usage

printString(..., screenwidth = 72)
printWarning(...)
printInfo(...)

Arguments

...  Text strings. These will be concatenated inside the function.

screenwidth  Width of the text field.
processStandards

Author(s)

Ron Wehrens and Pietro Franceschi

processStandards  Process input files containing raw data for pure standards.

Description

Peak picking and further processing for raw data of pure standards, including CAMERA processing. This function is not meant to be called directly - use createSTDdbLC or createSTDdbGC instead.

Usage

processStandards(stdInfo, settings, polarity = NULL, nSlaves)

Arguments

stdInfo  Object describing the pure standards: a data.frame containing, e.g., the name of the file, the name of the standard, descriptors like CAS or Chemspider IDs, etcetera.

settings  Settings list, containing sublists for peak picking and CAMERA grouping (GC-MS) or annotation (LC-MS).

polarity  Polarity of the analysis (used for CAMERA). Possible values are “positive” or “negative”. Ignored for GC-MS.

nSlaves  Number of cores to be used in peak picking.

Value

A list of CAMERA objects resulting from the analysis of the standard injections listed in the stdInfo table.

Author(s)

Ron Wehrens and Pietro Franceschi

See Also

xcmsSet, runCAMERA
### readStdInfo

**Read information of GC injections of standards from a csv file**

**Description**

The csv contains all information necessary to process a series of injections of standards (GC-MS). Required fields: Name, RTman, monoMW, stdFile, and an identifier such as CAS or ChemspiderID. At the moment, the system is completely based on CAS, although this may change in the future.

**Usage**

```r
readStdInfo(stdInfo, InputDir, sep = "", dec = ".", ...)  
```

**Arguments**

- `stdInfo` Input file in csv format, containing information on standards.
- `InputDir` Location of input files.
- `sep`, `dec`, ... optional arguments to `read.table`.

**Details**

In addition to reading all information on the chemical standards (which will eventually be transferred into an in-house database), the function checks whether some input files are unavailable, and whether some data files are not used. In the first case, an error is returned, in the second case a warning.

**Author(s)**

Ron Wehrens

**Examples**

```r
if (require(metaMSdata)) {
  ## this will lead to the completed version of the R object that is also
  ## available by typing "data(threeStdsInfo)", now containing the
  ## directory information that is not available in the RData object.
  input.file <- list.files(system.file("extdata", package = "metaMSdata"),
                          pattern = "csv", full.names = TRUE)
  threeStdsInfo <- readStdInfo(input.file,
                          system.file("extdata", package = "metaMSdata"),
                          sep = ";", dec = ",")
  ## only one of the files is used to set up the database, the others
  ## are for testing annotation
}
```
removeDoubleMasses  Remove double m/z entries in a pseudospectrum

Description
Since in nominal-mass GC data m/z values are rounded to whole numbers, in some cases an m/z value may occur multiple times - in that case the mean of the intensities is kept (and the retention times are averaged). removeDoubleMasses takes a list of spectra in the form of a three-column matrix, (mz, I, rt), summing the intensities and averaging the retention times for multiple identical masses. Not meant to be called directly by the user.

Usage
removeDoubleMasses(spclist)

Arguments
spclist A list of spectra, each one consisting of a three-column matrix (mz, I, rt).

Value
The function returns a list of spectra, where all "double" peaks have been averaged.

Author(s)
Ron Wehrens

runCAMERA The CAMERA element in the metaMS pipeline

Description
Run CAMERA package with settings from the settings list (see FEMsettings). Works both for LC and GC. Not meant to be called directly by the user.

Usage
runCAMERA(xset, chrom = c("LC", "GC"), settings, polarity, quick = TRUE)

Arguments
xset For LC, an xcmsSet object; for GC, a list of xcmsSet objects.
chrom The type of chromatography. Either "LC" or "GC".
settings The subset of settings contained into the "CAMERA" element of the settings list.
polarity The polarity of the injection, used by CAMERA to look for common adducts.
quick Only relevant for LC data. If TRUE, CAMERA runs only groupFWHM and findIsotopes. If FALSE, correlations between EICs are calculated and used for grouping (which can take some time).
Details

In the case of LC the function is used in data analysis and DB creation. In the first case, it increases the level of the annotation and it, by default, run with quick = TRUE. For DB creation, the grouping of the features into "pcgroups" (features with similar retention time) is used to choose the features to be included into the database. In this case camera is run with quick = FALSE: quick determines whether or not the correlation among the extracted ion chromatograms should be used to "validate" the pcgroups.

For GC data, only the grouping done by groupFWHM is performed: basically this clusters peaks with a similar retention time.

Value

An annotated xcmsSet object (an object of class CAMERA).

Author(s)

Ron Wehrens and Pietro Franceschi

See Also

annotate, groupFWHM

runGC

Wrapper for processing of GC-MS data files

Description

Main function of the pipeline for GC-MS data processing. It includes XCMS peak detection, definition of pseudospectra, and compound identification by comparison to a database of standards. The function also takes care of removal of artefacts like column bleeding and plasticizers, and definition of unknowns, consistently present across samples.

Usage

runGC(files, xset, settings, rtrange = NULL, DB = NULL, removeArtefacts = TRUE, findUnknowns = nexp >= mcs, returnXset = FALSE, RIstandards = NULL, nSlaves = 0)

Arguments

files input files, given as a vector of strings containing the complete paths. All formats supported by XCMS can be used.

xset alternatively, one can present a list of xcmsSet objects for whom CAMERA grouping has been done. In this case, only the annotation process will be done. If both files and xset are given, the former takes precedence.

settings a nested list of settings, to be used at individual steps of the pipeline.

rtrange part of the chromatograms that is to be analysed. If given, it should be a vector of two numbers indicating minimal and maximal retention time (in minutes).
runGC

- **DB**: Database containing the spectra of the pure standards. At least the following fields should be present: Name, std_rt, pspectrum and monoMW.

- **removeArtefacts**: Logical, whether or not to remove patterns identified as (e.g.) column bleeding. Only performed if a database containing such patterns is available.

- **findUnknowns**: Logical, whether to find patterns without identification that are present consistently in several samples. The default is to use TRUE if the number of samples is larger than the min.class.size setting in the 'betweenSamples' metaSetting.

- **returnXset**: Logical: should the XCMS output be returned? If yes, this is a list of xcmsSet objects, one element for each input file.

- **RIstandards**: A two-column matrix containing for the standards defining the RI scale both retention times and retention indices. If not given, no RI values will be calculated and retention times will be used instead.

- **nSlaves**: Number of cores to be used in peak picking.

**Value**

A list with the following elements:

- **PeakTable**: Data.frame containing annotation information. Every line is a feature, i.e. a pseudospectrum. The first columns are used to give information about these features, a.o. compound name, CAS and Chemspider IDs, etcetera. The last of these meta-information columns is always the one giving the retention time: "rt". After that, columns correspond to input files, and give measures of intensities for every single one of the features. If a feature is not detected in a sample, this is indicated with "0" (zero).

- **PseudoSpecra**: A list of pseudospectra in msp format, in the same order as the rows in the PeakTable.

- **xset**: Optionally, the xcmsSet object is returned, which can be useful for more detailed inspection of the results. It can also be used as an input for runGC, e.g., to test different annotation settings independently of the xcms/CAMERA part.

- **sessionInfo**: The output of sessionInfo() to keep track of the sw version used for the processing.

**Author(s)**

Ron Wehrens

**References**


**See Also**

msp, treat.DB, runCAMERA, peakDetection, matchSamples2DB, matchSamples2Samples, getAnnotationMat, addRI
Examples

```r
## analysis of an xset object
data(threeStdsDB)
data(FEMsettings)

data(GCresults) ## pre-compiled results
names(GCresults)

## Not run:
if (require(metaMSdata)) {
  ## object GCresults is created by
cdfdir <- system.file("extdata", package = "metaMSdata")
cdffiles <- list.files(cdfdir, pattern = "_GC_",
    full.names = TRUE, ignore.case = TRUE)
GCresults <- runGC(files = cdffiles, settings = TSQXLS.GC, DB = DB,
    returnXset = TRUE)

  ## to start directly from the XCMS/CAMERA results and not include
  ## peak picking in the pipeline, simply provide the "xset" argument
  ## rather than the "files" argument.
  ## no annotation database
  result.noannot <- runGC(xset = GCresults$xset, settings = TSQXLS.GC)
}

## End(Not run)
```

runLC

Wrapper for processing of LC-MS data files

Description

Main function of the pipeline for LC-MS data processing. It includes XCMS peak detection, grouping and alignment, CAMERA and feature annotation by comparison to a database of standards. The function also calculates the mass tolerance on the bases of the ion intensity and ion mass.

Usage

```r
runLC(files, xset, settings, rtrange = NULL, mzrange = NULL, DB = NULL,
  polarity = "positive", errf = NULL, returnXset = FALSE,
  intensity = "into", nSlaves = 0)
```

Arguments

- **files**: input files, given as a vector of strings containing the complete paths. All formats supported by XCMS can be used.
- **xset**: alternatively, one can present an object of class `xsAnnotate`, instead of the input files. In that case all of the XCMS and CAMERA tools: peak picking, CAMERA grouping, retention time correction and filling of missing peaks will be considered already done, and only annotation will be performed. This can be useful if one wants to compare different settings for annotation. If both files and xset are given, the former takes precedence.
settings
nested list of settings, to be used at individual steps of the pipeline. See the help of FEMsettings for details.

rtrange
An optional vector to slice the retention time range (in minutes).

mzrange
An optional vector to slice the mass spectrum

DB
database containing the spectra of the pure standards. For the description refer to the LCDBtest help.
polarity
The polarity of the analysis used for CAMERA annotation. Either "positive" or "negative".

errf
A model used to calculate the mass tolerance in ppm for each features on the bases of its mass, retention time and intensity. For further details refer to the help of AnnotateTable.

returnXset
logical: should the XCMS output be returned? If yes, this is a a list of xcmsSet objects, one element for each input file.

intensity
The intensity measure used in the output peaktabale. The available intensities are the ones provided by xcms. The default one is the total intensity (integral) of the feature on the detected chromatographic peak.

nSlaves
Number of cores to be used in peak picking.

Details
The mzrange and rtrange parameters are used to subset the mass and retention times considered in the analysis, reducing possible alignment problems at the extremes.
The error function calculates the expected m/z tolerance for feature annotation based on the mz and I values of each feature. To have a more complete description of the process refer to the help of AnnotateTable and the literature reference. An example is provided as well. Note that the use of "lm" is only one of the possible choices, but all kind of functional approximations working with the predict function could be used. If the error function is not provided the mass tolerance will be fixed to the value defined in the settings list.

Value
A list with three elements:

PeakTable
data.frame containing annotation information. Every line is a feature. The first columns are used to give information about these features, annotation, CAMERA, Chemspider IDs, etcetera. The last of these meta-information columns is always the one giving the retention time: "rt". After that, columns correspond to input files, and give measures of intensities for every single one of the features.

Annoation
The complete output of the AnnotateTable function.

Settings
The settings used in the pipeline.

xset
optionally, the xcmsSet/CAMERA object is returned, which can be useful for more detailed inspection of the results.

sessionInfo
The output of sessionInfo() to keep track of the sw version used for the processing

Author(s)
Pietro Franceschi
threeStdsDB

**Information on three chemical standards measured in GC-MS (liquid injection)**

**Description**

User information needed to build up an in-house database, the external database for cross-checking mass spectra, and the final database obtained with `createSTDdbGC` for three chemical standards.

**References**


**Examples**

data(LCresults)
names(LCresults)

```r
## Not run:
## load the settings for the analysis
data(FEMsettings)

## load the annotation DB
data(LCDBtest)

## load the Synapt Q-TOF error function
data(errf)

results.xset <- runLC(xset = LCresults$xset, settings = Synapt.RP,
                       DB = LCDBtest$DB)

## to start directly from the CDF files and include peak picking in the
## pipeline, simply provide the "files" argument rather than the "xset" argument

if (require(metaMSdata)) {
  ## get the path
cdfpath <- system.file("extdata", package = "metaMSdata")

  ## files
  files <- list.files(cdfpath, ".RP\.", full.names=TRUE)

  ## <---------- Use the Synapt Q-TOF error function ---------- >
  result.adaptive <- runLC(files, settings = Synapt.RP,
                           DB = LCDBtest$DB, errf = errf)

  ## <-------- Run the analysis with a fixed mass tolerance -------->
  result.fixed <- runLC(files, settings = Synapt.RP,
                        DB = LCDBtest$DB)
}

## End(Not run)
```
**Usage**

```r
data(threeStdsDB)
data(threeStdsInfo)
data(threeStdsNIST)
```

**Details**

Raw GC-MS data for the three standards, linalool, methyl salicylate and ethyl hexanoate, are available in package metaMSdata. Manual information required to build up an in-house database should be presented in the form of a data.frame, an example of which is `stdInfo`. Presenting this information to `createSTDdbGC` leads to processing of the raw data, and cross-checking with an external database containing mass spectra. An excerpt of the NIST database, containing only spectra for these three compounds, is available in `smallDB`. The final database that is then obtained can be inspected in object `DB`, which is a simple list of tags and values. For use as a reference database, several of these fields are mandatory. Currently these are `Name`, `monoMW`, `pspectrum` and `std.rt`.

**Source**

Georg Weingart

**Examples**

```r
data(threeStdsNIST)
length(smallDB)
data(threeStdsInfo)
stdInfo
data(threeStdsDB)
par(mfrow = c(3,1))
sapply(DB, function(x) plotPseudoSpectrum(x$pspectrum, main = x$Name))
```

---

**treat.DB**

*Scaling of pseudospectra in an msp object*

**Description**

This function transforms the “raw” data in an msp DB object into preprocessed data. Even if no scale and no mass.weight is applied, the intensities are still changed: scaled to unit length.

**Usage**

```r
treat.DB(DB, scale.p = c("sqrt", "none"), mass.weight = TRUE, isMSP = TRUE)
```

**Arguments**

- **DB**
  A database of spectra in the original intensity units.
- **scale.p**
  scale intensities with a square-root function, or leave them as they are. Default is to use scaling.
- **mass.weight**
  Logical: if TRUE, higher masses are given more weight.
- **isMSP**
  Logical: if TRUE, then the spectra are stored in slot `pspectrum`; otherwise the spectra are simply the list elements of `DB`. 

Value

The function returns the database, where intensities are scaled.

Author(s)

Ron Wehrens

Examples

data(threeStdsNIST) ## provides object smallDB, excerpt from NIST DB
smallDB.scaled <- treat.DB(smallDB)
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