Package ‘phenoTest’

April 26, 2017

Type Package

Title Tools to test association between gene expression and phenotype in a way that is efficient, structured, fast and scalable. We also provide tools to do GSEA (Gene set enrichment analysis) and copy number variation.

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Description Tools to test correlation between gene expression and phenotype in a way that is efficient, structured, fast and scalable. GSEA is also provided.

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Depends R (>= 2.12.0), Biobase, methods, annotate, Heatplus, BMA, ggplot2

Imports survival, limma, Hmisc, gplots, Category, AnnotationDbi, hopach, biomaRt, GSEABase, genefilter, xtable, annotate, mgcv, SNPchip, hgu133a.db, HTSanalyzeR, ellipse

Suggests GSEABase, KEGG.db, GO.db

Enhances parallel, org.Ce.eg.db, org.Mm.eg.db, org.Rn.eg.db, org.Hs.eg.db, org.Dm.eg.db

LazyLoad yes

biocViews Microarray, DifferentialExpression, MultipleComparison, Clustering, Classification

NeedsCompilation no

R topics documented:

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Test correlation between gene expression and phenotype.

Description

Test correlation between gene expression and phenotype.
phenoTest-package

Details
barplotSignatures

Summary plots for gene signature vs phenotype association

Description

Summarizes the univariate relationships between genes in one or more signatures and several phenotype variables, as summarized in epheno objects (which can be created with the ExpressionPhenoTest function).

By default barplotSignifSignatures performs a binomial test (binom.test from package stats) for each signature to see if the number of up up regulated and down regulated genes is different enough to be statistically different. When a reference gene set is provided we test if the proportions of up and down regulated genes of each gene set is different from the proportions in the reference gene set. This has been done with a chi-square test. When a reference gene set is provided and parameter testUpDown is TRUE (by default its FALSE) the number of genes corresponding to up and down regulated are compared with those of the reference gene set separately.

Usage

barplotSignatures(x, signatures, referenceSignature, alpha=.05, p.adjust.method='none', ylab, cex.text=1, ...)
barplotSignifSignatures(x, signatures, referenceSignature, testUpDown=FALSE, simulate.p.value = FALSE, B = 10^4, p.adjust.method='none', alpha=.05, ylab, ylim=ylim, cex.text=1, ...)

Arguments

x epheno object, as returned by ExpressionPhenoTest.

signatures List with each element corresponding to a signature. The gene names in each signature must match those in epheno.

referenceSignature If specified, the average fold change in each signature is compared to the average fold change in the signature referenceSignature.

testUpDown If set to TRUE, bars corresponding to up and down-regulated genes are compared with those of referenceSignature separately. This argument is ignored if referenceSignature is not specified.
barplotSignatures-methods

Methods for Function `barplotSignatures` in Package `phenoTest`

Description

Methods for function `barplotSignatures` in Package `phenoTest`. For more information read the function’s manual.

cex.text Character expansion for the text indicating the P-values. Ignored if referenceSignature is missing.
alpha Confidence levels for barplot error bars.
p.adjust.method P-value adjustment method, passed on to p.adjust.
simulate.p.value A logical indicating whether chi-square p-values should be computed by Monte Carlo simulation (passed on to chisq.test).
B Integer specifying the number of replicates in the Monte Carlo simulation (passed on to chisq.test).
ylab y-axis labels
ylim y-axis limits
... Other arguments to be passed on to boxplot.

Value

When a single signature is provided as input, a single plot assessing the association of that signature with all phenotype variables is created. If several signatures are provided, one separate plot is created for each phenotype variable.

Author(s)

Evarist Planet

Examples

```r
# create epheno
data(epheno)

# construct two signatures
data(epheno)
sign1 <- sample(featureNames(epheno))[1:20]
sign2 <- sample(featureNames(epheno))[1:15]
mySignature <- list(sign1,sign2)
names(mySignature) <- c('My first signature','My preferred signature')

# plot
barplotSignifSignatures(epheno[,Relapse],mySignature,alpha=0.05)
```
**ClusterPhenoTest**

Test association of clusters with phenotype.

**Description**

Test the associations between clusters that each sample belongs to (based on gene expression) and each phenotype.

**Usage**

`ClusterPhenoTest(x, cluster, vars2test, B=10^4, p.adjust.method='none')`
Arguments

x ExpressionSet with phenotype information stored in pData(x).
cluster variable of class character or factor telling at which cluster each sample belongs to.
vars2test list with components 'continuous', 'categorical', 'ordinal' and 'survival' indicating which phenotype variables should be tested. 'continuous', 'categorical' and 'ordinal' must be character vectors, 'survival' a matrix with columns named 'time' and 'event'. The names must match names in names(pData(x)).
B An integer specifying the number of replicates used in the chi-square Monte Carlo test (passed on to chisq.test).
p.adjust.method Method for P-value adjustment, passed on to p.adjust.

Details

Test association between the provided clusters and each phenotype.

For variables in vars2test\$continuous and vars2test\$ordinal a Kruskal-Wallis Rank Sum test is used; for vars2test\$categorical a chi-square test (with exact p-value if simulate.p.value is set to TRUE); for vars2test\$survival a Cox proportional hazards likelihood-ratio test.

Author(s)

David Rossell

Examples

#load data
data(eset)
eset

#construct vars2test
survival <- matrix(c("Relapse","Months2Relapse"),ncol=2,byrow=TRUE)
colnames(survival) <- c('event','time')
#add positive to have more than one category
pData(eset)[1:20,\'lymph.node.status\'] <- 'positive'
vars2test <- list(survival=survival,categorical=\'lymph.node.status\')
vars2test

#first half of the samples will be one cluster and the rest the other cluster
cluster <- c(rep('Cluster1',floor(ncol(eset)/2)),rep('Cluster2',ncol(eset)-floor(ncol(eset)/2)))

#test association
ClusterPhenoTest(eset,cluster,vars2test=vars2test)
epheno-class

Usage

data(epheno)

Format

The format is: Formal class 'epheno' [package "phenoTest"] with 8 slots ...

Examples

data(epheno)
## maybe str(epheno) ; plot(epheno) ...

Description

Object obtained with the ExpressionPhenoTest function. Contains FC, HR and pvals from testing
expression values of each gene against phenotypic variables.

Objects from the Class

Objects can be created by calls of the form new("epheno", assayData, phenoData, featureData, exprs, ...).
epheno-class

Slots

- `p.adjust.method`: Object of class "character" containing the multiple testing adjustment method used (if one was used).
- `approach`: Object of class "character" containing 'frequentist' or 'bayesian' depending on the user's selection.
- `assayData`: Object of class "AssayData" that is inherited from the ExpressionSet object used to create the epheno object.
- `phenoData`: Object of class "AnnotatedDataFrame" that contains information about the variables stored in the experimentData slot such as their class (continuous, categorical, etc) or type (mean, summaryDif, pval, etc).
- `featureData`: Object of class "AnnotatedDataFrame" that is inherited from the ExpressionSet object used to create the epheno object.
- `experimentData`: Object of class "MIAME" that is inherited from the ExpressionSet object used to create the epheno object.
- `annotation`: Object of class "character" that is inherited from the ExpressionSet object used to create the epheno object.
- `protocolData`: Object of class "AnnotatedDataFrame" that is inherited from the ExpressionSet object used to create the epheno object.
- `__classVersion__`: Object of class "Versions" that is inherited from the ExpressionSet object used to create the epheno object.

Extends

Class "ExpressionSet", directly. Class "eSet", by class "ExpressionSet", distance 2. Class "VersionedBiobase", by class "ExpressionSet", distance 3. Class "Versioned", by class "ExpressionSet", distance 4.

Methods

- `[ signature(x = "epheno", i = "ANY", j = "ANY")`: inherited from the ExpressionSet class.
- `dim signature(x = "epheno")`: inherited from the ExpressionSet class.
- `export2CSV signature(x = "epheno")`: ...
- `getFc signature(x = "epheno")`: getter for the fold changes.
- `getHr signature(x = "epheno")`: getter for the hazard ratios.
- `getMeans signature(x = "epheno")`: getter for the means.
- `getSignif signature(x = "epheno")`: getter for the pvalues or posterior probabilities.
- `getPvals signature(x = "epheno")`: getter for the pvalues.
- `getPostProbs signature(x = "epheno")`: getter for the posterior probabilities.
- `getSummaryDif signature(x = "epheno")`: getter that returns hazard ratios, fold changes and pvalues.
- `gseaSignatures signature(x = "epheno", signatures = "list")`: Used to compute GSEA. Please read the gseaSignatures manual.
- `logFcHr signature(x = "epheno")`: getter for the log of fold changes and hazard ratios.
- `p.adjust.method signature(x = "epheno")`: getter for the p value adjustment method that has been used.
- `phenoClass signature(x = "epheno")`: Returns the class off all variables.
- `phenoNames signature(x = "epheno")`: Returns the names of the tested phenotypes.
- `show signature(object = "epheno")`: Shows a brief overview of the object.
epheno2html

Author(s)
Evarist Planet

Examples
showClass("epheno")

epheno2html Create html files and plots from an epheno object.

Description
Creates html files and plots using an epheno object, which stores the association between a list of variables and gene expression.

Usage
epheno2html(x, epheno, outputdir, prefix = "", genelimit = 50, categories = 3, withPlots = TRUE, mc.cores = 1)

Arguments
x An object of class ExpressionSet (used to generate the epheno object) containing expression levels in exprs(x), phenotype information in pData(x) and annotation in annotation(x).
epheno an object produced by ExpressionPhenoTest. this object will contain univariate association between a list of phenotype variables and gene expression as well as p-values.
outputdir where to place files.
prefix will be used to add a text to the beginning of the files that will be created.
genelimit maximum number of genes on the list.
categories Number of categories used for continuous variables. It has to be the same as the one used for ExpressionPhenoTest.
withPlots when FALSE no plots will be produced. Makes the process faster.
mc.cores number of cores that will be used to run the process.

Author(s)
Evarist Planet

Examples
#Example on building homology tables for human.
#mart <- useMart("ensembl", "hsapiens_gene_ensembl")
#homol.symbol <- getLDS(attributes = c("entrezgene"),
  # mart = mart, attributesL = c("external_gene_id"),
  # martL = mart, filters = "entrezgene", values = entrezid)
#mart <- useMart("ensembl", "hsapiens_gene_ensembl")
#homol.geneName <- getLDS(attributes = c("entrezgene"),
  # mart = mart, attributesL = c("description"), martL = mart,
  # filters = "entrezgene", values = entrezid)
<table>
<thead>
<tr>
<th>eset</th>
<th>Example data.</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Description</strong></td>
<td>Example data of class ExpressionSet.</td>
</tr>
<tr>
<td><strong>Usage</strong></td>
<td>data(eset)</td>
</tr>
<tr>
<td><strong>Format</strong></td>
<td>The format is: Formal class 'ExpressionSet' [package &quot;Biobase&quot;] with 7 slots .@ assayData:environment: 0x1050d9390&gt;.@ phenoData:Formal class 'AnnotatedDataFrame' [package &quot;Biobase&quot;] with 4 slots .@ varMetadata :data.frame:; 7 obs. of 1 variable: .. .@ labelDescription: chr [1:7] NA NA NA NA ... .. .@ data :data.frame:; 286 obs. of 7 variables: .. .. ..$ PID : int [1:286] 3 5 6 7 8 9 11 14 15 17 ... .. .. ..$ GEOaccession : Factor w/ 286 levels &quot;GSM36777&quot;,&quot;GSM36778&quot;...: 17 20 21 22 24 25 58 59 60 61 ... .. .. ..$ lymph.node.status: chr [1:286] &quot;negative&quot; &quot;negative&quot; &quot;negative&quot; &quot;negative&quot; ... .. .. ..$ Months2Relapse : int [1:286] 101 118 9 106 37 125 109 14 99 137 ... .. .. ..$ ER.Status : num [1:286] 0 1 1 0 1 0 1 0 1 0 ... .. .. ..$ BrainRelapse : int [1:286] 0 0 0 0 0 0 0 0 0 0 ... .. .. ..@ dimLabels : chr [1:2] &quot;sampleNames&quot; &quot;sampleColumns&quot; .. .. ..@ <strong>classVersion</strong> Formal class 'Versions' [package &quot;Biobase&quot;] with 1 slots .@ featureData:Formal class 'AnnotatedDataFrame' [package &quot;Biobase&quot;] with 4 slots .@ varMetadata :data.frame:; 16 obs. of 3 variables: .. .. ..$ Column: chr [1:16] &quot;ID&quot; &quot;GB_ACC&quot; &quot;SPOT_ID&quot; &quot;Species Scientific Name&quot; ... .. .. ..$ Description: Facto...</td>
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nucleotide binding // inferred from electronic annotat|__truncated__,..: 23 26 27 40 81 18 39 71
74 69 ... ..@ dimLabels : chr [1:2] "featureNames" "featureColumns" .. ..@ __classVer-
sion__:Formal class 'Versions' [package "Biobase"] with 1 slots .. .. ..@ .Data:List of 1 ..
 .. ... .. ..$ : int [1:3] 1 1 0 ..@ experimentData :Formal class 'MIAME' [package "Biobase"]
with 13 slots .. .. ..@ name : chr "" .. .. ..@ lab : chr "" .. .. ..@ contact : chr "" .. .. ..@ title
: chr "" .. .. ..@ abstract : chr "" .. .. ..@ url : chr "" .. .. ..@ pubMedIds : chr "" .. .. ..@ samples : list() .. .. ..@ hybridizations : list() .. .. ..@ normControls : list() .. .. ..@ preprocess-
ing : list() .. .. ..@ other : list() .. .. ..@ ..@ __classVersion__:Formal class 'Versions' [package
"Biobase"] with 1 slots .. .. .. .. ..@ .Data:List of 1 .. .. .. .. .. ..$ : int [1:3] 1 1 0 ..@ annotation
: chr "hgu133a" ..@ protocolData :Formal class 'AnnotatedDataFrame' [package "Biobase"] with
4 slots ... .. ..@ varMetadata : 'data.frame': 0 obs. of 1 variable: .. .. .S labelDescription: chr(0)
... .. ..@ data : 'data.frame': 286 obs. of 0 variables .. .. ..@ dimLabels : chr [1:2] "sampleNames"
"sampleColumns" .. .. .. .. ..@ __classVersion__:Formal class 'Versions' [package "Biobase"] with 1
slots .. .. .. .. ..@ .Data:List of 1 .. .. .. .. .. ..$ : int [1:3] 1 1 0 ..@ __classVersion__:Formal class
'Versions' [package "Biobase"] with 1 slots .. .. .. .. .. ..@ .Data:List of 1 .. .. .. .. .. .. .. .. .. .. ..$ : int [1:3] 2 10 0 .. .. .. .. .. .. .. .. .. .. ..$: int [1:3] 1 0 0

References
Has been obtained from GEO (GSE2034). Only first 1000 probesets where stored (the rest has been removed).

Examples
data(eset)
## maybe str(eset) ; plot(eset) ...

eset.genelevel

Example data.

Description
Example data of class ExpressionSet with one probeset per gene.

Usage
data(eset.genelevel)

Format
The format is: Formal class 'ExpressionSet' [package "Biobase"] with 7 slots .@ assayData:<environment: 0x1050d9390> ..@ phenoData :Formal class 'AnnotatedDataFrame' [package "Biobase"]
with 4 slots ... ...@ varMetadata : 'data.frame': 7 obs. of 1 variable: .. .. .. ..$ labelDescription: chr [1:7] NA NA NA NA NA NA ... .. ..@ data : 'data.frame': 286 obs. of 7 variables: .. .. .. ..$ GEOaccession : Factor w/ 286 levels "GSM36777","GSM36778",...
: 17 20 21 22 24 25 58 59 60 61 ... .. .. .. ..$ lymph.node.status: chr [1:286] "negative" "negative" "negative" "negative" ... .. .. .. ..$ Months2Relapse : int [1:286] 101 118 9 106 125 109 14 99 137 ... .. .. .. ..$ Relapse : int [1:286] 0 0 1 0 1 0 0 0 0 ... .. .. .. ..$ BrainRelapse : int [1:286] 0 0 0 0 0 0 0 0 0 ... .. .. ..@ dimLabels : chr [1:2] "sampleNames" "sampleColumns" .. .. ..@ __classVersion__:Formal class 'Versions' [package "Biobase"] with 1 slots .. .. .. .. ..@ .Data:List of 1 .. .. .. .. .. ..$ : int [1:3] 1 1 0 ..@ featureData :Formal class 'AnnotatedDataFrame' [package
"Biobase"] with 4 slots ... @ varMetadata : 'data.frame': 16 obs. of 3 variables: ... . `$ Column : chr [1:16] "ID" "GB_ACC" "SPOT_ID" "Species Scientific Name" ... . `$ Description : Factor with 15 levels ","A gene symbol, when one is available (from UniGene)."...: 3 5 15 13 12 1 11 1 10 14 11 11 11 11 1 11 1 ... . `$ labelDescription: chr [1:16] NA NA NA NA ... .@ data : 'data.frame': 1000 obs. of 16 variables: ... . `$ GB_ACC : Factor with 21129 levels "AF052179","AF061832"...: 93 30 95 97 25 24 96 99 28 20 ... . `$ SPOT_ID : chr [1:1000] NA NA NA NA ... . `$ Species.Scientific.Name : Factor with 2 levels "Homo sapiens"...: 1 1 1 1 1 1 1 1 1 1 ... . `$ Annotation.Date : Factor with 2 levels "Jul 11, 2007"...: 1 1 1 1 1 1 1 1 1 1 ... . `$ Sequence.Type : Factor with 4 levels "Consensus sequence"...: 2 2 2 2 2 2 2 2 2 2 ... . `$ Sequence.Source : Factor with 3 levels "Affymetrix Proprietary Database"...: 1 2 1 2 1 2 1 1 2 1 ... . `$ Target.Description : Factor with 21363 levels "Consensus includes gb:A1656011 /FAEST /DB_XREF=gi:4739990 /DB_XREF=est:tt42e08.x1 /CLONE=IMAGE:2243462 /UG=Hs.116875 KIAA0156"...: 16 13 18 20 8 7 19 22 11 4 ... . `$ Representative/Public.ID : Factor with 14208 levels "ADP-ribosylation factor 1"...: 35 66 46 60 44 97 66 24 33 ... . `$ Gene.Symbol : Factor with 13074 levels "NM_000661 // NM_001024921"...: 37 45 41 52 1 37 45 41 52 1 ... . `$ Gene.Ontology.Biological.Process: Factor with 7245 levels ","0000049 // tRNA binding // non-traceable author statement /// 0000166 // nucleotide binding // inferred from genetic annotation ... . `$ Gene.Ontology.Cellular.Component: Factor with 4148 levels ","0000074 // regulation of progression through cell cycle // traceable author statement /// 0000049 // nucleic acid binding // inferred from genetic annotation ... . `$ Gene.Ontology.Molecular.Function: Factor with 7314 levels ","0000049 // tRNA binding // non-traceable author statement /// 0000166 // nucleotide binding // inferred from genetic annotation ... . `$ Gene.Ontology.Molecular.Function: Factor with 7314 levels ","0000049 // tRNA binding // non-traceable author statement /// 0000166 // nucleotide binding // inferred from genetic annotation ... . `$ Gene.Ontology.Molecular.Function: Factor with 7314 levels ","0000049 // tRNA binding // non-traceable author statement /// 0000166 // nucleotide binding // inferred from genetic annotation ... . `$ Gene.Ontology.Molecular.Function: Factor with 7314 levels ","0000049 // tRNA binding // non-traceable author statement /// 0000166 // nucleotide binding // inferred from genetic annotation ...

References

Has been obtained from GEO (GSE2034). Only first 1000 probesets where stored (the rest has been removed). After that the expressionSet was filtered to keep only one probeset per gene. We used the nsFilter function from package genefilter to accomplish this task.

Examples

```r
data(eset.genelevel)
## maybe str(eset.genelevel) ; plot(eset.genelevel) ...
```
eset2genelevel  
Filter ExpressionSet to keep one probe set per gene.

Description

Only one probe set per gene will be kept and entrezid will be used as gene identifier. nsFilter from package genefilter is used to select the probe set. The selected probe set is the one with higher interquartile range.

Usage

eset2genelevel(x)

Arguments

x  an object of class ExpressionSet.

Author(s)

Evarist Planet

See Also

genefilter::nsFilter

Examples

#data(eset)
#library(hgu133a.db)
#x <- eset2genelevel(eset)
#x
#head(featureNames(x))

export2CSV  
Export object to comma-separated text file.

Description

Saves object as comma-separated text file (CSV), using write.csv.

Usage

export2CSV(x, file, row.names=FALSE, ...)

Arguments

x  object to be exported. Currently methods for objects of class epheno (produced with ExpressionPhenoTest function) are implemented.

file  Name of the file where the results are to be saved

row.names  Passed on to write.csv

...  Other arguments to be passed on to write.csv
Description

Methods for function `export2CSV` in Package `phenoTest`

Methods

signature(x = "epheno") Exports summary differences (fold changes, hazard ratios), p-values and gene annotation (when available) to a CSV (comma separated value) file

ExpressionPhenoTest

Tests univariate association between a list of phenotype variables and gene expression.

Description

Tests univariate association between a list of phenotype variables and gene expression.

Usage

ExpressionPhenoTest(x, vars2test, adjustVars, p.adjust.method=’BH’,continuousCategories=3,mc.cores,approach=’frequentist’)
Details

If approach is ‘frequentist’: - The effect of both continuous, categorical and ordinal phenotype variables on gene expression levels are tested via lmFit. - For ordinal variables a single coefficient is used to test its effect on gene expression (trend test), which is then used to obtain a P-value (means for each category are reported in the output). - Gene expression effects on survival are tested via Cox proportional hazards model, as implemented in function ‘coxph’.

If approach is bayesian posterior probabilities are computed comparing the BIC of a model with the variable of interest as explanatory variable against the BIC of the same model without the variable of interest as explanatory variable.

Value

The output is an epheno object, which basically extends an ExpressionSet object. The means, fold changes, standardized hazard ratios and p-values are stored in the experimentData slot which is accessible with the exprs method. Information about the kind of information of each variable can be found in the phenoData slot which is accessible with the pData method.

There are several methods that can be used to access the information stored in an epheno object. For more information please type one of the following: getFc(x), getHr(x), getMeans(x), getSignif, getPvals(x), ...

Author(s)

David Rossell

References


Examples

```r
#load eset
data(eset)
eset

#prepare vars2test
survival <- matrix(c("Relapse","Months2Relapse"),ncol=2,byrow=TRUE)
colnames(survival) <- c('event','time')
vars2test <- list(survival=survival)

#run ExpressionPhenoTest
epheno <- ExpressionPhenoTest(eset,vars2test,p.adjust.method='none')
epheno
```

findCopyNumber

Find copy number regions using expression data in a similar way ACE does.

Description

Given enrichment scores between two groups of samples and the chromosomical positions of those enrichment scores this function finds areas where the enrichment is bigger/lower than expected if the positions were assigned at random. Plots of the regions and positions of the enriched regions are provided.
Usage

findCopyNumber(x, minGenes = 15, B = 100, p.adjust.method = "BH",
                pvalcutoff = 0.05, exprScorecutoff = NA, mc.cores = 1, useAllPerm = F,
genome = "hg19", chrLengths, sampleGenome = TRUE, useOneChr = FALSE,
useIntegrate = TRUE, plot=TRUE, minGenesPerChr=100)

Arguments

x
An object of class data.frame with gene or probe identifiers as row names and
the following columns: es (the enrichment score), chr (the chromosome where
the gene or probe belong to) and pos (position in the chromosome in megabases).
It can be obtained (from an epheno object) with the function getEsPositions.

minGenes
Minimum number of genes in a row that have to be enriched to mark the region
as enriched. Has to be bigger than 2.

B
Number of permutations that will be computed to calculate pvalues. If
useAllPerm is FALSE this value has to be bigger than 100. If useAllPerm is TRUE the com-
putations are much more expensive, therefore it is not recommended to use a B
bigger than 100.

p.adjust.method
P value adjustment method to be used. p.adjust.methods provides a list of avail-
able methods.

pvalcutoff
All genes with an adjusted p value lower than this parameter will be considered
enriched.

exprScorecutoff
Genes with a smoothed score that is not bigger (lower if the given number is
negative) than the specified value will not be considered significant.

mc.cores
Number of cores to be used in the computation. If mc.cores is bigger than 1 the
multicore library has to be loaded.

useAllPerm
If FALSE for each gene only permutations of genes that are in an area with
similar density (similar number of genes close to them) are used to compute
pvalues. If TRUE all permutations are used for each gene.

We recommend to use the option FALSE after having observed that the enrich-
ment can depend on the number of genes that are in the area.

We recommend to use the option TRUE if the positions of the enrichment score
are equidistant. Take into account that this option is much slower and needs less
permutations, therefore a smaller B is preferred.

See details for more info.

genome
Genome that will be used to draw cytobands.

chrLengths
An object of class numeric containing chromosome names as names. This
names have to be the same as the ones used in x$chr If missing the last po-
tion is used.

sampleGenome
If positions are sampled over the hole genome (across chromosomes) or within
each chromosome. This is TRUE by default.

useOneChr
Use only one chromosome to build the distribution under the null hypothesis
that genes/probes are not enriched. By default this is FALSE. The chromosome
that is used is chosen as follows: after removing small chromosomes we select
the one closest to the median quadratic distance to 0. Setting this parameter to
TRUE decreases processing time.
If we want to use integrate or pnorm to compute pvalues. The first does not assume any distribution for the distribution under the null hypothesis, the second assumes it is normally distributed.

If FALSE the function will make no plots.

Chromosomes with less than minGenesPerChr will be removed from the analysis.

Details

Enrichment scores can be either log fold changes, log hazard ratios, log variability ratios or any other score.

Within each chromosome a smoothed score for each gene is obtained via generalized additive models, the smoothing parameter for each chromosome being chosen via cross-validation. The obtained smoothing parameter of each chromosome will be used in permutations.

We assessed statistical significance by permuting the positions through the whole genome. If useAllPerm is FALSE for each gene the permutations of genes that are in an area with similar density (distance to tenth gene) are used to compute pvalues. We observed that genes with similar densities tend to have similar smoothed scores. If we set 1000 permutations ($B=1000$) scores are permuted through the whole genome 10 times (1000/10). For each smoothed score the permutations of the 100 smoothed scores with most similar density (distance to tenth gene) are used. Therefore each smoothed score will be compared to 1000 smoothed scores obtained from permutations.

If scores are at the same distance in the genome from each other (for instance when we have a score every fixed certain bases) the option useAllPerm=TRUE is recommended. In this case every smoothed score is compared to all smoothed scores obtained via permutations. In this case having 20,000 genes and setting the parameter $B=10$ would mean that the scores are permuted 10 times times through the whole genome, obtaining 200,000 permuted smoothed scores. Each observed smoothed score will be tested against the distribution of the 200,000 permuted smoothed scores.

Only regions with as many genes as told in minGenes being statistically significant (pvalue lower than parameter pvalcutoff) after adjusting pvalues with the method specified in p.adjust.method will be selected as enriched. If exprScorecutoff is different from NA, a gene to be statistically significant will need (additionally to the pvalue cutoff) to have a smoothed score bigger (lower if exprScorecutoff is negative) than the specified value.

Value

Plots all chromosomes and marks the enriched regions. Also returns a data.frame containing the positions of the enriched regions. This output can be passed by to the genesInArea function to obtain the names of the genes that are in each region.

Author(s)

Evarist Planet

See Also

getEsPositions, genesInArea

Examples

data(epheno)
phenoNames(epheno)
mypos <- getEsPositions(epheno,'Relapse')
genesInArea

Find genes that are in given areas.

Description

Combine the output of getEsPositions and findCopyNumber to see which genes are in the enriched areas.

Given areas of enrichment (obtained with findCopyNumber) and a set of genes or probes and their positions in the genome (obtained with getEsPositions) the function tells which genes fall in each area.

Usage

genesInArea(x, regions)

Arguments

x
An object of class data.frame with gene or probe identifiers as row names and the following columns: es (the enrichment score), chr (the chromosome where the gene or probe belong to) and pos (position in the chromosome in megabases). It can be obtained with the function getEsPositions.

regions
This is usually the output of findCopyNumber function.

Author(s)

Evarist Planet

See Also

genesInArea, findCopyNumber

test
get gseaSignatures’ elements

Subtract element’s of a gseaSignaturesSign or gseaSignaturesVar object (obtained using the gseaSignatures function).

Description

getEs returns ES (enrichment scores) getEsSim returns simulated ES (needed to compute pvals), getNes returns NES (normalized enrichment scores) and getFcHr returns the fold changes or hazard used to compute the ES, simulated ES and NES.

Usage

getEs(x)
getEsSim(x)
getNes(x)
geFcHr(x)

Arguments

x an gseaSignaturesSign or gseaSignaturesVar object. Those objects are obtained using the gseaSignatures function.

Author(s)

Evarist Planet

dgetEsPositions

Obtain chromosome positions for each gene.

Description

Given an object of class epheno obtain the gene positions on the genome.

Usage

dgetEsPositions(epheno, phenoName, organism = “human”, logEs = T, center = FALSE)

Arguments

epheno An object of class epheno usually obtained with ExpressionPhenoTest
phenoName The phenotype that we want to use. Has to be in phenoNames(epheno)
organism Has to be ‘human’ or ‘mouse’. The default is ‘human’.
logEs If the values have to be log scaled.
center If the values have to be genome centered. If TRUE the genome average will be substracted to every value.
**getGo**

**Details**

The output will usually be passed to findCopyNumber.

**Value**

An object of class `data.frame` will be returned containing 3 variables: `es` (enichment score for fold change or hazard ratio), `chr` (chromosome), `pos` (position in Mb). epheno's featureNames will be used as row names.

**Author(s)**

Evarist Planet

**Examples**

```r
data(epheno)
phenoNames(epheno)
mypos <- getEsPositions(epheno, 'Relapse')
head(mypos)
```

---

**getGo**

Create a list of gene sets based on GO pathways terms.

**Description**

This function creates a list of gene sets based on GO pathways terms. It is species-specific, and returns a list of gene sets, each of which is a character vector of Entrez gene identifiers.

This function is a wrapper to the function `GOGeneSets` from

**Usage**

```r
getGo(species = "Dm", ontologies = "MF")
```

**Arguments**

- **species**
  
a single character value specifying the species: "Dm" ("Drosophila_melanogaster"), "Hs" ("Homo_sapiens"), "Rn" ("Rattus_norvegicus"), "Mm" ("Mus_musculus") or "Ce" ("Caenorhabditis_elegans").

- **ontologies**
  
a single character value or a character vector specifying an ontology or multiple ontologies. The current version provides the following choices: "BP", "CC" and "MF"

**Details**

This function relies on the following packages: GSEABase, GO.db.

**Value**

a list of gene sets, with names as GO pathway IDs. Each gene set is a character vector of Entrez gene identifiers.
getKegg

Create a list of gene sets based on KEGG pathways terms.

Description

This function creates a list of gene sets based on KEGG pathways terms. It is species-specific, and returns a list of gene sets, each of which is a character vector of Entrez gene identifiers.

This function is a wrapper to the function KeggGeneSets from package HTSanalyzeR.

Usage

getKegg(species = "Dm")

Arguments

species a single character value specifying the species: "Dm" ("Drosophila_melanogaster"), "Hs" ("Homo_sapiens"), "Rn" ("Rattus_norvegicus"), "Mm" ("Mus_musculus") or "Ce" ("Caenorhabditis_elegans").

Details

This function relies on the following packages: GSEABase, KEGG.db.

Value

a list of gene sets, with names as KEGG pathway IDs. Each gene set is a character vector of Entrez gene identifiers.

Author(s)

Evarist Planet.

See Also

getGo
getters for the epheno object

Examples

```r
#library(KEGG.db)
#library(org.Hs.eg.db)
#kegg.Hs <- getKegg('Hs')
#str(kegg.Hs)
#kegg.Hs[1:2]
```

textual content:

`getters for the epheno object`  

*Getters for the epheno object:*

**Description**

getFc gets the fold changes. getHr gets the hazard ratios. getMeans gets the means. getPvals gets the p values. getPostProbs get the posterior probabilities. getSignif gets the pvalues or the posterior probabilities depending on the approach (frequentist or bayesian) that was used when the epheno object was created. getSummaryDif gets fold changes and hazard ratios. logFcHr gets the fold changes and hazard ratios after log scaling. p.adjust.method gets the p value adjustment method that was used when creating the object. phenoClass returns a data.frame telling the class (ordinal, continuous, categorical or survival) of each phenotype. phenoNames gets the phenotype names. approach gets the approach that was used (either frequentist or bayesian).

**Usage**

```r
getFc(x)
getHr(x)
getMeans(x)
getSignif(x)
getPvals(x)
getPostProbs(x)
getSummaryDif(x)
logFcHr(x)
p.adjust.method(x)
phenoClass(x)
phenoNames(x)
approach(x)
```

**Arguments**

`x` epheno object

**Author(s)**

Evarist Planet
getVars2test  

Get phenotypic variables that were tested.

Description

Returns an object containing the names of the variables that were tested when the epheno object was created. Will return an object of class list. Variables of the same type (categorical, survival, etc) will be in the same slot of the list. The slot names are the types of the variables.

Author(s)

Evarist Planet

Examples

data(epheno)
getVars2test(epheno)

getVars2test-methods

Methods for Function getVars2test in Package ‘phenoTest’

Description

Methods for function getVars2test in Package ‘phenoTest’. For more information read the function’s manual.

Methods

signature(x = "epheno") Method for an object of class epheno.

gsea

GSEA (Gene Set Enrichment Analysis).

Description

Computes the enrichment scores and simulated enrichment scores for each variable and signature. An important parameter of the function is logScale. Its default value is TRUE which means that by default the provided scores (i.e. fold changes, hazard ratios) will be log scaled. Remember to change this parameter to FALSE if your scores are already log scaled. The getEs, getEsSim, getFc, getHr and getFcHr methods can be used to access each subobject. For more information please visit the man pages of each method.

It also computes the NES (normalized enrichment score), p values and fdr (false discovery rate) for all variables and signatures. For an overview of the output use the summary method.

In case of providing gene sets which have more than 10 distinct lengths an approximation of the calculation of the enrichment score simulations (ESM) will be computed. The value of the ESM only depends on the length of the gene set. Therefore we compute the ESM over a grid of possible gene set lengths which are representative of the lengths of the provided gene sets. Then we fit a generalized additive model model with cubic splines to predict the NES value based on the length of every gene set. This provides a much faster approach that can be very useful when we need to run the software over a huge number of gene sets.
Usage

gsea(x, gsets, logScale=TRUE, absVals=FALSE, averageRepeats=FALSE, B=1000, mc.cores=1, test="perm", p.adjust.method="none", pval.comp.method="original", pval.smooth.tail=TRUE, minGenes=10, maxGenes=500, center=FALSE)

Arguments

x ePhenoTest, numeric or matrix object containing scores (hazard ratios or fold changes).
gsets character or list object containing the names of the genes that belong to each signature.
logScale if values should be log scaled.
absVals if TRUE fold changes and hazard ratios that are negative will be turned into positive before starting the process. This is useful when genes can go in both directions.
averageRepeats if x is of class numeric and has repeated names (several measures for some individual names) we can average the measures of the same names.
B number of simulations to perform.
mc.cores number of processors to use.
test the test that will be used. 'perm' stands for the permutation based method, 'wilcox' stands for the wilcoxon test (this is the fastest one) and 'ttestperm' stands for permutation t test.
p.adjust.method p adjustment method to be used. Common options are 'BH', 'BY', 'bonferroni' or 'none'. All available options and their explanations can be found on the p.adjust function manual.
pval.comp.method the p value computation method. Has to be one of 'signed' or 'original'. The default one is 'original'. See details for more information.
pval.smooth.tail if we want to estimate the tail of the distribution where the pvalues will be generated.
minGenes gene sets with less than minGenes genes will be removed from the analysis.
maxGenes gene sets with more than maxGenes genes will be removed from the analysis.
center if we want to center scores (fold changes or hazard ratios). The following is will be done: x = x-mean(x).

Details

The following preprocessing was done on the provided scores (i.e. fold changes, hazard ratios) to avoid errors during the enrichment score computation: -When having two scores with the same name its average was used. -Zeros were removed. -Scores without names (which can not be in any signature) removed. -Non complete cases (i.e. NAs, NaNs) were removed. ES score was calculated for each signature and variable (see references). If parameter test is 'perm' the signature was permuted and the ES score was recalculated (this happened B times for each variable, 1000 by default). If test is 'wilcox' a wilcoxon test in which we test the fact that the average value of the genes that do belong to our signature is different from the average value of the genes that do not
belong to our signature will be performed. If test is ‘tperm’ a permutation t-test will be used. Take into account that the final plot will be different when ‘wilcox’ is used.

The simulated enrichment scores and the calculated one are used to find the p value. P value calculation depends on the parameter pval.comp.method. The default value is ‘original’. In ‘original’ we are simply computing the proportion of absolute simulated ES which are larger than the observed absolute ES. In ‘signed’ we are computing the proportion of simulated ES which are larger than the observed ES (in case of having positive enrichment score) and the proportion of simulated ES which are smaller than the observed ES (in case of having negative enrichment score).

Author(s)

Evarist Planet

References


See Also

gsea.go, gsea.kegg

Examples

#load epheno object
data(epheno)
epheno

#we construct two signatures
sign1 <- sample(featureNames(epheno))[1:20]
sign2 <- sample(featureNames(epheno))[50:75]
mySignature <- list(sign1,sign2)
names(mySignature) <- c('My first signature','My preferred signature')

#run gsea functions
gseaData <- gsea(x=epheno,gsets=mySignature,B=100,mc.cores=1)
my.summary <- summary(gseaData)
my.summary
#plot(gseaData)

---

**gsea.kegg**  
*Perform Gene Set Enrichment Analysis (GSEA) of Gene Ontologies (GO) and Kegg gene sets.*

**Description**

The function obtains the GO or Kegg gene sets and performs GSEA analysis as implemented in the gsea function.
Usage

gsea.go(x, species='Hs', ontologies='MF', logScale=TRUE, absVals=FALSE, 
averageRepeats=FALSE, B=1000, mc.cores=1, test="perm", 
p.adjust.method="none", pval.comp.method="original", 
pval.smooth.tail=TRUE, minGenes=10, maxGenes=500, center=FALSE)
gsea.kegg(x, species='Hs', logScale=TRUE, absVals=FALSE, 
averageRepeats=FALSE, B=1000, mc.cores=1, test="perm", 
p.adjust.method="none", pval.comp.method="original", 
pval.smooth.tail=TRUE, minGenes=10, maxGenes=500, center=FALSE)

Arguments

x ePhenoTest, numeric or matrix object containing scores (hazard ratios or fold changes).

species a single character value specifying the species: "Dm" ("Drosophila_melanogaster"), "Hs" ("Homo_sapiens"), "Rn" ("Rattus_norvegicus"), "Mm" ("Mus_musculus") or "Ce" ("Caenorhabditis_elegans").

ontologies a single character value or a character vector specifying an ontology or multiple ontologies. The current version provides the following choices: "BP", "CC" and "MF".

logScale if values should be log scaled.

absVals if TRUE fold changes and hazard ratios that are negative will be turned into positive before starting the process. This is useful when genes can go in both directions.

averageRepeats if x is of class numeric and has repeated names (several measures for some individual names) we can average the measures of the same names.

B number of simulations to perform.

mc.cores number of processors to use.

test the test that will be used. 'perm' stands for the permutation based method, 'wilcox' stands for the wilcoxon test (this is the fastest one) and 'ttperm' stands for permutation t test.

p.adjust.method p adjustment method to be used. Common options are 'BH', 'BY', 'bonferroni' or 'none'. All available options and their explanations can be found on the p.adjust function manual.

pval.comp.method the p value computation method. Has to be one of 'signed' or 'original'. The default one is 'original'. See details for more information.

pval.smooth.tail if we want to estimate the tail of the ditribution where the pvalues will be generated.

minGenes gene sets with less than minGenes genes will be removed from the analysis.

maxGenes gene sets with more than maxGenes genes will be removed from the analysis.

center if we want to center scores (fold changes or hazard ratios). The following is will be done: \( x = x - \text{mean}(x) \).
Details

This function relies on the following packages: GSEABase, GO.db.

For more information about how the gene sets are obtained see the man page of the functions getGo and/or getKegg. For more information about the implemented GSEA see the man page of the function gsea.

Value

a list of gene sets, with names as GO pathway IDs. Each gene set is a character vector of Entrez gene identifiers.

Author(s)

Evarist Planet.

See Also

getGo

Examples

###load libs
#library(KEGG.db)
#library(org.Hs.eg.db)

###get data
#data(eset.genelevel)
#eset.genelevel

###prepare vars2test
#survival <- matrix(c("Relapse","Months2Relapse"),ncol=2,byrow=TRUE)
#colnames(survival) <- c('event','time')
#vars2test <- list(survival=survival,categorical='ER.Status')

###run ExpressionPhenoTest
#epheno <- ExpressionPhenoTest(eset.genelevel,vars2test,p.adjust.method='none')
#epheno

###run gsea with kegg gene sets.
#gseaData <- gsea.kegg(epheno[,1],'Hs')
#summary(gseaData)
#plot(gseaData[[1]],gseaData[[2]],selGsets='hsa04062')
Usage

\[
gsea2html(gseaData, epheno, variable, title = "", path, file, digits = 3, plotEs = FALSE, limit=100)
\]

Arguments

- **gseaData**: an object of class `gseaData`.
- **epheno**: the object of class `epheno` that was used to create `gseaData`.
- **variable**: variable that we are interested in.
- **title**: title that will be shown on top of the table.
- **path**: directory where we want to store the html files.
- **file**: filename.
- **digits**: Number of decimal digits that will be shown on the table.
- **plotEs**: if this is TRUE enrichment score plots will be plotted instead of normalized enrichment score plots.
- **limit**: maximum number of gene sets that will be exported.

Details

This function produces a browseable version of the table that we can obtain with `summary(gseaData)`. We will obtain one plot per NES (or ES) and we will be able to see which genes belong to each gene set and the values they have in the `epheno` object.
gseaData-class

Author(s)
Evarist Planet

Examples

#WITH PROBESET AS IDENTIFIER
data(eset)
data(epheno)

set.seed(777)
sign1 <- sample(featureNames(eset))[1:20]
sign2 <- sample(featureNames(eset))[1:50]
mySignature <- list(sign1,sign2)
names(mySignature) <- c('My first signature','Another signature')
mySignature

mygsea <- gsea(x=epheno[,1],gsets=mySignature,B=100,p.adjust='BH')
summary(mygsea)

#following line has been commented to prevent the creation of files
#gsea2html(gseaData=mygsea,epheno=epheno,variable=phenoNames(epheno)[1],title='My test',path='~/Desktop',file='myGSEA.html')

#WITH ENTREZID AS IDENTIFIER
data(eset.genelevel)
eset.genelevel

set.seed(777)
sign1 <- sample(featureNames(eset.genelevel))[1:20]
sign2 <- sample(featureNames(eset.genelevel))[1:50]
mySignature.genelevel <- list(sign1,sign2)
names(mySignature.genelevel) <- c('My first signature','Another signature')
mySignature.genelevel

epheno.genelevel <- ExpressionPhenoTest(eset.genelevel,vars2test=list(categorical='lymph.node.status'))
mygsea.genelevel <- gsea(x=epheno.genelevel,gsets=mySignature.genelevel,B=100,p.adjust='BH')
summary(mygsea.genelevel)

#following line has been commented to prevent the creation of files
#gsea2html(gseaData=mygsea.genelevel,epheno=epheno.genelevel,variable=phenoNames(epheno.genelevel),title='My test (at genelevel)')

gseaData-class

Class "gseaData"

Description
This class is an ES (enrichment score) and ES.sim (simulated enrichment score) container that will be used in the GSEA (Gene Set Enrichment Analysis) process. There is one container for every gene signature.

Objects from the Class
Objects can be created by calls of the form new("gseaData", ...).
gseaSignatures

Slots

.Data: Object of class "list".
gseaSignaturesSign: Object of class "gseaSignaturesSign" or "gseaSignaturesVar".
gseaSignificanceSign: Object of class "gseaSignificanceSign" or "gseaSignificanceVar".

Extends

Class "list", from data part. Class "vector", by class "list", distance 2. Class "AssayData", by class "list", distance 2.

Methods

getEs signature(x = "gseaData"): Returns the enrichment scores.
getEsSim signature(x = "gseaData"): Returns the simulated enrichment scores (the ones obtained after permutations).
getFcHr signature(x = "gseaData"): Returns the fold change and/or the hazard ratio that were used to compute the enrichment scores.

Author(s)

Evarist Planet

Examples

showClass("gseaSignaturesSign")

Usage

gseaSignatures(x, gsets, logScale=TRUE, absVals=FALSE, averageRepeats=FALSE, B=1000, mc.cores=1, test='perm', minGenes=10, maxGenes=500, center=FALSE)

Description

This function has been deprecated. You could better use gsea instead.

This function computes the first step in the process of obtaining a GSEA-like plot. It computes the enrichment scores and simulated enrichment scores for each variable and signature. The output will usually be used as input for the gseaSignificance function. An important parameter of the function is logScale. Its default value is TRUE which means that by default the provided scores (i.e. fold changes, hazard ratios) will be log scaled. Remember to change this parameter to FALSE if your scores are already log scaled. The getEs, getEsSim, getFc, getHr and getFcHr methods can be used to access each subobject. For more information please visit the man pages of each method.
gseaSignatures

Arguments

- **x**: ePhenoTest, numeric or matrix object containing hazard ratios or fold changes.
- **gsets**: character or list object containing the names of the genes that belong to each signature.
- **logScale**: if values should be log scaled.
- **absVals**: if TRUE, fold changes and hazard ratios that are negative will be turned into positive before starting the process. This is useful when genes can go in both directions.
- **averageRepeats**: if x is of class numeric and has repeated names (several measures for some individual names) we can average the measures of the same names.
- **B**: number of simulations to perform.
- **mc.cores**: number of processors to use.
- **test**: the test that will be used. ‘perm’ stands for the permutation based method, ‘wilcox’ stands for the wilcoxon test (this is the fastest one) and ‘ttperm’ stands for permutation t test.
- **minGenes**: gene sets with less than minGenes genes will be removed from the analysis.
- **maxGenes**: gene sets with more than maxGenes genes will be removed from the analysis.
- **center**: if we want to center scores (fold changes or hazard ratios). The following is will be done: x = x - mean(x).

Details

The following preprocessing was done on the provided scores (i.e. fold changes, hazard ratios) to avoid errors during the enrichment score computation: - When having two scores with the same name its average was used. - Zeros were removed. - Scores without names (which can not be in any signature) removed. - Non complete cases (i.e. NAs, NaNs) were removed. ES score was calculated for each signature and variable (see references). If parameter test is ‘perm’ the signature was permuted and the ES score was recalculated (this happened B times for each variable, 1000 by default). If test is ‘wilcox’ a wilcoxon test in which we test the fact that the average value of the genes that do belong to our signature is different from the average value of the genes that do not belong to our signature was performed. If test is ‘ttperm’ a permutation t-test will be used. Take into account that the final plot will be different when ‘wilcox’ is used.

Author(s)

Evarist Planet

References


Examples

```r
# load epheno object
data(epheno)
epheno

# we construct two signatures
sign1 <- sample(featureNames(epheno))[1:20]
```
sign2 <- sample(featureNames(epheno))[50:75]
mySignature <- list(sign1,sign2)
names(mySignature) <- c('My first signature','My preferred signature')

#run gsea functions
#my.gseaSignatures <- gseaSignatures(x=epheno,signatures=mySignature,B=100,mc.cores=1)
#my.gseaSignificance <- gseaSignificance(my.gseaSignatures)
#my.summary <- summary(my.gseaSignificance)
#my.summary
#plot(my.gseaSignatures,my.gseaSignificance)

---

**gseaSignatures-class**  
Class "gseaSignatures" ES and EsSim container.

**Description**  
This object contains de ES (enrichment scores) and simulated ES that will be used in the GSEA (Gene Set Enrichment Analysis) process.

**Objects from the Class**  
Objects can be created by calls of the form `new("gseaSignatures", ...)`.  

**Slots**  
- `.Data`: Object of class "list".  
- `es`: Object of class "numeric" Contains the observed enrichment scores. The ones that were computed from the data without permuting anything.  
- `es.sim`: Object of class "numeric" Contains the enrichment score that were obtained after permutations.  
- `signature`: Object of class "numeric" The subset of genes we are interested in.

**Extends**  
Class "list", from data part. Class "vector". by class "list", distance 2. Class "AssayData", by class "list", distance 2.

**Methods**  
No methods defined with class "gseaSignatures" in the signature.

**Author(s)**  
Evarist Planet

**Examples**  
```r
showClass("gseaSignatures")
```
Methods for Function `gseaSignatures` in Package 'phenoTest'

**Description**

Methods for function `gseaSignatures` in Package 'phenoTest'. For more information read the function’s manual.

**Methods**

- `signature(x = "ANY", signatures = "character")` Method for signature of class character.
- `signature(x = "ANY", signatures = "GeneSet")` Method for signature of class character.
- `signature(x = "epheno", signatures = "list")` Method for an epheno object and several signatures stored in an object of class list.
- `signature(x = "matrix", signatures = "GeneSetCollection")` Method for a matrix object and several signatures stored in an object of class GeneSetCollection.
- `signature(x = "epheno", signatures = "GeneSetCollection")` Method for an epheno object and several signatures stored in an object of class GeneSetCollection.
- `signature(x = "numeric", signatures = "GeneSetCollection")` Method for a numeric object and several signatures stored in an object of class GeneSetCollection.
- `signature(x = "matrix", signatures = "list")` Method for a matrix object and several signatures stored in an object of class list.
- `signature(x = "numeric", signatures = "list")` Method for a numeric object and several signatures stored in an object of class list.

**gseaSignaturesSign-class**

Class "gseaSignaturesSign"

**Description**

This class is an ES (enrichment score) and ES.sim (simulated enrichment score) container that will be used in the GSEA (Gene Set Enrichment Analysis) process. There is one container for every gene signature.

**Objects from the Class**

Objects can be created by calls of the form `new("gseaSignaturesSign", ...).`
Slots

- **.Data**: Object of class "list".
- **gseaSignatures**: Object of class "gseaSignatures" This is the object that will contain the ES and ES.sim.
- **es.sim.gam**: Object of class "matrix" enrichment scores computed with the gam method.
- **fc.hr**: Object of class "character" fold change or hazard ratio used to compute the enrichment scores.
- **s**: Object of class "logical" The subset of genes we are interested in.
- **test**: Object of class "character" The statistical test that will be used.

Extends

- Class "list", from data part. Class "vector", by class "list", distance 2. Class "AssayData", by class "list", distance 2.

Methods

- **getEs** signature(x = "gseaSignaturesSign"): Returns the enrichment scores.
- **getEsSim** signature(x = "gseaSignaturesSign"): Returns the simulated enrichment scores (the ones obtained after permutations).
- **getFcHr** signature(x = "gseaSignaturesSign"): Returns the fold change and/or the hazard ratio that were used to compute the enrichment scores.
- **gseaSignificance** signature(x = "gseaSignaturesSign"): This is the next step in the process of performing GSEA. This function will test if the gene sets are enriched.

Author(s)

- Evarist Planet

Examples

- `showClass("gseaSignaturesSign")`

---

**gseaSignaturesVar-class**

*Class "gseaSignaturesVar"

Description

This class is an ES (enrichment score) and ES.sim (simulated enrichment score) container that will be used in the GSEA (Gene Set Enrichment Analysis) process. There is one container for every phenotype. Every one of this containers (of class gseaSignaturesSign) is a container itself and has the enrichment scores of all signatures. GseaSignaturesVar contains one element per phenotype (phenotypic variable). Every one of this elements is of class gseaSignaturesSign and contains one element per signature.

Objects from the Class

Objects can be created by calls of the form `new("gseaSignaturesVar", ...).`
gseaSignificance

Slots

.Data: Object of class "list".

  gseaSignatures: Object of class "gseaSignaturesSign". This object contains the enrichment scores and other elements that will be used in the GSEA process.

Extends

Class "list", from data part. Class "vector", by class "list", distance 2. Class "AssayData", by class "list", distance 2.

Methods

  getEs signature(x = "gseaSignaturesVar"): Returns the enrichment scores.

  getEsSim signature(x = "gseaSignaturesVar"): Returns the simulated enrichment scores (the ones obtained after permutations).

  getFcHr signature(x = "gseaSignaturesVar"): Returns the fold change and/or the hazard ratio that were used to compute the enrichment scores.

  gseaSignificance signature(x = "gseaSignaturesVar"): This is the next step in the process of performing GSEA. This function will test if the gene sets are enriched.

Author(s)

  Evarist Planet

Examples

  showClass("gseaSignaturesVar")

---

Description

This function has been deprecated. You could better use gsea instead.

This function performs the second step in the process of obtaining a GSEA-like plot. It computes the NES (normalized enrichment score), p values and fdr (false discovery rate) for all variables and signatures. A gseaSignaturesSign or gseaSignaturesVar object will be needed as input (these objects can be obtained with the gseaSignatures function). For an overview of the output use the summary method. The next step after using the gseaSignificance function would be using the plot method.

Usage

  gseaSignificance(x, p.adjust.method='none', pval.comp.method='original', pval.smooth.tail=TRUE)
Arguments

x  gseaSignaturesSign or gseaSignaturesVar object obtained with the gseaSignatures method. This object contains the enrichment scores, the simulated enrichment scores and the fold changes or hazard ratios.

p.adjust.method  
p adjustment method to be used. Common options are 'BH', 'BY', 'bonferroni' or 'none'. All available options and their explanations can be found on the p.adjust function manual.

pval.comp.method  
the p value computation method. Has to be one of 'signed' or 'original'. The default one is 'original'. See details for more information.

pval.smooth.tail  
if we want to estimate the tail of the distribution where the pvalues will be generated.

Details

The simulated enrichment scores and the calculated one are used to find the p value. P value calculation depends on the parameter pval.comp.method. The default value is 'original'. In 'original' we are simply computing the proportion of absolute simulated ES which are larger than the observed absolute ES. In 'signed' we are computing the proportion of simulated ES which are larger than the observed ES (in case of having positive enrichment score) and the proportion of simulated ES which are smaller than the observed ES (in case of having negative enrichment score).

Author(s)

Evarist Planet

References


Examples

# for examples see the help file of gseaSignatures: ?gseaSignatures

---

gseaSignificanceSign-class

Class "gseaSignificanceSign"

Description

This object contains the results of the test of enrichment that was performed on each gene set. There is one container for every gene signature.
Objects from the Class

Objects can be created by calls of the form `new("gseaSignificanceSign", ...)`.

Slots

.Data: Object of class "list".
gseaSignificance: Object of class "matrix" Contains the statistics. Use the `summary` method to access this information.
p.adjust.method: Object of class "character" The p-value adjustment method that was used.

Extends

Class "list", from data part. Class "vector", by class "list", distance 2. Class "AssayData", by class "list", distance 2.

Methods

No methods defined with class "gseaSignificanceSign" in the signature.

Author(s)

Evarist Planet

Examples

`showClass("gseaSignificanceSign")`

---

**gseaSignificanceVar-class**

*Class "gseaSignificanceVar"*

**Description**

This object contains the results of the test of enrichment that was performed on each gene set and phenotype. There is one container for every phenotype. Every one of this containers (of class `gseaSignificanceSign`) is a container itself and has the results of the tests for all signatures. `GseaSignificanceVar` contains one element per phenotype (phenotypic variable). Every one of this elements is of class `gseaSignificanceSign` and contains one element per signature.

Objects from the Class

Objects can be created by calls of the form `new("gseaSignificanceVar", ...)`.

Slots

.Data: Object of class "list".
gseaSignificance: Object of class "gseaSignificanceSign" This object contains the results of the tests.
**heatmapPhenoTest** 39

**Extends**

Class "list", from data part. Class "vector", by class "list", distance 2. Class "AssayData", by class "list", distance 2.

**Methods**

No methods defined with class "gseaSignificanceVar" in the signature.

**Author(s)**

Evarist Planet

**Examples**

```
showClass("gseaSignificanceVar")
```

**Description**

Show the associations between clusters that each sample belongs to and each phenotype in a heatmap and/or a Kaplan-Meier plot.

**Usage**

```
heatmapPhenoTest(x, signatures, vars2test, probes2genes = FALSE, 
filterVar, filteralpha = 0.05, distCol = "pearson", nClust = 2, distRow 
= "cor", p.adjust.method = "none", simulate.p.value = FALSE, B = 10^5, 
linkage = "average", equalize = FALSE, center = TRUE, col, survCol, 
heat.kaplan="both", ...)
```

**Arguments**

- **x**: ExpressionSet with phenotype information stored in pData(x).
- **signatures**: Either character vector or list of character vectors with gene sets to be used to draw heatmaps (gene names should match those in featureNames(x)). A separate heatmap will be produced for each element in the list.
- **vars2test**: list with components 'continuous', 'categorical', 'ordinal' and 'survival' indicating which phenotype variables should be tested. 'continuous', 'categorical' and 'ordinal' must be character vectors, 'survival' a matrix with columns named 'time' and 'event'. The names must match names in names(pData(x)).
- **probes2genes**: If set to TRUE a single probe is selected for each gene. nSFilter is used to select the probe with highest inter-quartile range.
- **filterVar**: If specified, only genes with significant differences in the variable filterVar will be displayed in the heatmap. Note that this option will not affect the sample clustering, as this is obtained using both significant and non-significant genes.
- **filteralpha**: Significance level for the filtering based on filterVar.
heatmapPhenoTest

distCol  Distance metric used to cluster columns (e.g. patients/samples). Can take any value accepted by dist. Pearson and Spearman correlations are also allowed. Write 'spearman' or 'pearson' to use them.

nClust  Number of desired clusters.

distRow  Distance metric used to cluster rows (e.g. genes). Can take any value accepted by distancematrix.

p.adjust.method  Method for P-value adjustment, passed on to p.adjust.

simulate.p.value  If set to FALSE the chi-square test p-values are computed using asymptotics, otherwise a simulation is used (see chisq.test for details).

B  An integer specifying the number of replicates used in the chi-square Monte Carlo test (passed on to chisq.test).

linkage  Linkage used for clustering. Must be either 'complete', 'average' or 'minimum'.

equalize  Should color codes be equalized between genes, i.e. all genes present the same range of colors. Passed on to heatmap_plus.

center  centering is done by subtracting the column means (omitting NAs).

col  Color scheme to be used for heatmap. Defaults to a green/red scheme designed to look nice for microarray data.

survCol  Colors for the Kaplan-Meier survival curves.

heat.kaplan  can be "heat" if we want to plot a heatmap, "kaplan" if we want to plot a kaplan-meier or "both" if we want both of them.

...  Other arguments for the survival plot, e.g. lty etc.

Details

Makes two clusters of samples based on the expression levels of the genes from the given signature and plots a heatmap and/or a Kaplan-Meier showing the association between belonging to one cluster or the other and each phenotype.

For variables in vars2test\$continuous and vars2test\$ordinal a Kruskal-Wallis Rank Sum test is used; for vars2test\$categorical a chi-square test (with exact p-value if simulate.p.value is set to TRUE); for var2test\$survival a Cox proportional hazards likelihood-ratio test.

Author(s)

David Rossell

Examples

# load data
data(eset)
eset

# construct vars2test
survival <- matrix(c("Relapse","Months2Relapse"),nrow=2,byrow=TRUE)
colnames(survival) <- c('event','time')
vars2test <- list(survival=survival)
vars2test

# construct a signature
```
sign <- sample(featureNames(eset))[1:20]

# make plot
heatmapPhenoTest(eset, sign, vars2test=vars2test, heat.kaplan='heat')
heatmapPhenoTest(eset, sign, vars2test=vars2test, heat.kaplan='kaplan')
```

---

Methods for Function `heatmapPhenoTest` in Package `phenoTest`

**Description**

Methods for function `heatmapPhenoTest` in Package `phenoTest`. For more information read the function’s manual.

**Methods**

- `signature(x = "ExpressionSet", signatures = "character")` Method for an `ExpressionSet` object and one signature stored in an object of class character.
- `signature(x = "ExpressionSet", signatures = "list")` Method for an `ExpressionSet` object and several signatures stored in an object of class list.
- `signature(x = "ExpressionSet", signatures = "missing")` Method for an `ExpressionSet` object and no signatures.
- `signature(x = "ExpressionSet", signatures = "GeneSet")` Method for an `ExpressionSet` object and one signature stored in an object of class `GeneSet`.
- `signature(x = "ExpressionSet", signatures = "GeneSetCollection")` Method for an `ExpressionSet` object and several signatures stored in an object of class `GeneSetCollection`.

---

**pAdjust**

Adjust p values of an epheno object.

**Description**

Adjusts the p values of an epheno object. The `p.adjust` function will be used. For more information read the `p.adjust` function’s help.

**Usage**

```
pAdjust(x, method = "BH")
```

**Arguments**

- `x` an epheno object.
- `method` the correction method that will be used. See the `p.adjust` help for more info about the methods.

**Author(s)**

Evarist Planet
Examples

```r
# load epheno object
data(epheno)
epheno

# Adjust p-value
p.adjust.method(epheno)
epheno <- pAdjust(epheno, method="BH")
p.adjust.method(epheno)
```

---

**pAdjust-methods**

Methods for Function `pAdjust` in Package ‘phenoTest’

**Description**

Methods for function `pAdjust` in Package ‘phenoTest’. This function adjusts the p-values of an epheno object. For more information read the function’s manual.

**Methods**

`signature(x = "epheno")` Adjusts the p-values of an epheno object.

**Author(s)**

Evarist Planet

---

**pca**

Principal components plot.

**Description**

Creates a Principal Components plot where we can show paired samples, and confidence intervals for the mean of every group of interest. We can also choose the component or components we want to plot.

**Usage**

`pca(x, group, group2, pair, names, ellipse = FALSE, main = "", components = c(1, 2))`

**Arguments**

- `x` An object of class ExpressionSet.
- `group` Variable in `pData(x)` that contains the groups of interest. Samples of the same group will be plotted with the same color.
- `group2` Variable in `pData(x)` that contains secondary groups of interest. Sample of the same secondary group of interest will be plotted with the same symbol.
- `pair` Variable in `pData(x)` that contains the information about the pairs of data. Those pairs will be joined by a line.
**Description**

Builds a GSEA plot using a gseaData object. gseaData object can be obtained with the gsea function.

**Usage**

`plot.gseaData(x, selGsets, selVars, ...)`

**Arguments**

- `x` this has to be of class gseaData
- `selGsets` object of class character containing the names of the gene sets that we want to plot.
- `selVars` object of class character containing the names of the variables that we want to plot.
- `...` Arguments to be passed to `plot`.

**Author(s)**

Evarist Planet
The `plot.gseaSignatures` function builds a GSEA plot using a `gseaSignatures` object (one of `gseaSignaturesSign` or `gseaSignaturesVar` obtained with the `gseaSignatures` function) and a `gseaSignificance` object (one of `gseaSignificanceSign` or `gseaSignificanceVar` obtained with the `gseaSignificance` function).

**Usage**

```r
plot.gseaSignatures(x, gseaSignificance, es.ylim, nes.ylim, es.nes="both", ...)```

**Arguments**

- `x`: object of class `gseaSignaturesSign` or `gseaSignaturesVar`.
- `gseaSignificance`: object of class `gseaSignificanceSign` or `gseaSignificanceVar`.
- `es.ylim`: ylim values for the ES plot.
- `nes.ylim`: ylim values for the NES plot.
- `es.nes`: can be "es" if we want to plot enrichment score, "nes" if we want to plot normalised enrichment scores or "both" if we want to plot them both.
- `...`: Arguments to be passed to `plot`.

**Author(s)**

Evarist Planet

**References**


**Examples**

# for examples see the help file of gseaSignatures: ?gseaSignatures
Methods for Function show in Package ‘methods’.

Methods

Methods for function show in Package ‘methods’.

Methods

```r
signature(object = "AnnotatedDataFrame") Will show an object of class AnnotatedDataFrame.
signature(object = "ANY") Will show an object of class ANY.
signature(object = "classRepresentation") Will show an object of class classRepresentation.
signature(object = "container") Will show an object of class container.
signature(object = "epheno") Will show an object of class epheno.
signature(object = "eSet") Will show an object of class eSet.
signature(object = "genericFunction") Will show an object of class genericFunction.
signature(object = "gseaSignaturesSign") Will show an object of class gseaSignaturesSign.
signature(object = "gseaSignaturesVar") Will show an object of class gseaSignaturesVar.
signature(object = "gseaSignificanceSign") Will show an object of class gseaSignificanceSign.
signature(object = "gseaSignificanceVar") Will show an object of class gseaSignificanceVar.
signature(object = "LargeDataObject") Will show an object of class LargeDataObject.
signature(object = "MethodDefinition") Will show an object of class MethodDefinition.
signature(object = "MethodWithNext") Will show an object of class MethodWithNext.
signature(object = "MIAME") Will show an object of class MIAME.
signature(object = "namedList") Will show an object of class namedList.
signature(object = "ObjectsWithPackage") Will show an object of class ObjectsWithPackage.
signature(object = "oldClass") Will show an object of class oldClass.
signature(object = "ScalarCharacter") Will show an object of class ScalarCharacter.
signature(object = "ScalarObject") Will show an object of class ScalarObject.
signature(object = "signature") Will show an object of class signature.
signature(object = "TestResults") Will show an object of class TestResults.
signature(object = "traceable") Will show an object of class traceable.
signature(object = "Versioned") Will show an object of class Versioned.
signature(object = "Versions") Will show an object of class Versions.
signature(object = "VersionsNull") Will show an object of class VersionsNull.
```
smoothCoxph  

Plots the Cox proportional hazard smoothed by gene expression level.

Description

Builds a plot showing how hazard behaves over different levels of expression of a given gene. Confidence intervals are also provided.

Usage

smoothCoxph(time, event, x, xlim, ylim, xlab, ylab, logrisk=TRUE, ...)

Arguments

time  variable where time to survival is stored.
event variable where survival event is stored.
x numeric containing the expression levels of a given gene.
xlim xlim for the plot.
ylim ylim for the plot.
xlab xlab for the plot.
ylab ylab for the plot.
logrisk logrisk if we want to compute risk or logrisk estimates. By default this is TRUE, which has a better behaviour under small sample sizes.
... other arguments that will be passed to plot.

Author(s)

David Rossell.

Examples

#load eset
data(eset)

#make plot
smoothCoxph(pData(eset)$Months2Relapse,pData(eset)$Relapse,exprs(eset)[25,])

summary.gseaData  

Obtain a data.frame with the pvalues and fdr for all signatures and variables of a gseaData object.

Description

Builds a data.frame object that can easily be written to a csv file containing the ES, NES, pval.ES, pval.NES and FDR.
summary.gseaSignificance

Usage

summary.gseaData(object,...)

Arguments

object    object of class gseaData,
...          Arguments to be passed to summary.

Author(s)

Evarist Planet

References


See Also

summary.gseaSignificanceSign, summary.gseaSignificanceVar

Examples

# for examples see the help file of gseaSignatures: ?gsea

summary.gseaSignificance

Obtain a data.frame with the pvalues and fdr for all signatures and variables of a gseaSignificanceSign or gseaSignificanceVar object.

Description

Builds a data.frame object that can easily be written to a csv file containing the ES, NES, pval.ES, pval.NES and FDR.

Usage

summary.gseaSignificanceSign(object,...)

Arguments

object    object of class gseaSignificanceSign or gseaSignificanceVar.
...          Arguments to be passed to summary.

Author(s)

Evarist Planet

References

write.html

Examples

# for examples see the help file of gseaSigntaures: ?gseaSignatures

write.html Write a data.frame to an html file.

Description

Creates an html file with links and plots from a table.

Usage

write.html(x, links, tiny.pic, tiny.pic.size = 100, title = "", file, digits = 3)

Arguments

x Object of class data.frame.
links Object of class list with one item per column in x. If we want the ith column of x to have links to a site or local file we will have to write those links into the ith element of links.
tiny.pic Object of class list with one item per column in x. If we want the ith column of x to show plots instead of text we will have to write the path to the plots into the ith element of links.
tiny.pic.size size of the pictures if any.
title Title that will be shown on top of the html file.
file path and name of the file that will be created.
digits number of digits that will be shown in numeric columns of x.

Author(s)

Evarist Planet

See Also

write.csv, write.table, htmlpage

Examples

## Code has been commented to avoid the creation of files
##
#(x <- data.frame(gene.symbol=c('AARS','ABCF1','ABLIM1'),value=c(2.054,30.024,5.0221),plot=rep('Open',3)))
#tiny.pic <- links <- vector('list',length=ncol(x))
#links[[1]] <- paste('http://www.genecards.org/index.php?path=/Search/keyword/',x[,1])
#for (i in 1:nrow(x)) {
#  png(paste('~/Desktop/',x[i,1],'.png',sep=''))
#  plot(1:3,log(1:3))
#  dev.off()
#}
#tiny.pic[[3]] <- links[[3]] <- paste(x[,1],'.png',sep='')
#write.html(x,links=links,tiny.pic=tiny.pic,file='~/Desktop/x.html',title='My html test')
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