plateCore-package

plateCore: A Bioconductor package for high throughput analysis of flow cytometry data

Description

plateCore is a Bioconductor packaged created to make processing and analysis of large, complex flow datasets in R easier. High throughput flow studies are often run in a 96 or 384-well plate format, with a number of different samples, controls, and antibodies-dye conjugates present on the plate. Analyzing the output from the cytometer requires keeping track of the contents of each well, matching sample wells with control wells, gating each well/channel separately, making the appropriate plots, and summarizing the results. plateCore extends the flowCore and flowViz packages to work on flowPlate objects that represent these large flow datasets. For those familiar with flowCore and flowViz, the gating (filtering), transformation, and other data manipulations for flowPlates are very similar to flowSets.

Details

Package: plateCore
Type: Package
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Author(s)

Errol Strain, Florian Hahne, and Perry Haaland Maintainer: Errol Strain <estrain@gmail.com>

References

Insert flowCore and flowViz publications.

Examples

library(plateCore)
data(plateCore)

## Get the lymphocytes
applyControlGates

rectGate <- rectangleGate("FSC-H"=c(300,700),"SSC-H"=c(50,400))
pbmcPlate <- Subset(pbmcPlate, rectGate)

## Create a flowPlate object from the platePBMC and the wellAnnotation
fp <- flowPlate(pbmcPlate,wellAnnotation,plateName="P1")

---

applyControlGates  
Apply control gates to a flowPlate

Description

Once setControlGates has been used to create gates for a flowPlate object, gates are applied to test samples using applyControlGates. The applyControlGates function is separated from setControlGates since gates may need be changed outside of setControlGates.

Usage

applyControlGates(data, gateType="Negative-Control", ...)

Arguments

data  A flowPlate dataset.
gateType  The type of gate to be applied to the flowPlate. Currently only "Negative-Control" gates are supported.
...  optional arguments

Value

Returns a flowPlate where the wellAnnotation now contains additional columns corresponding to total number of events in a well (Total.Count), the percentage of cells above background (Percent.Positive), and the number of positive cells (Positive.Count).

Author(s)

Errol Strain

See Also

See Also setControlGates

Examples

library(plateCore)
data(plateCore)

## Get the lymphocytes
rectGate <- rectangleGate("FSC-H"=c(300,700),"SSC-H"=c(50,400))
pbmcPlate <- Subset(pbmcPlate, rectGate)

# Create a flowPlate from the sample data in plateCore
fp <- flowPlate(pbmcPlate,wellAnnotation,plateName="P1")
# Create a set of negative control gates and then apply them
fp <- setControlGates(fp,gateType="Negative.Control")
fp <- applyControlGates(fp,gateType="Negative.Control")

# Percent Positive and Counts columns are now in the wellAnnotation
head(wellAnnotation(fp))

compensate

Compensate a flowPlate to correct for the effects of spillover.

Description
Flow samples are often stained with multiple types of fluorophores. Unfortunately, the emission spectra for these different fluorophores often overlap, and the signals must be corrected before proceeding with the analysis. Compensate adjusts for spillover using the method implemented in the package flowCore. Unlike flowCore, compensate only adjusts for the dyes/fluorophores listed in wellAnnotation.

Usage
```r
## S4 method for signature 'flowPlate,ANY'
compensate(x, spillover)
```

Arguments
- `x`: A flowPlate
- `spillover`: The compensation matrix where the row and column names match the fluorescence channels of the flowPlate.

Value
Returns a compensated flowPlate.

Author(s)
Errol Strain

See Also
See Also `compensation-class`

Examples
```r
library(plateCore)
data(plateCore)

# Create the compensation matrix
comp.mat <- spillover(x=compensationSet,unstained=sampleNames(compensationSet)[5],
patt=".*H",fsc="FSC-H",ssc="SSC-H",method="median")

## Get the lymphocytes
rectGate <- rectangleGate("FSC-H"=c(300,700),"SSC-H"=c(50,400))
```
compensationSet

pbmcPlate <- Subset(pbmcPlate, rectGate)

# Create a flowPlate from the sample data in plateCore
fp <- flowPlate(pbmcPlate, wellAnnotation, plateName = "P1")

# apply the compensation matrix
fp <- compensate(fp, comp.mat)

---

compensationSet  Sample Compensation Data Set

Description

Sample Compensation Data Set

Usage

data(plateCore)

Format

The format is an object of class flowSet composed of 5 flowFrames. The flowSet consists of 4 stained and one unstained flowFrames. Peripheral Blood Mononucleocytes (PBMCs) were stained with FITC (Fluorescein isothiocyanate), PE (phycoerythrin), PerCp (Peridinin-chlorophyll), and APC (Allophycocyanin).

Author(s)

Errol Strain

Source

Sample data set from BD FACS CAP analysis.

See Also

See Also compensation-class

Examples

library(plateCore)
data(plateCore)

# Create the compensation matrix
comp.mat <- spillover(x=compensationSet, unstained=sampleNames(compensationSet)[5],
patt=".*H", fsc="FSC-H", ssc="SSC-H", method="median")
densityplot  

One-dimensional density plots for flowPlates

Description

This function is a modified version of densityplot from the flowViz package that allows for multiple flowFrames per panel. flowViz densityplot plots the density curves in a one per panel style, while the flowPlate densityplot can overlay densities.

Usage

```r
## S4 method for signature 'formula,flowPlate'
densityplot(x, data, xlab,
            prepanel=prepanel.densityplot.flowPlate,
            panel = panel.densityplot.flowPlate,
            as.table=TRUE,
            filterResult=NULL,
            ...)```

Arguments

- `x`: A formula describing the layout of the plots.
- `data`: A flowPlate.
- `xlab`: Label for the x-axis
- `prepanel`: Lattice-flowViz prepanel function.
- `panel`: Lattice-flowViz panel function.
- `as.table`: Defaults to table layout.
- `filterResult`: FilterResult can either take the character string "Negative.Control" and have the negative control wells added to the panels, or if filterResult is a flowFrame then the density curve for the flowFrame will be added to each panel.
- `...`: Optional arguments Other arguments are identical to densityPlot from flowViz.

See Also

`flowViz::densityplot`

Examples

```r
# Load the plateCore package and data
library(plateCore)
data(plateCore)

# Create a flowPlate from the sample data in plateCore
fp <- flowPlate(pbmcPlate,wellAnnotation,plateName="P1")

# Overlay the first 3 flowFrames. If the groups argument was
# omitted, then the flowFrames would be combined into a single
# density curve.
densityplot(~ `FSC-H`, fp[1:3], groups=name, auto.key=TRUE)
```
Correct for the effects of cell size (FSC) on autofluorescence

Description

The fixAutoFl function uses the method of Hahne et al. 2006 (Genome Biology) to fit a robust, log-log linear regression to the fluorescence channel of interest versus forward scatter (FSC). The current implementation scales the corrected data so the median fluorescence intensity (MFI) is the same before and after fixAutoFl.

Usage

fixAutoFl(fp,fsc="FSC.A",chanCols,unstain,...)

Arguments

fp A flowPlate
chanCols Selected channels to correct for autofluorescence.
unstain Name(s) of the unstained samples. The function will try to find samples with Sample.Type="Unstained" if no names are given. If there are multiple unstained samples the function will average the slopes.
fsc Name of the FSC parameter.
... optional arguments

Value

Returns a flowPlate with autofluorescence due to cell size (FSC) corrected.

Author(s)

Errol Strain

Examples

library(plateCore)
data(plateCore)

## Get the lymphocytes
rectGate <- rectangleGate("FSC-H":c(300,700),"SSC-H":c(500,400))
pbmcPlate <- Subset(pbmcPlate, rectGate)

## Create a flowPlate object from the platePBMC and the wellAnnotation
fp <- flowPlate(pbmcPlate,wellAnnotation,plateName="P1")

fp <- fixAutoFl(fp,fsc="FSC-H",chanCols=c("FL1-H","FL2-H","FL3-H","FL4-H"))
flowPlate

Create a flowPlate

Description

Constructor for a flowPlate object. sampleNames for the flowSet should match the Well.Id column of wellAnnotation. Well.Ids must be unique to sampleNames, which is usually ensured by using the 3 character designations for wells (e.g. "A01","A02",...,"H12").

Usage

flowPlate(data, wellAnnotation, plateName, ...)

Arguments

data flowSet object to be made into a flowPlate
wellAnnotation data.frame describing the layout and contents of the flowPlate
plateName Name of the flowPlate, should be unique within the set flowPlates under consideration
... optional arguments

Value

Returns a flowPlate object.

Author(s)

Errol Strain

Examples

library(plateCore)
data(plateCore)

## Get the lymphocytes
rectGate <- rectangleGate("FSC-H"=c(300,700),"SSC-H"=c(50,400))
pbmcPlate <- Subset(pbmcPlate, rectGate)

## Create a flowPlate object from the platePBMC and the wellAnnotation
fp <- flowPlate(pbmcPlate, wellAnnotation, plateName="P1")

## Subset the flowPlate, creating another flowPlate
fpSmall <- fp["A01"]

## Extract a flowFrame from a flowPlate
ff <- fp["A01"]

## Retrieve sample names from flowPlate
sampNames <- sampleNames(fp)

## Retrieve the annotatedDataFrame describing the flowPlate
adf <- phenoData(fp)
The flowPlate class.

Description

flowPlates are the basic data containers for the plateCore package. A flowPlate is essentially a flowSet-class, plus a data.frame describing the layout of the plate and contents of individual wells.

Slots

plateName: A character string containing the name of the plate.

plateSet: A flowSet-class containing FCS event data. Prior to creating a flowPlate, the FCS files are first read into a flowSet-class using read.flowSet.

wellAnnotation: A data.frame describing the layout of the plate. Each row describes one channel for a well.

Methods

[, [[ Subsetting. x[i] where i is either a scalar or character corresponding to a sample name, returns a flowPlate object, and x[[i]] a flowFrame object.

Usage:

flowSet[i]  
flowSet[[i]]

Author(s)

Errol Strain, Florian Hahne, Perry Haaland

Examples

library(plateCore)
data(plateCore)

##Look at the wellAnnotation
wellAnnotation[1:4,]

## Get the lymphocytes
rectGate <- rectangleGate("FSC-H"=c(300,700),"SSC-H"=c(50,400))
pbmcPlate <- Subset(pbmcPlate, rectGate)

## Create a flowPlate object from the platePBMC and the wellAnnotation
fp <- flowPlate(pbmcPlate,wellAnnotation,plateName="P1")

## Subset the flowPlate, creating another flowPlate
fpSmall <- fp["A01"]

## Extract a flowFrame from a flowPlate
ff <-fp["A01"]]
fpbind

Merge multiple flowPlates into a single virtual flowPlate

Description

A function to combine multiple flowPlates into a single flowPlate object. The plateName identifiers for the flowPlates must be unique within the set to be bound, otherwise the bind will fail.

Usage

fpbind(p1, p2, ...)

Arguments

p1 First flowPlate
p2 Second flowPlate
... Additional flowPlates

Value

Returns a flowPlate

Author(s)

Errol Strain

Examples

library(plateCore)
data(plateCore)

# Get the lymphocytes
rectGate <- rectangleGate("FSC-H"=c(300,700),"SSC-H"=c(50,400))
pbmcpPlate <- Subset(pbmcpPlate, rectGate)

# Create a flowPlate object from the platePBMC and the wellAnnotation
fp1 <- flowPlate(pbmcpPlate,wellAnnotation,plateName="P1")
fp2 <- flowPlate(pbmcpPlate,wellAnnotation,plateName="P2")

# Combine the plates.
virtPlate <- fpbind(fp1,fp2)
getGroups

Retrieve Negative control groups from a flowPlate

Description
Retrieve a list of negative control-based groups from a flowPlate, based on the information in wellAnnotation.

Usage
getGroups(data, type="Negative.Control", chan, ...)

Arguments
data A flowPlate dataset.
type Currently only Negative.Control groups are supported.
chan Fluorescence channel of interest ("FL1-H", "PE-H", etc.)
... optional arguments

Value
Returns a list of groups, where each group contains a single negative control well and the associated test well for a particular channel.

Author(s)
Errol Strain

Examples
library(plateCore)
data(plateCore)
# Create a flowPlate from the sample data in plateCore
fp <- flowPlate(pbmcPlate, wellAnnotation, plateName="P1")
# Create a set of negative control gates and then apply them
negCon <- getGroups(fp, chan="FL1-H")
negCon[1:2]
gutterPlot  

Description  
A Quality Control plot to check the number of events in each channel that are at either their minimum or maximum value. A large number of these events may indicate a problem with the sample.

Usage  

Arguments  
fp  A flowPlate.  
chans  Channels of interest to show on the gutterPlot.  
...  optional arguments  

Value  
Creates a plot where the x-axis is the different wells in a flowPlate, and the y-axis is the fraction of events at the boundary.

Author(s)  
Jon Gosink and Errol Strain

Examples  
library(plateCore)  
data(plateCore)  

### Create a flowPlate  
fp <- flowPlate(pbmcPlate,wellAnnotation,"p1001")  
gutterPlot(fp,chans=c("FSC-H","SSC-H","FL1-H","FL2-H","FL3-H","FL4-H"))

mfiPlot  

Description  
A Quality Control plot that shows the MFI Ratio versus the percentage of positive cells in a flowPlate. The robust logistic regression is performed using gmlrob from the robustbase package.

Usage  
mfiPlot(fp, thresh=2, Sample.Type="Test", Events="Percentage", ...)

mfiPlot
Arguments

- **fp**: A flowPlate.
- **thresh**: Points more than "thresh" number of standard deviations away from the best fit line will be colored red.
- **Sample.Type**: Type of sample to show on plot. Defaults to "Test".
- **Events**: The robust logistic regression can be performed using either the percentage of events above the negative control gate ("Percentage") or the actual number of events above the gate ("Actual").
- ... optional arguments to plot and points.

Value

Creates a plot where the x-axis is MFI Ratio and the y-axis is the percentage of cells above the negative control gate.

Author(s)

Errol Strain

Examples

```r
library(plateCore)
data(plateCore)

## Get the lymphocytes
rectGate <- rectangleGate("FSC-H"=c(300,700),"SSC-H"=c(50,400))
pbmcPlate <- Subset(pbmcPlate, rectGate)

# Create a flowPlate from the sample data in plateCore
fp <- flowPlate(pbmcPlate,wellAnnotation,plateName="P1")

# Create a set of negative control gates and then apply them
fp <- setControlGates(fp,gateType="Negative.Control")
fp <- applyControlGates(fp,gateType="Negative.Control")

# Compute summary statistics
fp <- summaryStats(fp)

## Create an MFI plot
mfiPlot(fp,thresh=2.5,xlab="MFI Ratio (Test MFI / Isotype MFI)",xlim=c(0.1,250),ylab="Percentage of cells above the isotype gate",pch=23)
```

Description

This function should not be called directly, use `densityplot`.
panel.densityplot.flowPlate

Usage

panel.densityplot.flowPlate(x, frames, channel, wellAnnotation, groups=NULL, subscripts, col = superpose.symbol$col, col.points = col, col.line = col, filterResult=NULL, ...)

Arguments

x character
frames flowFrames
channel channel of interest
wellAnnotation wellAnnotation data.frame
groups density plot groups parameter
subscripts densityplot subscripts parameter
col densityplot col parameter
col.points densityplot col.points parameter
col.line densityplot col.line parameter
filterResult densityplot filterResult parameter
... optional arguments

Author(s)

Errol Strain

See Also

See Also densityplot

Examples

# Load the plateCore package and data
library(plateCore)
data(plateCore)

# Create a flowPlate from the sample data in plateCore
fp <- flowPlate(pbmcPlate,wellAnnotation,plateName="P1")

# Overlay the first 3 flowFrames. If the groups argument was
# omitted, then the flowFrames would be combined into a single
# density curve.
densityplot(~ `FSC-H`, fp[1:3], groups=name, auto.key=TRUE)
panel.xyplot.flowPlate

Lattice-flowViz style panel function for flowPlate xyplot.

Description

This function should not be called directly, use `xyplot`.

Usage

```r
panel.xyplot.flowPlate(x, frames, channel.x, channel.y, channel.x.name, channel.y.name, filter = NULL, filterResults = NULL, displayFilter = TRUE, pch, smooth, wellAnnotation = NULL, col = superpose.symbol$col, ...)
```

Arguments

- `x`: character
- `frames`: flowFrames
- `channel.x`: xyplot channel.x parameter
- `channel.y`: xyplot channel.y parameter
- `channel.x.name`: xyplot channel.x.name parameter
- `channel.y.name`: xyplot channel.y.name parameter
- `filter`: xyplot filter parameter
- `filterResults`: xyplot filterResults parameter
- `displayFilter`: xyplot displayFilter parameter
- `pch`: xyplot pch parameter
- `smooth`: xyplot smooth parameter
- `wellAnnotation`: wellAnnotation data.frame
- `col`: xyplot col parameter
- `...`: optional arguments

Author(s)

Errol Strain

See Also

See Also `xyplot`
Examples

library(plateCore)
data(plateCore)

# Create a flowPlate from the sample data in plateCore
fp <- flowPlate(pbmcPlate, wellAnnotation, plateName="P1")

## Create a rectangle filter
rectGate <- rectangleGate("FSC-H"=c(300,700),"SSC-H"=c(50,400))

xyplot("SSC-H" ~ "FSC-H" | as.factor(name),
fp[1], smooth=FALSE, filter=rectGate, displayFilter=FALSE)

pbmcPlate

Description

One 96-well plate from a BD FACS CAP analysis of Peripheral Blood Mononucleocyte (PMBC) cells.

Usage

data(plateCore)

Format

The format is an object of class flowSet composed of 96 flowFrames. Each flowFrame corresponds to one well from the plate.

Details

BD FACS CAP (http://www.bd.com/technologies/discovery_platform/BD_FACS_CAP.asp) is a platform for screening a large number of antibodies (200+) on human samples. Antibodies are arrayed 3-per well on a 96-well plate, along with the appropriate controls. In this experiment, an early version of FACS CAP was used to screen PBMCs from 2 donors for 189 different human cell surface markers. The complete data set is available from the FICCS site shown below. The pbmcPlate include with plateCore is a lymphocyte enriched subset of one of the replicate plates for donor 1.

Source

Complete dataset available at http://www.ficcs.org/software.html#Data_Files, the Flow Informatics and Computational Cytometry Society website (FICCS)

References

Add reference for plateCore paper (when/if published).
plateSet

Get the flowSet from a flowPlate object

Description
A function to retrieve the flowSet from a flowPlate.

Usage
plateSet(fp, ...)

Arguments
fp A flowPlate
... optional arguments

Value
Returns a flowSet

Author(s)
Errol Strain

Examples
library(plateCore)
data(plateCore)

## Get the lymphocytes
rectGate <- rectangleGate("FSC-H"=c(300,700),"SSC-H"=c(50,400))
pbmcPlate <- Subset(pbmcPlate, rectGate)

## Create a flowPlate object from the platePBMC and the wellAnnotation
fp <- flowPlate(pbmcPlate, wellAnnotation, plateName="P1")

## Retrieves the flowSet
fs <- plateSet(fp)

plotPlate

Description
Make a row vs. column plot of a plate, where the wells are colored according to some value of choice (number of events, median signal intensity, percent positive, etc.).

Usage
plotPlate(fp, x = NA, method = "median", main, col, values, width = 1, na.action = "zero", ...)
prepanel.densityplot.flowPlate

Arguments

fp
A flowPlate.
x
A character indicating the variable of interest. Valid choices are "events", any single channel name (e.g. FSC-H, SSC-H, FL1-H, etc.), or vector of channel names if the method is mahalanobis.

method
Valid choices are mean, median, sd, mad, mahalanobis, or one of the numeric columns in the wellAnnotation data.frame (e.g. Percent.Positive, Positive.Count, MFI, MFI.Ratio)

main
Main text for the plot

col
Character vector of two colors.

values
Optional list of values, with names corresponding to sampleNames, that will be used for plotting.

width
Width of the well.

na.action
Handling of NA values, either "zero" or "omit".

...
optional arguments

Value
Plots the plate to the standard output.

Author(s)
Jon Gosink and Errol Strain

References
The original version of this plot came from the prada package.

Examples

library(plateCore)
data(plateCore)

## Create a flowPlate
fp <- flowPlate(pbmcPlate,wellAnnotation,"p1001")

plotPlate(transform("FL1-H"=log10) %on% fp,x="FL1-H",method="mean",col=c("yellow", "darkblue"))

prepanel.densityplot.flowPlate
Lattice-flowViz style panel function for flowPlate densityplot.

Description
This function should not be called directly, use densityplot.

Usage

prepanel.densityplot.flowPlate(x, frames, channel,...)
Arguments

- **x**  
  Character
- **frames**  
  flowFrames
- **channel**  
  Character string for channel name.
- ...  
  optional arguments

Author(s)

- Errol Strain

See Also

- See Also `densityplot`

Examples

```r
# Load the plateCore package and data
library(plateCore)
data(plateCore)

# Create a flowPlate from the sample data in plateCore
fp <- flowPlate(pbmcPlate,wellAnnotation,plateName="P1")

# Overlay the first 3 flowFrames. If the groups argument was
# omitted, then the flowFrames would be combined into a single
# density curve.
densityplot(~ `FSC-H`, fp[1:3], groups=name, auto.key=TRUE)
```

Description

This function should not be called directly, use `xyplot`.

Usage

```r
prepanel.xyplot.flowPlate(x, frames, channel.x, channel.y,...)
```

Arguments

- **x**  
  Character
- **frames**  
  flowFrames
- **channel.x**  
  Character string for channel name.
- **channel.y**  
  Character string for channel name.
- ...  
  optional arguments
setControlGates

Create control gates for a flowPlate

Description

A function to estimate the threshold between positive and negative cells. This threshold corresponds to a one-dimensional gate, and cells above the gate are considered positive. The default value of numMads=5 generally works well on the linear scale, but will need to be adjusted for transformed data. If each well contains a large number of events for the cell type of interest (>1000), then using the 99.5th quantile usually gives similar values.

Usage

setControlGates(data, gateType, threshType="MAD", numMads=5, isoquantile=.995, ...)

Arguments

data A flowPlate
gateType The type of gate to be set. Currently only "Negative.Control" gates are supported.
threshType Values can be either "MAD", for median absolute deviation based gating, or "isoQuant" for quantile based gating.
numMads Number of median absolute deviations above the median to set the initial gate.
isoquantile Quantile setting for "isoQuant" threshType.
... optional arguments.

Value

Returns a flowPlate
Subset

Author(s)

Errol Strain

Examples

library(plateCore)
data(plateCore)

## Get the lymphocytes
rectGate <- rectangleGate("FSC-H"=c(300,700),"SSC-H"=c(50,400))
pbmcPlate <- Subset(pbmcPlate, rectGate)

# Create a flowPlate from the sample data in plateCore
fp <- flowPlate(pbmcPlate,wellAnnotation,plateName="P1")

# Create a set of negative control gates and then apply them
fp <- setControlGates(fp,gateType="Negative.Control")

# There should now be a Negative.Control.Gate column in wellAnnotation
head(wellAnnotation(fp))

Description

Select a subset of events in a flowPlate. If a flowPlate and filter are supplied, then this function calls
the Subset function from flowCore. Additionally, the plateCore version of Subset also makes it easy
to filter individual flowFrames and keep the flowPlate structure.

Usage

## S4 method for signature 'flowPlate,ANY'
Subset(x, subset, select=NULL,...)

Arguments

x A flowPlate
subset A filter object
select An optional vector of either sample names or Well.IDs.
... optional arguments

Value

Returns a flowPlate

Author(s)

Errol Strain
Examples

```r
library(plateCore)
data(plateCore)

# Create a flowPlate from the sample data in plateCore
fp <- flowPlate(pbmcPlate, wellAnnotation, plateName="P1")

## Create a rectangle filter
rectGate <- rectangleGate("FSC-H"=c(300,700),"SSC-H"=c(50,400))

## Apply the filter only to sample A01. The other flowFrames
## are not filtered.
fp <- Subset(fp, rectGate, 'A01')
```

---

**summaryStats**  
*Compute summary statistics on a flowPlate*

**Description**

This function computes the median fluorescence intensity (MFI) and the MFI ratio (ratio of test well MFI to negative control MFI) for each well/channel in a flowPlate. The predicted percent positive (Predict.PP) and gate score (Gate.Score) come from a robust logistic regression of the MFI ratio to either the percentage of positive cells or the actual count of positive cells. Predict.PP is the estimated percent positive based on the MFI ratio and Gate.Score is the number of standardized residuals the sample data point is away from the best fit line. The glmrob function from the robustbase package is used for the regression. Results from summaryStats are stored in the wellAnnotation data.frame.

**Usage**

```r
summaryStats(data, Events="Percentage", ...)
```

**Arguments**

- **data**  
  A flowPlate

- **Events**  
  The robust logistic regression can be performed using either the percentage of events above the negative control gate ("Percentage") or the actual number of events above the gate ("Actual").

- **...**  
  optional arguments

**Value**

Returns a flowPlate

**Author(s)**

Errol Strain
### Examples

```r
library(plateCore)
data(plateCore)

## Get the lymphocytes
rectGate <- rectangleGate("FSC-H"=c(300,700),"SSC-H"=c(50,400))
pbmcPlate <- Subset(pbmcPlate, rectGate)

# Create a flowPlate from the sample data in plateCore
fp <- flowPlate(pbmcPlate, wellAnnotation, plateName="P1")

# Create a set of negative control gates and then apply them
fp <- setControlGates(fp, gateType="Negative.Control")
fp <- applyControlGates(fp, gateType="Negative.Control")

# Compute summary statistics
fp <- summaryStats(fp)

# There should now be MFI and MFI.ratio columns in the wellAnnotation
head(wellAnnotation(fp))
```

---

**wellAnnotation**  
*Retrieve a data.frame describing the content of a flowPlate*

**Description**

wellAnnotation returns the tall data.frame describing the layout of a flowPlate, where each row corresponds to one well-channel.

**Usage**

```r
wellAnnotation(fp, ...)
```

**Arguments**

- `fp`: A flowPlate dataset.
- `...`: optional arguments

**Value**

Returns a data.frame.

**Author(s)**

Errol Strain
xypot

Examples

library(plateCore)
data(plateCore)

# Create a flowPlate from the sample data in plateCore
fp <- flowPlate(pbmcPlate,wellAnnotation,plateName="P1")

# Look at the top of wellAnnotation
head(wellAnnotation(fp))

xypot

Scatter plots (dotplots) for flowPlates.

Description

A function to create dotplots, and smoothed scatter plots, from flowPlates. This function is a slightly modified version of xyplot from flowViz. The flowPlate xyplot allows users to overlay plots of test samples versus controls, and makes creating informative flowStrips easier. Refer to the documentation for xyplot from flowViz and lattice for more detailed information.

Usage

## S4 method for signature 'formula,flowPlate'
xypot(x, data, xlab, ylab,
as.table = TRUE,
prepanel = prepanel.xyplot.flowPlate,
panel = panel.xyplot.flowPlate,
pch = ".", smooth = TRUE,
filter = NULL,
filterResults = NULL,
displayFilter = TRUE,
flowStrip=NULL,
flowStripCex=1,
strip=function(...,style=1) strip.default(...,style=1),
...)

Arguments

x A formula describing the layout of the plots. Plots for flowPlates usually condition on either as.factor(name) or as.factor(Well.Id) since only one flowFrame can be shown on each panel (with the exception of Negative.Control overlays).
data A flowPlate.
xlab Label for x-axis.
ylab Label for y-axis.
as.table Defaults to table layout.
prepanel Lattice-flowViz prepanel function.
panel Lattice-flowViz panel function.
pch Plotting character.
smooth: Plot a smoothed scatterplot by default.
filter: A flowCore filter to apply to each flowFrame.
filterResults: If filterResults="Negative.Control", the negative control wells corresponding to a test well are overlayed in the test well plots.
displayFilter: Defaults to displaying filter on the plot.
flowStrip: Character vector indicating additional information to be printed on the strip. Values can include any combination of "Well.Id","MFI","MFI.Ratio", and "Percent.Positive".
flowStripCex: Font size for the flowStrip.
strip: Lattice strip function.

Author(s)
Errol Strain

See Also
flowViz::xyplot

Examples

library(plateCore)
data(plateCore)

# Create a flowPlate from the sample data in plateCore
fp <- flowPlate(pbmcPlate,wellAnnotation,plateName="P1")

## Create a rectangle filter
rectGate <- rectangleGate("FSC-H"=c(300,700),"SSC-H"=c(50,400))

xyplot("SSC-H" ~ "FSC-H" | as.factor(name),
fp[1], smooth=FALSE, filter=rectGate, displayFilter=FALSE)

Methods for Function %on% in Package 'plateCore'

Description

This operator is used to construct a transformFilter that first applies a transformList to the data before applying the filter operation to a flowPlate.

Author(s)
Errol Strain
Examples

library(plateCore)
data(plateCore)

# Create a flowPlate from the sample data in plateCore
fp <- flowPlate(pbmcPlate,wellAnnotation,plateName="P1")

## Create a rectangle filter
rectGate <- rectangleGate("FSC-H"=c(300,700),"SSC-H"=c(50,400))

xyplot("FL1-H" ~ "FSC-H" | as.factor(name),
transform("FL1-H"=log10) %on% fp, smooth=FALSE, filter=rectGate, displayFilter=FALSE)
Index

*Topic datasets
  compensationSet, 5
  pbmcPlate, 16
*Topic dplot
  densityplot, 6
  xyplot, 24
*Topic methods
  %on%, 25
  applyControlGates, 3
  compensate, 4
  densityplot, 6
  fixAutoFl, 7
  flowPlate, 8
  flowPlate-class, 9
  fpbind, 10
  getGroups, 11
  gutterPlot, 12
  mfiPlot, 12
  panel.densityplot.flowPlate, 13
  panel.xyplot.flowPlate, 15
  plateSet, 17
  plotPlate, 17
  prepanel.densityplot.flowPlate, 18
  prepanel.xyplot.flowPlate, 19
  setControlGates, 20
  Subset, 21
  summaryStats, 22
  wellAnnotation, 23
  xyplot, 24
*Topic package
  plateCore-package, 2
  [,flowPlate,ANY-method (flowPlate-class), 9
  ][,flowPlate,ANY-method (flowPlate-class), 9
  %on%,ANY,flowPlate-method (%on%), 25
  %on%-methods (%on%), 25
  %on%, 25
  applyControlGates, 3
  applyControlGates,flowPlate-method (applyControlGates), 3
  applyControlGates-methods (applyControlGates), 3
  compensate, 4
  compensate,flowPlate,ANY-method (compensate), 4
  compensate-method(compensate), 4
  compensationSet, 5
  densityplot, 6, 13, 14, 18, 19
  densityplot,formula,flowPlate,ANY,ANY,ANY,missing-method (densityplot), 6
  densityplot,formula,flowPlate-method (densityplot), 6
  densityplot-method (densityplot), 6
  fixAutoFl, 7
  fixAutoFl,flowPlate-method (fixAutoFl), 7
  fixAutoFl-method (fixAutoFl), 7
  flowFrame, 9
  flowPlate, 8, 8
  flowPlate,flowSet-method (flowPlate), 8
  flowPlate-class, 9
  flowPlate-method (flowPlate), 8
  flowViz::densityplot, 6
  flowViz::xyplot, 25
  fpbind, 10
  fpbind,flowPlate,flowPlate-method (fpbind), 10
  fpbind-method (fpbind), 10
  getGroups, 11
  getGroups,flowPlate-method (getGroups), 11
  getGroups-method (getGroups), 11
  gutterPlot, 12
  gutterPlot,flowPlate-method (gutterPlot), 12
  gutterPlot-method (gutterPlot), 12
  mfiPlot, 12
  mfiPlot,flowPlate-method (mfiPlot), 12
  mfiPlot-method (mfiPlot), 12
  panel.densityplot.flowPlate, 13
  panel.xyplot.flowPlate, 15
  pbmcPlate, 16
phenoData, flowPlate-method (flowPlate), 8
plateCore (plateCore-package), 2
plateCore-package, 2
plateSet, 17
plateSet, flowPlate-method (plateSet), 17
plateSet-method (plateSet), 17
plotPlate, 17
plotPlate, flowPlate-method (plotPlate), 17
plotPlate-method (plotPlate), 17
prepanel.densityplot.flowPlate, 18
prepanel.xyplot.flowPlate, 19
read.flowSet, 9
sampleNames, flowPlate-method (flowPlate), 8
setControlGates, 3, 20
setControlGates, flowPlate-method (setControlGates), 20
setControlGates-method (setControlGates), 20
Subset, 21
Subset, flowPlate, ANY-method (Subset), 21
Subset, flowPlate-method (Subset), 21
Subset-method (Subset), 21
summaryStats, 22
summaryStats, flowPlate-method (summaryStats), 22
summaryStats-method (summaryStats), 22
wellAnnotation, 23
wellAnnotation, flowPlate-method (wellAnnotation), 23
wellAnnotation-method (wellAnnotation), 23
xyplot, 15, 19, 20, 24
xyplot, formula, flowPlate-method (xyplot), 24
xyplot-method (xyplot), 24