Package ‘stemHypoxia’

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**Type** Package

**Title** Differentiation of Human Embryonic Stem Cells under Hypoxia gene expression dataset by Prado-Lopez et al. (2010)

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**Description**

**Author** Cristobal Fresno and Elmer A. Fernandez

**Maintainer** Cristobal Fresno <cristobalfresno@gmail.com>

**License** GPL (>=2)

**Depends** R (>= 2.14.1)

**URL**
http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE37761,

**biocViews**
ExperimentData, Tissue, StemCell, Homo_sapiens_Data,
CancerData, MicroarrayData, TissueMicroarrayData, GEO

**NeedsCompilation** no

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**Description**

Differentiation of Human Embryonic Stem Cells under Hypoxia gene expression dataset by Prado-Lopez et al. 2010.
Usage

data(stemHypoxia)

Format

Experimental design data.frame with samples and different treatment conditions: time points and oxygen levels.

- **time**: time where the sample was processed (0, 0.5, 1, 5, 10 or 15 days).
- **oxygen**: level of oxygen available in the conditioned medium (1, 5 or 21 %)
- **samplename**: acronym to describe chips’s treatment. Note that the first two are control conditions.

Details

This dataset represents the study published by Prado-Lopez et al. 2010.

- **Summary**: Early development of mammalian embryos occurs in an environment of relative hypoxia. Nevertheless, human embryonic stem cells (hESC), which are derived from the inner cell mass of blastocyst, are routinely cultured under the same atmospheric conditions (21% O2) as somatic cells. We hypothesized that O2 levels modulate gene expression and differentiation potential of hESC, and thus, we performed gene profiling of hESC maintained under normoxic or hypoxic (1% or 5% O2) conditions. Our analysis revealed that hypoxia downregulates expression of pluripotency markers in hESC but increases significantly the expression of genes associated with angio- and vasculogenesis including vascular endothelial growth factor and angiopoietin-like proteins. Consequently, we were able to efficiently differentiate hESC to functional endothelial cells (EC) by varying O2 levels; after 24 hours at 5% O2, more than 50% of cells were CD34+. Transplantation of resulting endothelial-like cells improved both systolic function and fractional shortening in a rodent model of myocardial infarction. Moreover, analysis of the infarcted zone revealed that transplanted EC reduced the area of fibrous scar tissue by 50%. Thus, use of hypoxic conditions to specify the endothelial lineage suggests a novel strategy for cellular therapies aimed at repair of damaged vasculature in pathologies such as cerebral ischemia and myocardial infarction.

Source


References


Examples

```r
# load the dataset
data(stemHypoxia)

# Exploring the experimental design data.frame
dim(design) #22 8
```
## Show the head of the data.frame and its summary

```r
head(design)
summary(design)
```

### M  
*Gene expression data matrix*

---

### Description

Differentiation of Human Embryonic Stem Cells under Hypoxia gene expression dataset by Prado-Lopez et al. 2010.

### Usage

```r
data(stemHypoxia)
```

### Format

Agilent-014850 Whole Human Genome Microarray 4x44K G4112F, rma gene expression matrix, where row stands for genes and columns for treatments (except the first two) respectively.

- **Gene_ID**: manufacturer feature id a.k.a AGILENT_OLOGO_ID.
- **GeneName**: gene symbol of the corresponding feature.
- **Additional columns**: treatment expression (see `design`)

### Details

This dataset represents the study published by Prado-Lopez et al. 2010.

- **Summary**: Early development of mammalian embryos occurs in an environment of relative hypoxia. Nevertheless, human embryonic stem cells (hESC), which are derived from the inner cell mass of blastocyst, are routinely cultured under the same atmospheric conditions (21% O2) as somatic cells. We hypothesized that O2 levels modulate gene expression and differentiation potential of hESC, and thus, we performed gene profiling of hESC maintained under normoxic or hypoxic (1% or 5% O2) conditions. Our analysis revealed that hypoxia downregulates expression of pluripotency markers in hESC but increases significantly the expression of genes associated with angio- and vasculogenesis including vascular endothelial growth factor and angiopoietin-like proteins. Consequently, we were able to efficiently differentiate hESC to functional endothelial cells (EC) by varying O2 levels; after 24 hours at 5% O2, more than 50% of cells were CD34+. Transplantation of resulting endothelial-like cells improved both systolic function and fractional shortening in a rodent model of myocardial infarction. Moreover, analysis of the infarcted zone revealed that transplanted EC reduced the area of fibrous scar tissue by 50%. Thus, use of hypoxic conditions to specify the endothelial lineage suggests a novel strategy for cellular therapies aimed at repair of damaged vasculature in pathologies such as cerebral ischemia and myocardial infarction.

### Source

References


Examples

```r
## load the dataset
data(stemHypoxia)

## Exploring the expression matrix M
dim(M) #48736 features x 28 sample + ID + Symbol
head(M) #Just to see some intensity values

##Boxplot of gene expressions
## Not run: boxplot(M[,-(1,2)])
```

---

Description

Differentiation of Human Embryonic Stem Cells under Hypoxia gene expression dataset by Prado-Lopez et al. 2010.

Usage

```r
data(stemHypoxia)
```

Format

This package contains two data.frames with the associated experimental data as follows:

- `M`: the microarray gene expression matrix (see `M`).
- `design`: the lm like experimental design object, describing the treatment structure (see `design`).

Details

This dataset represents the study published by Prado-Lopez et al. 2010.

Summary: Early development of mammalian embryos occurs in an environment of relative hyptem cells (hESC), which are derived from the inner cell mass of blastocyst, are routinely cultured under the same atmospheric conditions (21% O2) as somatic cells. We hypothesized that O2 levels modulate gene expression and differentiation potential of hESC, and thus, we performed gene profiling of hESC maintained under normoxic or hypoxic (1% or 5% O2) conditions. Our analysis revealed that hypoxia downregulates expression of pluripotency markers in hESC but increases significantly the expression of genes associated with angio- and
vasculogenesis including vascular endothelial growth factor and angiopoietin-like proteins. Consequently, we were able to efficiently differentiate hESC to functional endothelial cells (EC) by varying O2 levels; after 24 hours at 5% O2, more than 50% of cells were CD34+. Transplantation of resulting endothelial-like cells improved both systolic function and fractional shortening in a rodent model of myocardial infarction. Nevertheless, human embryonic stem cells (hESC), which are derived from the inner cell mass of blastocyst, are routinely cultured under the same atmospheric conditions (21% O2) as somatic cells. We hypothesized that O2 levels modulate gene expression and differentiation potential of hESC, and thus, we performed gene profiling of hESC maintained under normoxic or hypoxic (1% or 5% O2) conditions. Our analysis revealed that hypoxia downregulates expression of pluripotency markers in hESC but increases significantly the expression of genes associated with angiogenesis and vasculogenesis including vascular endothelial growth factor and angiopoietin-like proteins. Consequently, we were able to efficiently differentiate hESC to functional endothelial cells (EC) by varying O2 levels; after 24 hours at 5% O2, more than 50% of cells were CD34+. Transplantation of resulting endothelial-like cells improved both systolic function and fractional shortening in a rodent model of myocardial infarction. Moreover, analysis of the infarcted zone revealed that transplanted EC reduced the area of fibrous scar tissue by 50%. Thus, use of hypoxic conditions to specify the endothelial lineage suggests a novel strategy for cellular therapies aimed at repair of damaged vasculature in pathologies such as cerebral ischemia and myocardial infarction.

Source


References


See Also

M and design man pages for a complete description.
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