# Visualisation tools for next-generation sequencing

#### Simon Anders



EBI is an Outstation of the European Molecular Biology Laboratory.



Exploring and checking alignment with alignment viewers

Using genome browsers

Getting an overview over the whole data with Hilbert curve visualization

 Displaying peaks alongside feature annotation with the "GenomeGraph" package



#### Alignment Viewers: SAMtools tview

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Heng Li (Sanger Institute) et al.

# Alignment viewers: MaqView



Jue Ruan (Beijing Genomics Institute) et al.



#### Alignment Viewer: MapView

#### 🚵 paired aln chr12 part - MapView 3.0 File View Statistics Help Fast Positioning Col: 20642 \$ ማል ምምምም ልምም እስከ እንግ በግር የሚያስት እስከ የሚያስት እስከ የሚያስ የሚያስት የ CTTATACGATTATATAAGTCTATTTTTTGTCCCTATATTTTTT CTCTGTGGGTTGTGTTTAAATGGCTGGAAAATAAGTC ATTTTGTATACCTAT FATACGATTATA TATATTTT Row: â 1 GTGGGTTGTGTTTAAATGGCTGGAAAATAAGTCTGTTTTGTATA CTAT Reference Position: 20697 ACGATTATATAAGTCTATTT ATTT Coverage SUM:27 CTCT TGGGCTGTGTTTAAATGGCTGGAAAATAAGTCTATTTGTATAC CGGTTATATAAGTCTATTTTTG ATTTT A:2 T:14 CTTTGTGGGTT TGTTTAAATGGCTGGATAATAAGTCTATTTTGTATACCTATT CGGTTATATAAGTCTATTTTTTGTCCC TTTT G:11 C:0 AAATGGCTGGAAAATAAGTCTATTTTGTATACCTATT CTCTGTGGGCTGTGT N:O CGATTATATAAGTCTATTTTTCGTCCCTGTATT **SNP** Detection Variant Frequency:0.4400 CTCTGTGGGTTGTGTTT GGCT GGAAAAT AAGTCTATTTTGTATACATAT TTATATAAGTCTATTTTTTATCCCTATATTTT CTCTGTGGGGCTGTGTTTA GGCTGGAAAATAAGTCTATTTTGTATACATAT TCTTGGGCCA CTTATACGATTATATAAGTCTATTTTTTGTCCCTATATT Solexa quality score 🐱 CTCTGTGGGTTGTGTTTAAATGGCT GAAAATAAGTCTATTTTGTATACCTAT TCTTGGGCCATGCCT ACGATTATATAATTCTATTTTTTGTCCCTA TCTTGGGCCAT GGTTATNTAAGTCTATTTTTTGTCCCTATATTTT CTCTGTGGGCTGTGTTTAAA GCTGGAAAATAAGTCTATTTTGTATACCTAA Quality 20 20 Score TCTTGGGCCATG CTCTGTGGGGGTGTGTTTAAAT CTGGAAAATAAGTCTATTTTGTATACCTATT GATTATATAAGTCTATTTTTTGTOCCTATATTTT CTCTGTGGGGCTGTGTTTAAAT CTGGAAAATAAGTCTATTTTGTATACATATT TCTTGGGCCATG ATTATATAAGTNTATTTTTTGTCCCTATATTTT Variant >= 0.4000 CTCTGTGGGTTGTGTTTAAATG CTGGAAAATAANTCTATTTTGTATACCTAT TCTTGGGCCATG Frequency CTCTGTGGGGCTGTGTTTAAATG TAGAAAATAAGTCTATTTTGTATACCTAT TCTTAGGCCATGC Coverage TCTTGGGCCATGCCTTA 12 🗘 CTCTGTGGGTTGTGTTTAAATGGCTG AAATAAGTCTATTTTGTATACCTATT SUM >= CTCTGTGGGTTGTGTTTGAATGGCTGGA AATAAGTCTATTTTGTATACCTAT TCTTGGGCCATGCCTTAT TCTTGGGCCATGCCTTATACGA CTCTGTGGGTTGTGTTTAAATGGCTGGAA AGTCTATTTTGTATACCTAT **SNP** Detection CT CT GT GG GT T GT GT T T AAAT G G CT G G AAAA TCTATTTTGTATACCTAT TCTTGGGCCATGCCTTATACGATT No quality control CT CT GT GG GT T GT GT TT AAAAG GC TAGAAAAT A TTTGTATACCTAT1 TCTTGGGCCATGCCTTATACGATTATATA IFAST MODEL CTCTGTGGGCTGTGTGTTTAAATGGCTGGAAAATA TCTTGGGCCATGCCTTATACGGTTATATA TTTGTATACCTATT CT CT GT GGGT TGTGT TT AAATGGCT GGAAAAT AAG ATACCTATT TCTTGGGCCATGCCTTATACGATTATATAAGTCT Total: 18,356 CT CT GCGGGCTGTGTTT AAATGGCT GGAAAATAAGT TACCTATT TCTTGGGCCATGCCTTATACGATTATATAAGTCTA 22 \$ Current: TACATAT CTCTGTGGGCTGTGTTTAAATGGCTGGATAATAAGTC TCTTGGGCCATGCCTTATACGATTATATAAGTCTA ACCTATT CTCTGTGGGCTGTGTTTAAATAGCTGGAAAATAAGTC1 TCTTAGGCCATGCCTTATACGATTATATAAGTCTAT **Results list view** TCTTGGGCCATGCCTTATACGATTATATAGGCCTATTTTCGGT CTCTGTGGGTTGTGTTTAAAAGGCTAGAAAATAAGTC1 CT CT GT GGGCT GT GT GT TT AAAT GGCT GG AAAAT AAGT CT AT CTTGGGCCATGCCTTATACGATTATATAAGTATATTTTCGTCC CT CT GT GG GG CT GT GT GT GT GT GT GG GA A A T A A GT CT A T T T TGGGCCATGCCTTATACGATTATATAAGTCTATTTTTTGTCCCT CT CT GT GGGT TGT GT GT TT AAA AGGCT AGAAAAT AAGT CT AT TT T GGGCCATGCCTTATACGATTATATAAGTCTATTTTTCGTCCCTA Structure Variation CT CT GT GG GT T GT GT TT AAA AG GC T GG AA AAT AA GT CT AT TT T G CTGTGGGCTGTGTTTAAATGGCTGGAAAATAAGTCTATTTTGTA Distance>= v 5000 😂 P=20738 Q=23 PD=238 GGGTTGTGTTCAAATGGCTGGAAAATAAGTCTATTTTGTATTCC GGCTGTGTTTAAATGGCTGGAAAATAAGTCTATTTTGTATACCT Total: 15,376 GGTTGTGTTTAAATGGCTGGAAAATAAGTCTATTTTGTATACCT SV Detection GCTGTGGTTAAATGGCTGGAAAATAAGTCTATTTTGTATACCTA CTGTGTTTAAATGGCTGGAAAATAAGTCTATTTTGTATACC1 **Results list view** TGTGTTTAAATGGCTGGAAAATAAGTCTATTTTGTAT TGTGTTTAAATGGCTGGAAAATAAGTCTATTT < > ~ Selection Reference Position: 20697 Coverage SUM:27 A:2 T:14 G:11 C:0 N:0 Variant Frequency:0.4400 Details Statistics ON: 11,863,620 (A:3,386,857 T:3,379,903 G:2,545,229 C:2,551,630 N:1) 2,567,678 [+]1,284,493 [-]1,283,185 [contains 'N]40,791 Status: - 35 20642 - 20752 Row: --- Col: ---

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#### Hua Bao (Sun Yat-Sen University, Guangzhou) et al.

#### SAMtools pileup format

I	25514	G	G	42	0	25	5	^:.	ссссс
I	25515	т	т	42	0	25	5	••••	CC?CC
I	25516	А	G	48	48	25	7	GGGGG <b>^:</b> G <b>`:</b> g	CCCCCC5
I	25517	G	G	51	0	25	8	·····,^:, <sup>-</sup>	CCCCCC1?
I	25518	т	т	60	0	25	11	,,^:.^:,	CCCCCC3A<:;
I	25519	т	т	60	0	25	11	• • • • • • • • • • • • • • • • • • • •	CCCCCC>A@AA
I	25520	G	G	60	0	25	11	• • • • • • • • • • • •	CCCACC>A@ <a< td=""></a<>
I	25521	т	т	60	0	25	11	• • • • • • , , • , ,	CCCCCC?ACAA
I	25522	А	A	60	0	25	11	• • • • • • , , • , ,	CCCCCC>ACAA
I	25523	А	A	72	0	25	15	·····// ··/ ··· ······················	CCCCCC;ACAAC??C
I	25524	С	С	72	0	25	15	•••••	CCCCCC6< <a?c=9c< td=""></a?c=9c<>
I	25525	С	С	56	0	24	18		CCCCCC>ACA?C=AC<
I	25526	А	A	81	0	24	18	•••••	CCCCCC>ACAACAACA
I	25527	A	A	56	0	24	18	••••••G	CCCCCC?ACAA@A?CA

Fields: chromosome, position, reference base, consensus base, consensus quality, SNP quality, maximum mapping quality, coverage, base pile-up, base quality pile-up



#### Coverage vectors



#### <-- Solexa reads, aligned to genome

#### <-- coverage vector

Figure taken from Zhang et al., PLoS Comp. Biol. 2008



- A coverage vector (or "pile-up" vector) is an integer vector with on element per base pair in a chromosome, tallying the number of reads (or fragments) mapping onto each base pair.
- It is the essential intermediate data type in assays like ChIP-Seq or RNA-Seq

- Visualising coverage vectors is non-trivial, but essential for
  - quality control
  - hypothesis forming
  - etc.



#### Example: Histone modifications

- Barski et al. (Cell, 2007) have studied histone modification in the human genome with ChIP-Seq
- I use their data for H3K4me1 and H3K4me3 as example data.

• (Each data set is from two or three Solexa lanes)



#### Coverage vector for a full chromosome (chr10)



H3K4me3







#### Zoom in





#### Genome browser tracks

#### Tracks may contain

- Features (intervals with name)
  - without score
  - with score
- vectors (continuously varying score)

Standard formats for genome browser tracks

- BED
- GFF
- Wiggle fixedStep and variableStep



# Displaying tracks alongside annotation

- Either, upload your track file to a web-base browser
  - UCSC genome browse
  - Ensembl genome browser
- or use a stand-alone browser on your desktop computer
  - Integrated Genome Browser (IGB) [Genoviz]
  - Argo Genome Browser [Broad Institute]
  - Artemis [Sanger Institute]

Displaying large amounts of data requires patience and lots of RAM. Not all tools handle it well.







rtracklayer: Bioconductor package by M. Lawrence (FHCRC)

- import and export BED, Wiggle, and GFF files
- manipulate track data and get sub-views
- directly interact with a genome browser (UCSC or Argo) to drive displaying of track data



#### Difference between the track formats

- Formats for feature-by-feature data:
  - BED
  - GFF
- Formats for base-by-base scores
  - Wiggle
  - BedGraph
- Wiggle has three sub-types:
  - [BED-like]
  - variableStep
  - fixedStep



#### Wiggle format: variableStep and fixedStep

```
browser position chr19:59304200-59310700
browser hide all
track type=wiggle 0 name="varStepTrack" description="varStep example" \
    visibility=full autoScale=off viewLimits=0.0:25.0 color=50,150,255 \
    yLineMark=11.76 yLineOnOff=on priority=10
variableStep chrom=chr19 span=150
59304701 10.0
59304901 12.5
59305401 15.0
59305601 17.5
59305901 20.0
59306081 17.5
59306301 15.0
59307871 10.0
track type=wiggle 0 name="fixedStepTrack" description="fixedStep examle"
fixedStep chrom=chr19 start=59307401 step=300 span=200
1000
 900
 800
 700
 600
                                  All coordinates 1-based!
 500
 400
 300
 200
                                                                  EMBL-E
 100
```

#### bedGraph format

```
track type=bedGraph name="BedGraph Track"
chr19 59302000 59302300 -1.0
chr19 59302300 59302600 -0.75
chr19 59302600 59302900 -0.50
chr19 59302900 59303200 -0.25
chr19 59303200 59303500 0.0
chr19 59303500 59303800 0.25
chr19 59303800 59304100 0.50
chr19 59304100 59304400 0.75
chr19 59304400 59304700 1.00
```

All coordinates 0-based, half-open!

Specs: See UCSC Genome Browser web site



# Back to the bird's eyes view



H3K4me3





# H3K4me

We need a way to get a general overview on the data without either not seeing any details not getting lost in them.

A possible solution: Hilbert curve visualisation

S. An.: "Visualisation of genomic data with the Hilbert curve", Bioinformatics, Vol. 25 (2009) pp. 1231-1235



#### The Hilbert curve





#### What is hidden in here?





#### Hilbert plot of the constructed example vector













# Hilbert curve: Approaching the limit





#### Coverage vector for a full chromosome (chr10)



H3K4me3



350 "pu.me3.tbl" 300 250 200 150 100 50 0 4e+07 1e+08 2e+07 6e+07 8e+07 1.2e+08 1.4e+0 0

chrom. 10



# Hilbert plot of the coverage vectors



#### H3K4me1 (mono-methylation)

H3K4me3 (tri-methylation) EMBL-EBI



#### HilbertVis





#### HilbertVis

 stand-alone tool to display GFF, Wiggle, Maq map http://www.ebi.ac.uk/huber-srv/hilbert/ ( or Google for "hilbertvis")

• R package to display any long R vector

- either via commands for batch processing Bioconductor package "HilbertVis"
- or via GUI for exploring

Bioconductor package "HilbertVisGUI"



# Three-colour Hilbert plot



Overlay of the previous plots and exon density

red: mono-methylation green: tri-methylation

blue: exons



#### Other uses: Array-CGH



Log fold-changes between two *Arabidopsis* eco-types, chromosome 2

[Data courtesy of M. Seiffert, IPK Gatersleben



#### Other uses: Conservation scores



Human chromosome 10: Comparing phastCons conservation scores with exon density





#### GenomeGraphs

GenomeGraphs: Bioconductor package by S. Durrinck, UCB

• Load gene models from Ensembl via BiomaRt and plots them, alongside experimental data



#### GenomeGraphs: Code for sample plot

```
library(GenomeGraphs)
library(HilbertVis)
```

```
mart <- useMart("ensembl", dataset = "mmusculus_gene_ensembl")
start <- 57000000
end <- 59000000
plusStrand <- makeGeneRegion( chromosome = 10,
   start = start, end = end, strand = "+", biomart = mart )
minusStrand <- makeGeneRegion( chromosome = 10,
   start = start, end = end, strand = "-", biomart = mart )
genomeAxis <- makeGenomeAxis( )</pre>
```



#### GenomeGraphs: Code for sample plot, cont'd

```
track.ctcf <- makeBaseTrack(</pre>
   base = seq( start, end, length.out = 10000 ),
   value = shrinkVector(
      as.vector( cov.ctcf$chr10[start:end] ), 10000 ),
   dp = DisplayPars( lwd = 0.5, color="red", ylim=c(0, 50) ) )
track.gfp <- makeBaseTrack(</pre>
   base = seq( start, end, length.out = 10000 ),
   value = shrinkVector(
      as.vector( cov.gfp$chr10[start:end] ), 10000 ),
   dp = DisplayPars( lwd = 0.5, color="blue", ylim=c(0, 50) ) )
gdPlot( list( `plus`=plusStrand, `CTCF`=track.ctcf,
   genomeAxis, `GFP`=track.gfp, `minus`=minusStrand ) )
```





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