

# Package ‘BiSeq’

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**Description** The BiSeq package provides useful classes and functions to handle and analyze targeted bisulfite sequencing (BS) data such as reduced-representation bisulfite sequencing (RRBS) data. In particular, it implements an algorithm to detect differentially methylated regions (DMRs). The package takes already aligned BS data from one or multiple samples.

**License** LGPL-3

**biocViews** Genetics, Sequencing, MethylSeq, DNAMethylation

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---

annotateGRanges	<i>Annotates a GRanges object by means of a second GRanges object</i>
-----------------	---

---

### Description

Each genomic location of object is checked for overlapping with genomic ranges of regions. In case of an overlapping, this genomic location is marked as TRUE, or with the identifier of respective the regions object (if any).

### Usage

```
annotateGRanges(object, regions, name, regionInfo)
```

### Arguments

object	A GRanges.
regions	A GRanges.
name	A string specifying the name of the metadata column with the overlapping information to be added to object. Usually the type of the regions object, e.g. 'Promoter'.

**regionInfo**      **OPTIONAL:** A string or integer specifying the metadata column of regions object containing the identifiers, e.g. entrez gene IDs of the promoters. If not specified, the genomic locations of object are labeled as TRUE (overlap) or FALSE (no overlap).

### Details

If multiple ranges of regions overlap with a genomic region in object, the identifier names of the overlapping regions are separated by ','.

### Value

A GRanges object similar to object containing an additional metadata column with the overlapping information.

### Author(s)

Katja Hebestreit

### See Also

GRanges-class

### Examples

```
# load detected DMRs:
data(DMRs)

# annotate the DMRs with a GRanges object:
data(promoters)
DMRs.anno <- annotateGRanges(object = DMRs,
                             regions = promoters,
                             name = 'Promoter',
                             regionInfo = 'acc_no')

DMRs.anno
```

---

betaRegression

*A function to estimate and test a group factor within a beta regression*

---

### Description

This function models the methylation level within a beta regression. The first independent variable in formula is tested to be unequal to zero.

### Usage

```
betaRegression(formula, link, object, mc.cores, ...)
```

**Arguments**

formula	Symbolic description of the model. For the first independent variable the P value (Wald test) and the effect on methylation is returned. For details see below.
link	A character specifying the link function in the mean model ( $\mu$ ). Currently, "logit", "probit", "cloglog", "log", "loglog" are supported.
object	A BSrel object.
mc.cores	Passed to mclapply.
...	Other parameters passed to the betareg function.

**Details**

See betareg function for details.

mclapply

**Value**

A data.frame containing the position, chromosome, P value, estimated methylation level in group 1 and group 2 and methylation difference of group 1 and group 2.

**Author(s)**

Katja Hebestreit

**References**

Hebestreit, K., Dugas, M., and Klein HU. Detection of significantly differentially methylated regions in targeted bisulfite sequencing Data. In preparation. *Bioinformatics*. 2013 Jul 1;29(13):1647-53.

See also reference list in the documentation of [betareg](#).

**See Also**

betareg

**Examples**

```
# load RRBS data, subset to save time, find CpG clusters and smooth methylation data:
data(rrbs)
rrbs.small <- rrbs[1:1000,]
rrbs.clust.unlim <- clusterSites(object = rrbs.small,
                                groups = colData(rrbs)$group,
                                perc.samples = 4/5,
                                min.sites = 20,
                                max.dist = 100)

ind.cov <- totalReads(rrbs.clust.unlim) > 0
quant <- quantile(totalReads(rrbs.clust.unlim)[ind.cov], 0.9)
rrbs.clust.lim <- limitCov(rrbs.clust.unlim, maxCov = quant)

# with a small subset to save calculation time:
rrbs.part <- rrbs.clust.lim[1:100,]

predictedMeth <- predictMeth(object=rrbs.part)
```

```
betaResults <- betaRegression(formula = ~group, link = "probit",
                             object = predictedMeth, type="BR")
```

---

betaResults                      *The output of betaRegression*

---

### Description

Please see the package vignette for description.

### Usage

```
data(betaResults)
```

### Format

A data frame with 4276 observations on the following 10 variables:

chr a factor with levels chr1 chr2

pos a numeric vector

p.val a numeric vector

meth.group1 a numeric vector

meth.group2 a numeric vector

meth.diff a numeric vector

estimate a numeric vector

std.error a numeric vector

pseudo.R.sqrt a numeric vector

cluster.id a character vector

### Examples

```
data(betaResults)
head(betaResults)
```

---

betaResultsNull                      *The output of betaRegression for resampled data*

---

### Description

Please see the package vignette for description.

### Usage

```
data(betaResultsNull)
```

**Format**

A data frame with 4276 observations on the following 10 variables.

chr a factor with levels chr1 chr2  
 pos a numeric vector  
 p.val a numeric vector  
 meth.group1 a numeric vector  
 meth.group2 a numeric vector  
 meth.diff a numeric vector  
 estimate a numeric vector  
 std.error a numeric vector  
 pseudo.R.sqrt a numeric vector  
 cluster.id a character vector

**Examples**

```
data(betaResultsNull)
head(betaResultsNull)
```

---

binomLikelihoodSmooth *Calculates local likelihood estimations for binomial random variables*

---

**Description**

For a given set of binomial random variables with 1-dimensional coordinates, this function calculates the local likelihood estimation of the success probability  $p$  at a given point. For this purpose, a weighted likelihood estimation with weights obtained by a triangular kernel with given bandwidth is used. This can be used to predict values at points where no variable has been observed and/or to smooth observations using neighboured observations.

**Usage**

```
binomLikelihoodSmooth(pred.pos, pos, m, n, h)
```

**Arguments**

pred.pos	A vector of positions where $p$ should be estimated.
pos	A vector of positions where binomial variables have been observed.
m	A vector of length pos with the number of successful experiments.
n	A vector of length pos with the number of experiments.
h	The bandwidth of the kernel.

**Details**

For a given position  $x$ , the weighted likelihood for parameter  $p$

$$L(p; m, n, w) = \prod_{i=1}^k B(m_i | n_i, p)^{w_i}$$

is maximized.  $B$  denotes the binomial probability function. The weights  $w_i$  are calculated using a triangular kernel with bandwidth  $h$ :

$$w_i = K(x_i) = (1 - (|x - x_i|)/h) \mathbf{1}_{(|x - x_i|)/h \leq 1}$$

**Value**

A vector of length `pred.pos` giving the local likelihood estimation of the success probability  $p$  at the given positions.

**Author(s)**

Hans-Ulrich Klein

**See Also**

[predictMeth](#)

**Examples**

```
n = rpois(100, lambda=10)
E = c(rep(0.4, 30), rep(0.8, 40), rep(0.1, 30))
m = rbinom(100, n, E)
pos = 1:100
p_10 = binomLikelihoodSmooth(pos, pos, m, n, h=10)
p_20 = binomLikelihoodSmooth(pos, pos, m, n, h=20)

## Not run: plot(x=pos, y=m/n)
points(x=pos, y=p_10, col="green")
lines(x=pos, y=p_10, col="green")
points(x=pos, y=p_20, col="red")
lines(x=pos, y=p_20, col="red")
## End(Not run)
```

---

BSraw-class

*Class to contain raw Bisulfite Sequencing (BiSeq) Data*

---

**Description**

The BSraw class is derived from `RangedSummarizedExperiment` and contains a `SimpleList` of matrices named `methReads` and `totalReads` as assays.

**Objects from the Class**

Objects can be created by calls of the form `BSraw(metadata = list(), rowRanges, colData = DataFrame(row.names=colnames(methReads)), methReads, totalReads, ...)`.

However, one will most likely create a BSraw object when use [readBismark](#) to load data.

**Slots**

**metadata:** An optional list of arbitrary content describing the overall experiment.

**rowRanges:** Object of class "GRanges" containing the genome positions of CpG-sites covered by bisulfite sequencing. **WARNING:** The accessor for this slot is rowRanges, not rowRanges!

**colData:** Object of class "DataFrame" containing information on variable values of the samples.

**assays:** Object of class SimpleList of two matrices, named totalReads and methReads. The matrix totalReads contains the number of reads spanning a CpG-site. The rows represent the CpG sites in rowRanges and the columns represent the samples in colData. The matrix methReads contains the number of methylated reads spanning a CpG-site.

**Extends**

Class "[RangedSummarizedExperiment](#)", directly.

**Methods**

**totalReads** signature(x = "BSraw"): Gets the totalReads slot.

**totalReads<-** signature(x = "BSraw", value = "matrix"): Sets the totalReads slot.

**methReads** signature(x = "BSraw"): Gets the methReads slot.

**methReads<-** signature(x = "BSraw", value = "matrix"): Sets the methReads slot.

**combine** signature(x = "BSraw", y = "BSraw"): Combines two BSraw objects.

**Author(s)**

Katja Hebestreit

**See Also**

[RangedSummarizedExperiment](#), [BSrel-class](#), [readBismark](#)

**Examples**

```
showClass("BSraw")

## How to create a BSraw object by hand:
metadata <- list(Sequencer = "Sequencer", Year = "2013")
rowRanges <- GRanges(seqnames = "chr1",
                    ranges = IRanges(start = c(1,2,3), end = c(1,2,3)))
colData <- DataFrame(group = c("cancer", "control"),
                    row.names = c("sample_1", "sample_2"))
totalReads <- matrix(c(rep(10L, 3), rep(5L, 3)), ncol = 2)
methReads <- matrix(c(rep(5L, 3), rep(5L, 3)), ncol = 2)
BSraw(metadata = metadata,
      rowRanges = rowRanges,
      colData = colData,
      totalReads = totalReads,
      methReads = methReads)

## A more realistic example can be loaded:
data(rrbs)
rrbs

head(totalReads(rrbs))
```



```
head(methReads(rrbs))
```

---

**BSrel-class***Class to contain Bisulfite Sequencing (BiSeq) Data*

---

### Description

The BSrel class is derived from RangedSummarizedExperiment and contains a SimpleList of one matrix named methLevel as assays.

### Objects from the Class

Objects can be created by calls of the form `BSrel(metadata = list(), rowRanges, colData = DataFrame(row.names=colnames(methLevel)), methLevel, ...)`.

However, one will most likely create a BSraw object when use [readBismark](#) to load data.

### Slots

**metadata:** An optional list of arbitrary content describing the overall experiment.

**rowRanges:** Object of class "GRanges" containing the genome positions of CpG-sites covered by bisulfite sequencing. WARNING: The accessor for this slot is rowRanges, not rowRanges!

**colData:** Object of class "DataFrame" containing information on variable values of the samples.

**assays:** Object of class SimpleList of a matrix, named methLevel containing the methylation levels (between 0 and 1) per CpG site. The rows represent the CpG sites in rowRanges and the columns represent the samples in colData.

### Extends

Class "[RangedSummarizedExperiment](#)", directly.

### Methods

**methLevel** signature(x = "BSrel"): Gets the methLevel slot.

**methLevel<-** signature(x = "BSrel", value = "matrix"): Sets the methLevel slot.

**combine** signature(x = "BSrel", y = "BSrel"): Combines two BSrel objects.

### Author(s)

Katja Hebestreit

### See Also

[RangedSummarizedExperiment](#), [BSraw-class](#), [readBismark](#)

**Examples**

```
showClass("BSrel")

## How to create a BSrel object by hand:
metadata <- list(Sequencer = "Sequencer", Year = "2013")
rowRanges <- GRanges(seqnames = "chr1",
                     ranges = IRanges(start = c(1,2,3), end = c(1,2,3)))
colData <- DataFrame(group = c("cancer", "control"),
                    row.names = c("sample_1", "sample_2"))
methLevel <- matrix(c(rep(0.5, 3), rep(1, 3)), ncol = 2)
BSrel(metadata = metadata,
       rowRanges = rowRanges,
       colData = colData,
       methLevel = methLevel)

# Or get a BSrel object out of a BSraw object:
data(rrbs)
rrbs.rel <- rawToRel(rrbs)
```

---

clusterSites

*Assigns CpG cluster memberships on CpG sites within BSraw objects*


---

**Description**

Within a BSraw object clusterSites searches for agglomerations of CpG sites across all samples. In a first step the data is reduced to CpG sites covered in `round(perc.samples*ncol(object))` samples, these are called 'frequently covered CpG sites'. In a second step regions are detected where not less than `min.sites` frequently covered CpG sites are sufficiently close to each other (`max.dist`). Note, that the frequently covered CpG sites are considered to define the boundaries of the CpG clusters only. For the subsequent analysis the methylation data of all CpG sites within these clusters are used.

**Usage**

```
clusterSites(object, groups, perc.samples, min.sites, max.dist,
            mc.cores, ...)
```

**Arguments**

<code>object</code>	A BSraw.
<code>groups</code>	OPTIONAL. A factor specifying two or more sample groups within the given object. See Details.
<code>perc.samples</code>	A numeric between 0 and 1. Is passed to <code>filterBySharedRegions</code> .
<code>min.sites</code>	A numeric. Clusters should comprise at least <code>min.sites</code> CpG sites which are covered in at least <code>perc.samples</code> of samples, otherwise clusters are dropped.
<code>max.dist</code>	A numeric. CpG sites which are covered in at least <code>perc.samples</code> of samples within a cluster should not be more than <code>max.dist</code> bp apart from their nearest neighbors.
<code>mc.cores</code>	Passed to <code>mclapply</code> Default is 1.
<code>...</code>	Further arguments passed to the <code>filterBySharedRegions</code> function. closer than

## Details

There are three parameters that are important: `perc.samples`, `min.sites` and `max.dist`. For example, if `perc.samples=0.5`, the algorithm detects all CpG sites that are covered in at least 50% of the samples. Those CpG sites are called frequently covered CpG sites. In the next step the algorithm determines the distances between neighbored frequently covered CpG sites. When they are closer than (or close as) `max.dist` base pairs to each other, those frequently covered CpG sites and all other, less frequently covered CpG sites that are in between, belong to the same cluster. In the third step, each cluster is checked for the number of frequently covered CpG sites. If this number is less than `min.sites`, the cluster is discarded.

In other words: 1. The `perc.samples` parameter defines which are the frequently covered CpG sites. 2. The frequently covered CpG sites determine the boundaries of the clusters, depending on their distance to each other. 3. Clusters are discarded if they have too less frequently covered CpG sites.

If argument `group` is given, `perc.samples`, or `no.samples`, are applied for all group levels.

## Value

A BSraw object reduced to CpG sites within CpG cluster regions. A `cluster.id` metadata column on the `rowRanges` assigns cluster memberships per CpG site.

## Author(s)

Katja Hebestreit

## See Also

[filterBySharedRegions](#), `mclapply`

## Examples

```
data(rrbs)
rrbs.clust <- clusterSites(object = rrbs, groups = colData(rrbs)$group,
                          perc.samples = 4/5, min.sites = 20,
                          max.dist = 100)
```

---

<code>clusterSitesToGR</code>	<i>A function to obtain a GRanges object of CpG clusters from BSraw and BSrel objects</i>
-------------------------------	---

---

## Description

This function allows to get the start and end positions of CpG clusters from a BSraw or BSrel object, when there is a `cluster.id` column in the `rowRanges` slot.

## Usage

```
clusterSitesToGR(object)
```

## Arguments

`object` A BSraw or BSrel object with a `cluster.id` column in the `rowRanges` slot. Usually the output of `clusterSites`.

**Value**

An object of class GRanges is returned.

**Author(s)**

Katja Hebestreit

**See Also**

[clusterSites](#)

**Examples**

```
data(rrbs)
rrbs.clustered <- clusterSites(rrbs)

clusterSitesToGR(rrbs.clustered)
```

---

compareTwoSamples      *Detects DMRs by comparing two samples*

---

**Description**

Determines the differences of (smoothed) methylation levels between two samples and aggregates the sites surpassing a minimum difference to DMRs.

**Usage**

```
compareTwoSamples(object, sample1, sample2, minDiff, max.dist)
```

**Arguments**

object	A BSrel.
sample1	A numeric or character specifying the first sample to be used.
sample2	A numeric or character specifying the second sample to be used.
minDiff	A numeric greater than 0 and smaller or equal to 1.
max.dist	Numeric. The maximum distance between two CpG sites (or grid points) with absolute methylation differences greater or equal than minDiff in a DMR. If grid points are used: should be the same as grid.dist in predictMeth.

**Details**

This function determines the differences between the methylation levels of sample1 and sample2 for each site. Successive sites with methylation differences smaller or equal to minDiff are summarized.

**Value**

A GRanges object.

**Author(s)**

Katja Hebestreit

**See Also**[predictMeth](#)**Examples**

```
data(rrbs)
rrbs <- rrbs[, c(1,6)]
CpG.clusters <- clusterSites(object = rrbs, perc.samples = 1,
                             min.sites = 20, max.dist = 100)
predictedMeth <- predictMeth(object = CpG.clusters)
DMRs <- compareTwoSamples(predictedMeth, sample1 = 1, sample2 = 2,
                           minDiff = 0.3, max.dist = 100)
```

---

`covBoxplots`*Creates boxplots of coverages per sample*

---

**Description**

A boxplot per sample is plotted for the coverages of CpG-sites. It is constrained to CpG-sites which are covered in the respective sample (coverage  $\neq 0$  and not NA).

**Usage**

```
R
covBoxplots(object, ...)
```

**Arguments**

<code>object</code>	A BSraw.
<code>...</code>	Other graphical parameters passed to the boxplot function.

**Author(s)**

Katja Hebestreit

**See Also**[boxplot](#)**Examples**

```
data(rrbs)
covBoxplots(rrbs)
```

---

covStatistics	<i>Prints a short summary of coverage statistics per sample</i>
---------------	---

---

**Description**

This function produces information per samples about 1.) the covered CpG-sites 2.) the median of their coverages.

**Usage**

```
covStatistics(object)
```

**Arguments**

object            A BiSeq object.

**Author(s)**

Katja Hebestreit

**Examples**

```
data(rrbs)
covStatistics(rrbs)
```

---

DMRs	<i>The output of findDMRs</i>
------	-------------------------------

---

**Description**

Please see the package vignette for description.

**Usage**

```
data(DMRs)
```

**Format**

A GRanges of the chromosomes, start and end positions of the detected DMRs together with information (in the metadata columns) on DMRs: median.p, codemedian.meth.group1, codemedian.meth.group2, median.meth.diff.

**Examples**

```
data(DMRs)
head(DMRs)
```

---

`estLocCor`*Estimates the correlations of the z-scores*

---

**Description**

For each location the correlation of this location's z-score to  $\bar{Z}$  of its CpG cluster is estimated.

**Usage**

```
estLocCor(vario.sm)
```

**Arguments**

`vario.sm`      Output of `smoothVariogram`.

**Value**

A list:

`variogram`      A variogram matrix, usually created by `smoothVariogram` beforehand.  
`pValsList`      A list of the test results per CpG cluster.  
`sigma.cluster`      The standard deviations of z-scores within each cluster.  
`Z.cluster`      The arithmetic means of the z-scores for each cluster.  
`length.cluster`      The widths (number of pase pairs) of each cluster.

**Author(s)**

Katja Hebestreit

**References**

Yoav Benjamini and Ruth Heller (2007): False Discovery Rates for Spatial Signals. *American Statistical Association*, 102 (480): 1272-81.

**See Also**

[makeVariogram](#), [smoothVariogram](#)

**Examples**

```
data(betaResultsNull)

vario <- makeVariogram(betaResultsNull)
vario.sm <- smoothVariogram(vario, sill = 1)

locCor <- estLocCor(vario.sm)
```

---

filterByCov	<i>Filters regions (or single CpGs) of a BSraw object with a minimum coverage</i>
-------------	---

---

### Description

This method reduces a BSraw object to its regions (or single CpGs) with a minimum number of reads.

### Usage

```
filterByCov(object, minCov, global)
```

### Arguments

object	A BSraw.
minCov	Minimum number of reads overlapping the CpG sites.
global	A logical indicating whether the regions should achieve the minimum coverage in each sample. If <code>global = TRUE</code> the filtered object will consist of the regions achieving the minimum coverage in all samples. If <code>global = FALSE</code> (default) this function filters the regions for each sample separately, irrespectively of the coverages in other samples. <code>totalReads</code> and <code>methReads</code> are set to zero, if the minimum coverage is not obtained. Regions covered too sparse in all samples are dropped.

### Value

A BSraw object containing the CpGs or regions achieving the minimum coverage in all (if `global=TRUE`) or at least one (if `global=FALSE`) samples.

### Author(s)

Katja Hebestreit

### See Also

[filterBySharedRegions](#)

### Examples

```
data(rrbs)
rrbs.reduced <- filterByCov(object=rrbs, minCov=10, global=TRUE)
```





---

`findDMRs`*Aggregates CpG sites to DMRs*

---

**Description**

This function aggregates CpG sites to DMRs on the basis of their P values.

**Usage**

```
findDMRs(test.out, alpha, max.dist, diff.dir)
```

**Arguments**

<code>test.out</code>	An object returned by <code>betaRegression</code> .
<code>alpha</code>	OPTIONAL. A DMR contains CpG sites with P values smaller or equal than <code>alpha</code> .
<code>max.dist</code>	Numeric. The maximum distance between two P values smaller than <code>alpha</code> in a DMR. Should be the same as <code>grid.dist</code> in <code>predictMeth</code> .
<code>diff.dir</code>	Logical. Should DMRs be separated if the direction of methylation differences changes? If TRUE (default), then resulting DMRs will consist of sites which are all hypomethylated, or hypermethylated respectively.

**Value**

A GRanges object storing the start and end positions of the DMRs with information in metadata columns:

<code>median.p</code>	median of P values
<code>median.meth.group1</code>	median of modeled methylation level of group1.
<code>median.meth.group2</code>	median of modeled methylation level of group2.
<code>median.meth.diff</code>	median of difference of modeled methylation levels of group1 and group2.

**Author(s)**

Katja Hebestreit

**See Also**

[predictMeth](#), [betaRegression](#)

**Examples**

```
## Variogram under Null hypothesis (for resampled data):
data(vario)

plot(vario$variogram$v)
vario.sm <- smoothVariogram(vario, sill=0.9)
```

```

# auxiliary object to get the pValsList for the test
# results of interest:
data(betaResults)
vario.aux <- makeVariogram(betaResults, make.variogram=FALSE)

# Replace the pValsList slot:
vario.sm$pValsList <- vario.aux$pValsList

## vario.sm contains the smoothed variogram under the Null hypothesis as
## well as the p Values that the group has an effect on DNA methylation.

locCor <- estLocCor(vario.sm)

clusters.rej <- testClusters(locCor, FDR.cluster = 0.1)

clusters.trimmed <- trimClusters(clusters.rej, FDR.loc = 0.05)

DMRs <- findDMRs(clusters.trimmed, max.dist=100, diff.dir=TRUE)

```

---

globalTest	<i>Test whether at least one CpG is differentially methylated in a given genomic region</i>
------------	---

---

### Description

This method is a wrapper for conveniently invoking the `globaltest` method `gt` on a `BSrel-class` object. The `globaltest` can be applied to test against a high dimensional alternative in various regression models. E.g., it can be used to test whether at least one CpG is differentially methylated between two groups.

### Usage

```
globalTest(response, alternative, ...)
```

### Arguments

response	The response vector of the regression model. May be supplied as a vector or as a formula object. In the latter case, the right hand side of response defines the null hypothesis. The default null hypothesis is $\sim 1$ , i.e. only an intercept.
alternative	An object of <code>BSrel-class</code> defining the alternative. The CpGs are used as explanatory variable in the alternative regression model. The null hypothesis is that the coefficients of all CpGs are zero. If response is given as formula, <code>colData(alternative)</code> is used to obtain the respective data.
...	Other arguments passed to the <code>gt</code> method. The argument <code>subsets</code> can be given as <code>GRanges-class</code> object. Then, the <code>globaltest</code> is applied for each region using only the CpGs lying within the respective region. This is useful for, e.g., testing all promoter regions within function call.

### Details

For details see the documentation of the `gt` method in package `globaltest`.

**Value**

The function returns an object of class `gt` object. Several operations and diagnostic plots for this class are provided by the `globaltest` package.

**Author(s)**

Hans-Ulrich Klein

**References**

Goeman, J. J., van de Geer, S. A., and van Houwelingen, J. C. (2006). Testing against a high-dimensional alternative. *Journal of the Royal Statistical Society Series B- Statistical Methodology*, 68(3):477-493.

**See Also**

`link{gt}`, `link{BSrel}`

**Examples**

```
data(rrbs)
rrbs <- rawToRel(rrbs)
regions <- GRanges(IRanges(start=c(850000, 1920000, 500), end=c(879000, 1980000, 600)),
  seqnames=c("chr1", "chr2", "chr3"))

globalTest(group~1, rrbs)
globalTest(group~1, rrbs, subsets=regions)
```

---

limitCov

*Limits the coverage of a BSraw object*

---

**Description**

Number of methylated and unmethylated reads of a CpG site with coverage above `maxCov` are reduced such that the methylation level remains unchanged.

**Usage**

```
limitCov(object, maxCov)
```

**Arguments**

<code>object</code>	A <code>BSraw</code> .
<code>maxCov</code>	The maximum number of reads a CpG should have. All coverages above this threshold are limited. (Default is 50)

**Details**

This function might be useful prior to the use of `predictMeth` to limit the weights of CpGs with extremely high coverages. See `binomLikelihoodSmooth` for details.

**Value**

A BSraw object.

**Author(s)**

Katja Hebestreit

**See Also**

[predictMeth](#), [binomLikelihoodSmooth](#)

**Examples**

```
data(rrbs)

rrbs.clust.unlim <- clusterSites(object = rrbs,
                                groups = colData(rrbs)$group,
                                perc.samples = 4/5,
                                min.sites = 20, max.dist = 100)

covBoxplots(rrbs.clust.unlim)

# 90% quantile of coverage is 39x
quantile(totalReads(rrbs.clust.unlim)[totalReads(rrbs.clust.unlim)>0],
         0.9)

rrbs.clust.lim <- limitCov(rrbs.clust.unlim, maxCov = 39)
covBoxplots(rrbs.clust.lim)
```

---

logisticRegression      *Estimates and tests a group factor*

---

**Description**

It is used to fit a linear model on the log odds of each (smoothed) methylation level. The first independent variable in formula is tested to be unequal to zero.

**Usage**

```
logisticRegression(formula, link, object, mc.cores)
```

**Arguments**

formula	An object of class formula. For the first independent variable the p-value and the effect on methylation is returned.
link	A character specifying the link function. Currently, "logit", "probit", "cloglog", "log", "loglog" are supported.
object	A BSrel object.
mc.cores	Passed to mclapply.

**Value**

A data.frame containing the position, chromosome, P value, estimated methylation level in group 1 and group 2 and methylation difference of group 1 and group 2.

**Author(s)**

Katja Hebestreit

**See Also**

mclapply, glm

**Examples**

```
data(predictedMeth)

logisticResults <- logisticRegression(formula = ~group, link = "logit",
                                     object = predictedMeth)
```

---

makeVariogram	<i>Variogram estimator.</i>
---------------	-----------------------------

---

**Description**

A function which estimates the variogram of the z-scores in the given data frame.

**Usage**

```
makeVariogram(test.out, make.variogram, sample.clusters, max.dist)
```

**Arguments**

test.out	A data frame. Usually the output of betaRegression. Must contain columns chr, pos, p.val and cluster.id.
make.variogram	A logical. Default is TRUE.
sample.clusters	Can speed up variogram estimation significantly. Default is NULL, and all data is used to estimate the variogram. If set to numeric, the variogram will be estimated on the basis of the data of randomly selected sample.clusters only. Especially useful if there are many clusters.
max.dist	Can speed up variogram estimation significantly. The variogram is estimated for distances until this threshold. Default is 500 base pairs, since the variogram usually does not change for distances larger than 100 base pairs, because methylation of CpG sites further away are not correlated anymore. Especially useful if there are large clusters.

**Details**

For each CpG site the z-score is determined by `qnorm(1 - P value)`. The variogram of the z-scores of locations  $k$  and  $l$  within one cluster is estimated robustly by

$$2\hat{\gamma}(h) = [\text{median}(Z_k - Z_l)^2 : (s_k, s_l) \in N(h)] / .455$$

**Value**

A list:

- |           |  |
|-----------|--|
| variogram | A list of two: A matrix, called <code>v</code> with columns <code>h</code> and <code>v</code> , and a numeric, called <code>h.est</code> . <code>v</code> comprises the data that was used to estimate the variogram. <code>h.est</code> comprises the distances seen in the data. If <code>sample.clusters=NULL</code> , <code>h.est</code> is identical to <code>v\$h</code> . |
| pValsList | A list of data frames. Each data frame corresponds to a CpG cluster and contains same information as <code>test.out</code> plus the columns <code>z.score</code> and <code>pos.new</code> (position corresponding to the respective CpG cluster).  |

**Author(s)**

Katja Hebestreit

**References**

Yoav Benjamini and Ruth Heller (2007): False Discovery Rates for Spatial Signals. *American Statistical Association*, 102 (480): 1272-81.

**See Also**

[betaRegression](#)

**Examples**

```
data(betaResults)
vario <- makeVariogram(betaResults)
plot(vario$variogram$v)
```

---

plotBindingSites      *Plots the mean methylation of given regions*

---

**Description**

`plotBindingSites` takes several genomic regions (e.g. protein binding sites), centers them such that the position 0 refers to the center of each region and finally calculates the mean methylation of all regions for each given sample. If several samples are given, the median of the samples' methylation values and optionally other quantiles are plotted.

**Usage**

```
plotBindingSites(object, regions, width, groups, quantiles, bandwidth, ...)
```

**Arguments**

object	An object of class BSraw or BSrel.
regions	Regions given a GRanges object. The regions may have different widths.
width	The width of the genomic region that is plotted. Default value is the width of the largest given region.
groups	An optional factor defining two or more groups within the given object. The mean methylation is then plotted for each group separately.
quantiles	Other quantiles to be plotted besides the median. Default are the 25% and the 75% quantiles.
bandwidth	The bandwidth of the kernel smoother used for smoothing methylation values. Default value is 1/8 width.
...	Other graphical parameters passed to the plot function.

**Details**

First, all regions were expanded or shrunk to the given width by adding or removing base pairs symmetrically at both ends of the regions (not by scaling). A new coordinate system is centered at the middle of the equally sized regions. Next, the relative methylation values for each sample are averaged across all regions. That means, if there are several CpGs from different regions lying the same position, the mean methylation value is calculated for that position. Then, the median of these methylation values across all samples is calculated. Optionally, other quantiles are calculated, too. The median of the methylation is then plotted for each position after smoothing using a gaussian kernel with the given bandwidth.

If the given regions correspond to binding sites of a certain protein, the plot can be used to discover whether the protein induces changes in the DNA methylation in the proximity of its binding sites.

**Author(s)**

Hans-Ulrich Klein

**See Also**

[BSraw-class](#), [BSrel-class](#)

**Examples**

```
data(rrbs)
data(promoters)
plotBindingSites(object=rrbs,
                 regions=promoters,
                 width=4000,
                 groups=colData(rrbs)$group)
```



---

plotMeth	<i>Plots raw and smoothed methylation data for a given region</i>
----------	---

---

### Description

This function plots the raw and the smoothed methylation data for one sample and a given region. The smoothed data is shown as a line (one line per CpG cluster) and the raw data is shown as points with color intensities proportional to the coverage.

### Usage

```
plotMeth(object.raw, object.rel, region, col.lines, lwd.lines, col.points, ...)
```

### Arguments

<code>object.raw</code>	A BSraw with only one sample.
<code>object.rel</code>	A BSrel with only one sample.
<code>region</code>	A GRanges of length one.
<code>col.lines</code>	OPTIONAL. The color for the line representing the smoothed methylation values.
<code>lwd.lines</code>	OPTIONAL. The line width for the line representing the smoothed methylation values.
<code>col.points</code>	OPTIONAL. The color for the points representing the raw methylation levels.
<code>...</code>	Other graphical parameters passed to the plot function.

### Author(s)

Katja Hebestreit

### See Also

[plotSmoothMeth](#), [plot](#)

### Examples

```
data(rrbs)
data(predictedMeth)

region <- GRanges(seqnames="chr1",
                  ranges=IRanges(start = 875200,
                                 end = 875500))

plotMeth(object.raw = rrbs[,6],
         object.rel = predictedMeth[,6],
         region = region)
```

---

`plotMethMap`*Plots methylation values of multiple samples in a given region*

---

### Description

A heatmap like plot is generated showing the relative methylation of single CpG sites. Samples are clustered hierarchically.

### Usage

```
plotMethMap(object, region, groups, intervals, ...)
```

### Arguments

<code>object</code>	A <code>BSraw</code> or <code>BSrel</code> object storing the methylation values.
<code>region</code>	A <code>GRanges</code> object giving the region of interest.
<code>groups</code>	OPTIONAL. A factor that will be encoded by a color bar.
<code>intervals</code>	OPTIONAL. A logical indicating whether neighboured CpG sites should be placed side by side (if <code>FALSE</code> ) or whether the intervals between CpG sites should be preserved (if <code>TRUE</code> ).
<code>...</code>	Further arguments passed to the heatmap function.

### Details

The relative methylation values are passed to the heatmap function. Default colors are green (not methylated), black and red (methylated). To ensure that a relative methylation of 0 corresponds to green, 0.5 to black and 1 to red, the default value for the `zlim` argument of the `heatmap` function is set to `c(0, 1)`. And the default for the `scale` parameter is set to "none".

If argument `intervals` is set to `TRUE`, `region` should not be too large (< 1kb) and respect the resolution of your screen.

### Author(s)

Hans-Ulrich Klein

### See Also

`heatmap`, `BSraw-class`, `BSrel-class`, `filterBySharedRegions`, `filterByCov`

### Examples

```
data(rrbs)
data(predictedMeth)
data(DMRs)

plotMethMap(rrbs, region = DMRs[4], groups = colData(rrbs)[, "group"])

plotMethMap(predictedMeth, region = DMRs[4],
             groups = colData(rrbs)[, "group"], intervals = FALSE)
```

---

plotSmoothMeth	<i>Plots smoothed methylation values for a bunch of samples and a given region</i>
----------------	--

---

### Description

This function plots the smoothed methylation data as lines for a given region and all given samples. It is also possible to average the data for groups of samples.

### Usage

```
plotSmoothMeth(object.rel, region, groups, group.average, ...)
```

### Arguments

object.rel	A BSrel.
region	A GRanges of length one.
groups	OPTIONAL. A factor defining two or more sample groups within the given object.
group.average	OPTIONAL. A logical. If TRUE, then the data is averaged for the groups given in groups. Default is FALSE..
...	Other graphical parameters passed to the plot function.

### Author(s)

Katja Hebestreit

### See Also

[plotMeth](#), [plot](#)

### Examples

```
data(predictedMeth)
data(DMRs)

plotSmoothMeth(object.rel = predictedMeth,
               region = DMRs[3] + 200,
               groups = colData(predictedMeth)$group,
               col=c("magenta", "blue"))
legend("topright",
      lty=1,
      legend=levels(colData(predictedMeth)$group),
      col=c("magenta", "blue"))
```

---

predictedMeth	<i>The output of predictMeth</i>
---------------	----------------------------------

---

**Description**

Please see the package vignette for description.

**Usage**

```
data(predictedMeth)
```

**Format**

A BSrel object with the smoothed methylation data.

**Examples**

```
data(predictedMeth)
show(predictedMeth)
```

---

predictMeth	<i>Predicts methylation levels along CpG sites or for a grid of sites in CpG clusters.</i>
-------------	--

---

**Description**

Uses local regression to predict methylation levels per sample.

**Usage**

```
predictMeth(object, h, grid.dist, mc.cores)
```

**Arguments**

object	A BSraw with a cluster.id metadata column on the rowRanges, usually the output of clusterSites.
h	Bandwidth in base pairs. Large values produce a smoother curve. Default is 80.
grid.dist	OPTIONAL. If numeric, than methylation values are predicted at intervals of grid.dist base pairs. By default, methylation is smoothed at each CpG site.
mc.cores	Passed to mclapply. Default is 1.

**Details**

Uses binomLikelihoodSmooth with pos = CpG position, m = number methylated reads and n = number of reads. pred.pos corresponds to all CpG positions, or to the grid sites respectively, within the CpG clusters.

**Value**

A BSrel object containing the predicted methylation levels in the methLevel slot.

**Author(s)**

Katja Hebestreit

**See Also**[clusterSites](#), [binomLikelihoodSmooth](#), [mclapply](#)**Examples**

```
data(rrbs)

rrbs.clust.unlim <- clusterSites(object = rrbs,
                                groups = colData(rrbs)$group,
                                perc.samples = 4/5,
                                min.sites = 20, max.dist = 100)

ind.cov <- totalReads(rrbs.clust.unlim) > 0
quant <- quantile(totalReads(rrbs.clust.unlim)[ind.cov], 0.9)
rrbs.clust.lim <- limitCov(rrbs.clust.unlim, maxCov = quant)

# with a small subset to save calculation time:
rrbs.part <- rrbs.clust.lim[1:100,]

predictedMeth <- predictMeth(object=rrbs.part)
```

---

promoters

*A GRanges of promoters of the human genome*

---

**Description**

Please see the package vignette for description.

**Usage**

```
data(promoters)
```

**Format**

A GRanges object with the chromosomes, start and end positions of defined human promoter regions together with an accession number stored in a metadata column.

**Examples**

```
data(promoters)
head(promoters)
```

---

rawToRel	<i>Converts a BSraw object to a BSrel object</i>
----------	--

---

**Description**

Determines the methLevel matrix via: `methReads(object) / totalReads(object)`.

**Usage**

```
rawToRel(object)
```

**Arguments**

object            A BSraw.

**Value**

A BSrel.

**Author(s)**

Katja Hebestreit

**See Also**

[BSraw-class](#) [BSrel-class](#)

**Examples**

```
data(rrbs)
rrbs.rel <- rawToRel(rrbs)
```

---

readBismark	<i>Reads cytosine methylation stati determined by Bismark</i>
-------------	---

---

**Description**

Bismark is a bisulfite read mapper and methylation caller. This method reads Bismark's output files and returns a BSraw object.

**Usage**

```
readBismark(files, colData)
```

**Arguments**

files            A character pointing to cov files created by Bismark's `methylation_extractor` and `bismark2bedGraph`; see Details. This can be a compressed file (see [file](#)).

colData         Samples' names plus additional sample information as character, `data.frame` or `DataFrame`.

## Details

Input files are created with Bismark as follows (from the command line):

```
bismark_methylation_extractor -s --comprehensive test_sample.sam  
bismark2bedGraph -o CpG_context_test_sample.bedGraph CpG_context_test_sample.txt
```

This will output two files, a .bedGraph and a .cov file. We will import the CpG\_context\_test\_sample.cov using readBismark.

The colData argument should specify the sample names as character. Alternatively, a data.frame or DataFrame can be given. Then, the row names are used as sample names and the data frame is passed to the final BSraw object.

## Value

A BSraw object storing coverage and methylation information.

## Author(s)

Hans-Ulrich Klein

## References

<http://www.bioinformatics.bbsrc.ac.uk/projects/bismark/>

## See Also

[BSraw-class](#)

## Examples

```
file <- system.file("extdata", "CpG_context_test_sample.cov", package = "BiSeq")  
rrbs <- readBismark(file,  
                   colData= DataFrame(row.names="sample_1"))
```

---

rrbs

*RRBS data of APL patient samples and controls.*

---

## Description

RRBS data of the CpG sites CpG sites from genomic regions on p arms of chromosome 1 and 2 covered in at least one sample. Data was obtained from 5 APL patient samples and 5 control samples (APL in remission). RRBS data was preprocessed with the Bismark software version 0.5.

## Usage

```
rrbs
```

## Format

A [BSraw-class](#) object.

**Source**

Schoofs T, Rohde C, Hebestreit K, Klein HU, Goellner S, Schulze I, Lerdrup M, Dietrich N, Agrawal-Singh S, Witten A, Stoll M, Lengfelder E, Hofmann WK, Schlenke P, Buechner T, Hansen K, Berdel WE, Rosenbauer F, Dugas M, Mueller-Tidow C (2012). DNA methylation changes are a late event in acute promyelocytic leukemia and coincide with loss of transcription factor binding. *Blood*.

**References**

Krueger F, Andrews SR. Bismark: a flexible aligner and methylation caller for Bisulfite-Seq applications. *Bioinformatics*. 2011;27:1571-1572.

**Examples**

```
data(rrbs)
show(rrbs)
```

---

smoothVariogram	<i>Smoothes variogram</i>
-----------------	---------------------------

---

**Description**

Nonparametric smoothing with kernel regression estimators and adaptable bandwidth for variogram smoothing.

**Usage**

```
smoothVariogram(variogram, sill, bandwidth)
```

**Arguments**

variogram	A list or a matrix. Usually the output of makeVariogram.
sill	A numeric. The sill (upper bound) of the variogram. See Details.
bandwidth	A numeric vector of same length as the variogram (number of rows). Default: seq(10, 1000, length.out=nrow(variogram)). See Details.

**Details**

It is necessary to smooth the variogram. Especially for greater  $h$  the variogram tends to oscillate strongly. This is the reason why the default bandwidth increases with increasing  $h$ . Nevertheless, the smoothed variogram may further increase or decrease after a horizontal part (sill). This is mostly due to the small number of observations for high distances. To wipe out this bias it is useful to set the smoothed variogram to a fixed value above a certain  $h$ , usually the mean value of the horizontal part. If a smoothed value  $v.sm$  is greater than `sill` for distance  $h_{range}$ , this  $v.sm$  and all other smoothed values with  $h > h_{range}$  are set to `sill`. Internally, the function `lokerns` from package `lokerns` is used for smoothing.

**Value**

The variogram matrix (or a list with the variogram matrix) with an additional column of the smoothed  $v$  values.



**Author(s)**

Katja Hebestreit

**See Also**[makeVariogram](#), [lokerns](#)**Examples**

```
data(vario)

# Find out the sill (this is more obvious for larger data sets):
plot(vario$variogram$v)

vario.sm <- smoothVariogram(vario, sill = 0.9)

plot(vario$variogram$v)
lines(vario.sm$variogram[,c("h", "v.sm")],
      col = "red")
```

---

**summarizeRegions***Aggregates methylation information of single CpG sites*

---

**Description**

This method summarizes the methylation states of single CpG sites to a single methylation state for a given genomic region.

**Usage**

```
summarizeRegions(object, regions, outputAll)
```

**Arguments**

<code>object</code>	An <code>BSraw</code> or <code>BSrel</code> object.
<code>regions</code>	A <code>GRanges</code> object storing the genomic regions.
<code>outputAll</code>	A logical. If <code>outputAll = TRUE</code> , all regions will be returned. If <code>FALSE</code> (default), regions are dropped if their coverage is zero.

**Details**

When the given object is of class `BSraw-class`, all (methylated) reads of all CpG site lying within a region are summed up and assign as total number of (methylated) reads to that region. It is recommended to use `limitCov` before applying `summarizeRegions` to an `BSraw-class` object in order to avoid an excessive influence of a single CpG site on the methylation value of a region. When the given object is of class `BSrel-class`, the mean relative methylation of all CpGs within a region is assign to that region.

The `rowRanges` slot of the returned object is the given object regions with all columns preserved.

**Value**

An BSraw or an BSrel object storing methylation information about the given regions.

**Author(s)**

Hans-Ulrich Klein

**See Also**

[BSraw-class](#), [BSrel-class](#), [limitCov](#)

**Examples**

```
data(rrbs)
rrbs.clustered <- clusterSites(rrbs)
regions <- clusterSitesToGR(rrbs.clustered)

rrbs <- limitCov(rrbs, maxCov=50)
rrbsRegion <- summarizeRegions(rrbs, regions)
totalReads(rrbsRegion)
```

---

testClusters

*Tests CpG clusters*

---

**Description**

CpG clusters are tested with a cluster-wise FDR level.

**Usage**

```
testClusters(locCor, FDR.cluster)
```

**Arguments**

locCor	Output of estLocCor.
FDR.cluster	A numeric. The WFDR (weighted FDR) level at which the CpG clusters should be tested. Default is 0.05.

**Details**

CpG clusters containing at least one differentially methylated location are detected.

**Value**

A list is returned:

FDR.cluster	Chosen WFDR (weighted FDR) for clusters.
CpGs.clust.reject	A list of the CpG sites together with test results within clusters that were rejected.

CpGs.clust.not.reject	A list of the CpG sites together with test results within clusters that were not rejected.
clusters.reject	A GRanges of the clusters that were rejected.
clusters.not.reject	A GRanges of the clusters that were not rejected.
sigma.clusters.reject	The standard deviations for z-scores within each rejected cluster.
variogram	The variogram matrix.
m	Number of clusters tested.
k	Number of clusters rejected.
u.1	Cutoff point of the largest P value rejected.

**Author(s)**

Katja Hebestreit

**References**

Yoav Benjamini and Ruth Heller (2007): False Discovery Rates for Spatial Signals. *American Statistical Association*, 102 (480): 1272-81.

**See Also**

[estLocCor](#), [trimClusters](#)

**Examples**

```
## Variogram under Null hypothesis (for resampled data):
data(vario)

plot(vario$variogram$v)
vario.sm <- smoothVariogram(vario, sill=0.9)

# auxiliary object to get the pValsList for the test
# results of interest:
data(betaResults)
vario.aux <- makeVariogram(betaResults, make.variogram=FALSE)

# Replace the pValsList slot:
vario.sm$pValsList <- vario.aux$pValsList

## vario.sm contains the smoothed variogram under the Null hypothesis as
## well as the p Values that the group has an effect on DNA methylation.

locCor <- estLocCor(vario.sm)

clusters.rej <- testClusters(locCor, FDR.cluster = 0.1)
```

---

trimClusters	<i>Trims CpG clusters</i>
--------------	---------------------------

---

**Description**

CpG clusters rejected in a previous step are trimmed.

**Usage**

```
trimClusters(clusters.rej, FDR.loc)
```

**Arguments**

clusters.rej	Output of testClusters.
FDR.loc	Location-wise FDR level. Default is 0.2.

**Details**

Not differentially methylated CpG sites are removed within the CpG clusters rejected by testClusters.

**Value**

A data.frame containing the differentially methylated CpG sites.

**Author(s)**

Katja Hebestreit

**References**

Yoav Benjamini and Ruth Heller (2007): False Discovery Rates for Spatial Signals. American Statistical Association, 102 (480): 1272-81.

**See Also**

[testClusters](#)

**Examples**

```
## Variogram under Null hypothesis (for resampled data):
data(vario)

plot(vario$variogram$v)
vario.sm <- smoothVariogram(vario, sill=0.9)

# auxiliary object to get the pValsList for the test
# results of interest:
data(betaResults)
vario.aux <- makeVariogram(betaResults, make.variogram=FALSE)

# Replace the pValsList slot:
vario.sm$pValsList <- vario.aux$pValsList
```

```
## vario.sm contains the smoothed variogram under the Null hypothesis as
## well as the p Values that the group has an effect on DNA methylation.

locCor <- estLocCor(vario.sm)

clusters.rej <- testClusters(locCor, FDR.cluster = 0.1)

clusters.trimmed <- trimClusters(clusters.rej, FDR.loc = 0.05)
```

---

vario	<i>Output of makeVariogram</i>
-------	--------------------------------

---

### Description

Please see the package vignette for description.

### Usage

```
data(vario)
```

### Format

A list consisting of the variogram (a matrix) and the pValsList (a list of the data frames of test results).

### Examples

```
data(vario)
names(vario)
```

---

writeBED	<i>Writes BSraw and BSrel data to a bed file suitable for the IGV</i>
----------	---

---

### Description

The created bed files contains an entry for each CpG site. Strand information, relative methylation and absolute number of reads covering the CpG sites are stored. The relative methylation is indicated by colors: green via black to red for unmethylated to methylated.

### Usage

```
writeBED(object, name, file)
```

### Arguments

object	A BSraw or BSrel object.
name	Track names (sample names) written to the bed file's header.
file	Character vector with names of the bed file.

**Details**

The written bed file contains the following extra information:

1. score: the relative methylation of the CpG site
2. name: the coverage of the CpG site
3. itemRgb: a color value visualizing the methylation score

A separate bed file is created for each sample in the given object. The lengths of the arguments `name` and `file` should equal the number of samples.

**Value**

Nothing. Bed files are written.

**Author(s)**

Hans-Ulrich Klein

**See Also**

[readBismark](#)

**Examples**

```
data(rrbs)
s1 <- rrbs[,1]
out <- tempfile(), fileext = ".bed")
writeBED(s1, name = colnames(s1), file = out)
```

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