Package ‘flowWorkspace’

January 27, 2020

Type Package

Title Infrastructure for representing and interacting with gated and ungated cytometry data sets.

Version 3.35.8

Date 2011-06-10

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Description This package is designed to facilitate comparison of automated gating methods against manual gating done in flowJo. This package allows you to import basic flowJo workspaces into BioConductor and replicate the gating from flowJo using the flowCore functionality. Gating hierarchies, groups of samples, compensation, and transformation are performed so that the output matches the flowJo analysis.

License file LICENSE

License_restricts_use yes

LazyLoad yes

Imports Biobase, BiocGenerics, cytolib (>= 1.9.7), lattice, latticeExtra, XML, ggplot2, graph, graphics, grDevices, methods, stats, stats4, utils, RBGL, tools, Rgraphviz, data.table, dplyr, Rcpp, stringr, scales, matrixStats, RcppParallel, RProtoBufLib, digest, flowCore(>= 1.53.4), ncdfFlow(>= 2.25.4)


Suggests testthat, flowWorkspaceData (>= 2.23.1), knitr, ggcyto, parallel, CytoML, openCyto

LinkingTo Rcpp, BH(>= 1.62.0-1), RProtoBufLib(>= 1.9.4), cytolib(>= 1.9.8), Rhol5lib, RcppArmadillo, RcppParallel(>= 4.4.2-1)

VignetteBuilder knitr
biocViews  ImmunoOncology, FlowCytometry, DataImport, Preprocessing,
          DataRepresentation

SystemRequirements  GNU make, C++11

Encoding  UTF-8

RoxygenNote  7.0.2

git_url  https://git.bioconductor.org/packages/flowWorkspace

git_branch  master

git_last_commit  425b47ca

git_last_commit_date  2020-01-21

Date/Publication  2020-01-26

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flowWorkspace-package

Import and replicate flowJo workspaces and gating schemes using flowCore.

Description

Import flowJo workspaces into R. Generate the flowJo gating hierarchy and gates using flowCore functionality. Transform and compensate data in accordance with flowJo settings. Plot gates, gating hierarchies, population statistics, and compare flowJo vs flowCore population summaries.

Details

Package:     flowWorkspace
Type:        Package
Version:     0.5.40
Date:        2011-03-04
License:     Artistic 2.0
LazyLoad:    yes
Depends:     R (>= 2.16.0), Rcpp (>= 0.9.9)
**asinhtGml2_trans**

**Author(s)**

Greg Finak, Mike Jiang

**References**

http://www.rglab.org/

---

**asinhtGml2_trans  Inverse hyperbolic sine transformation.**

**Description**

Used to construct inverse hyperbolic sine transform object.

**Usage**

asinhtGml2_trans(..., n = 6, equal.space = FALSE)

**Arguments**

- ...  parameters passed to asinh_Gml2
- n  desired number of breaks (the actual number will be different depending on the data range)
- equal.space  whether breaks at equal-spaced intervals

**Value**

asinhtGml2 transformation object

**Examples**

trans.obj <- asinhtGml2_trans(equal.space = TRUE)
data <- 1:1e3
brks.func <- trans.obj[["breaks"]]
brks <- brks.func(data)
brks # fasinhs space displayed at raw data scale

#transform it to verify it is equal-spaced at transformed scale
trans.func <- trans.obj[["transform"]]
brks.trans <- trans.func(brks)
brks.trans
asinh_Gml2

Description

hyperbolic sine/inverse hyperbolic sine transform function constructor. It is simply a special form of flowjo_fasinh with length set to 1 and different default values for parameters t,m,a.

Usage

asinh_Gml2(T = 262144, M = 4.5, A = 0, inverse = FALSE)

Arguments

- **T**: numeric the maximum value of input data
- **M**: numeric the full width of the transformed display in asymptotic decades
- **A**: numeric Additional negative range to be included in the display in asymptotic decades
- **inverse**: whether to return the inverse function

Value

fasinh/fsinh transform function

Examples

trans <- asinh_Gml2()
data.raw <- c(1,1e2,1e3)data.trans <- trans(data.raw)data.trans

inverse.trans <- asinh_Gml2(inverse = TRUE)inverse.trans(data.trans)

booleanFilter-class

A class describing logical operation (& or |) of the reference populations

Description

booleanFilter class inherits class expressionFilter and exists for the purpose of methods dispatching.

Usage

booleanFilter(expr, ..., filterId = "defaultBooleanFilter")

char2booleanFilter(expr, ..., filterId = "defaultBooleanFilter")
cf_get_h5_file_path

Arguments

expr expression
... further arguments to the expression
filterId character identifier

See Also

add GatingHierarchy

Examples

# "4+/TNFa+" and "4+/IL2+" are two existing gates
# note: no spaces between node names and & , ! operators
booleanFilter("4+/TNFa+&!4+/IL2+")

# programmatically
n1 <- "4+/TNFa+
 n2 <- "4+/IL2+
 exprs <- paste0(n1, "&", n2)
call <- substitute(booleanFilter(v), list(v = as.symbol(exprs)))
eval(call)

Description

Return the file path of the underlying h5 file

Usage

cf_get_h5_file_path(cf)

Arguments

cf cytoframe object

Details

For the in-memory version of cytoframe, it returns an empty string. This can be used to check whether it is on-disk format.

See Also

Other cytoframe/cytoset IO functions: cf_write_h5(), cs_get_h5_file_path(), load_cytoframe_from_fcs(), load_cytoframe_from_h5(), load_cytoset_from_fcs()
**cf_write_h5**  
*Save the cytoframe as h5 format*

**Description**

Save the cytoframe as h5 format

**Usage**

cf_write_h5(cf, filename)

**Arguments**

cf        cytoframe object
filename   the full path of the output h5 file

**See Also**

Other cytoframe/cytoset IO functions: cf_get_h5_file_path(), cs_get_h5_file_path(), load_cytoframe_from_fcs(), load_cytoframe_from_h5(), load_cytoset_from_fcs()

---

**clone**  
*clone a GatingSet*

**Description**

clone a GatingSet

**Usage**

clone(x, ...)

gs_clone(x, h5_dir = tempdir())

gs_copy_tree_only(x)

**Arguments**

x         A GatingSet
...

h5_dir = tempdir() the directory to store the h5-based flow data matrix

**Details**

Note that the regular R assignment operation on a GatingSet object does not return the copy as one would normally expect because the GatingSet contains environment slots (and external pointer for GatingSet), which require deep-copying. So make sure to use this clone method in order to make a copy of existing object.
compensate

Value

A copy of a given GatingSet.

Examples

## Not run:
#gs is a GatingSet
gs2 <- gs_clone(gs) #gs2 is independent from gs and have its own copy of both gating trees and flow data
gs3 <- gs_copy_tree_only(gs) #gs3 has its own copy of gating trees but share the same flow data with original gs

## End(Not run)

compensate compensate the flow data associated with the GatingSet

Description

The compensation is saved in the GatingSet and can be retrieved by gh_get_compensations.

Usage

compensate(x, spillover)

Arguments

x GatingSet, GatingSetList, cytoframe, or cytoset
spillover compensation object or spillover matrix or a list of compensation objects

Value

a GatingSet, GatingSetList, cytoframe, or cytoset object with the underlying flow data compensated.

Examples

## Not run:
cfile <- system.file("extdata", "compdata", "compmatrix", package="flowCore")
comp.mat <- read.table(cfile, header=TRUE, skip=2, check.names = FALSE)
## create a compensation object
comp <- compensation(comp.mat, compensationId="comp1")
#add it to GatingSet
gs <- compensate(gs, comp)

## End(Not run)
compute_timestep  

**Description**
compute time step from fcs keyword

**Usage**
```r
compute_timestep(kw, unit.range, timestep.source = c("TIMESTEP", "BTIM"))
```

**Arguments**
- `kw`: list of keywords
- `unit.range`: the actual measured time unit range
- `timestep.source`: either "TIMESTEP" or "BTIM". prefer to STIMESTEP keyword when it is non NULL

convert_legacy_gs  

**Description**
convert the legacy(mixed with R and C++ files) GatingSet archive to the new format(C++ only)

**Usage**
```r
convert_legacy_gs(from, to)
convert_legacy_gslist(from, to)
```

**Arguments**
- `from`: the old archive path
- `to`: the new archive path

**Examples**
```r
## Not run:
convert_legacy_gs(old_gs_path, new_gs_path)
## End(Not run)
```
cs_add_sample

Add a cytoframe to a cytoset

Description

Add a cytoframe to a cytoset

Usage

\[\text{cs_add_sample}(cs, sn, fr)\]

cs_get_h5_file_path

Return the file path of the underlying h5 files

Description

Return the file path of the underlying h5 files

Usage

\[\text{cs_get_h5_file_path}(x)\]

See Also

Other cytoframe/cytoset IO functions: cf_get_h5_file_path(), cf_write_h5(), load_cytoframe_from_fcs(), load_cytoframe_from_h5(), load_cytoset_from_fcs()

cytoframe

cytoframe: A reference class for efficiently managing the data representation of a flowFrame

Description

This class serves the same purpose as the flowFrame class: to store quantitative data on cell populations from a single FCS run. The primary difference is in the underlying representation of the data. While flowFrame objects store the underlying data matrix in the exprs slot as an R object, cytoframe objects store the matrix (as well as the data from the other slots) in a 'C' data structure that is accessed through an external pointer. This allows for greater optimization of data operations including I/O, parsing, transformation, and gating.
Details

From the user's standpoint, interacting with a cytoframe is very similar to interacting with a flowframe, with one important difference. While operations such as subsetting or copying a flowframe using the standard R assignment operator (<-) will perform a deep copy of the data in its slots, the same operations on a cytoframe will produce a view to the same underlying data as the original object. This means that changes made to the cytoframe resulting from subsetting or copying will affect the original cytoframe. If a deep copy of the underlying data is desired, the `realize_view` method will accomplish this.

Because the cytoframe class inherits from flowFrame, the flowFrame slots are present but not utilized. Thus, attempting to access them directly will yield empty data structures. However, the `exprs`, `parameters`, or `description` methods work in a manner similar to a flowFrame by accessing the same information from the underlying data structure.

Methods

There are separate documentation pages for most of the methods listed here which should be consulted for more details.

[ Subsetting. Returns an object of class cytoframe. The syntax for subsetting is similar to that of data.frames. In addition to the usual index vectors (integer and logical by position, character by parameter names), cytoframes can be subset via `filterResult` and `filter` objects.

Usage:
```r
cytoframe[i,j]
cytoframe[filter,]
cytoframe[filterResult,]
```

Note that the value of argument `drop` is ignored when subsetting cytoframes.

$ Subsetting by channel name. This is similar to subsetting of columns of data.frames, i.e., `frame$FSC.H` is equivalent to `frame[, "FSC.H"]`. Note that column names may have to be quoted if they are not valid R symbols (e.g. `frame$"FSC-H"`).

`exprs, exprs<-` exprs returns an object of class matrix containing the measured intensities. Rows correspond to cells, columns to the different measurement channels. The `colnames` attribute of the matrix is supposed to hold the names or identifiers for the channels. The `rownames` attribute would usually not be set.

`exprs<-` replaces the raw data intensities. The replacement value must be a numeric matrix with colnames matching the parameter definitions. Implicit subsetting is allowed (i.e. less columns in the replacement value compared to the original cytoframe, but all have to be defined there).

Usage:
```r
exprs(cytoframe)
exprs(cytoframe) <- value
```

`head, tail` Show first/last elements of the raw data matrix

Usage:
```r
head(cytoframe)
tail(cytoframe)
```

`description, description<-` Extract or replace the whole list of annotation keywords obtained from the metadata of the original FCS file. Usually one would only be interested in a subset of keywords, in which case the `keyword` method is more appropriate. The optional `hideInternal` parameter can be used to exclude internal FCS parameters starting with $.

Usage:
```r
description(cytoframe)
description(cytoframe) <- value
```
```r
description(cytoframe)
description(cytoframe) <- value

keyword, keyword<- Extract all entries or a single entry from the annotations by keyword or replace the entire list of key/value pairs with a new named list. See keyword for details.
Usage:
keyword(cytoframe)
keyword(cytoframe, character)
keyword(cytoframe) <- list(value)

parameters, parameters<- Extract parameters and return an object of class AnnotatedDataFrame containing information about each column of the cytoframe, or replace such an object. This information will generally be filled in by load_cytoframe_from_fcs or similar functions using data from the FCS keywords describing the parameters. To access the actual parameter annotation, use pData(parameters(cytoframe)). Replacement is only valid with AnnotatedDataFrames containing all varLabels name, desc, range, minRange and maxRange, and matching entries in the name column to the colnames of the exprs matrix. See parameters for more details.
Usage:
parameters(cytoframe)
parameters(cytoframe) <- value

show Display details about the cytoframe object.

summary Return descriptive statistical summary (min, max, mean and quantile) for each channel
Usage:
summary(cytoframe)

plot Basic plots for cytoframe objects. If the object has only a single parameter this produces a histogram. For exactly two parameters we plot a bivariate density map (see smoothScatter and for more than two parameters we produce a simple splom plot. To select specific parameters from a flowFrame for plotting, either subset the object or specify the parameters as a character vector in the second argument to plot. The smooth parameters lets you toggle between density-type smoothScatter plots and regular scatterplots. For far more sophisticated plotting of flow cytometry data, see the flowViz package.
Usage:
plot(cytoframe,...)
plot(cytoframe, character,...)
plot(cytoframe, smooth=FALSE,...)

ncol, nrow, dim Extract the dimensions of the data matrix.
Usage:
ncol(cytoframe)
nrow(cytoframe)
dim(cytoframe)

featureNames, colnames, colnames<-. colnames and featureNames are synonyms, they extract parameter names (i.e., the colnames of the data matrix). For colnames there is also a replacement method. This will update the name column in the parameters slot as well.
Usage:
featureNames(cytoframe)
colnames(cytoframe)
colnames(cytoframe) <- value
```
**names**  Extract pretty formatted names of the parameters including parameter descriptions.

*Usage:*

names(cytoframe)

**identifier**  Extract GUID of a cytoframe. Returns the file name if no GUID is available. See identifier for details.

*Usage:*

identifier(cytoframe)

**range**  Get instrument or actual data range of the cytoframe. Note that instrument dynamic range is not necessarily the same as the range of the actual data values, but the theoretical range of values the measurement instrument was able to capture. The values of the dynamic range will be transformed when using the transformation methods for cytoframes.

*Parameters:*

- x: cytoframe object.
- type: Range type. either "instrument" or "data". Default is "instrument"

*Usage:*

range(x,type = "data")

**each_row, each_col**  Apply functions over rows or columns of the data matrix. These are convenience methods. See each_col for details.

*Usage:*

each_row(cytoframe,function,...)
each_col(cytoframe,function,...)

**transform**  Apply a transformation function on a cytoframe object. This uses R’s transform function by treating the cytoframe like a regular data.frame. flowCore provides an additional inline mechanism for transformations (see %on%) which is strictly more limited than the out-of-line transformation described here.

*Usage:*

transform(cytoframe,translist,...)

**filter**  Apply a filter object on a cytoframe object. This returns an object of class filterResult, which could then be used for subsetting of the data or to calculate summary statistics. See filter for details.

*Usage:*

filter(cytoframe,filter)

**split**  Split cytoframe object according to a filter, a filterResult or a factor. For most types of filters, an optional flowSet=TRUE parameter will create a flowSet rather than a simple list. See split for details.

*Usage:*

split(cytoframe,filter,flowSet=FALSE,...)
split(cytoframe,filterResult,flowSet=FALSE,...)
split(cytoframe,factor,flowSet=FALSE,...)

**Subset**  Subset a cytoframe according to a filter or a logical vector. The same can be done using the standard subsetting operator with a filter, filterResult, or a logical vector as first argument.

*Usage:*

Subset(cytoframe,filter)
Subset(cytoframe,logical)
**cbind2** Not yet implemented.

Expand a cytoframe by the data in a numeric matrix of the same length. The matrix must have column names different from those of the cytoframe. The additional method for numerics only raises a useful error message.

*Usage:*

```r
cbind2(cytoframe, matrix)
cbind2(cytoframe, numeric)
```

**compensate** Apply a compensation matrix (or a `compensation` object) on a cytoframe object. This returns a compensated cytoframe.

*Usage:*

```r
compensate(cytoframe, matrix)
compensate(cytoframe, data.frame)
```

**decompensate** Not yet implemented.

Reverse the application of a compensation matrix (or a `compensation` object) on a cytoframe object. This returns a decompensated cytoframe.

*Usage:*

```r
decompensate(cytoframe, matrix)
decompensate(cytoframe, data.frame)
```

**spillover** Extract spillover matrix from description slot if present. It is equivalent to `keyword(x,c("spillover", "SPILL"))` thus will simply return a list of keywords value for "spillover" and "SPILL".

*Usage:*

```r
spillover(cytoframe)
```

**realize_view** Returns a new cytoframe with its own copy of the underlying data (a deep copy). The optional filepath argument accepts a string to specify a full filename for storing the new copy of the data in h5 format.

*Usage:*

```r
realize_view(cytoframe, filepath)
```

**Author(s)**

F. Hahne, B. Ellis, P. Haaland and N. Le Meur

**See Also**

`flowSet`, `read.FCS`

---

**cytoset**

`cytoset`: a reference class for efficiently managing the data representation of a `flowSet`.

**Description**

This class is a container for a set of cytoframe objects, analogous to a `flowSet`. 

---

[100x761]cytoset
[118x733]cbind2 Not yet implemented.
[140x721]Expand a cytoframe by the data in a numeric matrix of the same length. The matrix must have column names different from those of the cytoframe. The additional method for numerics only raises a useful error message.
[140x697]Usage:
[143x666]cbind2(cytoframe,matrix)
[143x651]cbind2(cytoframe,numeric)
[118x632]compensate Apply a compensation matrix (or a compensation object) on a cytoframe object. This returns a compensated cytoframe.
[140x620]Usage:
[143x589]compensate(cytoframe,matrix) compensate(cytoframe,data.frame)
[118x571]decompensate Not yet implemented.
[140x559]Reverse the application of a compensation matrix (or a compensation object) on a cytoframe object. This returns a decompensated cytoframe.
[140x547]Usage:
[143x516]decompensate(cytoframe,matrix) decompensate(cytoframe,data.frame)
[118x531]spillover Extract spillover matrix from description slot if present. It is equivalent to keyword(x,c("spillover", "SPILL")) thus will simply return a list of keywords value for "spillover" and "SPILL".
[140x485]Usage:
[143x455]spillover(cytoframe)
[118x436]realize_view Returns a new cytoframe with its own copy of the underlying data (a deep copy). The optional filepath argument accepts a string to specify a full filename for storing the new copy of the data in h5 format.
[140x381]Usage:
[143x381]realize_view(cytoframe,filepath)

**Author(s)**

F. Hahne, B. Ellis, P. Haaland and N. Le Meur

**See Also**

`flowSet`, `read.FCS`

---

| cytoset | cytoset: a reference class for efficiently managing the data representation of a `flowSet` |

**Description**

This class is a container for a set of cytoframe objects, analogous to a `flowSet`. 

---
Details

Similar to the distinction between the cytoframe and flowFrame classes, the primary difference between the cytoset and flowSet classes is in the underlying representation of the data. Because cytoset is a reference class, copying or subsetting a cytoset object will return a cytoset pointing to the same underlying data. A deep copy of the data can be obtained via the `realize_view` method.

There is one notable exception to the typical behavior of most methods returning a cytoframe. The standard extraction operator ([[[]]]) will by default perform a deep copy of the subset being extracted and return a flowFrame. This is for the sake of compatibility with existing user scripts.

Creating Objects

Objects can be created using `cytoset()` and then adding samples by providing a cytoframe and sample name to `cs_add_sample`:

```r
cs <- cytoset()
cs_add_sample(cs, "Sample Name", cytoframe)
```

The safest and easiest way to create cytosets directly from FCS files is via the `load_cytoset_from_fcs` function, and there are alternative ways to specify the files to read. See the separate documentation for details.

Methods

[, [[ Subsetting. x[i] where i is a scalar, returns a cytoset object, and x[[i]] a flowFrame object. In this respect the semantics are similar to the behavior of the subsetting operators for lists. x[i,j] returns a cytoset for which the parameters of each cytoframe have been subset according to j. x[[i,j]] returns the subset of a single flowFrame for all parameters in j.

The reason for the default behavior for the extraction operator [[]] of returning a flowFrame rather than cytoframe is for backwards compatibility with existing user scripts. This behavior can be overridden to instead return a cytoframe with the additional `returnType` argument.

Usage:
```r
cytoset[i]
cytoset[i,j]
cytoset[[i]]
cytoset[[i,returnType = "cytoframe"]]
```

\$ Subsetting by frame name. This will return a single cytoframe object. Note that names may have to be quoted if they are not valid R symbols (e.g. cytoset$"sample 1")

`colnames`, `colnames<-` - Extract or replace the character object with the (common) column names of all the data matrices in the cytoframes.

Usage:
```r
colnames(cytoset)
colnames(cytoset) <- value
```

`identifier`, `identifier<-` - Extract or replace the name item from the environment.

Usage:
```r
identifier(cytoset)
identifier(cytoset) <- value
```
**phenoData**, **phenoData**<-  Extract or replace the AnnotatedDataFrame containing the phenotypic data for the whole data set. Each row corresponds to one of the cytoframes. The sampleNames of phenoData (see below) must match the names of the cytoframes in the frames environment.

*Usage:*

```r
phenoData(cytoset)
phenoData(cytoset) <- value
```

**pData**, **pData**<-  Extract or replace the data frame (or columns thereof) containing actual phenotypic information from the phenoData of the underlying data.

*Usage:*

```r
pData(cytoset)
pData(cytoset)$someColumn <- value
```

**varLabels**, **varLabels**<-  Not yet implemented.

Extract and set varLabels in the AnnotatedDataFrame of the phenoData of the underlying data.

*Usage:*

```r
varLabels(cytoset)
varLabels(cytoset) <- value
```

**sampleNames**  Extract and replace sample names from the phenoData. Sample names correspond to frame identifiers, and replacing them will also replace the GUID for each cytoframe. Note that sampleName needs to be unique.

*Usage:*

```r
sampleNames(cytoset)
sampleNames(cytoset) <- value
```

**keyword**  Extract or replace keywords specified in a character vector or a list from the description slot of each frame. See **keyword** for details.

*Usage:*

```r
keyword(cytoset,list(keywords))
keyword(cytoset,keywords)
keyword(cytoset) <- list(foo="bar")
```

**length**  number of cytoframe objects in the set.

*Usage:*

```r
length(cytoset)
```

**show**  display object summary.

**summary**  Return descriptive statistical summary (min, max, mean and quantile) for each channel of each cytoframe

*Usage:*

```r
summary(cytoset)
```

**fsApply**  Apply a function on all frames in a cytoset object. Similar to `sapply`, but with additional parameters. See separate documentation for details.

*Usage:*

```r
fsApply(cytoset,function,...)
fsApply(cytoset,function,use.exprs=TRUE,...)
```

**compensate**  Apply a compensation matrix on all frames in a cytoset object. See separate documentation for details.

*Usage:*

```r
compensate(cytoset,matrix)
```
transform  Apply a transformation function on all frames of a cytoset object. See separate documentation for details.

Usage:
transform(cytoset,...)

filter  Apply a filter object on a cytoset object. There are methods for filters, filterSets and lists of filters. The latter has to be a named list, where names of the list items are matching sampleNames of the cytoset. See filter for details.

Usage:
filter(cytoset,filter)
filter(cytoset,list(filters))

split  Split all cytoset objects according to a filter, filterResult or a list of such objects, where the length of the list has to be the same as the length of the cytoset. This returns a list of cytoframes or an object of class cytoset if the flowSet argument is set to TRUE. Alternatively, a cytoset can be split into separate subsets according to a factor (or any vector that can be coerced into factors), similar to the behaviour of split for lists. This will return a list of cytosets. See split for details.

Usage:
split(cytoset,filter)
split(cytoset,filterResult)
split(cytoset,list(filters))
split(cytoset,factor)

Subset  Returns a cytoset of cytoframes that have been subset according to a filter or filterResult, or according to a list of such items of equal length as the cytoset.

Usage:
Subset(cytoset,filter)
Subset(cytoset,filterResult)
Subset(cytoset,list(filters))

rbind2  Not yet implemented.
Combine two cytoset objects, or one cytoset and one cytoframe object.

Usage:
rbind2(cytoset,cytoset)
rbind2(cytoset,cytoframe)

spillover  Compute spillover matrix from a compensation set. See separate documentation for details.

realize_view  Returns a new cytoset with its own copy of the underlying data (a deep copy). The optional filepath argument accepts a string to specify a full directory name for storing the new copies of the data from the FCS files in h5 format.

Usage:
realize_view(cytoset,filepath)

cs_add_sample  Adds a cytoframe to the cytoset with sample name given by a string.

Usage:
cs_add_sample(cytoset,"SampleName",cytoframe)
**Description**

These methods consist of a pair of methods to coerce a cytoframe to or from a flowFrame and another pair to coerce a cytoset to or from a flowSet.

**Usage**

```r
cytoframe_to_flowFrame(fr)
flowFrame_to_cytoframe(fr, ...)
cytoset_to_flowSet(cs)
flowSet_to_cytoset(fs, path = tempfile())
```

**Details**

The conversion between the two sets of data container classes mostly entails a conversion of the back-end representation of the data. cytoframe and cytoset objects contain flowFrame and flowSet objects respectively, so coercion of a cytoframe to flowFrame entails moving the data from the 'C'-level data structure to the corresponding exprs, description, and parameters slots. Coercion of a flowFrame to a cytoframe entails creation of the 'C'-level data structure from the flowFrame slots. The names of each of the methods are pretty self-explanatory.

**Methods**

```r
cytoframe_to_flowFrame(object = "cytoframe") Returns a flowFrame object coerced from a cytoframe object.
flowFrame_to_cytoframe(object = "flowFrame") Returns a cytoframe object coerced from a flowFrame object.
cytoset_to_flowSet(object = "cytoset") Returns a flowSet object coerced from a cytoset object.
flowSet_to_cytoset(object = "flowSet") Returns a cytoset object coerced from a flowSet object.
```

**Examples**

```r
library(flowCore)
data("GvHD")
fs <- GvHD[1]
cs <- flowSet_to_cytoset(fs)
cf <- cs[[1, returnType="cytoframe"]]
ff <- cytoframe_to_flowFrame(cf)
```
estimateLogicle  

*Compute logicle transformation from the flowData associated with a GatingHierarchy*

**Description**

See details in `estimateLogicle`

**Usage**

```r
estimateLogicle(x, channels, ...)
```

**Arguments**

- `x`: a GatingHierarchy
- `channels`: channels or markers for which the logicle transformation is to be estimated.
- `...`: other arguments

**Value**

transformerList object

**Examples**

```r
## Not run:
# gs is a GatingSet
trans.list <- estimateLogicle(gs[[1]], c("CD3", "CD4", "CD8"))
# trans.list is a transformerList that can be directly applied to GatinigSet
gs <- transform(gs, trans.list)
## End(Not run)
```

extract_cluster_pop_name_from_node

*Extract the population name from the node path It strips the parent path and cluster method name.*

**Description**

Extract the population name from the node path It strips the parent path and cluster method name.

**Usage**

```r
extract_cluster_pop_name_from_node(node, cluster_method_name)
```

**Arguments**

- `node`: population node path
- `cluster_method_name`: the name of the clustering method
Examples

```r
extract_cluster_pop_name_from_node("cd3/flowClust_pop1", "flowClust")
#returns "pop1"
```

---

`filter_to_list`  
*convert flowCore filter to a list It convert the flowCore gate to a list whose structure can be understood by underlying c++ data structure.*

### Description

*convert flowCore filter to a list*

It convert the flowCore gate to a list whose structure can be understood by underlying c++ data structure.

### Usage

```r
filter_to_list(x)
```

### Arguments

- `x`  
  filter a flowCore gate. Currently supported gates are: "rectangleGate", "polygonGate", "ellipsoidGate" and "booleanFilter"

### Value

* a list

---

`fix_channel_slash`  
*toggle the channel names between '/' and '_' character*

### Description

FlowJoX tends to replace '/' in the original channel names with '_' in gates and transformations. We need to do the same to the flow data but also need to change it back during the process since the channel names of the flowSet can’t be modified until the data is fully compensated.

### Usage

```r
fix_channel_slash(chnls, slash_loc = NULL)
```

### Arguments

- `chnls`  
  the channel names
- `slash_loc`  
  a list that records the locations of the original slash character within each channel name so that when restoring slash it won’t tamper the the original '_' character.

### Value

* the toggled channel names*
flowjo_biexp

*construct the flowJo-type biexponential transformation function*

---

**Description**

Normally it was parsed from flowJo xml workspace. This function provides the alternate way to construct the flowJo version of logicle transformation function within R.

**Usage**

```r
define <- flowjo_biexp(
  channelRange = 4096,
  maxValue = 262144,
  pos = 4.5,
  neg = 0,
  widthBasis = -10,
  inverse = FALSE
)
```

**Arguments**

- `channelRange`: numeric the maximum value of transformed data
- `maxValue`: numeric the maximum value of input data
- `pos`: numeric the full width of the transformed display in asymptotic decades
- `neg`: numeric Additional negative range to be included in the display in asymptotic decades
- `widthBasis`: numeric unkown.
- `inverse`: logical whether to return the inverse transformation function.

**Examples**

```r
trans <- flowjo_biexp()
data.raw <- c(-1, 1e3, 1e5)
data.trans <- trans(data.raw)
round(data.trans)
inv <- flowjo_biexp(inverse = TRUE)
round(inv(data.trans))
```

---

**flowjo_biexp_trans**

*flowJo biexponential transformation.*

---

**Description**

Used for constructing biexponential transformation object.

**Usage**

```r
flowjo_biexp_trans(..., n = 6, equal.space = FALSE)
flowJo_biexp_trans(...)```

---
flowjo_fasinh

Arguments

... parameters passed to flowJoTrans
n desired number of breaks (the actual number will be different depending on the data range)
equal.space whether breaks at equal-spaced intervals

Value

biexponential transformation object

Examples

library(flowCore)
data(GvHD)
fr <- GvHD[[1]]
data.raw <- exprs(fr)[, "FL1-H"]
trans.obj <- flowjo_biexp_trans(equal.space = TRUE)
brks.func <- trans.obj["breaks"]
brks <- brks.func(data.raw)
brks # biexp space displayed at raw data scale

#transform it to verify it is equal-spaced at transformed scale
trans.func <- trans.obj["transform"]
print(trans.func(brks))

flowjo_fasinh inverse hyperbolic sine transform function

Description

hyperbolic sine/inverse hyperbolic sine (flowJo-version) transform function constructor

Usage

flowjo_fasinh(m = 4, t = 12000, a = 0.7, length = 256)
flowjo_fsinh(m = 4, t = 12000, a = 0.7, length = 256)

Arguments

m numeric the full width of the transformed display in asymptotic decades
t numeric the maximum value of input data
a numeric Additional negative range to be included in the display in asymptotic decades
length numeric the maximum value of transformed data

Value

fasinh/fsinh transform function
Examples

trans <- flowjo_fasinh()
data.raw <- c(1, 1e2, 1e3)
data.trans <- trans(data.raw)
data.trans

inverse.trans <- flowjo_fsinh()
inverse.trans(data.trans)

flowjo_fasinh_trans  flowJo inverse hyperbolic sine transformation.

Description

Used to construct the inverse hyperbolic sine transform object.

Usage

flowjo_fasinh_trans(..., n = 6, equal.space = FALSE)
flowJo_fasinh_trans(...)

Arguments

... parameters passed to flowjo_fasinh
n desired number of breaks (the actual number will be different depending on the
data range)

Value

fasinh transformation object

Examples

trans.obj <- flowjo_fasinh_trans(equal.space = TRUE)
data <- 1:1e3
brks.func <- trans.obj["breaks"]
brks <- brks.func(data)
brks # fasinh space displayed at raw data scale

#transform it to verify it is equal-spaced at transformed scale
trans.func <- trans.obj["transform"]
round(trans.func(brks))
flowjo_log_trans

**flowjo_log_trans**  
*flog transform function*

**Description**

flog transform function constructor. It is different from flowCore version of logtGml2 in the way that it reset negative input so that no NAN will be returned.

**Usage**

```r
flowjo_log_trans(
  decade = 4.5,
  offset = 1,
  scale = 1,
  n = 6,
  equal.space = FALSE
)
```

**Arguments**

- **decade**: total number of decades (i.e. log(max)-log(min))
- **offset**: offset to the original input (i.e. min value)
- **scale**: the linear scale factor
- **inverse**: whether return the inverse function

**Value**

flog(or its inverse) transform function

**Examples**

```r
trans <- flowjo_log_trans()
data.raw <- c(1,1e2,1e3)
data.trans <- trans[["transform"]](data.raw)
data.trans

inverse.trans <- trans[["inverse"]]
inverse.trans(data.trans)

# negative input
data.raw <- c(-10,1e2,1e3)
data.trans <- trans[["transform"]](data.raw)
data.trans
inverse.trans(data.trans)# we lose the original value at lower end since flog can't restore negative value

# different
trans <- flowjo_log_trans(decade = 3, offset = 30)
data.trans <- trans[["transform"]](data.raw)
data.trans
inverse.trans <- trans[["inverse"]]
inverse.trans(data.trans)
```
flowWorkspace-deprecated

 Deprecated functions in package flowWorkspace.

Description

- `getStats` -> `gs(/gh)_pop_get_stats`
- `getProp` -> `gh_pop_get_proportion`
- `getTotal` -> `gh_pop_get_count`
- `getPopStats` -> `gs(/gh)_pop_get_stats`
- `getNodes` -> `gs_get_pop_paths`
- `getParent` -> `gs_pop_get_parent`
- `getChildren` -> `gs_pop_get_children`
- `getGate` -> `gs(/gh)_get_gate`
- `getIndices` -> `gh_pop_get_indices`
- `isGated` -> `gh_pop_is_gated`
- `isNegated` -> `gh_pop_is_negated`
- `isHidden` -> `gh_pop_is_hidden`
- `getData` -> `gs(/gh)_get_data`
- `getTransformations` -> `gh_get_transformations`
- `getCompensationMatrices` -> `gh_get_compensations`
- `plotGate` -> `autoplot`
- `setNode` -> `gs(/gh)_set_node_name / gs(/gh)_set_node_visible`
- `isNcdf` -> `gs_is_h5`
- `clone` -> `gs_clone`
- `recompute` -> `gs_recompute`
- `flowData` -> `gs_cyto_data`
- `flowData<` -> `gs_cyto_data<`
- `getLoglevel` -> `get_log_level`
- `setLoglevel` -> `set_log_level`
- `rbind2` -> `gslist_to_gs`
- `filterObject` -> `filter_to_list`
- `add` -> `gs_pop_add`
- `Rm` -> `gs_pop_remove`
- `copyNode` -> `gh_copy_gate`
- `openWorkspace` -> `open_flowjo_xml`
- `flowJo.flog` -> `flowjo_log_trans`
- `flowJoTrans` -> `flowjo_biexp`
- `flowJo_biexp_trans` -> `flowjo_biexp_trans`
- `flowJo.fasinh` -> `flowjo_fasinh`
flowWorkspace.par.init

workspace version is parsed from xml node '/Workspace/version' in flowJo workspace and matched with this list to dispatch to the one of the three workspace parsers

Description
workspace version is parsed from xml node '/Workspace/version' in flowJo workspace and matched with this list to dispatch to the one of the three workspace parsers

Usage
flowWorkspace.par.init()

flowWorkspace.par.set

flowWorkspace.par.set sets a set of parameters in the flowWorkspace package namespace.

Description
flowWorkspace.par.set gets a set of parameters in the flowWorkspace package namespace.

Usage
flowWorkspace.par.set(name, value)

flowWorkspace.par.get(name = NULL)
flow_breaks

Arguments

name The name of a parameter category to get or set.
value A named list of values to set for category name or a list of such lists if name is missing.

Details

It is currently used to add/remove the support for a specific flowJo versions (parsed from xml node '/Workspace/version' in flowJo workspace)

Examples

#get the flowJo versions currently supported
old <- flowWorkspace.par.get("flowJo_versions")

#ad the new version
old[["win"]]<-c(old[["win"]], "1.7")
flowWorkspace.par.set("flowJo_versions", old)
flowWorkspace.par.get("flowJo_versions")

flow_breaks Generate the breaks that makes sense for flow data visualization

Description

It is mainly used as helper function to construct breaks function used by 'trans_new'.

Usage

flow_breaks(x, n = 6, equal.space = FALSE, trans.fun, inverse.fun)

Arguments

x the raw data values
n desired number of breaks (the actual number will be different depending on the data range)
equal.space whether breaks at equal-spaced intervals
trans.fun the transform function (only needed when equal.space is TRUE)
inverse.fun the inverse function (only needed when equal.space is TRUE)

Value

either 10^n intervals or equal-spaced(after transformed) intervals in raw scale.
Example

```r
library(flowCore)
data(GvHD)
fr <- GvHD[[1]]
data.raw <- exprs(fr)[, "FL1-H"]
flow_breaks(data.raw)

trans <- logicleTransform()
inv <- inverseLogicleTransform(trans = trans)
myBrks <- flow_breaks(data.raw, equal.space = TRUE, trans = trans, inv = inv)
round(myBrks)
#to verify it is equally spaced at transformed scale
print(trans(myBrks))
```

**Description**

helper function to generate a trans objects Used by other specific trans constructor

**Usage**

```r
flow_trans(name, trans.fun, inverse.fun, equal.space = FALSE, n = 6)
```

**Arguments**

- `name`: transformation name
- `trans.fun`: the transform function (only needed when equal.space is TRUE)
- `inverse.fun`: the inverse function (only needed when equal.space is TRUE)
- `equal.space`: whether breaks at equal-spaced intervals
- `n`: desired number of breaks (the actual number will be different depending on the data range)

**GatingHierarchy-class**

**Description**

GatingHierarchy is a class for representing the gating hierarchy, which can be either imported from a flowJo workspace or constructed in R.

**Details**

There is a one-to-one correspondence between GatingHierarchy objects and FCS files in the flowJo workspace. Each sample (FCS file) is associated with its own GatingHierarchy. It is also more space efficient by storing gating results as logical/bit vector instead of copying the raw data.

Given a GatingHierarchy, one can extract the data associated with any subpopulation, extract gates, plot gates, and extract population proportions. This facilitates the comparison of manual gating methods with automated gating algorithms.
## GatingSet-class

### Description

GatingSet holds a set of GatingHierarchy objects, representing a set of samples and the gating scheme associated with each.

### Details

Objects stores a collection of GatingHierarchies and represent a group in a flowJo workspace. A GatingSet can have two “states”. After a call to `flowjo_to_gatingset(...,execute=FALSE)` , the workspace is imported but the data is not. Setting `execute` to `TRUE` is needed in order to load, transform, compensate, and gate the associated data. Whether or not a GatingHierarchy has been applied to data is encoded in the `flag` slot. Some methods will warn the user, or may not function correctly if the GatingHierarchy has not been executed. This mechanism is in place, largely for the purpose of speed when working with larger workspaces. It allows the use to load a workspace and subset desired samples before proceeding to load the data.

### Slots

- **pointer**: Object of class "externalptr". points to the gating hierarchy stored in C data structure.
- **transformation**: Object of class "list". a list of transformation objects used by GatingSet.

### See Also

- `GatingHierarchy`

### Examples

```r
## Not run:
require(flowWorkspaceData)
d<-system.file("extdata",package="flowWorkspaceData")
wsfile<-list.files(d,pattern="A2004Analysis.xml",full=TRUE)
library(CytoML)
ws <- open_flowjo_xml(wsfile);
G<-try(flowjo_to_gatingset(ws,path=d,name=1));
gh <- G[[1]]
gh_pop_compare_stats(gh);
gh_plot_pop_count_cv(gh)
nodes <- gs_get_pop_paths(gh)
thisNode <- nodes[4]
require(ggcyto)
autoplot(gh,thisNode);
gh_pop_get_gate(gh,thisNode);
gh_pop_get_data(gh,thisNode)
## End(Not run)
```
Examples

```r
## Not run:
require(flowWorkspaceData)
d <- system.file("extdata", package="flowWorkspaceData")
wsfile <- list.files(d, pattern="A2004Analysis.xml", full=TRUE)
library(CytoML)
ws <- open_flowjo_xml(wsfile);
G <- try(flowjo_to_gatingset(ws, execute=TRUE, path=d, name=1));
gs_plot_pop_count_cv(G);
## End(Not run)
```

GatingSet-methods
constructors for GatingSet

Description
construct a gatingset with empty trees (just root node)

Usage
GatingSet(x)

Arguments

- **x**
  - a flowSet, ncdfFlowSet, or cytoset

Examples

```r
## Not run:
# fdata could be a flowSet, ncdfFlowSet, or GatingSet
gs <- GatingSet(fdata)
## End(Not run)
```

GatingSetList-class
Class "GatingSetList"

Description
A list of of GatingSet objects. This class exists for method dispatching.

Usage
GatingSetList(x, samples = NULL)

Arguments

- **x**
  - a list of GatingSet
- **samples**
  - character vector specifying the order of samples. if not specified, the samples are ordered as the underlying stored order.
GatingSetList-class

Details

Objects store a collection of GatingSets, which usually has the same gating trees and markers. Most GatingSets methods can be applied to GatingSetList.

See Also

GatingSet GatingHierarchy

Examples

```r
# Not run:
#load several GatingSets from disk
gs_list <- lapply(list.files("../gs_toMerge", full=T) , function(this_folder){
  load_gs(this_folder)
})

gs_list is a list
gs_groups <- merge(gs_list)
#returns a list of GatingSetList objects
gslist2 <- gs_groups[[2]]
#gslist2 is a GatingSetList that contains multiple GatingSets and they share the same gating and data structure

gslist2 class(gslist2)
sampleNames(gslist2)

#reference a GatingSet by numeric index
gslist2[[1]]
#reference a GatingSet by character index
gslist2["30104.fcs"]

#loop through all GatingSets within GatingSetList
lapply(gslist2, sampleNames)

#subset a GatingSetList by [
sampleNames(gslist2[c(4,1)])
sampleNames(gslist2[c(1,4)])
gslist2[c("30104.fcs")]

#get flow data from it
gs_pop_get_data(gslist2)
#get gated flow data from a particular population
gs_pop_get_data(gslist2, "3+")

#extract the gates associated with one population
gs_pop_get_gate(gslist2, "3+")
gs_pop_get_gate(gslist2, 5)

#extract the pheno data
pData(gslist2[3:1])
#modify the pheno data
pd <- pData(gslist2)
pd$id <- 1:nrow(pd)
pData(gslist2) <- pd
pData(gslist2[3:2])

#plot the gate
```
get_log_level

autoplot(gslist2[1:2],5)

# remove certain gates by loop through GatingSets
gs_get_pop_paths(gslist2[[1]])
lapply(gslist2, function(gs) gs_pop_remove("Excl", gs = gs))

# extract the stats
gs_pop_get_count_fast(gslist2)
# extract statistics by using getQStats defined in QUALIFIER package
res <- getQStats(gslist2[c(4,2)], isMFI = F, isSpike = F, nslaves = 1)

# archive the GatingSetList
save_gslist(gslist2, path = "/rglab/workspace/flowIncubator/output/gslist", overwrite = T)
gslist2 <- load_gslist(path = "/rglab/workspace/flowIncubator/output/gslist")

# convert GatingSetList into one GatingSet by merge_list_to_gs
gs_merged2 <- merge_list_to_gs(gslist2)
gs_merged2

## End(Not run)
## Not run:
samleNames(gsA) # return A1, A2
samleNames(gsB) # return B1, B2
gs.list <- list(gsA, gsB)
gslist <- GatingSetList(gs.list)
samleNames(gslist) # return A1, A2, B1, B2

# set different order when create the GatingSetList
gslist <- GatingSetList(gs.list, samples = c("A1", "B1", "A2", "B2"))
samleNames(gslist) # return A1, B1, A2, B2

## End(Not run)

get_log_level
get/set the log level

Description
It is helpful sometime to get more detailed print out for the purpose of trouble shooting

Usage
get_log_level()
set_log_level(level = "none")

Arguments
level a character that represents the log level, can be value of c("none", "GatingSet", "GatingHierarchy", "Population", "gate") default is "none", which does not print any information from C parser.
Value

a character that represents the internal log level

Examples

get_log_level()
set_log_level(“Population”)
get_log_level()

gh_apply_to_new_fcs  

**constructors for GatingSet**

Description

construct object from existing gating hierarchy(gating template) and flow data

Usage

gh_apply_to_new_fcs(x, files, swap_cols = FALSE, ...)

Arguments

- `x`: GatingHierarchy
- `files`: fcs file paths
- `swap_cols`: for internal usage
- `...`: other arguments.
- `y`: sample names
- `path`: character specifies the path to the flow data (FCS files)

gh_copy_gate  

**Copy a node along with all of its descendant nodes to the given ancestor**

Description

Copy a node along with all of its descendant nodes to the given ancestor

Usage

gh_copy_gate(gh, node, to)

Arguments

- `gh`: GatingHierarchy
- `node`: the node to be copied
- `to`: the new parent node under which the node will be copied
**Examples**

```r
library(flowWorkspace)
dataDir <- system.file("extdata", package="flowWorkspaceData")
suppressMessages(gs <- load_gs(list.files(dataDir, pattern = "gs_manual", full = TRUE)))
gh <- gs[[1]]
old.parent <- gs_pop_get_parent(gh, "CD4")
new.parent <- "singlets"
gh_copy_gate(gh, "CD4", new.parent)
gs_get_pop_paths(gh)
```

---

**gh_get_cluster_labels**  
Retrieve the cluster labels from the cluster nodes

**Description**

Clustering results are stored as individual gated nodes. This helper function collect all the gating indices from the same clustering run (identified by 'parent' node and 'cluster_method_name" and merge them as a single factor.

**Usage**

```r
gh_get_cluster_labels(gh, parent, cluster_method_name)
```

**Arguments**

- `gh`  
  GatingHierarchy
- `parent`  
  the parent population/node name or path
- `cluster_method_name`  
  the name of the clustering method

---

**gh_get_compensations**  
Retrieve the compensation matrices from a GatingHierarchy

**Description**

Retrieve the compensation matrices from a GatingHierarchy.

**Usage**

```r
gh_get_compensations(x)
```

**Arguments**

- `x`  
  A GatingHierarchy object.

**Details**

Return all the compensation matrices in a GatingHierarchy.
gh_get_transformations

Value
A list of matrix representing the spillover matrix in GatingHierarchy

Examples
### Not run:
#Assume gh is a GatingHierarchy
gh_get_compensations(gh);

### End(Not run)

gh_get_transformations

Return a list of transformations or a transformation in a GatingHierarchy

Description
Return a list of all the transformations or a transformation in a GatingHierarchy

Usage
gh_get_transformations(
  x,
  channel = NULL,
  inverse = FALSE,
  only.function = TRUE,
  ...
)

Arguments
x A GatingHierarchy object
channel character channel name
inverse logical whether to return the inverse transformation function. Valid when
only.function is TRUE
only.function logical whether to return the function or the entire transformer object(see
scales package) that contains transform and inverse and breaks function.
...
other arguments equal.spaced logical passed to the breaks functio to determine
whether to break at 10^n or equally spaced intervals

Details
Returns a list of the transformations or a transformation in the flowJo workspace. The list is of
length L, where L is the number of distinct transformations applied to samples in the flowjo_workspace.
Each element of L is itself a list of length M, where M is the number of parameters that were trans-
formed for a sample or group of samples in a flowjo_workspace. For example, if a sample has
10 parameters, and 5 are transformed during analysis, using two different sets of transformations,
then L will be of length 2, and each element of L will be of length 5. The elements of L repre-
sent channel- or parameter-specific transformation functions that map from raw intensity values to
channel-space used by flowJo.
Value

Lists of functions (or transform objects when only.function is FALSE), with each element of the list representing a transformation applied to a specific channel/parameter of a sample.

Examples

```r
## Not run:
# Assume gh is a GatingHierarchy
ggplot(gh); # return a list transformation functions
ggplot(gh, inverse = TRUE); # return a list inverse transformation functions
ggplot(gh, channel = "FL1-H"); # only return the transformation associated with given channel
ggplot(gh, channel = "FL1-H", only.function = FALSE); # return the entire transform object
## End(Not run)
```

---

**gh_plot_pop_count_cv**  
Plot the coefficient of variation between xml and openCyto population statistics for each population in a gating hierarchy.

Description

This function plots the coefficient of variation calculated between the xml population statistics and the openCyto population statistics for each population in a gating hierarchy extracted from a xml Workspace.

Usage

```r
gh_plot_pop_count_cv(x, path = "auto", ...)
gs_plot_pop_count_cv(x, scales = list(x = list(rot = 90)), path = "auto", ...)
```

Arguments

- `x` A GatingHierarchy from or a GatingSet.
- `path` character see `gs_get_pop_paths`
- `...` Additional arguments to the barplot methods.
- `scales` list see `barchart`

Details

The CVs are plotted as barplots across panels on a grid of size m by n.

Value

Nothing is returned.

See Also

`gs_pop_get_count_fast`
Examples

```r
## Not run:
#G is a GatingHierarchy
gs_plot_pop_count_cv(G,4,4);

## End(Not run)
```

### gh_pop_compare_stats

**Description**

Compare the stats(count/freq) between the version parsed from xml and the one recalculated/gated from R

**Usage**

```r
gh_pop_compare_stats(x, path = "auto", ...)
```

**Arguments**

- `x`: GatingHierarchy
- `path`: see `gs_get_pop_paths`
- `...`: not used

### gh_pop_get_cluster_name

**Description**

check if a node is clustering node

**Usage**

```r
gh_pop_get_cluster_name(gh, node)
```

**Arguments**

- `gh`: GatingHierarchy
- `node`: the population/node name or path

**Value**

the name of the clustering method. If it is not cluster node, returns NULL.
**gh_pop_get_data**  
get gated flow data from a GatingHierarchy/GatingSet/GatingSetList

**Description**
get gated flow data from a GatingHierarchy/GatingSet/GatingSetList

**Usage**
gh_pop_get_data(obj, y = "root", inverse.transform = FALSE, ...)

**Arguments**
- **obj** A GatingHierarchy, GatingSet or GatingSetList object.
- **y** character the node name or full/(partial) gating path. If not specified, will return the complete flowFrame/flowSet at the root node.
- **inverse.transform** logical flag indicating whether to inverse transform the data
- **...** arguments passed to ncdFlow::

**Details**
Returns a flowFrame/flowSet containing the events in the gate defined at node y. Subset membership can be obtained using gh_pop_get_indices. Population statistics can be obtained using getPop and gh_pop_compare_stats. When calling gh_pop_get_data on a GatingSet, the trees representing the GatingHierarchy for each sample in the GaingSet are presumed to have the same structure. To update the data, use gs_cyto_data method.

**Value**
A flowFrame object if obj is a GatingHierarchy. A flowSet or ncdfflowSet if a GatingSet. A ncdfflowList if a GatingSetList.

**See Also**
gs_cyto_data gh_pop_get_indices gh_pop_compare_stats

**Examples**
```r
## Not run:
#G is a GatingSet
gedata(G, 3) # get a flowSet constructed from the third node / population in the tree.
gedata(G, "cd4")

# gh is a GatingHierarchy
gh_pop_get_data(gh)
```

## End(Not run)
**gh_pop_get_full_path**

convert the partial gating path to the full path

**Description**
convert the partial gating path to the full path

**Usage**

```r
gh_pop_get_full_path(gh, path)
```

**Arguments**

- `gh`: GatingHierarchy object
- `path`: the partial gating path

**Value**

the full gating path
**gh_pop_get_indices**  

*Get the membership indices for each event with respect to a particular gate in a GatingHierarchy*

---

**Description**

Returns a logical vector that describes whether each event in a sample is included or excluded by this gate.

**Usage**

```
gh_pop_get_indices(obj, y)
```

**Arguments**

- `obj`: A `GatingHierarchy` representing a sample.
- `y`: A character giving the name or full/(partial) gating path of the population / node of interest.

**Details**

Returns a logical vector that describes whether each event in the data file is included in the given gate of this `GatingHierarchy`. The indices are for all events in the file, and do not reflect the population counts relative to the parent but relative to the root. To get population frequencies relative to the parent one cross-tabulate the indices of `y` with the indices of its parent.

**Value**

A logical vector of length equal to the number of events in the FCS file that determines whether each event is or is not included in the current gate.

**Note**

Generally you should not need to use `gh_pop_get_indices` but the more convenient methods `gh_pop_get_proportion` and `gh_pop_compare_stats` which return population frequencies relative to the parent node. The indices returned reference all events in the file and are not directly suitable for computing population statistics, unless subsets are taken with respect to the parent populations.

**See Also**

- `gh_pop_compare_stats`

**Examples**

```r
## Not run:
#G is a gating hierarchy
#Return the indices for population 5 (topological sort)
gh_pop_get_indices(G, gs_get_pop_paths(G, tsort=TRUE)[5]);

## End(Not run)
```
**gh_pop_get_indices_mat**

*Return the single-cell matrix of 1/0 dichotomized expression*

**Description**

Return the single-cell matrix of 1/0 dichotomized expression

**Usage**

```r
gh_pop_get_indices_mat(gh, y)
```

**Arguments**

- `gh`: GatingHierarchy object
- `y`: character string containing the boolean or of node names. e.g. `"cd4|cd8"`

---

**gh_pop_get_proportion**  *Get count or proportion from populations*

**Description**

Get count or proportion from populations

**Usage**

```r
gh_pop_get_proportion(x, y, xml = FALSE)
gh_pop_get_count(x, y, xml = FALSE)
```

**Arguments**

- `x`: GatingHierarchy
- `y`: character node name or path
- `xml`: whether to extract xml stats or openCyto stats
**Description**

move a node along with all of its descendant nodes to the given ancestor

**Usage**

```r
gh_pop_move(gh, node, to)
```

**Arguments**

- **gh** GatingHierarchy
- **node** the node to be moved
- **to** the new parent node under which the node will be moved to

**Examples**

```r
library(flowWorkspace)
dataDir <- system.file("extdata", package="flowWorkspaceData")
suppressMessages(gs <- load_gs(list.files(dataDir, pattern = "gs_manual", full = TRUE)))
gh <- gs[[1]]
old.parent <- gs_pop_get_parent(gh, "CD4")
new.parent <- "singlets"
gh_pop_move(gh, "CD4", new.parent)
gs_pop_get_parent(gh, "CD4")
```

---

**Description**

It is useful when we want to alter the population at events level yet without removing or adding the existing gates.

**Usage**

```r
gh_pop_set_indices(obj, y, z)
```

**Arguments**

- **obj** GatingHierarchy object
- **y** character node name or path
- **z** logical vector as local event indices relative to node y
Examples

library(flowWorkspace)
dataDir <- system.file("extdata", package="flowWorkspaceData")
suppressMessages(gs <- load_gs(list.files(dataDir, pattern = "gs_manual", full = TRUE)))
gh <- gs[[1]]
# get pop counts
pop.stats <- gh_pop_get_stats(gh, nodes = c("CD3+", "CD4", "CD8"))
pop.stats

# subsample 30% cell events at CD3+ node
total.cd3 <- pop.stats[pop == "CD3+", count]
gInd <- seq_len(total.cd3)  # create integer index for cd3
gInd <- sample.int(total.cd3, size = total.cd3 * 0.3)  # randomly select 30%
# convert it to logical index
logicalInd <- rep(FALSE, total.cd3)
logicalInd[gInd] <- TRUE
# replace the original index stored at GatingHierarchy
gh_pop_set_indices(gh, "CD3+", logicalInd)
# check the updated pop counts
gh_pop_get_stats(gs[[1]], nodes = c("CD3+", "CD4", "CD8"))  # note that CD4, CD8 are not updated
# update all the descendants of CD3+
nodes <- gh_pop_get_descendants(gh, "CD3+")
for (node in nodes) suppressMessages(recompute(gh, node))
gh_pop_get_stats(gs[[1]], nodes = c("CD3+", "CD4", "CD8"))  # now all are update to date

gh_pop_set_xml_count  
save the event counts parsed from xml into c++ tree structure

Description

It is for internal use by the diva parser

Usage

gh_pop_set_xml_count(gh, node, count)

Arguments

gh  
GatingHierarchy

node  
the unique gating path that uniquely identifies a population node

count  
integer number that is events count for the respective gating node directly parsed from xml file

Examples

## Not run:
gh_pop_set_xml_count(gh, "CD3", 10000)
## End(Not run)
**gslist_to_gs**

*Merge a GatingSetList into a single GatingSet*

**Description**

Merge a GatingSetList into a single GatingSet

**Usage**

```r
gslist_to_gs(x, ...)
```

**Arguments**

- `x`  
  GatingSetList

- `...`  
  other arguments passed to `gslist_to_gs` method for `ncdfFlowList`

**gs_check_redundant_nodes**

*try to determine the redundant terminal(or leaf) nodes that can be removed*

**Description**

These leaf nodes make the gating trees to be different from one another and can be removed by the subsequent convenient call `gs_remove_redundant_nodes`.

**Usage**

```r
gs_check_redundant_nodes(x, path = "auto", ...)
```

**Arguments**

- `x`  
  GatingSet or list of groups(each group is a list of 'GatingSet'). When it is a list, it is usually the outcome from `gs_split_by_tree`.

- `path`  
  argumented passed to `gs_get_pop_paths`. The default value is "auto".

- `...`  
  other arguments passed to `gs_get_pop_paths`.

**Value**

a list of the character vectors indicating the nodes that are considered to be redundant for each group of GatingSets.

**Examples**

```r
## Not run:
gslist <- list(gs1, gs2, gs3, gs4, gs5)
gs_groups <- gs_split_by_tree(gslist)
toRm <- gs_check_redundant_nodes(gs_groups)
## End(Not run)
```
gs_cyto_data

Fetch or replace the flowData object associated with a GatingSet.

Description
Accessor method that gets or replaces the flowset/ncdfFlowSet object in a GatingSet or GatingHierarchy.

Usage

\[
gs\_cyto\_data(x, \ldots)
\]

\[
gs\_cyto\_data(x) \leftarrow \text{value}
\]

Arguments

- `x`: A GatingSet
- `value`: The replacement flowSet or ncdfFlowSet object
- `inverse.transform`: logical flag indicating whether to inverse transform the data

Details
Accessor method that sets or replaces the ncdfFlowSet object in the GatingSet or GatingHierarchy.

Value
the object with the new flowSet in place.

gs_get_compensation_internal

extract compensation object from GatingSet

Description
extract compensation object from GatingSet

Usage

\[
gs\_get\_compensation\_internal(gs, \text{sampleName})
\]

Arguments

- `gs`: GatingSet
- `sampleName`: sample name
gs_get_leaf_nodes

gs_get_leaf_nodes (x, ...)  
gs_get_leaf_nodes (x, ...)

Description

gs_get_leaf_nodes get all the leaf nodes

Usage

gs_get_leaf_nodes (x, ...)

Arguments

x GatingHierarchy/GatingSet object

... arguments passed to 'gs_get_pop_paths' method

Value

the leaf nodes

gs_get_pop_paths

Get the names of all nodes from a gating hierarchy.

Description

gs_get_pop_paths returns a character vector of names of the nodes (populations) in the GatingSet.

Usage

gs_get_pop_paths (  
  x,  
  y = NULL,  
  order = "regular",  
  path = "full",  
  showHidden = FALSE,
  ...  
)

gh_get_pop_paths (  
  x,  
  y = NULL,  
  order = "regular",  
  path = "full",  
  showHidden = FALSE,
  ...  
)

Arguments

x  
A GatingSet. Assuming the gating hierarchy are identical within the GatingSet, the Gating tree of the first sample is used to query the node information.

y  
A character not used.

order  
order=c("regular","tsort","bfs") returns the nodes in regular, topological or breadth-first sort order. "regular" is default.

path  
A character or numeric scalar. When numeric, it specifies the fixed length of gating path (length 1 displays terminal name). When character, it can be either 'full' (full path, which is default) or 'auto' (display the shortest unique gating path from the bottom of gating tree).

showHidden  
logical whether to include the hidden nodes

...  
Additional arguments.

Details

integer indices of nodes are based on regular order, so whenever need to map from character node name to integer node ID, make sure to use default order which is regular.

Value

gs_get_pop_paths returns a character vector of node/population names, ordered appropriately.

Examples

## Not run:
#G is a gating hierarchy
gs_get_pop_paths(G, path = 1)#return node names (without prefix)
gs_get_pop_paths(G,path = "full")#return the full path
gs_get_pop_paths(G, path = 2)#return the path as length of two
gs_get_pop_paths(G, path = "auto")#automatically determine the length of path
gs_pop_set_name(G,"L","lymph")

## End(Not run)

gs_get_singlecell_expression

Return the cell events data that express in any of the single populations defined in y

Description

Returns a list of matrix containing the events that expressed in any one of the populations defined in y
Usage

gs_get_singlecell_expression(
  x,
  nodes,
  other.markers = NULL,
  swap = FALSE,
  threshold = TRUE,
  marginal = TRUE,
  mc.cores = getOption("mc.cores", 1L),
  ...
)

gs_get_singlecell_expression_by_gate(...)

Arguments

x A GatingSet or GatingSetList object.
nodes character vector specifying different cell populations
other.markers character vector specifying the extra markers/channels to be returned besides the ones derived from "nodes" and "map" argument. It is only valid when threshold is set to FALSE.
swap logical indicates whether channels and markers of flow data are swapped.
threshold logical indicates whether to threshold the flow data by setting intensity value to zero when it is below the gate threshold.
marginal logical indicates whether to the gate is treaded as 1d marginal gate. Default is TRUE, which means markers are determined either by node name or by 'map' argument explained below. When FALSE, the markers are determined by the gate dimensions. and node name and 'map' argument are ignored.
mc.cores passed to mclapply. Default is 1, which means the process runs in serial mode. When it is larger than 1, parallel mode is enabled.
...
other arguments map a named list providing the mapping between node names (as specified in the gating hierarchy of the gating set) and channel names (as specified in either the desc or name columns of the parameters of the associated flowFrames in the GatingSet), see examples.
ignore.case whether to ignore case when match the marker names. Default is FALSE.

Value

A list of numerci matrices

Author(s)

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See Also

gh_pop_get_indices gs_pop_get_count_fast
Examples

```r
## Not run:
#G is a GatingSet
nodes <- c("4+/TNFa+", "4+/IL2+")
res <- gs_get_singlecell_expression(gs, nodes)
res[[1]]

# if it fails to match the given nodes to the markers, then try to provide the mapping between node and marker explicitly
res <- gs_get_singlecell_expression(gs, nodes, map = list("4+/TNFa+" = "TNFa", "4+/IL2+" = "IL2"))

# It can also operate on the 2d gates by setting marginal to FALSE
# The markers are no longer deduced from node names or supplied by map
# Instead, it retrieves the markers that are associated with the gates
nodes <- c("4+/TNFa+IFNg+", "4+/IL2+IL3+")
res <- gs_get_singlecell_expression(gs, nodes, marginal = FALSE)
#or simply call convenient wrapper
gs_get_singlecell_expression_by_gate(gs, nodes)

## End(Not run)
```

gs_is_h5

_determine the flow data associated with a Gating Hierarchy is based on ‘ncdfFlowSet’ or ‘flowSet’_

Description
determine the flow data associated with a Gating Hierarchy is based on ‘ncdfFlowSet’ or ‘flowSet’

Usage
gs_is_h5(x)
isNcdf(x)

Arguments

x  GatingHierarchy object

Value

logical
**gs_plot_diff_tree**

visualize the tree structure difference among the GatingSets

**Description**

visualize the tree structure difference among the GatingSets

**Usage**

```r
gs_plot_diff_tree(x, path = "auto", ...)
```

**Arguments**

- `x`: list of groups(each group is a list of 'GatingSet'). it is usually the outcome from `gs_split_by_tree`.
- `path`: passed to `getNodes`
- `...`: passed to `getNodes`

**Examples**

```r
## Not run:
gslist <- list(gs1, gs2, gs3, gs4, gs5)
gs_groups <- gs_split_by_tree(gslist)
gs_plot_diff_tree(gs_groups)
## End(Not run)
```

---

**gs_pop_add**

Create a GatingSet and add/remove the flowCore gate(or population) to/from a GatingHierarchy/GatingSet.

**Description**

GatingSet method creates a gatingset from a flowSet with the ungated data as the root node. add method add the flowCore gate to a GatingHierarchy/GatingSet. gs_pop_set_gate method update the gate of one population node in GatingHierarchy/GatingSet. Rm method Remove the population node from a GatingHierarchy/GatingSet. They are equivalent to the workFlow.add and Rm methods in flowCore package. recompute method does the actual gating after the gate is added,i.e. calculating the event indices according to the gate definition.

**Usage**

```r
gs_pop_add(gs, gate, validityCheck = TRUE, ...)
gs_pop_remove(gs, node, ...)
```
Arguments

- **gs**: A GatingSet
- **gate**: A filter or a list of filters to be added to the GatingSet.
- **validityCheck**: logical whether to check the consistency of tree structure across samples. default is TRUE. Can be turned off when speed is preferred to the robustness.

... some other arguments to specify how the gates are added to the gating tree.
- **names**: a character vector of length four, which specifies the population names resulted by adding a quadGate. The order of the names is clock-wise starting from the top left quadrant population.
- **parent**: a character scalar to specify the parent node name where the new gate to be added to, by default it is NULL, which indicates the root node
- **name**: a character scalar to specify the node name of population that is generated by the gate to be added.
- **recompute**: a logical flag
- **negated**: a logical scalar to specify whether the gate is negated, which means the the population outside of the gate will be kept as the result population. It is FALSE by default.

- **node**: A character identifies the population node in a GatingHierarchy or GatingSet to remove

Value

GatingSet method returns a GatingSet object with just root node. add method returns a population node ID (or four population node IDs when adding a quadGate) that uniquely identify the population node within a GatingHierarchy.

See Also

- `GatingSet-class`

Examples

```r
## Not run:
library(flowCore)
data(GvHD)
# select raw flow data
fs<-GvHD[[1:3]]

# transform the raw data
tf <- transformList(colnames(fs[[1]])[3:6], asinh, transformationId="asinh")
fs_trans<-transform(fs,tf)

# add transformed data to a gatingset
gs <- GatingSet(fs_trans)
gs
gs_get_pop_paths(gs[[1]]) # only contains root node

# add one gate
rg <- rectangleGate("FSC-H"=c(200,400), "SSC-H"=c(250, 400),
      filterId="rectangle")

nodeID<-gs_pop_add(gs, rg)# it is added to root node by default if parent is not specified
```
nodeID

gs_get_pop_paths(gs[[1]]) #the second population is named after filterId of the gate

#add a quadGate
qg <- quadGate("FL1-H"=2, "FL2-H"=4)
nodeIDs<-gs_pop_add(gs,qg,parent="rectangle")
nodeIDs #quadGate produces four population nodes

gs_get_pop_paths(gs[[1]]) #population names are named after dimensions of gate if not specified

#add a boolean Gate
bg<-booleanFilter("CD15 FITC-CD45 PE+\"|\"CD15 FITC+CD45 PE-\")
bg
nodeID2<-gs_pop_add(gs,bg,parent="rectangle")
nodeID2
gs_get_pop_paths(gs[[1]])

#do the actual gating
recompute(gs)

#plot one gate for one sample
autoplot(gs[[1]],"rectangle")

autoplot(gs[[1]],nodeIDs) #may be smoothed automatically if there are not enough events after gating

#plot gates across samples using lattice plot
autoplot(gs,nodeID)

#plot all gates for one sample
autoplot(gs[[1]]) #boolean gate is skipped by default
autoplot(gs[[1]],bool=TRUE)

#plot the gating hierarchy
plot(gs[[1]])

#remove one node causing the removal of all the descendants

  gs_pop_remove('rectangle', gs = gs)
  gs_get_pop_paths(gs[[1]])

## End(Not run)

---

### gs_pop_get_count_fast

*Return a table of population statistics for all populations in a GatingHierarchy/GatingSet or the population proportions or the total number of events of a node (population) in a GatingHierarchy*

**Description**

gs_pop_get_count_fast is more useful than getPop. Returns a table of population statistics for all populations in a GatingHierarchy/GatingSet. Includes the xml counts, openCyto counts and frequencies.

**Usage**

```
gs_pop_get_count_fast(
  x,
  statistic = c("freq", "count"),
  xml = FALSE,
  subpopulations = NULL,
)```
gs_pop_get_count_fast

format = c("long", "wide"),
path = "full",
...
)

gs_pop_get_count_with_meta(x, ...)

Arguments

x
a GatingSet or GatingSetList

statistic
character specifies the type of population statistics to extract. (only valid when format is "wide"). Either "freq" or "count" is currently supported.

xml
logical indicating whether the statistics come from xml (if parsed from xml workspace) or from openCyto.

subpopulations
character vector to specify a subset of populations to return. (only valid when format is "long")

format
character value of c("wide", "long") specifying whether to organize the output in long or wide format

path
character see gs_get_pop_paths

... additional arguments passed to gs_pop_get_count_fast

Details

gs_pop_get_count_fast returns a table population statistics for all populations in the gating hierarchy. The output is useful for verifying that the import was successful, if the xml and openCyto derived counts don’t differ much (i.e. if they have a small coefficient of variation.) for a GatingSet, returns a matrix of proportions for all populations and all samples

Value

gs_pop_get_count_fast returns a data.frame with columns for the population name, xml derived counts, openCyto derived counts, and the population proportions (relative to their parent population). a data.table of merged population statistics with sample metadata.

See Also

gs_get_pop_paths

Examples

## Not run:
#gh is a GatingHierarchy
gs_pop_get_count_fast(gh);
gh_pop_get_stats(gh, gs_get_pop_paths(gh, tsort=T)[5])

#gs is a GatingSet
gs_pop_get_count_fast(gs)
#optionally output in long format as a data.table
gs_pop_get_count_fast(gs, format = "long", path = "auto")
#only get stats for a subset of populations
gs_pop_get_count_fast(gs, format = "long", subpopulations = gs_get_pop_paths(gs)[4:6])
gs_pop_get_gate

Return the flowCore gate definition associated with a node in a GatingHierarchy/GatingSet.

Description

Return the flowCore gate definition object associated with a node in a GatingHierarchy or GatingSet object.

Usage

gh_pop_get_gate(obj, y)

gs_pop_get_gate(obj, y)

Arguments

obj A GatingHierarchy or GatingSet

y A character the name or full/(partial) gating path of the node of interest.

Value

A gate object from flowCore. Usually a polygonGate, but may be a rectangleGate. Boolean gates are represented by a "BooleanGate" S3 class. This is a list boolean gate definition that references populations in the GatingHierarchy and how they are to be combined logically. If obj is a GatingSet, assuming the trees associated with each GatingHierarchy are identical, then this method will return a list of gates, one for each sample in the GatingSet corresponding to the same population indexed by y.

See Also

gh_pop_get_data gs_get_pop_paths

Examples

## Not run: #gh is a GatingHierarchy
gh_pop_get_gate(gh, "CD3") #return the gate for the fifth node in the tree, but fetch it by name.

#G is a GatingSet
gs_pop_get_gate(G, "CD3") #return a list of gates for the fifth node in each tree

## End(Not run)
gs_pop_get_parent

Return the name of the parent population or a list of child populations of the current population in the GatingHierarchy

Description

Returns the name of the parent population or a character/numeric vector of all the children of the current population in the given GatingHierarchy.

Usage

gs_pop_get_parent(obj, y, ...)
gh_pop_get_parent(obj, y, ...)
gs_pop_get_children(obj, y, showHidden = TRUE, ...)
gh_pop_get_children(obj, y, showHidden = TRUE, ...)

Arguments

obj
A GatingHierarchy
y
a character/numeric the name or full/(partial) gating path or node indices of the node / population.
...
other arguments passed to gs_get_pop_paths methods
showHidden
logical whether to include the hidden children nodes.

Value

gs_pop_get_parent returns a character vector, the name of the parent population. gs_pop_get_children returns a character or numeric vector of the node names or node indices of the child nodes of the current node. An empty vector if the node has no children.

See Also

gs_get_pop_paths

Examples

## Not run:
#G is a gatinghierarchy
#return the name of the parent of the fifth node in the hierarchy.
gs_pop_get_parent(G,gs_get_pop_paths(G[[1]][5]))
n<-gs_get_pop_paths(G,tsort=T)[4];
gs_pop_get_children(G,n);#Get the names of the child nodes of the 4th node in this gating hierarchy.
gs_pop_get_children(G,4);#Get the ids of the child nodes

## End(Not run)
gs_pop_get_stats

Extract stats from populations (or nodes)

Description

Extract stats from populations (or nodes)

Usage

gs_pop_get_stats(x, ...)

gh_pop_get_stats(
  x,
  nodes = NULL,
  type = "count",
  xml = FALSE,
  inverse.transform = FALSE,
  stats.fun.arg = list(),
  ... )

Arguments

x          a GatingSet or GatingHierarchy

...        arguments passed to gs_get_pop_paths method.

nodes      the character vector specifies the populations of interest. default is all available nodes

type       the character vector specifies the type of pop stats or a function used to compute population stats. when character, it is expected to be either "count" or "percent". Default is "count" (total number of events in the populations). when a function, it takes a flowFrame object through 'fr' argument and return the stats as a named vector.

xml        whether to extract xml stats or openCyto stats

inverse.transform logical flag. Whether inverse transform the data before computing the stats.

stats.fun.arg a list of arguments passed to 'type' when 'type' is a function.

Value

a data.table that contains stats values (if MFI, for each marker per column) along with 'pop' column and 'sample' column (when used on a 'GatingSet')

Examples

## Not run:
dataDir <- system.file("extdata", package="flowWorkspaceData")
suppressMessages(gs <- load_gs(list.files(dataDir, pattern = "gs_manual", full = TRUE)))

# get stats all nodes
dt <- gs_pop_get_stats(gs) #default is "count"
nodes <- c("CD4", "CD8")
gs_pop_get_stats(gs, nodes, "percent")

# pass a build-in function
gs_pop_get_stats(gs, nodes, type = pop.MFI)

# compute the stats based on the raw data scale
gs_pop_get_stats(gs, nodes, type = pop.MFI, inverse.transform = TRUE)

# supply user-defined stats fun
pop.quantiles <- function(fr){
  chnls <- colnames(fr)
  res <- matrixStats::colQuantiles(exprs(fr), probs = 0.75)
  names(res) <- chnls
  res
}
gs_pop_get_stats(gs, nodes, type = pop.quantiles)

## End(Not run)

---

### gs_pop_get_stats_tfilter

```
Extract stats from populations(or nodes) within a restricted time
window
```

#### Description

Extract stats from populations(or nodes) within a restricted time window

#### Usage

```
gs_pop_get_stats_tfilter(x, ...)
```

```
gh_pop_get_stats_tfilter(
  x,
  nodes = NULL,
  type = "count",
  inverse.transform = FALSE,
  stats.fun.arg = list(),
  tfilter = NULL,
  ...
)
```

#### Arguments

- **nodes**: the character vector specifies the populations of interest. default is all available nodes
- **type**: the character vector specifies the type of pop stats or a function used to compute population stats. when character, it is expected to be either "count" or "percent". Default is "count" (total number of events in the populations). when a function, it takes a flowFrame object through ’fr’ argument and return the stats as a named vector.
gs_pop_set_gate

Description
update the population node with a flowCore-compatible gate object

Usage
gh_pop_set_gate(obj, y, value, negated = FALSE, ...)
gs_pop_set_gate(obj, y, value, ...)

Arguments
obj GatingHierarchy or GatingSet
y character node name or path
value filter or filterList or list of filter objects
negated logical see add
... other arguments

details
Usually recompute is followed by this call since updating a gate doesn’t re-calculate the cell events within the gate automatically. see filterObject for the gate types that are currently supported.

Examples
## Not run:
rg1 <- rectangleGate("FSC-H"=c(200,400), "SSC-H"=c(250, 400), filterId="rectangle")
rg2 <- rectangleGate("FSC-H"=c(200,400), "SSC-H"=c(250, 400), filterId="rectangle")
flist <- list(rg1,rg2)
names(flist) <- sampleNames(gs[1:2])
gs_pop_set_gate(gs[1:2], "lymph", flist)
recompute(gs[1:2], "lymph")

## End(Not run)
gs_pop_set_name

Update the name of one node in a gating hierarchy/GatingSet.

Description

`gh_pop_set_name/gs_pop_set_name` update the name of one node in a gating hierarchy/GatingSet.

Usage

```
gh_pop_set_name(x, y, value)
gs_pop_set_name(x, y, value)
```

Arguments

- **x**: GatingHierarchy
- **y**: pop name/path
- **value**: A character the name of the node

Examples

```r
## Not run:
#G is a gating hierarchy
gs_get_pop_paths(G[[1]]) # return node names
gh_pop_set_name(G, "L", "lymph")
## End(Not run)
```

gs_pop_set_visibility

hide/unhide a node

Description

hide/unhide a node

Usage

```
gh_pop_set_visibility(x, y, value)
gs_pop_set_visibility(x, y, value)
```

Arguments

- **x**: GatingHierarchy object
- **y**: character node name or path
- **value**: TRUE/FALSE to indicate whether to hide a node
Examples

```r
## Not run:
gh_pop_set_visibility(gh, 4, FALSE) # hide a node
gs_remove_redundant_channels
```

### Description
Removing these redundant channels can help standardize the channels across different GatingSet objects and make them mergable.

### Usage
```r
gs_remove_redundant_channels(gs, ...)
```

### Arguments
- **gs**: a GatingSet
- **...**: other arguments passed to gs_get_pop_paths method

### Value
A new GatingSet object that has redundant channels removed. Please note that this new object shares the same reference (or external pointers) with the original GatingSets.

Examples

```r
## Not run:
gs_new <- gs_remove_redundant_channels(gs)
```

### Description
It is usually called after `gs_split_by_tree` and `gs_check_redundant_nodes`. The operation is done in place through external pointers which means all the original GatingSets are modified.

### Usage
```r
gs_remove_redundant_nodes(x, toRemove)
```
gs_split_by_channels

split GatingSets into groups based on their flow channels

Arguments

x GatingSet or list of groups (each group is a list of 'GatingSet'). When it is a list, it is usually the outcome from gs_split_by_tree.

toRemove list of the node sets to be removed. its length must equals to the length of 'x'. When x is a list, toRemove is usually the outcome from gs_check_redundant_nodes.

Examples

## Not run:
gslist <- list(gs1, gs2, gs3, gs4, gs5)
gs_groups <- gs_split_by_tree(gslist)
toRm <- gs_check_redundant_nodes(gs_groups)
gs_remove_redundant_nodes(gs_groups, toRm)

#Now they can be merged into a single GatingSetList.
#Note that the original gs objects are all modified in place.
GatingSetList(gslist)

## End(Not run)
gs_split_by_tree  

**gs_split_by_tree**  
split GatingSets into groups based on their gating schemes Be careful that the splitted results still points to the original data set!!

Description

It allows isomorphism in Gating tree and ignore difference in hidden nodes i.e. tree is considered to be the same as long as gs_get_pop_paths(gh, path = "auto", showHidden = F) returns the same set

Usage

```r
gs_split_by_tree(x)
```

Arguments

- **x** a list of GatingSets or one GatingSet

Value

when x is a GatingSet, this function returns a list of sub-GatingSets When x is a list of GatingSets, it returns a list of list, each list itself is a list of GatingSets, which share the same gating tree.

Examples

```r
## Not run:
gslist <- list(gs1, gs2, gs3, gs4, gs5)
gs_groups <- gs_split_by_tree(gslist)
## End(Not run)
```

gs_update_channels  

**gs_update_channels**  
Update the channel information of a GatingSet (C++ part)

Description

It updates the channels stored in gates, compensations and transformations based on given mapping between the old and new channel names.

Usage

```r
gs_update_channels(gs, map, all = TRUE)
```

Arguments

- **gs** a GatingSet object
- **map** data.frame contains the mapping from old (case insensitive) to new channel names Note: Make sure to remove the '<' or '>' characters from 'old' name because the API tries to only look at the raw channel name so that the gates with both prefixed and non-prefixed names could be updated.
- **all** logical whether to update the flow data as well
Value

when 'all' is set to TRUE, it returns a new GatingSet but it still shares the same underling c++ tree structure with the original GatingSet otherwise it returns nothing (less overhead.)

Examples

```r
## Not run:
## this will update both "Qdot 655-A" and "<Qdot 655-A>"
gs <- gs_update_channels(gs, map = data.frame(old = c("Qdot 655-A"),
                                          new = c("QDot 655-A"))
)

## End(Not run)
```

---

**identifier-methods**  
*Retrieve/replace the GUID of a GatingSet or GatingSetList*

**Description**  
Retrieve or replace the GUID (globally unique identifier) for a GatingSet or GatingSetList

**Usage**

```r
identifier(object)

identifier(object) <- value
```

**Arguments**

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>object</td>
<td>a GatingSet or GatingSetList</td>
</tr>
<tr>
<td>value</td>
<td>string</td>
</tr>
</tbody>
</table>

**keyword**  
*Retrieve a specific keyword for a specific sample in a GatingHierarchy or or set of samples in a GatingSet or GatingSetList*

**Description**

Retrieve a specific keyword for a specific sample in a GatingHierarchy or or set of samples in a GatingSet or GatingSetList

**Usage**

```r
keyword(object, keyword)
```
lapply-methods

Arguments

object GatingHierarchy or GatingSet or GatingSetList
keyword character specifying keyword name. When missing, extract all keywords.
... other arguments passed to keyword-methods

Details

See keyword in Package ‘flowCore’

See Also

keyword-methods

Examples

## Not run:
# get all the keywords from all samples
keyword(G)
# get all the keywords from one sample
keyword(G[[1]])
# filter the instrument setting
keyword(G[[1]], compact = TRUE)
# get single keyword from all samples
keyword(G, "FILENAME")
# get single keyword from one sample
keyword(G[[1], "FILENAME"])

## End(Not run)

lapply-methods apply FUN to each sample (i.e. GatingHierarchy or cytoframe) in a GatingSet or cytoset

Description

sample names are used for names of the returned list

Usage

lapply(X, FUN, ...)

Arguments

X GatingSet or cytoset
FUN function to be applied to each sample in 'GatingSet' or 'cytoset'
... other arguments to be passed to 'FUN'
**length**  
*Methods to get the length of a GatingSet*

**Description**

Return the length of a GatingSet or GatingSetList object (number of samples).

**Usage**

```r
length(x)
show(object)
```

**Arguments**

- `x`: GatingSet object
- `object`: object

---

**load_cytoframe_from_fcs**  
*Read a single FCS file in to a cytoframe*

**Description**

Similar to `read.FCS`, this takes a filename for a single FCS file and returns a cytoframe.

**Usage**

```r
load_cytoframe_from_fcs(
  filename,
  transformation = "linearize",
  which.lines = NULL,
  alter.names = FALSE,
  column.pattern = NULL,
  invert.pattern = FALSE,
  decades = 0,
  is_h5 = FALSE,
  h5_filename = tempfile(fileext = ".h5"),
  min.limit = NULL,
  truncate_max_range = TRUE,
  dataset = NULL,
  emptyValue = TRUE,
  num_threads = 1,
  ignore.text.offset = FALSE,
  text.only = FALSE
)
```
load_cytoframe_from_fcs

Arguments

filename  Character of length 1: filename

transformation  An character string that defines the type of transformation. Valid values are linearize (default), linearize-with-PnG-scaling, or scale. The linearize transformation applies the appropriate power transform to the data. The linearize-with-PnG-scaling transformation applies the appropriate power transform for parameters stored on log scale, and also a linear scaling transformation based on the ‘gain’ (FCS $PnG$ keywords) for parameters stored on a linear scale. The scale transformation scales all columns to $[0,10^{\text{decades}}]$. defaulting to decades=0 as in the FCS4 specification. A logical can also be used: TRUE is equal to linearize and FALSE (or NULL) corresponds to no transformation. Also when the transformation keyword of the FCS header is set to "custom" or "applied", no transformation will be used.

which.lines  Numeric vector to specify the indices of the lines to be read. If NULL all the records are read, if of length 1, a random sample of the size indicated by which.lines is read in.

alter.names  boolean indicating whether or not we should rename the columns to valid R names using make.names. The default is FALSE.

column.pattern  An optional regular expression defining parameters we should keep when loading the file. The default is NULL.

invert.pattern  logical. By default, FALSE. If TRUE, inverts the regular expression specified in column.pattern. This is useful for indicating the channel names that we do not want to read. If column.pattern is set to NULL, this argument is ignored.

decades  When scaling is activated, the number of decades to use for the output.

is_h5  Boolean indicating whether the data should be stored in h5 format

h5_filename  String specifying a name for the h5 file if is_h5 is TRUE

min.limit  The minimum value in the data range that is allowed. Some instruments produce extreme artifactual values. The positive data range for each parameter is completely defined by the measurement range of the instrument and all larger values are set to this threshold. The lower data boundary is not that well defined, since compensation might shift some values below the original measurement range of the instrument. This can be set to an arbitrary number or to NULL (the default value), in which case the original values are kept.

truncate_max_range  logical type. Default is TRUE. can be optionally turned off to avoid truncating the extreme positive value to the instrument measurement range i.e. $PnR$.

dataset  The FCS file specification allows for multiple data segments in a single file. Since the output of load_cytoframe_from_cytoset is a single cytoframe we can’t automatically read in all available sets. This parameter allows to chose one of the subsets for import. Its value is supposed to be an integer in the range of available data sets. This argument is ignored if there is only a single data segment in the FCS file.

emptyValue  Boolean indicating whether or not we allow empty value for keyword values in TEXT segment. It affects how the double delimiters are treated. IF TRUE, The double delimiters are parsed as a pair of start and end single delimiter for an empty value. Otherwise, double delimiters are parsed one part of string as the keyword value. default is TRUE.

num_threads  Integer allowing for parallelization of the parsing operation by specifying a number of threads
ignore.text.offset
Whether to ignore the keyword values in TEXT segment when they don’t agree with the HEADER. Default is FALSE, which throws the error when such discrepancy is found. User can turn it on to ignore TEXT segment when he is sure of the accuracy of HEADER so that the file still can be read.

files
A vector of filenames

...Further arguments that get passed on to read.AnnotatedDataFrame, see details

Details
The function load_cytoframe_from_fcs works with the output of the FACS machine software from a number of vendors (FCS 2.0, FCS 3.0 and List Mode Data LMD). However, the FCS 3.0 standard includes some options that are not yet implemented in this function. If you need extensions, please let me know. The output of the function is an object of class cytoframe.

For specifications of FCS 3.0 see http://www.isac-net.org and the file ../doc/fcs3.html in the doc directory of the package.

The which.lines arguments allow you to read a subset of the record as you might not want to read the thousands of events recorded in the FCS file. It is mainly used when there is not enough memory to read one single FCS (which probably will not happen). It will probably take more time than reading the entire FCS (due to the multiple disk IO).

Value
An object of class cytoframe that contains the data, the parameters monitored, and the keywords and values saved in the header of the FCS file.

See Also
Other cytoframe/cytoset IO functions: cf_get_h5_file_path(), cf_write_h5(), cs_get_h5_file_path(), load_cytoframe_from_h5(), load_cytoset_from_fcs()

load_cytoframe_from_h5
Load the cytoframe from h5 format

Description
Load the cytoframe from h5 format

Usage
load_cytoframe_from_h5(filename, readonly = TRUE, on_disk = TRUE)

Arguments
filename
the full path of the output h5 file
readonly
logical flag indicating whether to open h5 data as readonly. Default is TRUE.
on_disk
logical flag indicating whether to keep the data on disk and load it on demand. Default is TRUE.
load_cytoset_from_fcs

Read one or several FCS files in to a cytoset

Description

Similar to read.flowSet, this takes a list of FCS filenames and returns a cytoset.

Usage

load_cytoset_from_fcs(
  files = NULL,
  path = ".",
  pattern = NULL,
  phenoData = NULL,
  descriptions,
  name.keyword,
  transformation = "linearize",
  which.lines = NULL,
  alter.names = FALSE,
  column.pattern = NULL,
  invert.pattern = FALSE,
  decades = 0,
  is_h5 = FALSE,
  min.limit = NULL,
  truncate_max_range = TRUE,
  dataset = NULL,
  emptyValue = TRUE,
  num_threads = 1,
  ignore.text.offset = FALSE,
  sep = "\t",
  as.is = TRUE,
  name,
  h5_dir = tempdir(),
  ...
)

Arguments

files Optional character vector with filenames.
path Directory where to look for the files.
pattern This argument is passed on to dir, see details.
phenoData An object of class AnnotatedDataFrame, character or a list of values to be extracted from the cytoframe object, see details.
descriptions Character vector to annotate the object of class cytoset.
name.keyword An optional character vector that specifies which FCS keyword to use as the sample names. If this is not set, the GUID of the FCS file is used for sample Names, and if that is not present (or not unique), then the file names are used.

transformation see load_cytoframe_from_fcs for details.

which.lines see load_cytoframe_from_fcs for details.

alter.names see load_cytoframe_from_fcs for details.

column.pattern see load_cytoframe_from_fcs for details.

invert.pattern see load_cytoframe_from_fcs for details.

decades see load_cytoframe_from_fcs for details.

is_h5 Boolean indicating whether the data should be stored in h5 format

min.limit see load_cytoframe_from_fcs for details.

truncate_max_range see load_cytoframe_from_fcs for details.

dataset see load_cytoframe_from_fcs for details.

emptyValue see load_cytoframe_from_fcs for details.

num_threads Integer allowing for parallelization of the parsing operation by specifying a number of threads

ignore.text.offset see load_cytoframe_from_fcs for details.

sep Separator character that gets passed on to read.AnnotatedDataFrame.

as.is Logical that gets passed on to read.AnnotatedDataFrame. This controls the automatic coercion of characters to factors in the phenoData.

name An optional character scalar used as name of the object.

h5_dir String specifying a name for the h5 directory for the h5 files if is_h5 is TRUE

... Further arguments that get passed on to read.AnnotatedDataFrame, see details.

Details
There are four different ways to specify the file from which data is to be imported:
First, if the argument phenoData is present and is of class AnnotatedDataFrame, then the file names are obtained from its sample names (i.e. row names of the underlying data.frame). Also column name will be generated based on sample names if it is not there. This column is mainly used by visualization methods in flowViz. Alternatively, the argument phenoData can be of class character, in which case this function tries to read a AnnotatedDataFrame object from the file with that name by calling read.AnnotatedDataFrame(file.path(path,phenoData),...{}).

In some cases the file names are not a reasonable selection criterion and the user might want to import files based on some keywords within the file. One or several keyword value pairs can be given as the phenoData argument in form of a named list.

Third, if the argument phenoData is not present and the argument files is not NULL, then files is expected to be a character vector with the file names.

Fourth, if neither the argument phenoData is present nor files is not NULL, then the file names are obtained by calling dir(path,pattern).

Value
An object of class cytoset.
load_meta

Flush/load meta data (keywords, pData, channels/markers) to/from disk (only valid for on-disk cytoset/cytoframe)

Description
Flush/load meta data (keywords, pData, channels/markers) to/from disk (only valid for on-disk cytoset/cytoframe)

Usage

```r
cf_flush_meta(cf)
cf_load_meta(cf)
```

Arguments

- `cf` cytoframe object

lock

Lock/Unlock the cytoset/cytoframe by turning on/off its read-only flag

Description
Lock/Unlock the cytoset/cytoframe by turning on/off its read-only flag

Usage

```r
cf_lock(cf)
cf_unlock(cf)
```

Arguments

- `cf` cytoframe object

- `cs` cytoset object

See Also

Other cytoframe/cytoset IO functions: `cf_get_h5_file_path()`, `cf_write_h5()`, `cs_get_h5_file_path()`, `load_cytoframe_from_fcs()`, `load_cytoframe_from_h5()`
logicleGml2_trans GatingML2 version of logicle transformation.

Description

The only difference from logicle_trans is it is scaled to c(0,1) range.

Usage

logicleGml2_trans(
  T = 262144,
  M = 4.5,
  W = 0.5,
  A = 0,
  n = 6,
  equal.space = FALSE
)

Arguments

T, M, W, A see logicleGml2
n desired number of breaks (the actual number will be different depending on the data range)
equal.space whether breaks at equal-spaced intervals

Value

a logicleGml2 transformation object

Examples

trans.obj <- logicleGml2_trans(equal.space = TRUE)
data <- 1:1e3
brks.func <- trans.obj["breaks"]
brks <- brks.func(data)
brks # logicle space displayed at raw data scale
#transform it to verify the equal-spaced breaks at transformed scale
print(trans.obj["transform"](brks))

logicle_trans logicle transformation.

Description

Used for construct logicle transform object.

Usage

logicle_trans(..., n = 6, equal.space = FALSE)
logtGml2_trans

Description

Used to construct GML 2.0 flog transformer object.

Usage

logtGml2_trans(t = 262144, m = 4.5, n = 6, equal.space = FALSE)

Arguments

t  top scale value
m  number of decades
n  desired number of breaks (the actual number will be different depending on the data range)
equal.space  whether breaks at equal-spaced intervals

Details

GML 2.0 standard log transform function constructor. The definition is as in the GML 2.0 standard section 6.2 "parametrized logarithmic transformation – flog" This deviates from standard only in the following way. Before applying the logarithmic transformation, non-positive values are assigned the smallest positive value from the input rather than having undefined values (NA) under the transformation.

Value

logtGml2 transformation object
Examples

```r
trans.obj <- logtGml2_trans(t = 1e3, m = 1, equal.space = TRUE)
data <- 1:1e3
brks.func <- trans.obj[['breaks']]brks <- brks.func(data)
brks # fasin space displayed at raw data scale
#transform it to verify it is equal-spaced at transformed scale
trans.func <- trans.obj[['transform']]
brks.trans <- trans.func(brks)
brks.trans
```

markernames

*Get/set the column(channel) or marker names*

Description

It simply calls the methods for the underlying flow data (flowSet/ncdfFlowSet/ncdfFlowList).

Usage

```r
markernames(object)
markernames(object) <- value
colnames(object)
colnames(object) <- value
```

Arguments

- `value` named character vector for markernames<-, regular character vector for colnames<-
- `x, object` GatingHierarchy/GatingSet/GatingSetList
- `do.NULL, prefix` not used.

Examples

```r
## Not run:
markers.new <- c("CD4", "CD8")
chnls <- c("<B710-A>", "<R780-A>")
names(markers.new) <- chnls
markernames(gs) <- markers.new
chnls <- colnames(gs)
chnls.new <- chnls
chnls.new[c(1, 4)] <- c("fsc", "ssc")
colnames(gs) <- chnls.new

## End(Not run)
```
merge_list_to_gs

**Description**

It also checks the consistency of the cyto data and gates.

**Usage**

```r
merge_list_to_gs(x, ...)
```

**Arguments**

- `x`: a list of GatingSets
- `...`: other arguments (not used)

ncFlowSet

*Fetch the flowData object associated with a GatingSet.*

**Description**

Deprecated by `flowData` method

Deprecated by `flowData` method

nodeflags

*The flags of gate nodes*

**Description**

`gh_pop_is_gated` checks if a node is already gated. `gh_pop_is_negated` checks if a node is negated. `gh_pop_is_hidden` checks if a node is hidden.

**Usage**

```r
gh_pop_is_gated(obj, y)
gh_pop_is_negated(obj, y)
gh_pop_is_hidden(obj, y)
gh_pop_is_bool_gate(obj, y)
```

**Arguments**

- `obj`: GatingHierarchy
- `y`: node/gating path
- `...`: not used
openWorkspace

*Description*

It is now moved along with entire flowJo parser to CytoML package

*Usage*

```r
openWorkspace(file, ...)
```

*Arguments*

- `file` xml file
- `...` other arguments

---

pData-methods

*Description*

Accessor method that gets or replaces the pData of the flowset/ncdfFlowSet object in a GatingHierarchy, GatingSet, or GatingSetList

*Usage*

```r
pData(object)
pData(object) <- value
```

*Arguments*

- `object` GatingSet or GatingSetList
- `value` data.frame The replacement of pData for flowSet or ncdfFlowSet object

*Value*

- a data.frame
plot-methods

plot-methods
plot a gating tree

Description
Plot a tree/graph representing the GatingHierarchy

Usage
plot(x, y, ...)

Arguments
x GatingHierarchy or GatingSet. If GatingSet, the first sample will be used to extract gating tree.
y missing or character specifies.
... other arguments:
  • boolean: TRUE|FALSE logical specifying whether to plot boolean gate nodes. Defaults to FALSE.
  • showHidden: TRUE|FALSE logical whether to show hidden nodes
  • layout: See layoutGraph in package Rgraphviz
  • width: See layoutGraph in package Rgraphviz
  • height: See layoutGraph in package Rgraphviz
  • fontsize: See layoutGraph in package Rgraphviz
  • labelfontsize: See layoutGraph in package Rgraphviz
  • fixedsize: See layoutGraph in package Rgraphviz

Examples
## Not run:
#gs is a GatingSet
plot(gs) # the same as plot(gs[[1]])
#plot a substree rooted from 'CD4'
plot(gs, "CD4")

## End(Not run)

plotGate-methods-defunct
Plot gates and associated cell population contained in a GatingHierarchy or GatingSet
Description

**Important:** The `plotGate` methods are now defunct and gates should instead be plotted using the `autoplot` method from the `ggcyto` package. The `plotGate` documentation has been left here to ease the transition.

When applied to a `GatingHierarchy`, `arrange` is set as `TRUE`, then all the gates associated with it are plotted as different panel on the same page. If `arrange` is `FALSE`, then it plots one gate at a time. By default, `merge` is set as `TRUE`, plot multiple gates on the same plot when they share common parent population and axis. When applied to a `GatingSet`, if `lattice` is `TRUE`, it plots one gate (multiple samples) per page, otherwise, one sample (with multiple gates) per page.

Usage

```r
plotGate(x, y, ...)  
```

Arguments

- `x` GatingSet or GatingHierarchy object
- `y` character the node name or full/(partial) gating path or numeric representing the node index in the `GatingHierarchy`, or `missing` which will plot all gates and one gate per page. It is useful for generating plots in a multi-page pdf. Nodes can be accessed with `gs_get_pop_paths`.
- `...` 
  - `bool` logical specifying whether to plot boolean gates.
  - `arrange.main` character The title of the main page of the plot. Default is the sample name. Only valid when `x` is `GatingHierarchy`
  - `arrange` logical indicating whether to arrange different populations/nodes on the same page via `arrangeGrob` call.
  - `merge` logical indicating whether to draw multiple gates on the same plot if these gates share the same parent population and same x,y dimensions/parameters;
  - `projections` list of character vectors used to customize x,y axis. By default, the x,y axis are determined by the respective gate parameters. The elements of the list are named by the population name or path (see `y`). Each element is a pair of named character specifying the channel name(or marker name) for x, y axis. Short form of channel or marker names (e.g. "APC" or "CD3") can be used as long as they can be uniquely matched to the dimensions of flow data. For example, `projections = list("lymph" = c(x = "SSC-A", y = "FSC-A"), "CD3" = c(x = "CD3", y = "SSC-A"))`
  - `par.settings` list of graphical parameters passed to `lattice`;
  - `gpar` list of grid parameters passed to `grid.layout`;
  - `lattice` logical deprecated;
  - `formula` formula a formula passed to `xyplot` function of `flowViz`, by default it is `NULL`, which means the formula is generated according to the x,y parameters associated with gate.
  - `cond` character the conditioning variable to be passed to `lattice` plot.
  - `overlayNode` names. These populations are plotted on top of the existing gates(defined by `y` argument) as the overlaid dots.
  - `overlay.symbol` a named (lattice graphic parameter) list that defines the symbol color and size for each overlaid population. If not given, we automatically assign the colors.
pop_add

- `keyLattice` legend parameter for overlay symbols.
- `default.y` character specifying y channel for `xyplot` when plotting a 1d gate. Default is "SSC-A" and session-wise setting can be stored by `flowWorkspace.par.set("plotGate", list(default.y = "FSC-A"))`.
- `type` character either "xyplot" or "densityplot". Default is "xyplot" and session-wise setting can be stored by `flowWorkspace.par.set("plotGate", list(type = "xyplot"))`.
- `fitGate` used to disable behavior of plotting the gate region in 1d density-plot. Default is FALSE and session-wise setting can be stored by `flowWorkspace.par.set("plotGate", list(fitGate = FALSE))`.
- `strip` logical specifies whether to show pop name in strip box, only valid when `x` is `GatingHierarchy`.
- `strip.text` either "parent" (the parent population name) or "gate" (the gate name).
- `raw.scale` logical whether to show the axis in raw(untransformed) scale. Default is TRUE and can be stored as session-wise setting by `flowWorkspace.par.set("plotGate", list(raw.scale = TRUE))`.
- `xlim`, `ylim` character can be either "instrument" or "data" which determines the x, y axis scale either by instrument measurement range or the actual data range. or numeric which specifies customized range. They can be stored as session-wise setting by `flowWorkspace.par.set("plotGate", list(lim = "instrument"))`.
- `...` path A character or numeric scalar passed to `gs_get_pop_paths` method (used to control how the gating/node path is displayed)
- ... The other additional arguments to be passed to `xyplot`.

Value

A `trellis` object if `arrange` is FALSE.

References


Examples

```r
## Not run:
#G is a GatingHierarchy
plotGate(G, gs_get_pop_paths(G)[5]); # plot the gate for the fifth node

## End(Not run)
```

pop_add

Add populations to a GatingHierarchy

Description

Add populations to a GatingHierarchy.
Usage

pop_add(gate, gh, ...)  
## S3 method for class 'filter'
pop_add(gate, gh, ...)  
## S3 method for class 'filters'
pop_add(gate, gh, names = NULL, ...)  
## S3 method for class 'quadGate'
pop_add(gate, gh, names = NULL, ...)  
## S3 method for class 'logical'
pop_add(gate, gh, parent, name, recompute, cluster_method_name = NULL, ...)  
## S3 method for class 'factor'
pop_add(gate, gh, name = NULL, ...)  
## S3 method for class 'logicalFilterResult'
pop_add(gate, gh, ...)  
## S3 method for class 'multipleFilterResult'
pop_add(gate, gh, name = NULL, ...)  
gh_pop_remove(gh, node, ...)

Arguments

gate a gate object that extends flowCore::filter or flowCore::filters
gh GatingHierarchy
... other arguments
names a character vector of length four, which specifies the population names resulted by adding a quadGate. The order of the names is clock-wise starting from the top left quadrant population.
parent a character scalar to specify the parent node name where the new gate to be added to, by default it is NULL, which indicates the root node
name the population name
recompute whether to recompute the gates
cluster_method_name when adding the logical vectors as the gates, the name of the cluster method can be used to tag the populations as the extra meta information associated with the gates.
node population name/path
prettyAxis

Determine tick mark locations and labels for a given channel axis

Description

Determine tick mark locations and labels for a given channel axis

Usage

prettyAxis(gh, channel)

Arguments

gh GatingHiearchy
channel character channel name

Value

when there is transformation function associated with the given channel, it returns a list of that contains positions and labels to draw on the axis other wise returns NULL

Examples

## Not run:
prettyAxis(gh, "<B710-A>")

## End(Not run)

recompute

Compute the cell events by the gates stored within the gating tree.

Description

Compute each cell event to see if it falls into the gate stored within the gating tree and store the result as cell count.

Usage

recompute(x, y="root", alwaysLoadData=FALSE, ...)

Arguments

x GatingSet or GatingSetList
y character node name or node path. Default "root". Optional.
alwaysLoadData logical. Specifies whether to load the flow raw data for gating boolean gates. Default 'FALSE'. Optional. Sometime it is more efficient to skip loading the raw data if all the reference nodes and parent are already gated. 'FALSE' will check the parent node and reference to determine whether to load the data. This check may not be sufficient since the further upstream ancestor nodes may not be gated yet. In that case, we allow the gating to fail and prompt user to recompute those nodes explicitly. When TRUE, then it forces data to be loaded to guarantee the gating process to be uninterrupted at the cost of unnecessary data IO.

... other arguments leaf.bool whether to compute the leaf boolean gate, default is TRUE.

Details

It is usually used immediately after add or gs_pop_set_gate calls.

rotate_gate

Simplified geometric rotation of gates associated with nodes

Description

Rotate a gate associated with a node of a GatingHierarchy or GatingSet. This method is a wrapper for rotate_gate that enables updating of the gate associated with a node of a GatingHierarchy or GatingSet.

rotate_gate calls gs_pop_set_gate to modify the provided GatingHierarchy or GatingSet directly so there is no need to re-assign its output. The arguments will be essentially identical to the flowCore method, except for the specification of the target gate. Rather than being called on an object of type flowCore:filter, here it is called on a GatingHierarchy or GatingSet object with an additional character argument for specifying the node whose gate should be transformed. The rest of the details below are taken from the FlowCore documentation.

Usage

rotate_gate(obj, y, deg = NULL, rot_center = NULL, ...)

Arguments

obj A GatingHierarchy or GatingSet object
y A character specifying the node whose gate should be modified
deg An angle in degrees by which the gate should be rotated in the counter-clockwise direction
rot_center A separate 2-dimensional center of rotation for the gate, if desired. By default, this will be the center for ellipsoidGate objects or the centroid for polygonGate objects. The rot_center argument is currently only supported for polygonGate objects.
... not used
This method allows for geometric rotation of filter types defined by simple geometric gates (ellipsoidGate, and polygonGate). The method is not defined for rectangleGate or quadGate objects, due to their definition as having 1-dimensional boundaries.

The angle provided in the deg argument should be in degrees rather than radians. By default, the rotation will be performed around the center of an ellipsoidGate or the centroid of the area encompassed by a polygonGate. The rot_center argument allows for specification of a different center of rotation for polygonGate objects (it is not yet implemented for ellipsoidGate objects) but it is usually simpler to perform a rotation and a translation individually than to manually specify the composition as a rotation around a shifted center.

See Also

transform_gate FlowCore::rotate_gate

Examples

## Not run:
# Rotates the original gate 15 degrees counter-clockwise
rotate_gate(gs, node, deg = 15)
# Rotates the original gate 270 degrees counter-clockwise
rotate_gate(gs, node, 270)

## End(Not run)

---

**sampleNames**

Get/update sample names in a GatingSet

**Description**

Return a sample names contained in a GatingSet

**Usage**

sampleNames(object)

sampleNames(object) <- value

**Arguments**

object a GatingSet

value character new sample names

**Details**

The sample names comes from pdata of fs.

**Value**

A character vector of sample names
Examples
## Not run:
#G is a GatingSet
sampleNames(G)
## End(Not run)

### sampleNames, character-method

Get sample names from a GatingSet archive folder

Description
retrive sample names by scanning h5 files from a GatingSet folder

Usage
## S4 method for signature 'character'
sampleNames(object)

Arguments
object a GatingSet folder

Value
A character vector of sample names

Examples
## Not run:
sampleNames(gd)
## End(Not run)

save_cytoset

save/load a cytoset to/from disk.

Description
Save/load a cytoset to/from the disk.

Usage
save_cytoset(cs, path, ...)
load_cytoset(path, ...)
save_gs

Arguments

- cs: A cytoset
- path: A character scalar giving the path to save/load the GatingSet to/from.
- ... other arguments passed to save_gs/load_gs

Value

load_cytoset returns a cytoset object

Examples

```r
## Not run:
#G is a GatingSet
save_cytoset(cs, outdir)
cs <- load_cytoset(outdir)

## End(Not run)
```

---

**Description**

Save/load a GatingSet/GatingSetList which is the gated flow data including gates and populations to/from the disk. The GatingSet object The internal C data structure (gating tree), ncdfFlowSet object(if applicable)

**Usage**

```r
save_gs(gs, path, cdf = c("copy", "move", "skip", "symlink", "link"), ...)
load_gs(path, h5_readonly = TRUE, select = character(), verbose = FALSE)
save_gslist(gslist, path, ...)
load_gslist(path)
```

**Arguments**

- path: A character scalar giving the path to save/load the GatingSet to/from.
- cdf: a character scalar. The valid options are: "copy", "move", "skip", "symlink", "link" specifying what to do with the cdf data file. Sometime it is more efficient to move or create a link of the existing cdf file to the archived folder. It is useful to "skip" archiving cdf file if raw data has not been changed.
- ... other arguments: not used.
- h5_readonly: whether to open h5 data as read-only. Default is TRUE
- select: an integer or character vector to select a subset of samples to load
verbose logical flag to optionally print the versions of the libraries that were used to
archive the GatingSet for troubleshooting purpose.

gslist A GatingSetList
G A GatingSet

Value
load_gs returns a GatingSet object load_gslist returns a GatingSetList object

See Also
GatingSet-class, GatingSetList-class

Examples
## Not run:
#G is a GatingSet
save_gs(G,path="tempFolder")
G1<-load_gs(path="tempFolder")

#G is a GatingSet
save_gslist(gslist1,path="tempFolder")
gslist2<-load_gslist(path="tempFolder")

## End(Not run)

scale_gate

Simplified geometric scaling of gates associated with nodes

Usage
scale_gate(obj, y, scale = NULL, ...)

Arguments
obj A GatingHierarchy or GatingSet object
y A character specifying the node whose gate should be modified
scale Either a numeric scalar (for uniform scaling in all dimensions) or numeric vector
specifying the factor by which each dimension of the gate should be expanded
(absolute value > 1) or contracted (absolute value < 1). Negative values will
result in a reflection in that dimension.
... not used
Details

This method allows uniform or non-uniform geometric scaling of filter types defined by simple geometric gates (quadGate, rectangleGate, ellipsoidGate, and polygonGate) Note that these methods are for manually altering the geometric definition of a gate. To easily transform the definition of a gate with an accompanying scale transformation applied to its underlying data, see ?gcyto::rescale_gate.

The scale argument passed to scale_gate should be either a scalar or a vector of the same length as the number of dimensions of the gate. If it is scalar, all dimensions will be multiplicatively scaled uniformly by the scalar factor provided. If it is a vector, each dimension will be scaled by its corresponding entry in the vector.

The scaling behavior of scale_gate depends on the type of gate passed to it. For rectangleGate and quadGate objects, this amounts to simply scaling the values of the 1-dimensional boundaries. For polygonGate objects, the values of scale will be used to determine scale factors in the direction of each of the 2 dimensions of the gate (scale_gate is not yet defined for higher-dimensional polytopeGate objects). Important: For ellipsoidGate objects, scale determines scale factors for the major and minor axes of the ellipse, in that order. Scaling by a negative factor will result in a reflection in the corresponding dimension.

See Also

transform_gate flowCore::scale_gate

Examples

```r
## Not run:
# Scales both dimensions by a factor of 5
scale_gate(gs, node, 5)

# Shrinks the gate in the first dimension by factor of 1/2
# and expands it in the other dimension by factor of 3
scale_gate(gs, node, c(0.5,3))

## End(Not run)
```

Description

Simplified geometric translation of gates associated with nodes

Shift the location of a gate associated with a node of a GatingHierarchy or GatingSet. This method is a wrapper for shift_gate that enables updating of the gate associated with a node of a GatingHierarchy or GatingSet.

shift_gate calls gs_pop_set_gate to modify the provided GatingHierarchy or GatingSet directly so there is no need to re-assign its output. The arguments will be essentially identical to the flowCore method, except for the specification of the target gate. Rather than being called on an object of type flowCore::filter, here it is called on a GatingHierarchy or GatingSet object with an additional character argument for specifying the node whose gate should be transformed. The rest of the details below are taken from the flowCore documentation.
**Usage**

```r
shift_gate(obj, y, dx=NULL, dy=NULL, center=NULL, ...)
```

**Arguments**

- `obj` A `GatingHierarchy` or `GatingSet` object
- `y` A character specifying the node whose gate should be modified
- `dx` Either a numeric scalar or numeric vector. If it is scalar, this is just the desired shift of the gate in its first dimension. If it is a vector, it specifies both dx and dy as (dx, dy). This provides an alternate syntax for shifting gates, as well as allowing shifts of `ellipsoidGate` objects in more than 2 dimensions.
- `dy` A numeric scalar specifying the desired shift of the gate in its second dimension.
- `center` A numeric vector specifying where the center or centroid should be moved (rather than specifying dx and/or dy)
- `...` not used

**Details**

This method allows for geometric translation of filter types defined by simple geometric gates (`rectangleGate`, `quadGate`, `ellipsoidGate`, or `polygonGate`). The method provides two approaches to specify a translation. For `rectangleGate` objects, this will shift the min and max bounds by the same amount in each specified dimension. For `quadGate` objects, this will simply shift the dividing boundary in each dimension. For `ellipsoidGate` objects, this will shift the center (and therefore all points of the ellipse). For `polygonGate` objects, this will simply shift all of the points defining the polygon.

The method allows two different approaches to shifting a gate. Through the dx and/or dy arguments, a direct shift in each dimension can be provided. Alternatively, through the center argument, the gate can be directly moved to a new location in relation to the old center of the gate. For `quadGate` objects, this center is the intersection of the two dividing boundaries (so the value of the boundary slot). For `rectangleGate` objects, this is the center of the rectangle defined by the intersections of the centers of each interval. For `ellipsoidGate` objects, it is the center of the ellipsoid, given by the mean slot. For `polygonGate` objects, the centroid of the old polygon will be calculated and shifted to the new location provided by center and all other points on the polygon will be shifted by relation to the centroid.

**See Also**

- `transform_gate`
- `flowCore::shift_gate`

**Examples**

```r
## Not run:
# Moves the entire gate +500 in its first dimension and 0 in its second dimension
shift_gate(gs, node, dx = 500)

# Moves the entire gate +250 in its first dimension and +700 in its second dimension
shift_gate(gs, node, dx = 500, dy = 700)

# Same as previous
shift_gate(gs, node, c(500, 700))

# Move the gate based on shifting its center to (700, 1000)
```
The tools to standardize the tree structures and channel names.

Description

- `gs_split_by_tree(x)`
- `gs_split_by_channels(x)`
- `gs_check_redundant_nodes(x)`
- `gs_remove_redundant_nodes(x, toRemove)`
- `gs_remove_redundant_channels(gs)`
- `gs_update_channels(gs, map, all = TRUE)`
- `gh_pop_move(gh, node, to)`
- `gs_pop_set_visibility(x, y, FALSE)`

Details

In order to merge multiple GatingSets into single `GatingSetList`, the gating trees and channel names must be consistent. These functions help removing the discrepancies and standardize the GatingSets so that they are mergable.

- `gs_split_by_tree` splits the GatingSets into groups based on the gating tree structures.
- `gs_split_by_channels` split GatingSets into groups based on their flow channels.
- `gs_check_redundant_nodes` returns the terminal(or leaf) nodes that makes the gating trees to be different among GatingSets and thus can be considered to remove as redundant nodes.
- `gs_remove_redundant_nodes` removes the terminal(or leaf) nodes that are detected as redundant by `gs_check_redundant_nodes`.
- `gs_remove_redundant_channels` remove the redundant channels that are not used by any gate defined in the GatingSet.
- `gs_update_channels` modifies the channel names in place. (Usually used to standardize the channels among GatingSets due to the letter case discrepancies or typo).
- `gh_pop_move` inserts a dummy gate to the GatingSet. Is is useful trick to deal with the extra non-leaf node in some GatingSets that can not be simply removed by `gs_remove_redundant_nodes`
- `gs_pop_set_visibility` hide a node/gate in a GatingSet. It is useful to deal with the non-leaf node that causes the tree structure discrepancy.
Description

pop.MFI computes and returns the median fluorescence intensity for each marker. They are typically used as the arguments passed to gh_pop_get_stats method to perform the sample-wise population stats calculations.

Usage

pop.MFI(fr)

Arguments

fr a flowFrame represents a gated population

Value

a named numeric vector

Description

subset the GatingSet/GatingSetList based on ’pData’

Usage

subset(x, subset, ...)

Arguments

x GatingSet or GatingSetList
subset logical expression(within the context of pData) indicating samples to keep. see subset
... other arguments. (not used)

Value

a codeGatingSet or GatingSetList object
swap_data_cols

Description

Swap the colnames Perform some validity checks before returning the updated colnames

Usage

swap_data_cols(cols, swap_cols)

Arguments

cols the original colname vector
swap_cols a named list specifying the pairs to be swapped

Value

the new colname vector that has some colnames swapped

Examples

library(flowCore)
data(GvHD)
fr <- GvHD[[1]]
colnames(fr)
new <- swap_data_cols(colnames(fr), list("FSC-H" = "SSC-H", "FL2-H" = "FL2-A"))
colnames(fr) <- new

transform

transform the flow data associated with the GatingSet

Description

The transformation functions are saved in the GatingSet and can be retrieved by gh_get_transformations. Currently only flowJo-type biexponential transformation(either returned by gh_get_transformations or constructed by flowJoTrans) is supported.

Usage

transform(`_data`, translist, ...)

Arguments

_data GatingSet or GatingSetList
translist expect a transformList object or a list of transformList objects(with names matched to sample names)
... other arguments passed to `transform` method for `ncdfFlowSet`(e.g. `ncdf-File`)
Value

a GatingSet or GatingSetList object with the underlying flow data transformed.

Examples

```r
## Not run:
library(flowCore)
data(GvHD)
fs <- GvHD[1:2]
gs <- GatingSet(fs)

# construct biexponential transformation function
biexpTrans <- flowjo.biexp_trans(channelRange=4096, maxValue=262144, pos=4.5, neg=0, widthBasis=-10)

# make a transformList object
chnls <- c("FL1-H", "FL2-H")
transList <- transformerList(chnls, biexpTrans)

# add it to GatingSet
gs_trans <- transform(gs, transList)

## End(Not run)
```

transformerList

Constructor for transformerList object

Description

Similar to transformList function, it constructs a list of transformer objects generated by trans_new method from scales so that the inverse and breaks functions are also included.

Usage

transformerList(from, trans)

Arguments

from channel names
trans a trans object or a list of trans objects constructed by trans_new method.

Examples

```r
library(flowCore)
library(scales)
# create transformer object from scratch
trans <- logicleTransform(w = 0.5, t = 262144, m = 4.5, a = 0)
inv <- inverseLogicleTransform(trans = trans)
trans.obj <- flow_trans("logicle", trans, inv, n = 5, equal.space = FALSE)

# or simply use convenient constructor
#trans.obj <- logicle_trans(n = 5, equal.space = FALSE, w = 0.5, t = 262144, m = 4.5, a = 0)
```
transformerList(c("FL1-H", "FL2-H"), trans.obj)

# use different transformer for each channel
trans.obj2 <- asinhGml2_trans()
transformerList(c("FL1-H", "FL2-H"), list(trans.obj, trans.obj2))

transform_gate

Simplified geometric transformations of gates associated with nodes

Description

Perform geometric transformations of a gate associated with a node of a GatingHierarchy or GatingSet. This method is a wrapper for transform_gate that enables updating of the gate associated with a node of a GatingHierarchy or GatingSet.

transform_gate calls gs_pop_set_gate to modify the provided GatingHierarchy or GatingSet directly so there is no need to re-assign its output. The arguments will be essentially identical to the flowCore method, except for the specification of the target gate. Rather than being called on an object of type flowCore::filter, here it is called on a GatingHierarchy or GatingSet object with an additional character argument for specifying the node whose gate should be transformed. The rest of the details below are taken from the flowCore documentation.

Usage

transform_gate(obj, y, scale = NULL, deg = NULL, rot_center = NULL, dx = NULL, dy = NULL, center = NULL, ...)

Arguments

obj A GatingHierarchy or GatingSet object

y A character specifying the node whose gate should be modified

scale Either a numeric scalar (for uniform scaling in all dimensions) or numeric vector specifying the factor by which each dimension of the gate should be expanded (absolute value > 1) or contracted (absolute value < 1). Negative values will result in a reflection in that dimension. For rectangleGate and quadGate objects, this amounts to simply scaling the values of the 1-dimensional boundaries. For polygonGate objects, the values of scale will be used to determine scale factors in the direction of each of the 2 dimensions of the gate (scale_gate is not yet defined for higher-dimensional polytopeGate objects). **Important:** For ellipsoidGate objects, scale determines scale factors for the major and minor axes of the ellipse, in that order.

deg An angle in degrees by which the gate should be rotated in the counter-clockwise direction.

rot_center A separate 2-dimensional center of rotation for the gate, if desired. By default, this will be the center for ellipsoidGate objects or the centroid for polygonGate objects. The rot_center argument is currently only supported for polygonGate objects. It is also usually simpler to perform a rotation and a translation individually than to manually specify the composition as a rotation around a shifted center.

dx Either a numeric scalar or numeric vector. If it is scalar, this is just the desired shift of the gate in its first dimension. If it is a vector, it specifies both dx and dy as (dx, dy). This provides an alternate syntax for shifting gates, as well as allowing shifts of ellipsoidGate objects in more than 2 dimensions.
transform_gate

dy  A numeric scalar specifying the desired shift of the gate in its second dimension.

center  A numeric vector specifying where the center or centroid should be moved (rather than specifying dx and/or dy)

...  Assignments made to the slots of the particular Gate-type filter object in the form "<slot_name> = <value>"

Details

This method allows changes to the four filter types defined by simple geometric gates (quadGate, rectangleGate, ellipsoidGate, and polygonGate) using equally simple geometric transformations (shifting/translation, scaling/dilation, and rotation). The method also allows for directly resetting the slots of each Gate-type object. Note that these methods are for manually altering the geometric definition of a gate. To easily transform the definition of a gate with an accompanying scale transformation applied to its underlying data, see ?ggcyto::rescale_gate.

First, transform_gate will apply any direct alterations to the slots of the supplied Gate-type filter object. For example, if "mean = c(1, 3)" is present in the argument list when transform_gate is called on a ellipsoidGate object, the first change applied will be to shift the mean slot to (1, 3). The method will carry over the dimension names from the gate, so there is no need to provide column or row names with arguments such as mean or cov for ellipsoidGate or boundaries for polygonGate.

transform_gate then passes the geometric arguments (dx, dy, deg, rot_center, scale, and center) to the methods which perform each respective type of transformation: shift_gate, scale_gate, or rotateGate. The order of operations is to first scale, then rotate, then shift. The default behavior of each operation follows that of its corresponding method but for the most part these are what the user would expect. A few quick notes:

• rotate_gate is not defined for rectangleGate or quadGate objects, due to their definition as having 1-dimensional boundaries.

• The default center for both rotation and scaling of a polygonGate is the centroid of the polygon. This results in the sort of scaling most users expect, with a uniform scale factor not distorting the shape of the original polygon.

See Also

flowCore::transform_gate

Examples

## Not run:
# Scale the original gate non-uniformly, rotate it 15 degrees, and shift it
transform_gate(gs, node, scale = c(2,3), deg = 15, dx = 500, dy = -700)

# Scale the original gate (in this case an ellipsoidGate) after moving its center to (1500, 2000)
transform_gate(gs, node, scale = c(2,3), mean = c(1500, 2000))

## End(Not run)
Bracket operators on GatingSet and GatingSetList objects

Description
- `[` subsets a GatingSet or GatingSetList using the familiar bracket notation
- `[[` extracts a GatingHierarchy object from a GatingSet or GatingSetList

Usage
- `x[i]`
- `x[[i]]`

Arguments
- `x` a GatingSet or GatingSetList
- `i` numeric or logical or character used as sample indices

Value
returns an object of the same type as `x` corresponding to the subset of indices in `i`, while `[[i]]` returns a single GatingHierarchy
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