Package ‘ChromSCape’

May 17, 2024

Title  Analysis of single-cell epigenomics datasets with a Shiny App

Version  1.14.0

Description  ChromSCape - Chromatin landscape profiling for Single Cells - is a ready-to-launch user-friendly Shiny Application for the analysis of single-cell epigenomics datasets (scChIP-seq, scATAC-seq, scCUT&Tag, ...) from aligned data to differential analysis & gene set enrichment analysis. It is highly interactive, enables users to save their analysis and covers a wide range of analytical steps: QC, preprocessing, filtering, batch correction, dimensionality reduction, visualization, clustering, differential analysis and gene set analysis.

License  GPL-3

biocViews  ShinyApps, Software, SingleCell, ChIPSeq, ATACSeq, MethylSeq, Classification, Clustering, Epigenetics, PrincipalComponent, SingleCell, ATACSeq, ChIPSeq, Annotation, BatchEffect, MultipleComparison, Normalization, Pathways, Preprocessing, QualityControl, ReportWriting, Visualization, GeneSetEnrichment, DifferentialPeakCalling

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URL  https://github.com/vallotlab/ChromSCape

BugReports  https://github.com/vallotlab/ChromSCape/issues

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methods, jsonlite, edgeR, stats, graphics, grDevices, utils, S4Vectors, SingleCellExperiment, SummarizedExperiment, msigdb, forcats, Rcpp, coop, matrixTests, DelayedArray

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**Author** Pacome Prompsy [aut, cre](https://orcid.org/0000-0003-4375-7583), Celine Vallot [aut](https://orcid.org/0000-0003-1601-2359)

**Maintainer** Pacome Prompsy <pacome.prompsy@curie.fr>

## Contents

annotation_from_merged_peaks ................................................ 5
annoToCol2 ................................................................. 6
anocol_binary .............................................................. 7
anocol_categorical .......................................................... 7
bams_to_matrixindexes ....................................................... 8
beds_to_matrixindexes ....................................................... 8
calculate_CNA .............................................................. 9
calculate_gain_or_loss ....................................................... 11
calculate_logRatio_CNA ...................................................... 12
call_macs2_merge_peaks ..................................................... 12
changeRange ............................................................... 13
CheA3_TF_nTargets .......................................................... 14
check_correct_datamatrix .................................................... 15
choose_cluster_scExp ....................................................... 15
choose_perplexity .......................................................... 16
col2hex ..................................................................... 16
colors_scExp ............................................................... 17
combine_datamatrix .......................................................... 18
combine_enrichmentTests ................................................... 18
comparable_variables ......................................................... 19
CompareedgeRGLM ............................................................ 20
CompareWilcoxon ............................................................. 21
concatenate_scBed_into_clusters .......................................... 22
consensus_clustering_scExp ................................................. 23
correlation_and_hierarchical_clust_scExp ................................. 24
count_coverage ............................................................. 25
<table>
<thead>
<tr>
<th>Function</th>
<th>Page</th>
</tr>
</thead>
<tbody>
<tr>
<td>create_project_folder</td>
<td>26</td>
</tr>
<tr>
<td>create_sample_name_mat</td>
<td>27</td>
</tr>
<tr>
<td>create_scDataset_raw</td>
<td>27</td>
</tr>
<tr>
<td>create_scExp</td>
<td>28</td>
</tr>
<tr>
<td>DA_custom</td>
<td>30</td>
</tr>
<tr>
<td>DA_one_vs_rest</td>
<td>31</td>
</tr>
<tr>
<td>DA_pairwise</td>
<td>32</td>
</tr>
<tr>
<td>define_feature</td>
<td>33</td>
</tr>
<tr>
<td>detect_samples</td>
<td>33</td>
</tr>
<tr>
<td>differential_activation</td>
<td>34</td>
</tr>
<tr>
<td>differential_analysis_scExp</td>
<td>35</td>
</tr>
<tr>
<td>distPearson</td>
<td>37</td>
</tr>
<tr>
<td>enrichmentTest</td>
<td>37</td>
</tr>
<tr>
<td>enrich_TF_ChEA3_genes</td>
<td>38</td>
</tr>
<tr>
<td>enrich_TF_ChEA3_scExp</td>
<td>38</td>
</tr>
<tr>
<td>exclude_features_scExp</td>
<td>40</td>
</tr>
<tr>
<td>feature_annotation_scExp</td>
<td>41</td>
</tr>
<tr>
<td>filter_correlated_cell_scExp</td>
<td>41</td>
</tr>
<tr>
<td>filter_genes_with_refined_peak_annotation</td>
<td>43</td>
</tr>
<tr>
<td>filter_scExp</td>
<td>43</td>
</tr>
<tr>
<td>find_clusters_louvain_scExp</td>
<td>44</td>
</tr>
<tr>
<td>find_top_features</td>
<td>45</td>
</tr>
<tr>
<td>generate_analysis</td>
<td>46</td>
</tr>
<tr>
<td>generate_count_matrix</td>
<td>49</td>
</tr>
<tr>
<td>generate_coverage_tracks</td>
<td>49</td>
</tr>
<tr>
<td>generate_feature_names</td>
<td>51</td>
</tr>
<tr>
<td>generate_report</td>
<td>51</td>
</tr>
<tr>
<td>gene_set_enrichment_analysis_scExp</td>
<td>52</td>
</tr>
<tr>
<td>getExperimentNames</td>
<td>54</td>
</tr>
<tr>
<td>getMainExperiment</td>
<td>55</td>
</tr>
<tr>
<td>get_color_dataframe_from_input</td>
<td>55</td>
</tr>
<tr>
<td>get_cyto_features</td>
<td>56</td>
</tr>
<tr>
<td>get_genomic_coordinates</td>
<td>57</td>
</tr>
<tr>
<td>get_most_variable_cyto</td>
<td>57</td>
</tr>
<tr>
<td>get_pathway_mat_scExp</td>
<td>58</td>
</tr>
<tr>
<td>gg_fill_hue</td>
<td>59</td>
</tr>
<tr>
<td>groupMat</td>
<td>59</td>
</tr>
<tr>
<td>H1proportion</td>
<td>60</td>
</tr>
<tr>
<td>has_genomic_coordinates</td>
<td>60</td>
</tr>
<tr>
<td>hclustAnnotHeatmapPlot</td>
<td>61</td>
</tr>
<tr>
<td>hg38.chromosomes</td>
<td>61</td>
</tr>
<tr>
<td>hg38.cytoBand</td>
<td>62</td>
</tr>
<tr>
<td>hg38.GeneTSS</td>
<td>63</td>
</tr>
<tr>
<td>imageCol</td>
<td>63</td>
</tr>
<tr>
<td>import_count_input_files</td>
<td>64</td>
</tr>
<tr>
<td>import_scExp</td>
<td>65</td>
</tr>
<tr>
<td>index_peaks_barcode_to_matrix_index</td>
<td>66</td>
</tr>
<tr>
<td>inter_correlation_scExp</td>
<td>66</td>
</tr>
</tbody>
</table>
intra_correlation_scExp ................................................. 67
launchApp ................................................................. 68
load_MSIgb ............................................................... 69
merge_MACS2_peaks ....................................................... 69
mm10.chromosomes ....................................................... 70
mm10.cytoBand ............................................................ 70
mm10.GeneTSS ............................................................. 71
normalize_scExp .......................................................... 71
num_cell_after_cor_filt_scExp ......................................... 72
num_cell_after_QC_filt_scExp .......................................... 73
num_cell_before_cor_filt_scExp ........................................ 73
num_cell_in_cluster_scExp .............................................. 74
num_cell_scExp ............................................................ 75
pca_irlba_for_sparseMatrix .............................................. 75
plot_cluster_consensus_scExp .......................................... 76
plot_correlation_PCA_scExp ............................................. 77
plot_coverage_BigWig ..................................................... 78
plot_differential_summary_scExp ....................................... 79
plot_differential_volcano_scExp ....................................... 79
plot_distribution_scExp .................................................. 80
plot_gain_or_loss_barplots ............................................. 81
plot_heatmap_scExp ....................................................... 82
plot_inter_correlation_scExp ......................................... 83
plot_intra_correlation_scExp .......................................... 84
plot_most_contributing_features ....................................... 84
plot_percent_active_feature_scExp .................................... 85
plot_pie_most_contributing_chr ....................................... 86
plot_reduced_dim_scExp ................................................. 87
plot_reduced_dim_scExp_CNA ........................................... 88
plot_top_TF_scExp ....................................................... 89
plot_violin_feature_scExp .............................................. 90
preprocessing_filtering_and_reduction ................................ 91
preprocess_CPM ............................................................ 92
preprocess_feature_size_only .......................................... 93
preprocess_RPKM .......................................................... 93
preprocess_TFIDF .......................................................... 94
preprocess_TPM ............................................................ 95
rawfile_ToBigWig .......................................................... 95
raw_counts_to_sparse_matrix ......................................... 96
read_count_mat_with_separated_chr_start_end ......................... 98
read_sparse_matrix ....................................................... 98
rebin_helper ............................................................... 99
rebin_matrix ............................................................. 100
reduce_dms_scExp ........................................................ 101
reduce_dim_batch_correction .......................................... 102
remove_chr_M_fun ........................................................ 102
remove_non_canonical_fun ............................................. 103
results_enrichmentTest ................................................. 103
**annotation_from_merged_peaks**

*Find nearest peaks of each gene and return refined annotation*

**Description**

Find nearest peaks of each gene and return refined annotation

**Usage**

```r
annotation_from_merged_peaks(scExp, odir, merged_peaks, geneTSS_annotation)
```

**Arguments**

- `scExp` A SingleCellExperiment object
- `odir` An output directory where to write the mergedpeaks BED file
- `merged_peaks` A list of GRanges object containing the merged peaks
- `geneTSS_annotation` A GRanges object with reference genes

**Value**

A data.frame with refined annotation
annotToCol2

Description

annotToCol2

Usage

annotToCol2(
  annotS = NULL,
  annotT = NULL,
  missing = c("", NA),
  anotype = NULL,
  maxnumcateg = 2,
  categCol = NULL,
  quantitCol = NULL,
  plotLegend = TRUE,
  plotLegendFile = NULL
)

Arguments

annotS A color matrix
annotT A color matrix
missing Convert missing to NA
anotype Annotation type
maxnumcateg Maximum number of categories
categCol Categorical columns
quantitCol Quantitative columns
plotLegend Plot legend ?
plotLegendFile Which file to plot legend ?

Value

A matrix of continuous or discrete colors

Examples

data("scExp")
annotToCol2(SingleCellExperiment::colData(scExp), plotLegend = FALSE)
anocol_binary

**Description**

Helper binary column for anocol function

**Usage**

```r
anocol_binary(anocol, anotype, plotLegend, annotS)
```

**Arguments**

- `anocol` The color feature matrix
- `anotype` The feature types
- `plotLegend` Plot legend?
- `annotS` A color matrix

**Value**

A color matrix similar to anocol with binary columns colored

anocol_categorical

**Description**

Helper binary column for anocol function

**Usage**

```r
anocol_categorical(anocol, categCol, anotype, plotLegend, annotS)
```

**Arguments**

- `anocol` The color feature matrix
- `categCol` Colors for categorical features
- `anotype` The feature types
- `plotLegend` Plot legend?
- `annotS` A color matrix

**Value**

A color matrix similar to anocol with binary columns colored
bams_to_matrix_indexes

Count bam files on interval to create count indexes

Description

Count bam files on interval to create count indexes

Usage

bams_to_matrix_indexes(dir, which, BPPARAM = BiocParallel::bpparam())

Arguments

dir       A directory containing single cell BAM files and BAI files
which     Genomic Range on which to count
BPPARAM   BPPARAM object for multiprocessing. See bpparam for more informations.
Will take the default BPPARAM set in your R session.

Value

A list containing a "feature index" data.frame and a count vector for non 0 entries, both used to form the sparse matrix

beds_to_matrix_indexes

Count bed files on interval to create count indexes

Description

Count bed files on interval to create count indexes

Usage

beds_to_matrix_indexes(dir, which, BPPARAM = BiocParallel::bpparam())

Arguments

dir       A directory containing the single cell BED files
which     Genomic Range on which to count
BPPARAM   BPPARAM object for multiprocessing. See bpparam for more informations.
Will take the default BPPARAM set in your R session.

Value

A list containing a "feature index" data.frame and a names of cells as vector both used to form the sparse matrix
**calculate_CNA**

Estimate copy number alterations in cytobands

**Description**

Cytobands are considered large enough in order that a variation at the cytoband level is not considered as an epigenetic event but as a genetic event, e.g. Copy Number Alterations. The function successively:

- Calculates the fraction of reads in each cytoband (FrCyto). See `calculate_cyto_mat`
- Calculates the log2-ratio FrCyto of each cell by the average FrCyto in normal cells. See `calculate_logRatio_CNA`
- Estimates if there was a gain or a loss of copy in each cytoband. See `calculate_gain_or_loss`

The corresponding matrices are accessible in the reducedDim slots "cytoBands", "logRatio_cytoBands" and "gainOrLoss_cytoBands" respectively.

**Usage**

```r
calculate_CNA(
  scExp,
  control_samples = unique(scExp$sample_id)[1],
  ref_genome = c("hg38", "mm10")[1],
  quantiles_to_define_gol = c(0.05, 0.95)
)
```

**Arguments**

- `scExp` : A SingleCellExperiment with "logRatio_cytoBand" reducedDim slot filled. See `calculate_logRatio_CNA`
- `control_samples` : Sample IDs of the normal sample to take as reference.
- `ref_genome` : Reference genome ('hg38' or 'mm10')
- `quantiles_to_define_gol` : Quantiles of normal log2-ratio distribution below/above which cytoband is considered to be a loss/gain. (c(0.05, 0.95)). See `calculate_gain_or_loss`

**Value**

The SCE with the fraction of reads, log2-ratio and gain or loss in each cytobands in each cells (of dimension cell x cytoband) in the reducedDim slots.
calculate_cyto_mat

Calculate Fraction of reads in each cytobands

description

Re-Count binned reads onto cytobands and calculate the fraction of reads in each of the cytoband in each cell. For each cell, the fraction of reads in any given cytoband is calculated. Cytobands are considered large enough in order that a variation at the cytoband level is not considered as an epigenetic event but as a genetic event, e.g. Copy Number Alterations.

Usage

calculate_cyto_mat(scExp, ref_genome = c("hg38", "mm10") [1])

Arguments

scExp A SingleCellExperiment with genomic coordinate as features (peaks or bins)
ref_genome Reference genome ('hg38' or 'mm10')

Value

The SCE with the fraction of reads in each cytobands in each cells (of dimension cell x cytoband ) in the reducedDim slot "cytoBand".

Examples

data("scExp")
scExp = calculate_cyto_mat(scExp, ref_genome="hg38")
SingleCellExperiment::reducedDim(scExp, "cytoBand")
calculate_gain_or_loss

Estimate the copy gains/loss of tumor vs normal based on log2-ratio of fraction of reads

Description

Given a SingleCellExperiment object with the slot "logRatio_cytoBand" containing the log2-ratio of the fraction of reads in each cytoband, estimate if the cytoband was lost or acquired a gain in a non-quantitative way. To do so, the quantiles distribution of the normal cells are calculated, and any cytoband below or above will be considered as a loss/gain. The False Discovery Rate is directly proportional to the quantiles.

Usage

calculate_gain_or_loss(scExp, controls, quantiles = c(0.05, 0.95))

Arguments

- **scExp**: A SingleCellExperiment with "logRatio_cytoBand" reducedDim slot filled. See `calculate_logRatio_CNA`
- **controls**: Sample IDs or Cell IDs of the normal sample to take as reference.
- **quantiles**: Quantiles of normal log2-ratio distribution below/above which cytoband is considered to be a loss/gain. (c(0.05,0.95))

Value

The SCE with the gain or loss in each cytobands in each cells (of dimension cell x cytoband) in the reducedDim slot "gainOrLoss_cytoBand".

Examples

data("scExp")
scExp = calculate_cyto_mat(scExp, ref_genome="hg38")
scExp = calculate_logRatio_CNA(scExp, controls=unique(scExp$sample_id)[1])
scExp = calculate_gain_or_loss(scExp, controls=unique(scExp$sample_id)[1])
SingleCellExperiment::reducedDim(scExp, "gainOrLoss_cytoBand")
### calculate_logRatio_CNA

*Calculate the log2-ratio of tumor vs normal fraction of reads in cytobands*

**Description**

Given a SingleCellExperiment object with the slot "cytoBand" containing the fraction of reads in each cytoband, calculates the log2-ratio of tumor vs normal fraction of reads in cytobands, cell by cell. If the average signal in normal sample in a cytoband is 0, set this value to 1 so that the ratio won't affect the fraction of read value.

**Usage**

```r
calculate_logRatio_CNA(scExp, controls)
```

**Arguments**

- `scExp` A SingleCellExperiment with "cytoBand" reducedDim slot filled.
  - see `calculate_cyto_mat`
- `controls` Sample IDs or Cell IDs of the normal sample to take as reference.

**Value**

The SCE with the log2-ratio of fraction of reads in each cytobands in each cells (of dimension cell x cytoband ) in the reducedDim slot "logRatio_cytoBand".

**Examples**

```r
data("scExp")
scExp = calculate_cyto_mat(scExp, ref_genome="hg38")
scExp = calculate_logRatio_CNA(scExp, controls=unique(scExp$sample_id)[1])
SingleCellExperiment::reducedDim(scExp, "logRatio_cytoBand")
```

---

### call_macs2_merge_peaks

*Calling MACS2 peak caller and merging resulting peaks*

**Description**

Calling MACS2 peak caller and merging resulting peaks
changeRange

Usage

```r
call_macs2_merge_peaks(
    affection,
    odir,
    p.value,
    format = c("scBED", "BAM")[1],
    ref,
    peak_distance_to_merge
)
```

Arguments

- **affectation**: Annotation data.frame with cell cluster and cell id information
- **odir**: Output directory to write MACS2 output
- **p.value**: P value to detect peaks, passed to MACS2
- **format**: File format, either "BAM" or "scBED"
- **ref**: Reference genome to get chromosome information from.
- **peak_distance_to_merge**: Distance to merge peaks

Value

A list of merged GRanges peaks

---

changeRange

Description

changeRange

Usage

```r
changeRange(v, newmin = 1, newmax = 10)
```

Arguments

- **v**: A numeric vector
- **newmin**: New min
- **newmax**: New max

Value

A matrix with values scaled between newmin and newmax
CheA3_TF_nTargets

A data.frame with the number of targets of each TF in ChEA3

Description

This data.frame was obtained by downloading datasets from ChEA3 database (https://maayanlab.cloud/chea3/) and merging targets for:

- ARCHS4_Coexpression
- ENCODE_ChIP-seq
- Enrichr_Queries
- GTEx_Coexpression
- Literature_ChIP-seq
- ReMap_ChIP-seq

Usage

data("CheA3_TF_nTargets")

Format

CheA3_TF_nTargets - a data.frame with 1632 rows (unique TFs) and 2 columns

References


The data.frame is composed of two columns:

- TF column containing the TF gene names (human)
- nTargets_TF containing the number of targets for this TF in the combined database.

Examples

data("CheA3_TF_nTargets")
head(CheA3_TF_nTargets)
**check_correct_datamatrix**

*Check if matrix rownames are well formatted and correct if needed*

**Description**

Throws warnings / error if matrix is in the wrong format

**Usage**

```r
check_correct_datamatrix(datamatrix_single, sample_name = "")
```

**Arguments**

- **datamatrix_single**
  A sparse matrix
- **sample_name**
  Matrix sample name for warnings

**Value**

A sparseMatrix in the right rownames format

**choose_cluster_scExp**

*Choose a number of clusters*

**Description**

This functions takes as input a SingleCellExperiment object and a number of cluster to select. It outputs a SingleCellExperiment object with each cell assigned to a correlation cluster in colData. Also calculates a hierarchical clustering of the consensus associations calculated by ConsensusClusterPlus.

**Usage**

```r
choose_cluster_scExp(
  scExp,
  nclust = 3,
  consensus = FALSE,
  hc_linkage = "ward.D"
)
```

**Arguments**

- **scExp**
  A SingleCellExperiment object containing consclust in metadata.
- **nclust**
  Number of cluster to pick (3)
- **consensus**
  Use consensus clustering results instead of simple hierarchical clustering ? (FALSE)
- **hc_linkage**
  A linkage method for hierarchical clustering. See cor. (’ward.D’)
Value

Returns a SingleCellExperiment object with each cell assigned to a correlation cluster in colData.

Examples

data("scExp")
scExp_cf = correlation_and_hierarchical_clust_scExp(scExp)
scExp_cf = choose_cluster_scExp(scExp_cf,nclust=3,consensus=FALSE)
table(scExp_cf$cell_cluster)

scExp_cf = consensus_clustering_scExp(scExp)
scExp_cf_consensus = choose_cluster_scExp(scExp_cf,nclust=3,consensus=TRUE)
table(scExp_cf_consensus$cell_cluster)

choose_perplexity  
Choose perplexity depending on number of cells for Tsne

Description

Choose perplexity depending on number of cells for Tsne

Usage

choose_perplexity(dataset)

Arguments

dataset  
A matrix of features x cells (rows x columns)

Value

A number between 5 and 30 to use in Rtsne function

col2hex  
Col2Hex

Description

Transform character color to hexadecimal color code.

Usage

col2hex(cname)
colors_scExp

Arguments

cname Color name

Value

The HEX color code of a particular color

Description

Adding colors to cells & features

Usage

colors_scExp(
    scExp,
    annotCol = "sample_id",
    color_by = "sample_id",
    color_df = NULL
)

Arguments

scExp A SingleCellExperiment Object
annotCol Column names to color
color_by If specifying color_df, column names to color
color_df Color data.frame to specify which color for which condition

Value

A SingleCellExperiment with additional "color" columns in colData

Examples

data("scExp")
sExp = colors_scExp(scExp,annotCol = c("sample_id",
    "total_counts"),
    color_by = c("sample_id","total_counts"))

# Specific colors using a manually created data.frame :
color_df = data.frame(sample_id=unique(scExp$sample_id),
    sample_id_color=c("red","blue","green","yellow"))
sExp = colors_scExp(scExp,annotCol="sample_id",
    color_by="sample_id",color_df=color_df)
**combine_datamatrix**

*Combine two matrices and emit warning if no regions are in common*

**Description**

Combine two matrices and emit warning if no regions are in common

**Usage**

```r
combine_datamatrix(datamatrix, datamatrix_single, file_names, i)
```

**Arguments**

- `datamatrix`: A sparse matrix or NULL if empty
- `datamatrix_single`: Another sparse matrix
- `file_names`: File name corresponding to the matrix for warnings
- `i`: file number

**Value**

A combined sparse matrix

---

**combine_enrichmentTests**

*Run enrichment tests and combine into list*

**Description**

Run enrichment tests and combine into list

**Usage**

```r
combine_enrichmentTests(
  diff,
  enrichment_qval,
  qval.th,
  logFC.th,
  min.percent,
  annotFeat_long,
  peak_distance,
  refined_annotation,
  GeneSets,
  GeneSetsDf,
  GenePool, 
  progress = NULL
)
```
comparable_variables

**Arguments**

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>diff</td>
<td>Differential list</td>
</tr>
<tr>
<td>enrichment_qval</td>
<td>Adjusted p-value threshold above which a pathway is considered significative list</td>
</tr>
<tr>
<td>qval.th</td>
<td>Differential analysis adjusted p.value threshold</td>
</tr>
<tr>
<td>logFC.th</td>
<td>Differential analysis log-fold change threshold</td>
</tr>
<tr>
<td>min.percent</td>
<td>Minimum fraction of cells having the feature active to consider it as significantly differential. (0.01)</td>
</tr>
<tr>
<td>annotFeat_long</td>
<td>Long annotation</td>
</tr>
<tr>
<td>peak_distance</td>
<td>Maximum gene to peak distance</td>
</tr>
<tr>
<td>refined_annotation</td>
<td>Refined annotation data.frame if peak calling is done</td>
</tr>
<tr>
<td>GeneSets</td>
<td>List of pathways</td>
</tr>
<tr>
<td>GeneSetsDf</td>
<td>Data.frame of pathways</td>
</tr>
<tr>
<td>GenePool</td>
<td>Pool of possible genes for testing</td>
</tr>
<tr>
<td>progress</td>
<td>A shiny Progress instance to display progress bar.</td>
</tr>
</tbody>
</table>

**Value**

A list of list of pathway enrichment data.frames for Both / Over / Under and for each cluster

---

**comparable_variables**  
*Find comparable variable scExp*

**Description**

Find comparable variable scExp

**Usage**

comparable_variables(scExp, allExp = TRUE)

**Arguments**

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>scExp</td>
<td>A SingleCellExperiment</td>
</tr>
<tr>
<td>allExp</td>
<td>A logical indicating wether alternative experiments comparable variables should also be fetch.</td>
</tr>
</tbody>
</table>

**Value**

A character vector with the comparable variable names
CompareedgeRGLM

*Description*

Creates a summary table with the number of genes under- or overexpressed in each group and outputs several graphical representations

*Usage*

```r
CompareedgeRGLM(
  dataMat = NULL,
  annot = NULL,
  ref_group = NULL,
  groups = NULL,
  featureTab = NULL,
  norm_method = "TMMwsp"
)
```

*Arguments*

- `dataMat` reads matrix
- `annot` selected annotation of interest
- `ref_group` List containing one or more vectors of reference samples. Name of the vectors will be used in the results table. The length of this list should be 1 or the same length as the groups list
- `groups` List containing the IDs of groups to be compared with the reference samples. Names of the vectors will be used in the results table
- `featureTab` Feature annotations to be added to the results table
- `norm_method` Which method to use for normalizing ("upperquantile")

*Value*

A dataframe containing the foldchange and p.value of each feature

*Author(s)*

Eric Letouze & Celine Vallot

*Examples*

```r
data("scExp")
scExp_cf = correlation_and_hierarchical_clust_scExp(scExp)
scExp_cf = choose_cluster_scExp(scExp_cf,nclust=2,consensus=FALSE)
featureTab = as.data.frame(SummarizedExperiment::rowRanges(scExp_cf))
```
rownames(featureTab) = featureTab$ID
ref_group = list("C1"=scExp_cf$cell_id[which(scExp_cf$cell_cluster=="C1")])
groups = list("C2"=scExp_cf$cell_id[which(scExp_cf$cell_cluster=="C2")])
myres = CompareedgeRGLM(as.matrix(SingleCellExperiment::counts(scExp_cf)),
annot=as.data.frame(SingleCellExperiment::colData(scExp_cf)),
ref_group=ref_group,groups=groups, featureTab=featureTab)

---

**Description**

CompareWilcox

**Usage**

```r
CompareWilcox(
  dataMat = NULL,
  annot = NULL,
  ref_group = NULL,
  groups = NULL,
  featureTab = NULL,
  block = NULL,
  BPPARAM = BiocParallel::bpparam()
)
```

**Arguments**

- `dataMat`: A raw count matrix
- `annot`: A cell annotation data.frame
- `ref_group`: List with cells in reference group(s)
- `groups`: List with cells in group(s) to test
- `featureTab`: data.frame with feature annotation
- `block`: Use a blocking factor to conteract batch effect ?
- `BPPARAM`: BPARAM object for multiprocessing. See bpparam for more informations. Will take the default BPARAM set in your R session.

**Value**

A dataframe containing the foldchange and p.value of each feature

**Author(s)**

Eric Letouze & Celine Vallot & Pacome Prompsy
Examples

data("scExp")
scExp_cf = correlation_and_hierarchical_clust_scExp(scExp)
scExp_cf = choose_cluster_scExp(scExp_cf,nclust=2,consensus=FALSE)
featureTab = as.data.frame(SummarizedExperiment::rowRanges(scExp_cf))
rownames(featureTab) = featureTab$ID
ref_group = list("C1"=scExp_cf$cell_id[which(scExp_cf$cell_cluster=="C1")])
groups = list("C2"=scExp_cf$cell_id[which(scExp_cf$cell_cluster=="C2")])
myres = CompareWilcox(as.matrix(SingleCellExperiment::normcounts(scExp_cf)),
annot=as.data.frame(SingleCellExperiment::colData(scExp_cf)),
ref_group=ref_group,groups=groups, featureTab=featureTab)

concatenate_scBed_into_clusters

Concatenate single-cell BED into clusters

Description

Concatenate single-cell BED into clusters

Usage

concatenate_scBed_into_clusters(affectation, files_list, odir)

Arguments

affectation Annotation data.frame containing cluster information
files_list Named list of scBED file paths to concatenate. List Names must match affectation$sample_id and basenames must match affectation$barcode.
odir Output directory to write concatenate pseudo-bulk BEDs.

Value

Merge single-cell BED files into cluster BED files. Ungzip file if BED is gzipped.
**Description**


**Usage**

```r
consensus_clustering_scExp(
  scExp,
  prefix = NULL,
  maxK = 10,
  reps = 100,
  pItem = 0.8,
  pFeature = 1,
  distance = "pearson",
  clusterAlg = "hc",
  innerLinkage = "ward.D",
  finalLinkage = "ward.D",
  plot_consclust = "pdf",
  plot_icl = "png"
)
```

**Arguments**

- `scExp`: A SingleCellExperiment object containing 'PCA' in reducedDims.
- `prefix`: character value for output directory. Directory is created only if plot_consclust is not NULL. This title can be an abosolute or relative path.
- `maxK`: integer value. maximum cluster number to evaluate. (10)
- `reps`: integer value. number of subsamples. (100)
- `pItem`: numerical value. proportion of items to sample. (0.8)
- `pFeature`: numerical value. proportion of features to sample. (1)
- `distance`: character value. 'pearson': (1 - Pearson correlation), 'spearman': (1 - Spearman correlation), 'euclidean', 'binary', 'maximum', 'canberra', 'minkowski' or custom distance function. ('pearson')
- `clusterAlg`: character value. cluster algorithm. 'hc' heirarchical (hclust), 'pam' for partitioning around medoids, 'km' for k-means upon data matrix, 'kmdist' ('hc') for k-means upon distance matrices (former km option), or a function that returns a clustering. ('hc')
**correlation_and_hierarchical_clust_scExp**

innerLinkage  hierarchical linkage method for subsampling. ('ward.D')
finalLinkage  hierarchical linkage method for consensus matrix. ('ward.D')
plot_consclust character value. NULL - print to screen, 'pdf', 'png', 'pngBMP' for bitmap png, helpful for large datasets. ('pdf')
plot_iCL     same as above for item consensus plot. ('png')

**Details**

This function takes as input a SingleCellExperiment object that must have 'PCA' in reducedDims and outputs a SingleCellExperiment object containing consclust list calculated cluster consensus and item consensus scores in metadata.

**Value**

Returns a SingleCellExperiment object containing consclust list, calculated cluster consensus and item consensus scores in metadata.

**References**


**Examples**

data("scExp")
scExp_cf = correlation_and_hierarchical_clust_scExp(scExp)
scExp_cf = consensus_clustering_scExp(scExp)

**Description**

Correlation and hierarchical clustering

Calculates cell to cell correlation matrix based on the PCA feature space and runs hierarchical clustering taking 1 - correlation scores as distance.

**Usage**

correlation_and_hierarchical_clust_scExp(scExp, hc_linkage = "ward.D")

**Arguments**

- `scExp` A SingleCellExperiment object, containing 'PCA' in reducedDims.
- `hc_linkage` A linkage method for hierarchical clustering. See cor. ('ward.D')
**count_coverage**

**Details**

This function takes as input a SingleCellExperiment object that must have PCA calculated and outputs a SingleCellExperiment object with correlation matrix and hierarchical clustering.

**Value**

Return a SingleCellExperiment object with correlation matrix & hierarchical clustering.

**Examples**

```r
data("scExp")
scExp_cf = correlation_and_hierarchical_clust_scExp(scExp)
```

---

**count_coverage**  
*Create a smoothed and normalized coverage track from a BAM file and given a bin GenomicRanges object (same as deepTools bamCoverage)*

**Description**

Normalization is CPM, smoothing is done by averaging on n_smoothBin regions left and right of any given region.

**Usage**

```r
count_coverage(
  input, 
  format = "BAM", 
  bins, 
  canonical_chr, 
  norm_factor, 
  n_smoothBin = 5, 
  ref = "hg38", 
  read_size = 101, 
  original_bins = NULL
)
```

**Arguments**

- `input`: Either a named list of character vector of path towards single-cell BED files or a sparse raw matrix of small bins (≤500bp). If a named list specifying `scBEDn` the names MUST correspond to the 'sample_id' column in your SingleCellExperiment object. The single-cell BED files names MUST match the barcode names in your SingleCellExperiment (column 'barcode'). The scBED files can be gzipped or not.
- `format`: File format, either "BAM" or "BED"
- `bins`: A GenomicRanges object of binned genome
create_project_folder

Create ChromSCape project folder

Description

Creates a project folder that will be recognizable by ChromSCape Shiny application.

Usage

create_project_folder(
  output_directory,
  analysis_name = "Analysis_1",
  ref_genome = c("hg38", "mm10")[1]
)

Arguments

output_directory
  Path towards the directory to create the 'ChromSCape_Analyses' folder and the analysis subfolder. If this path already contains the 'ChromSCape_Analyses' folder, will only create the analysis subfolder.

analysis_name
  Name of the analysis. Must only contain alphanumerical characters or '_'.

ref_genome
  Reference genome, either 'hg38' or 'mm10'.

Value

Creates the project folder and returns the root of the project.

Examples

dir = tempdir()
create_project_folder(output_directory = dir,
  analysis_name = "Analysis_1")
list.dirs(file.path(dir))
create_sample_name_mat

Create a sample name matrix

**Description**

Create a sample name matrix

**Usage**

```r
create_sample_name_mat(nb_samples, samples_names)
```

**Arguments**

- `nb_samples`: Number of samples
- `samples_names`: Character vector of sample names

**Value**

A matrix

create_scDataset_raw

Create a simulated single cell datamatrix & cell annotation

**Description**

Create a simulated single cell datamatrix & cell annotation

**Usage**

```r
create_scDataset_raw(
    cells = 300,
    features = 600,
    featureType = c("window", "peak", "gene"),
    sparse = TRUE,
    nsamp = 4,
    ref = "hg38",
    batch_id = factor(rep(1, nsamp))
)
```
Arguments

- **cells** Number of cells (300)
- **features** Number of features (600)
- **featureType** Type of feature (window)
- **sparse** Is matrix sparse? (TRUE)
- **nsamp** Number of samples (4)
- **ref** Reference genome (‘hg38’)
- **batch_id** Batch origin (factor((1,1,1,1)))

Value

A list composed of:
- *mat*: a sparse matrix following an approximation of the negative binomial law (adapted to scChIPseq)
- *annot*: a data.frame of cell annotation
- *batches*: an integer vector with the batch number for each cell

Examples

```r
# Creating a basic sparse 600 genomic bins x 300 cells matrix and annotation
l = create_scDataset_raw()
head(l$mat)
head(l$annot)
head(l$batches)

# Specifying number of cells, features and samples
l2 = create_scDataset_raw(cells = 500, features = 500, nsamp=2)

# Specifying species
mouse_l = create_scDataset_raw(ref="mm10")

# Specifying batches
batch_l = create_scDataset_raw(nsamp=4, batch_id = factor(c(1,1,2,2)))

# Peaks of different size as features
peak_l = create_scDataset_raw(featureType="peak")
head(peak_l$mat)

# Genes as features
gene_l = create_scDataset_raw(featureType="gene")
head(gene_l$mat)
```

**create_scExp**  
Wrapper to create the single cell experiment from count matrix and feature dataframe
create_scExp

Description

Create the single cell experiment from (sparse) datamatrix and feature dataframe containing feature names and location. Also optionally removes zero count Features, zero count Cells, non canonical chromosomes, and chromosome M. Calculates QC Metrics (scran).

Usage

create_scExp(
  datamatrix,
  annot,
  remove_zero_cells = TRUE,
  remove_zero_features = TRUE,
  remove_non_canonical = TRUE,
  remove_chr_M = TRUE,
  mainExpName = "main",
  verbose = TRUE
)

Arguments

datamatrix A matrix or sparseMatrix of raw counts. Features x Cells (rows x columns).
annot A data.frame containing informations on cells. Should have the same number of rows as the number of columns in datamatrix.
remove_zero_cells remove cells with zero counts ? (TRUE)
remove_zero_features remove cells with zero counts ? (TRUE)
remove_non_canonical remove non canonical chromosomes ?(TRUE)
remove_chr_M remove chromosomes M ? (TRUE)
mainExpName Name of the mainExpName e.g. 'bins', 'peaks'... ("default")
verbose (TRUE)

Value

Returns a SingleCellExperiment object.

Examples

raw <- create_scDataset_raw()
scExp = create_scExp(raw$mat, raw$annot)
scExp
DA_custom

**Differential Analysis Custom in 'One vs One' mode**

**Description**

Differential Analysis Custom in 'One vs One' mode

**Usage**

```r
DA_custom(
  affectation,
  by,
  counts,
  method,
  feature,
  block,
  ref,
  group,
  progress = NULL,
  BPPARAM = BiocParallel::bpparam()
)
```

**Arguments**

- **affectation**: An annotation data.frame with cell_id and
- **by**: A character specifying the column of the object containing the groups of cells to compare.
- **counts**: Count matrix
- **method**: DA method : Wilcoxon or EdgeR
- **feature**: Feature tables
- **block**: Blocking feature
- **ref**: If de_type is custom, the reference to compare (data.frame), must be a one-column data.frame with cell_clusters or sample_id as character in rows
- **group**: If de_type is custom, the group to compare (data.frame), must be a one-column data.frame with cell_clusters or sample_id as character in rows
- **progress**: A shiny Progress instance to display progress bar.
- **BPPARAM**: BPPARAM object for multiprocessing. See bpparam for more informations. Will take the default BPPARAM set in your R session.

**Value**

A list of results, groups compared and references
Differential Analysis in ‘One vs Rest’ mode

### Description

Differential Analysis in ‘One vs Rest’ mode

### Usage

```r
DA_one_vs_rest(
  affectation,
  by,
  counts,
  method,
  feature,
  block,
  progress = NULL,
  BPPARAM = BiocParallel::bpparam()
)
```

### Arguments

- **affectation**: An annotation data.frame with cell_id and cell_cluster columns
- **by**: A character specifying the column of the object containing the groups of cells to compare.
- **counts**: Count matrix
- **method**: DA method: Wilcoxon or EdgeR
- **feature**: Feature tables
- **block**: Blocking feature
- **progress**: A shiny Progress instance to display progress bar.
- **BPPARAM**: BPPARAM object for multiprocessing. See `bpparam` for more informations. Will take the default BPPARAM set in your R session.

### Value

A list of results, groups compared and references
DA_pairwise  

Run differential analysis in Pairwise mode

Description

Run differential analysis in Pairwise mode

Usage

```r
DA_pairwise(
  affectation,
  by,
  counts,
  method,
  feature,
  block,
  progress = NULL,
  BPPARAM = BiocParallel::bpparam()
)
```

Arguments

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>affectation</td>
<td>An annotation data.frame with cell_cluster and cell_id columns</td>
</tr>
<tr>
<td>by</td>
<td>A character specifying the column of the object containing the groups of cells to compare.</td>
</tr>
<tr>
<td>counts</td>
<td>Count matrix</td>
</tr>
<tr>
<td>method</td>
<td>DA method, Wilcoxon or edgeR</td>
</tr>
<tr>
<td>feature</td>
<td>Feature data.frame</td>
</tr>
<tr>
<td>block</td>
<td>Blocking feature</td>
</tr>
<tr>
<td>progress</td>
<td>A shiny Progress instance to display progress bar.</td>
</tr>
<tr>
<td>BPPARAM</td>
<td>BPARAM object for multiprocessing. See bpparam for more informations. Will take the default BPARAM set in your R session.</td>
</tr>
</tbody>
</table>

Value

A list of results, groups compared and references
**define_feature**  
*Define the features on which reads will be counted*

**Description**

Define the features on which reads will be counted

**Usage**

```r
define_feature(ref = c("hg38","mm10")[1],
  peak_file = NULL,
  bin_width = NULL,
  genebody = FALSE,
  extendPromoter = 2500)
```

**Arguments**

- `ref` Reference genome
- `peak_file` A bed file if counting on peaks
- `bin_width` A number of bins if dividing genome into fixed width bins
- `genebody` A logical indicating if feature should be counted in genebodies and promoter.
- `extendPromoter` Extension length before TSS (2500).

**Value**

A GRanges object

**Examples**

```r
gr_bins = define_feature("hg38", bin_width = 50000)
gr_genes = define_feature("hg38", genebody = TRUE, extendPromoter = 5000)
```

**detect_samples**  
*Heuristic discovery of samples based on cell labels*

**Description**

Identify a fixed number of common string (samples) in a set of varying strings (cells). E.g. in the set "Sample1_cell1","Sample1_cell2","Sample2_cell1","Sample2_cell2" and with nb_samples=2, the function returns "Sample1","Sample1","Sample2","Sample2".

**Usage**

```r
detect_samples(barcodes, nb_samples = 1)
```
**differential_activation**

**Arguments**

- **barcodes**
  Vector of cell barcode names (e.g. `Sample1_cell1`, `Sample1_cell2...`)
- **nb_samples**
  Number of samples to find

**Value**

character vector of sample names the same length as cell labels

**Examples**

```r
codes = c(paste0("HBCx22_BC", seq_len(100)),
paste0("mouse_sample_XX", 208:397))
samples = detect_samples(barcodes, nb_samples=2)
```

**Description**

Based on the statement that single-cell epigenomic dataset are very sparse, specifically when analysis small bins or peaks, we can define each feature as being ‘active’ or not simply by the presence or the absence of reads in this feature. This is the equivalent of binarize the data. When trying to find differences in signal for a feature between multiple cell groups, this function simply compare the percentage of cells ‘activating’ the feature in each of the group. The p-values are then calculated using a Pearson’s Chi-squared Test for Count Data (comparing the number of active cells in one group vs the other) and corrected using Benjamini-Hochberg correction for multiple testing.

**Usage**

```r
differential_activation(
  scExp,
  by = c("cell_cluster", "sample_id") [1],
  verbose = TRUE,
  progress = NULL
)
```

**Arguments**

- **scExp**
  A SingleCellExperiment object containing consclust with selected number of cluster.
- **by**
  Which grouping to run the marker enrichment ?
- **verbose**
  Print ?
- **progress**
  A shiny Progress instance to display progress bar.
differential_analysis_scExp

Details

To calculate the logFC, the percentage of activation of the features are corrected for total number of reads to correct for library size bias. For each cluster ('group') the function consider the rest of the cells as the reference.

Value

Returns a dataframe of differential activation results that contains the rowData of the SingleCellExperiment with additional logFC, q.value, group activation (fraction of cells active for each feature in the group cells), reference activation (fraction of cells active for each feature in the reference cells).

See Also

For Pearson’s Chi-squared Test for Count Data chisq.test. For other differential analysis see differential_analysis_scExp.

Examples

data("scExp")
res = differential_activation(scExp, by = "cell_cluster")
res = differential_activation(scExp, by = "sample_id")

differential_analysis_scExp

Runs differential analysis between cell clusters

Description

Based on clusters of cell defined previously, runs non-parametric Wilcoxon Rank Sum test to find significantly depleted or enriched features, in 'one_vs_rest' mode or 'pairwise' mode. In pairwise mode, each cluster is compared to all other cluster individually, and then pairwise comparisons between clusters are combined to find overall differential features using combineMarkers function from scran.

Usage

differential_analysis_scExp(
  scExp,
  de_type = c("one_vs_rest_fast", "one_vs_rest", "pairwise", "custom")[[1]],
  by = "cell_cluster",
  method = "wilcox",
  block = NULL,
  group = NULL,
  ref = NULL,
  prioritize_genes = nrow(scExp) > 20000,
  max_distanceToTSS = 1000,
  progress = NULL,
  BPPARAM = BiocParallel::bpparam()
)
Arguments

scExp  A SingleCellExperiment object containing consclust with selected number of
cluster.

de_type  Type of comparisons. Either 'one_vs_rest', to compare each cluster against all
others, or 'pairwise' to make 1 to 1 comparisons. ('one_vs_rest')

by  = A character specifying the column of the object containing the groups of cells
to compare. Exclusive with de_type == custom

method  Differential testing method, either 'wilcox' for Wilcoxon non-parametric testing
or 'neg.binomial' for edgerGLM based testing. ('wilcox')

block  Use batches as blocking factors? If TRUE, block will be taken as the column
"batch_id" from the SCE. Cells will be compared only within samples belonging
to the same batch.

group  If de_type = "custom", the sample / cluster of interest as a one-column data.frame.
The name of the column is the group name and the values are character either
cluster ("C1", "C2", ...) or sample_id.

ref  If de_type = "custom", the sample / cluster of reference as a one-column data.frame.
The name of the column is the group name and the values are character either
cluster ("C1", "C2", ...) or sample_id.

prioritize_genes  First filter by loci being close to genes? E.g. for differential analysis, it is more
relevant to keep features close to genes

max_distanceToTSS  If prioritize_genes is TRUE, the maximum distance to consider a feature close
to a gene.

progress  A shiny Progress instance to display progress bar.

BPPARAM  BPPARAM object for multiprocessing. See bpparam for more informations.
Will take the default BPPARAM set in your R session.

Details

This functions takes as input a SingleCellExperiment object with consclust, the type of compari-
on, either 'one_vs_rest' or 'pairwise', the adjusted p-value threshold (qval.th) and the fold-change
threshold (logFC.th). It outputs a SingleCellExperiment object containing a differential list.

Value

Returns a SingleCellExperiment object containing a differential list.

Examples

data("scExp")
scExp_cf = differential_analysis_scExp(scExp)
distPearson

Description

distPearson

Usage

distPearson(m)

Arguments

m\nA matrix

Value

A dist object

enrichmentTest

Description

enrichmentTest

Usage

enrichmentTest(gene.sets, mylist, possibleIds, sep = ";", silent = FALSE)

Arguments

gene.sets\nA list of reference gene sets
mylist\nA list of genes to test
possibleIds\nAll existing genes
sep\nSeparator used to collapse genes
silent\nSilent mode ?

Value

A dataframe with the gene sets and their enrichment p.value
enrich_TF_ChEA3_genes  Find the TF that are enriched in the differential genes using ChEA3 API

Description

Find the TF that are enriched in the differential genes using ChEA3 API

Usage

enrich_TF_ChEA3_genes(genes)

Arguments

genes  A character vector with the name of genes to enrich for TF.

Value

Returns a SingleCellExperiment object containing list of enriched Gene Sets for each cluster, either in depleted features, enriched features or simply differential features (both).

References


Examples

data(scExp)
enrich_TF_ChEA3_genes(head(unlist(strsplit(SummarizedExperiment::rowData(scExp)$Gene, split = ",", fixed = TRUE)), 15))

enrich_TF_ChEA3_scExp  Find the TF that are enriched in the differential genes using ChEA3 database

Description

Find the TF that are enriched in the differential genes using ChEA3 database
enrich_TF_ChEA3_scExp

Usage

enrich_TF_ChEA3_scExp(
  scExp,
  ref = "hg38",
  qval.th = 0.01,
  logFC.th = 1,
  min.percent = 0.01,
  peak_distance = 1000,
  use_peaks = FALSE,
  progress = NULL,
  verbose = TRUE
)

Arguments

scExp A SingleCellExperiment object containing list of differential features.
ref A reference annotation, either 'hg38' or 'mm10'. ('hg38')
qval.th Adjusted p-value threshold to define differential features. (0.01)
logFC.th Fold change threshold to define differential features. (1)
min.percent Minimum fraction of cells having the feature active to consider it as significantly
differential. (0.01)
peak_distance Maximum distanceToTSS of feature to gene TSS to consider associated, in bp.
(1000)
use_peaks Use peak calling method (must be calculated beforehand). (FALSE)
progress A shiny Progress instance to display progress bar.
verbose A logical to print message or not. (TRUE)

Value

Returns a SingleCellExperiment object containing list of enriched Gene Sets for each cluster, either
in depleted features, enriched features or simply differential features (both).

Examples

data("scExp")

scExp = enrich_TF_ChEA3_scExp(
  scExp,
  ref = "hg38",
  qval.th = 0.01,
  logFC.th = 1,
  min.percent = 0.01)
exclude_features_scExp

Remove specific features (CNA, repeats)

Description
Remove specific features (CNA, repeats)

Usage

```r
exclude_features_scExp(
  scExp,
  features_to_exclude,
  by = "region",
  verbose = TRUE
)
```

Arguments

- `scExp` A SingleCellExperiment object.
- `features_to_exclude` A GenomicRanges object or data.frame containing genomic regions or features to exclude or path towards a BED file containing the features to exclude.
- `by` Type of features. Either 'region' or 'feature_name'. If 'region', will look for genomic coordinates in columns 1-3 (chr,start,stop). If 'feature_name', will look for a genes in first column. ('region')
- `verbose` (TRUE)

Value

A SingleCellExperiment object without features to exclude.

Examples

```r
raw <- create_scDataset_raw()
scExp = create_scExp(raw$mat, raw$annot)
features_to_exclude = data.frame(chr=c("chr4","chr7","chr17"),
  start=c(500000,8000000,2000000),
  end=c(100000,16000000,25000000))
features_to_exclude = as(features_to_exclude,"GRanges")
scExp = exclude_features_scExp(scExp,features_to_exclude)
scExp
```
**feature_annotation_scExp**

*Add gene annotations to features*

**Description**

Add gene annotations to features

**Usage**

```r
feature_annotation_scExp(scExp, ref = "hg38", reference_annotation = NULL)
```

**Arguments**

- `scExp` A SingleCellExperiment object.
- `ref` Reference genome. Either 'hg38' or 'mm10'. ('hg38')
- `reference_annotation` A data.frame containing gene (or else) annotation with genomic coordinates.

**Value**

A SingleCellExperiment object with annotated rowData.

**Examples**

```r
raw <- create_scDataset_raw()
scExp = create_scExp(raw$mat, raw$annot)
scExp = feature_annotation_scExp(scExp)
head(SummarizedExperiment::rowRanges(scExp))

# Mouse
raw = create_scDataset_raw(ref = "mm10")
scExp = create_scExp(raw$mat, raw$annot)
scExp = feature_annotation_scExp(scExp, ref="mm10")
head(SummarizedExperiment::rowRanges(scExp))
```

---

**filter_correlated_cell_scExp**

*Filter lowly correlated cells*

**Description**

Remove cells that have a correlation score lower than what would be expected by chance with other cells.
Usage

```r
filter_correlated_cell_scExp(scExp, random_iter = 5,
corr_threshold = 99, percent_correlation = 1,
downsample = 2500, verbose = TRUE, n_process = 250,
BPPARAM = BiocParallel::bpparam())
```

Arguments

- `scExp` A SingleCellExperiment object containing 'Cor', a correlation matrix, in reducedDims.
- `random_iter` Number of random matrices to create to calculate random correlation scores. (50)
- `corr_threshold` Quantile of random correlation score above which a cell is considered to be 'correlated' with another cell. (99)
- `percent_correlation` Percentage of the cells that any cell must be 'correlated' to in order to not be filtered. (1)
- `downsample` Number of cells to calculate correlation filtering threshold ? (2500)
- `verbose` Print messages ? (TRUE)
- `n_process` Number of cell to proceed at a time. Increase this number to increase speed at memory cost
- `BPPARAM` BPPARAM object for multiprocessing. See bpparam for more informations. Will take the default BPPARAM set in your R session.

Details

This functions takes as input a SingleCellExperiment object that must have correlation matrix calculated and outputs a SingleCellExperiment object without lowly correlated cells. TSNE is recalculated.

Value

Returns a SingleCellExperiment object without lowly correlated cells. The calculated correlation score limit threshold is saved in metadata.

Examples

```r
data("scExp")
dim(scExp)
scExp_cf = filter_correlated_cell_scExp(scExp,
corr_threshold = 99, percent_correlation = 1)
dim(scExp_cf)
```
filter_genes_with_refined_peak_annotation

*Filter genes based on peak calling refined annotation*

**Description**

Filter genes based on peak calling refined annotation

**Usage**

```r
filter_genes_with_refined_peak_annotation(
  refined_annotation,
  peak_distance,
  signific,
  over,
  under
)
```

**Arguments**

- `refined_annotation` A data.frame containing each gene distance to real peak
- `peak_distance` Minimum distance to an existing peak to accept a given gene
- `signific` Indexes of all significantly differential genes
- `over` Indexes of all significantly overexpressed genes
- `under` Indexes of all significantly underexpressed genes

**Value**

List of significantly differential, overexpressed and underexpressed genes close enough to existing peaks

---

filter_scExp

*Filter cells and features*

**Description**

Function to filter out cells & features from SingleCellExperiment based on total count per cell, number of cells 'ON' in features and top covered cells that might be doublets.
find_clusters_louvain_scExp

Usage

filter_scExp(
    scExp,
    min_cov_cell = 1600,
    quant_removal = 95,
    min_count_per_feature = 10,
    verbose = TRUE
)

Arguments

scExp A SingleCellExperiment object.

min_cov_cell Minimum counts for each cell. (1600)

quant_removal Centile of cell counts above which cells are removed. (95)

min_count_per_feature Minimum number of reads per feature (10).

verbose (TRUE)

Value

Returns a filtered SingleCellExperiment object.

Examples

raw <- create_scDataset_raw()
scExp = create_scExp(raw$mat, raw$annot)
scExp. = filter_scExp(scExp)

# No feature filtering (all features are valuable)
scExp. = filter_scExp(scExp,min_count_per_feature=30)

# No cell filtering (all features are valuable)
scExp. = filter_scExp(scExp,min_cov_cell=0,quant_removal=100)

find_clusters_louvain_scExp

Build SNN graph and find cluster using Louvain Algorithm

Description

Build SNN graph and find cluster using Louvain Algorithm
Usage

```
find_clusters_louvain_scExp(
  scExp,  
  k = 10,  
  resolution = 1,  
  use.dimred = "PCA",  
  type = c("rank", "number", "jaccard")[3],  
  BPPARAM = BiocParallel::bpparam()  
)
```

Arguments

- **scExp**: A SingleCellExperiment with PCA calculated
- **k**: An integer scalar specifying the number of nearest neighbors to consider during graph construction.
- **resolution**: A numeric specifying the resolution of clustering to pass to igraph::cluster_louvain function.
- **use.dimred**: A string specifying the dimensionality reduction to use.
- **type**: A string specifying the type of weighting scheme to use for shared neighbors.
- **BPPARAM**: BPPARAM object for multiprocessing. See bpparam for more informations. Will take the default BPPARAM set in your R session.

Value

A SingleCellExperiment containing the vector of clusters (named C1, C2 ....)

Examples

```
data('scExp')

scExp = find_clusters_louvain_scExp(scExp, k = 10)
```

Description

Find the top most covered features that will be used for dimensionality reduction. Optionally remove non-top features.
Usage

```r
find_top_features(
    scExp,  # A SingleCellExperiment.
    n = 20000,  # Either an integer indicating the number of top covered regions to find or a character vector of the top percentile of features to keep (e.g. 'q20' to keep top 20% features).
    keep_others = FALSE,  # Logical indicating if non-top regions are to be removed from the SCE or not (FALSE).
    prioritize_genes = FALSE,  # First filter by loci being close to genes? E.g. for differential analysis, it is more relevant to keep features close to genes
    max_distanceToTSS = 10000,  # If prioritize_genes is TRUE, the maximum distance to consider a feature close to a gene.
    verbose = TRUE  # Print?
)
```

Arguments

- `n`: Either an integer indicating the number of top covered regions to find or a character vector of the top percentile of features to keep (e.g. 'q20' to keep top 20% features).
- `keep_others`: Logical indicating if non-top regions are to be removed from the SCE or not (FALSE).
- `prioritize_genes`: First filter by loci being close to genes? E.g. for differential analysis, it is more relevant to keep features close to genes
- `max_distanceToTSS`: If prioritize_genes is TRUE, the maximum distance to consider a feature close to a gene.
- `verbose`: Print?

Value

A SCE with top features

Examples

```r
data(scExp)
scExp_top = find_top_features(scExp, n = 4000, keep_others = FALSE)
```

---

**generate_analysis**

*Generate a complete ChromSCape analysis*

Description

Generate a complete ChromSCape analysis
Usage

generate_analysis(input_data_folder,  
analysis_name = "Analysis_1",  
output_directory = "/",  
input_data_type = c("scBED", "DenseMatrix", "SparseMatrix", "scBAM")][1],  
rebin_sparse_matrix = FALSE,  
feature_count_on = c("bins", "genebody", "peaks")][1],  
feature_count_parameter = 50000,  
ref_genome = c("hg38", "mm10")][1],  
run = c("filter", "CNA", "cluster", "consensus", "peak_call", "coverage",  
       "DA", "GSA", "report")][c(1,3,6,7,8,9)],  
min_reads_per_cell = 1000,  
max_quantile_read_per_cell = 99,  
n_top_features = 40000,  
norm_type = "CPM",  
subsample_n = NULL,  
exclude_regions = NULL,  
n_clust = NULL,  
corr_threshold = 99,  
percent_correlation = 1,  
maxK = 10,  
qval.th = 0.1,  
logFC.th = 1,  
enrichment_qval = 0.1,  
doBatchCorr = FALSE,  
batch_sels = NULL,  
control_samples_CNA = NULL,  
genes_to_plot = c("Krt8", "Krt5", "Tgfb1", "Foxq1", "Cdkn2b",  
                   "Cdkn2a", "chr7:15000000-20000000")
)

Arguments

input_data_folder  
Directory containing the input data.

analysis_name  
Name given to the analysis.

output_directory  
Directory where to create the analysis and the HTML report.

input_data_type  
The type of input data.

feature_count_on  
For raw data type, on which features to count the cells.

feature_count_parameter  
Additional parameter corresponding to the ‘feature_count_on’ parameter. E.g. 
for ‘bins’ must be a numeric, e.g. 50000, for ‘peaks’ must be a character containing path towards a BED peak file.
rebin_sparse_matrix
A boolean specifying if the SparseMatrix should be rebinned on features (see feature_count_on and feature_count_parameter).

ref_genome
The genome of reference.

run
What steps to run. By default runs everything. Some steps are required in order to run downstream steps.

min_reads_per_cell
Minimum number of reads per cell.

max_quantile_read_per_cell
Upper quantile above which to consider cells doublets.

n_top_features
Number of features to keep in the analysis.

norm_type
Normalization type.

subsample_n
Number of cells per condition to downsample to, for performance principally.

exclude_regions
Path towards a BED file containing CNA to exclude from the analysis (optional).

n_clust
Number of clusters to force choice of clusters.

corr_threshold
Quantile of correlation above which two cells are considered as correlated.

percent_correlation
Percentage of the total cells that a cell must be correlated with in order to be kept in the analysis.

maxK
Upper cluster number to rest for ConsensusClusterPlus.

qval.th
Adjusted p-value below which to consider features differential.

logFC.th
Log2-fold-change above/below which to consider a feature depleted/enriched.

enrichment_qval
Adjusted p-value below which to consider a gene set as significantly enriched in differential features.

doBatchCorr
Logical indicating if batch correction using fastMNN should be run.

batch_sels
If doBatchCorr is TRUE, a named list containing the samples in each batch.

control_samples_CNA
If running CopyNumber Analysis, a character vector of the sample names that are 'normal'.

genes_to_plot
A character vector containing genes of interest of which to plot the coverage.

Value

Creates a ChromSCape-readable directory and saved objects, as well as a multi-tabbed HTML report resuming the analysis.

Examples

## Not run:
generate_analysis("/path/to/data/", "Analysis_1")

## End(Not run)
generate_count_matrix  Generate count matrix

Description

Generate count matrix

Usage

generate_count_matrix(cells, features, sparse, cell_names, feature_names)

Arguments

cells  Number of cells
features  Number of features
sparse  Is matrix sparse ?
cell_names  Cell names
feature_names  Feature names

Value

A matrix or a sparse matrix

generate_coverage_tracks  Generate cell cluster pseudo-bulk coverage tracks

Description

Generate cell cluster pseudo-bulk coverage tracks. First, scBED files are concatenated into cell clusters contained in the 'by' column of your SingleCellExperiment object. To do so, for each sample in the given list, the barcodes of each cluster are grepped and BED files are merged into pseudo-bulk of clusters (C1,C2,...). Two cells from different can have the same barcode ID as cell affectation is done sample by sample. Then coverage of pseudo-bulk BED files is calculated by averaging & smoothing reads on small genomic window (150bp per default). The pseudo bulk BED and BigWigs coverage tracks are writtend to the output directory. This functionality is not available on Windows as it uses the 'cat' and 'gzip' utilities from Unix OS.
Usage

generate_coverage_tracks(
  scExp_cf,
  input,
  odir,
  format = "scBED",
  ref_genome = c("hg38", "mm10")[[1]],
  bin_width = 150,
  n_smoothBin = 5,
  read_size = 101,
  quantile_for_peak_calling = 0.85,
  by = "cell_cluster",
  progress = NULL
)

Arguments

scExp_cf  
A SingleCellExperiment with cluster selected. (see choose_cluster_scExp). It is recommended having a minimum of ~100 cells per cluster in order to obtain smooth tracks.

input  
Either a named list of character vector of path towards single-cell BED files or a sparse raw matrix of small bins (<500bp). If a named list specifying scBED the names MUST correspond to the 'sample_id' column in your SingleCellExperiment object. The single-cell BED files names MUST match the barcode names in your SingleCellExperiment (column 'barcode'). The scBED files can be gzipped or not.

odir  
The output directory to write the cumulative BED and BigWig files.

format  
File format, either "raw_mat", "BED" or "BAM"

ref_genome  
The genome of reference, used to constrain to canonical chromosomes. Either 'hg38' or 'mm10'. 'hg38' per default.

bin_width  
The width of the bin to create the coverage track. The smaller the greater the resolution & runtime. Default to 150.

n_smoothBin  
Number of bins left & right to average ('smooth') the signal on. Default to 5.

read_size  
The estimated size of reads. Default to 101.

quantile_for_peak_calling  
The quantile to define the threshold above which signal is considered as a peak.

by  
A character specifying a categorical column of scExp_cf metadata by which to group cells and generate coverage tracks and peaks.

progress  
A Progress object for Shiny. Default to NULL.

Value

Generate coverage tracks (.bigwig) for each group in the SingleCellExperiment "by" column.
generate_feature_names

Generate feature names

Description

Generate feature names

Usage

generate_feature_names(featureType, ref, features)

Arguments

featureType Type of feature
ref Reference genome
features Number of features to generate

Value

A character vector of feature names

generate_report From a ChromSCape analysis directory, generate an HTML report.

Description

From a ChromSCape analysis directory, generate an HTML report.
Usage
generate_report(  
  ChromSCape_directory,  
  prefix = NULL,  
  run = c("filter", "CNA", "cluster", "consensus", "peak_call", "coverage", "DA", "GSA", "report")[c(1, 3, 6, 7, 8, 9)],  
  genes_to_plot = c("Krt8", "Krt5", "Tgfb1", "Foxq1", "Cdkn2b", "Cdkn2a", "chr7:15000000-20000000"),  
  control_samples_CNA = NULL
)

Arguments
ChromSCape_directory
  Path towards the ChromSCape directory of which you want to create the report. The report will be created at the root of this directory.

prefix
  Name of the analysis with the filtering parameters (e.g. Analysis_3000_100000_99_uncorrected). You will find the prefix in the Filtering_Normalize_Reduce subfolder.

run
  Which steps to report ("filter", "CNA", "cluster", "consensus", "peak_call", "coverage", "DA", "GSA", "report"). Only indicate steps that were done in the analysis. By default do not report CNA, consensus and peak calling.

genes_to_plot
  For the UMAP, which genes do you want to see in the report.

control_samples_CNA
  If running the Copy Number Alteration (CNA) part, which samples are the controls

Value
  Generate an HTML report at the root of the analysis directory.

Examples
  ## Not run:
  generate_analysis("/path/to/data/", "Analysis_1")
  
  ## End(Not run)

---
gene_set_enrichment_analysis_scExp

*Runs Gene Set Enrichment Analysis on genes associated with differential features*
Description

This function takes previously calculated differential features and runs hypergeometric test to look for enriched gene sets in the genes associated with differential features, for each cell cluster. This function takes as input a SingleCellExperiment object with consclust, the type of comparison, either 'one_vs_rest' or 'pairwise', the adjusted p-value threshold (qval.th) and the fold-change threshold (logFC.th). It outputs a SingleCellExperiment object containing a differential list.

Usage

gene_set_enrichment_analysis_scExp(
  scExp,
  enrichment_qval = 0.1,
  ref = "hg38",
  GeneSets = NULL,
  GeneSetsDf = NULL,
  GenePool = NULL,
  qval.th = 0.01,
  logFC.th = 1,
  min.percent = 0.01,
  peak_distance = 1000,
  use_peaks = FALSE,
  GeneSetClasses = c("c1_positional", "c2_curated", "c3_motif", "c4_computational", "c5_GO", "c6_oncogenic", "c7_immunologic", "hallmark"),
  progress = NULL
)

Arguments

scExp A SingleCellExperiment object containing list of differential features.
enrichment_qval Adjusted p-value threshold for gene set enrichment. (0.1)
ref A reference annotation, either 'hg38' or 'mm10'. ('hg38')
GeneSets A named list of gene sets. If NULL will automatically load MSigDB list of gene sets for specified reference genome. (NULL)
GeneSetsDf A dataframe containing gene sets & class of gene sets. If NULL will automatically load MSigDB dataframe of gene sets for specified reference genome. (NULL)
GenePool The pool of genes to run enrichment in. If NULL will automatically load GenCode list of genes for specified reference genome. (NULL)
qval.th Adjusted p-value threshold to define differential features. (0.01)
logFC.th Fold change threshold to define differential features. (1)
min.percent Minimum fraction of cells having the feature active to consider it as significantly differential. (0.01)
peak_distance Maximum distanceToTSS of feature to gene TSS to consider associated, in bp. (1000)
use_peaks Use peak calling method (must be calculated beforehand). (FALSE)
getExperimentNames

GeneSetClasses  Which classes of MSIGdb to look for.
progress        A shiny Progress instance to display progress bar.

Value

Returns a SingleCellExperiment object containing list of enriched Gene Sets for each cluster, either
in depleted features, enriched features or simply differential features (both).

Examples

data("scExp")

# Usually recommending qval.th = 0.01 & logFC.th = 1 or 2
## Not run: scExp.cf = gene_set_enrichment_analysis_scExp(scExp,
  qval.th = 0.4, logFC.th = 0.3)
## End(Not run)

gene_set_enrichment_analysis(scExp, GeneSetClasses, progress)

getExperimentNames(scExp)

getExperimentNames  Get experiment names from a SingleCellExperiment

Description

Get experiment names from a SingleCellExperiment

Usage

gene_set_enrichment_analysis(scExp)

Arguments

scExp            A SingleCellExperiment with named mainExp and altExps.

Value

Character vector of unique experiment names

Examples

data(scExp)
getExperimentNames(scExp)
getMainExperiment  Get Main experiment of a SingleCellExperiment

Description
Get Main experiment of a SingleCellExperiment

Usage
getMainExperiment(scExp)

Arguments
scExp  A SingleCellExperiment with named mainExp and altExps.

Value
The swapped SingleCellExperiment towards "main" experiment

Examples
data(scExp)
getMainExperiment(scExp)

get_color_dataframe_from_input
Get color dataframe from shiny::colorInput

Description
Get color dataframe from shiny::colorInput

Usage
get_color_dataframe_from_input(
  input,
  levels_selected,
  color_by = c("sample_id", "total_counts"),
  input_id_prefix = "color_"
)

Arguments

input Shiny input object
levels_selected Names of the features
color_by Which feature color to retrieve
input_id_prefix Prefix in front of the feature names

Value

A data.frame with the feature levels and the colors of each level of this feature.

get_cyto_features

Map features onto cytobands

Description

Map the features of a SingleCellExperiment onto the cytobands of a given genome. Some features might not be mapped to any cytobands (e.g. if they are not in the canonical chromosomes), and are removed from the returned object.

Usage

get_cyto_features(scExp, ref_genome = c("hg38", "mm10") [1])

Arguments

scExp A SingleCellExperiment with genomic coordinate as features (peaks or bins)
ref_genome Reference genome ("hg38" or "mm10")

Details

The cytobands are an arbitrary cutting of the genome that dates back to staining metaphase chromosomes with Giemsa.

Value

A data.frame of the SCE features with their corresponding cytoband name

Examples

data("scExp")
matching_cyto = get_cyto_features(scExp, ref_genome="hg38")
get_genomic_coordinates

Get SingleCellExperiment’s genomic coordinates

Description
Get SingleCellExperiment’s genomic coordinates

Usage
get_genomic_coordinates(scExp)

Arguments
scExp A SingleCellExperiment object.

Value
A GRanges object of genomic coordinates.

Examples
raw <- create_scDataset_raw()
scExp = create_scExp(raw$mat, raw$annot)
feature_GRanges = get_genomic_coordinates(scExp)

get_most_variable_cyto

Retrieve the cytobands with the most variable fraction of reads

Description
Given a SingleCellExperiment object with the slot "cytoBand" containing the fraction of reads in each cytoband, calculates the variance of each cytoband and returns a data.frame with the top variables cytobands. Most cytobands are expected to be unchanged between normal and tumor samples, therefore focusing on the top variable cytobands enable to focus on the most interesting regions.

Usage
get_most_variable_cyto(scExp, top = 50)

Arguments
scExp A SingleCellExperiment with "cytoBand" reducedDim slot filled.
top Number of cytobands to return (50).
get_pathway_mat_scExp

Value
A data.frame of the top variable cytoBands and their variance

Examples

```r
data("scExp")
scExp = calculate_cyto_mat(scExp, ref_genome="hg38")
get_most_variable_cyto(scExp, top=50)
```

get_pathway_mat_scExp  Get pathway matrix

Description
Get pathway matrix

Usage

```r
get_pathway_mat_scExp(
  scExp,
  pathways,
  max_distanceToTSS = 1000,
  ref = "hg38",
  GeneSetClasses = c("c1_positional", "c2_curated", "c3_motif", "c4_computational",
                     "c5_GO", "c6_oncogenic", "c7_immunologic", "hallmark"),
  progress = NULL
)
```

Arguments

scExp  A SingleCellExperiment
pathways  A character vector specifying the pathways to retrieve the cell count for.
max_distanceToTSS  Numeric. Maximum distance to a gene’s TSS to consider a region linked to a gene. (1000)
ref  Reference genome, either mm10 or hg38
GeneSetClasses  Which classes of MSIGdb to load
progress  A shiny Progress instance to display progress bar.

Value
A matrix of cell to pathway
gg_fill_hue

Examples

data(scExp)
mat = get_pathway_mat_scExp(scExp, pathways = "KEGG_T_CELL_RECEPTOR_SIGNALING_PATHWAY")

gg_fill_hue  gg_fill_hue

Description

gg_fill_hue

Usage

gg_fill_hue(n)

Arguments

n num hues

Value

A color in HEX format

groupMat  groupMat

Description

groupMat

Usage

groupMat(mat = NA, margin = 1, groups = NA, method = "mean")

Arguments

mat A matrix
margin By row or columns ?
groups Groups
method Method to group

Value

A grouped matrix
H1proportion

Description

H1proportion

Usage

H1proportion(pv = NA, lambda = 0.5)

Arguments

pv P.value vector
lambda Lambda value

Value

H1 proportion value

has_genomic_coordinates

Description

Does SingleCellExperiment has genomic coordinates in features?

Usage

has_genomic_coordinates(scExp)

Arguments

scExp A SingleCellExperiment object

Value

TRUE or FALSE
Examples

```r
raw <- create_scDataset_raw()
scExp = create_scExp(raw$mat, raw$annot)
has_genomic_coordinates(scExp)
raw_genes = create_scDataset_raw(featureType="gene")
scExp_gene = create_scExp(raw_genes$mat, raw_genes$annot)
has_genomic_coordinates(scExp_gene)
```

### Description

hclustAnnotHeatmapPlot

### Usage

```r
hclustAnnotHeatmapPlot(
  x = NULL,
  hc = NULL,
  hmColors = NULL,
  anocol = NULL,
  xpos = c(0.1, 0.9, 0.114, 0.885),
  ypos = c(0.1, 0.5, 0.5, 0.6, 0.62, 0.95),
  dendro.cex = 1,
  xlab.cex = 0.8,
  hmRowNames = FALSE,
  hmRowNames.cex = 0.5
)
```

### Arguments

- **x**: A correlation matrix
- **hc**: An hclust object
- **hmColors**: A color palette
- **anocol**: A matrix of colors
- **xpos**: Xpos
- **ypos**: Ypos
- **dendro.cex**: Size of denro names
- **xlab.cex**: Size of x label
- **hmRowNames**: Write rownames ?
- **hmRowNames.cex**: Size of rownames ?
Value

A heatmap

----

**hg38.chromosomes**

*Data.frame of chromosome length - hg38*

**Description**

This data frame provides the length of each "canonical" chromosomes of Homo Sapiens genome build hg38.

**Usage**

```r
data("hg38.chromosomes")
```

**Format**

- **hg38.chromosomes** - a data frame with 24 rows and 3 variables:
  - **chr**  Chromosome - character
  - **start**  Start of the chromosome (bp) - integer
  - **end**  End of the chromosome (bp) - integer

----

**hg38.cytoBand**

*Data.frame of cytoBand location - hg38*

**Description**

This data frame provides the location of each cytoBands of Homo Sapiens genome build hg38.

**Usage**

```r
data("hg38.cytoBand")
```

**Format**

- **hg38.cytoBand** - a data frame with 862 rows and 4 variables:
  - **chr**  Chromosome - character
  - **start**  Start of the chromosome (bp) - integer
  - **end**  End of the chromosome (bp) - integer
  - **cytoBand**  Name of the cytoBand - character
Description

This dataframe was extracted from Gencode v25 and report the Transcription Start Site of each gene in the Homo Sapiens genome build hg38.

Usage

data("hg38.GeneTSS")

Format

hg38.GeneTSS - a data frame with 24 rows and 3 variables:

chr  Chromosome - character
start Start of the gene (TSS) - integer
end  End of the gene - integer
gene Gene symbol - character

Description

imageCol

Usage

imageCol(
  matcol = NULL,
  strat = NULL,
  xlab.cex = 0.5,
  ylab.cex = 0.5,
  drawLines = c("none", "h", "v", "b")[1],
  ...
)

Arguments

matcol  A matrix of colors
strat   Strat
xlab.cex X label size
ylab.cex Y label size
drawLines Draw lines ?
...     Additional parameters
import_count_input_files

*Import and count input files depending on their format*

**Description**

Import and count input files depending on their format

**Usage**

```r
import_count_input_files(
  files_dir_list,
  file_type,
  which,
  ref,
  verbose,
  progress,
  BPPARAM = BiocParallel::bpparam()
)
```

**Arguments**

- `files_dir_list` A named list of directories containing the input files.
- `file_type` Input file type.
- `which` A GRanges object of features.
- `ref` Reference genome.
- `verbose` Print ?
- `progress` A progress object for Shiny.
- `BPPARAM` BPPARAM object for multiprocessing. See `bpparam` for more informations. Will take the default BPPARAM set in your R session.

**Value**

A list with the feature indexes data.frame containing non-zeroes entries in the count matrix and the cell names
import_scExp

Read single-cell matrix(ces) into scExp

Description

Combine one or multiple matrices together to create a sparse matrix and cell annotation data.frame.

Usage

import_scExp(file_paths, remove_pattern = "", temp_path = NULL)

Arguments

file_paths A character vector of file names towards single cell epigenomic matrices (features x cells) (must be .txt / .tsv)
remove_pattern A string pattern to remove from the sample names. Can be a regexp.
temp_path In case matrices are stored in temporary folder, a character vector of path towards temporary files. (NULL)

Value

A list containing:

- datamatrix: a sparseMatrix of features x cells
- annot_raw: an annotation of cells as data.frame

Examples

mat1 = mat2 = create_scDataset_raw()$mat
tmp1 = tempfile(fileext = ".tsv")
tmp2 = tempfile(fileext = ".tsv")
write.table(as.matrix(mat1),file=tmp1,sep = "\t",
row.names = TRUE,col.names = TRUE,quote = FALSE)
write.table(as.matrix(mat2),file=tmp2, sep = "\t",
row.names = TRUE,col.names = TRUE,quote = FALSE)
file_paths = c(tmp1,tmp2)
out = import_scExp(file_paths)
index_peaks_barcode_to_matrix_indexes

Read index-peaks-barcodes trio files on interval to create count indexes

Description

Read index-peaks-barcodes trio files on interval to create count indexes

Usage

index_peaks_barcode_to_matrix_indexes(
    feature_file,
    matrix_file,
    barcode_file,
    binarize = FALSE
)

Arguments

feature_file A file containing the features genomic locations
matrix_file A file containing the indexes of non-zero values and their value (respectively i,j,x, see sparseMatrix)
barcode_file A file containing the barcode ids
binarize Binarize matrix ?

Value

A list containing a "feature index" data.frame, name_cells, and a region GenomicRange object used to form the sparse matrix

inter_correlation_scExp

Calculate inter correlation between cluster or samples

Description

Calculate inter correlation between cluster or samples
**Usage**

```r
inter_correlation_scExp(
  scExp_cf,  
  by = c("sample_id", "cell_cluster")[[1]],
  reference_group = unique(scExp_cf[[by]])[[1]],
  other_groups = unique(scExp_cf[[by]]),
  fullCor = TRUE
)
```

**Arguments**

- `scExp_cf`: A `SingleCellExperiment`
- `by`: On which feature to calculate correlation ("sample_id" or "cell_cluster")
- `reference_group`: Reference group to calculate correlation with. Must be in accordance with "by".
- `other_groups`: Groups on which to calculate correlation (can contain multiple groups, and also `reference_group`). Must be in accordance with "by".
- `fullCor`: A logical specifying if the correlation matrix was calculated on the entire set of cells (TRUE).

**Value**

A data.frame of average inter-correlation of cells in other_groups with cells in reference_group

**Examples**

```r
data(scExp)
inter_correlation_scExp(scExp)
```

---

**intra_correlation_scExp**

*Calculate intra correlation between cluster or samples*

**Description**

Calculate intra correlation between cluster or samples

**Usage**

```r
intra_correlation_scExp(
  scExp_cf,  
  by = c("sample_id", "cell_cluster")[[1]],
  fullCor = TRUE
)
```
launchApp

Arguments

- **scExp_cf**: A SingleCellExperiment
- **by**: On which feature to calculate correlation ("sample_id" or "cell_cluster")
- **fullCor**: Logical specifying if the correlation matrix was run on the entire number of cells or on a subset.

Value

A data.frame of cell average intra-correlation

Examples

```r
data(scExp)
intra_correlation_scExp(scExp, by = "sample_id")
intra_correlation_scExp(scExp, by = "cell_cluster")```

launchApp

Launch ChromSCape

Description

Main function to launch ChromSCape in your favorite browser. You can pass additional parameters that you would pass to shiny::runApp (runApp)

Usage

```r
launchApp(launch.browser = TRUE, ...)
```

Arguments

- **launch.browser**: Wether to launch browser or not
- **...**: Additional parameters passed to runApp

Value

Launches the shiny application

Examples

```r
## Not run:
launchApp()
## End(Not run)
```
load_MSIGdb

Load and format MSIGdb pathways using msigdb package

Description
Load and format MSIGdb pathways using msigdb package

Usage
load_MSIGdb(
  ref,
  GeneSetClasses = c("c1_positional", "c2_curated", "c3_motif", "c4_computational",
                    "c5_GO", "c6_oncogenic", "c7_immunologic", "hallmark")
)

Arguments
ref Reference genome, either mm10 or hg38
GeneSetClasses Which classes of MSIGdb to load

Value
A list containing the GeneSet (list), GeneSetDf (data.frame) and GenePool character vector of all possible genes

merge_MACS2_peaks
Merge peak files from MACS2 peak caller

Description
Merge peak files from MACS2 peak caller

Usage
merge_MACS2_peaks(peak_file, peak_distance_to_merge, min_peak_size = 200, ref)

Arguments
peak_file A character specifying the path towards the peak file (BED or bedGraph format)
peak_distance_to_merge Maximum distance to merge two peaks
min_peak_size An integer specifying the minimum size of peaks
ref Reference genome

Value
Peaks as GRanges
**Description**

This data frame provides the length of each "canonical" chromosomes of Mus Musculus (Mouse) genome build mm10.

**Usage**

data("mm10.chromosomes")

**Format**

mm10.chromosomes - a data frame with 24 rows and 3 variables:

- **chr**: Chromosome - character
- **start**: Start of the chromosome (bp) - integer
- **end**: End of the chromosome (bp) - integer

---

**Description**

This data frame provides the location of each cytoBands of Homo Sapiens genome build mm10.

**Usage**

data("mm10.cytoBand")

**Format**

mm10.cytoBand - a data frame with 862 rows and 4 variables:

- **chr**: Chromosome - character
- **start**: Start of the chromosome (bp) - integer
- **end**: End of the chromosome (bp) - integer
- **cytoBand**: Name of the cytoBand - character
Description

This dataframe was extracted from Gencode v25 and report the Transcription Start Site of each gene in the Mus Musculus genome build mm10 (Mouse).

Usage

data("mm10.GeneTSS")

Format

mm10.GeneTSS - a data frame with 24 rows and 3 variables:

  chr  Chromosome name - character
  start  Start of the gene (TSS) - integer
  end  End of the gene - integer
  gene  Gene symbol - character

normalize_scExp

Description

Normalize counts

Usage

normalize_scExp(
    scExp,
    type = c("CPM", "TFIDF", "RPKM", "TPM", "feature_size_only")
  )

Arguments

  scExp  A SingleCellExperiment object.
  type  Which normalization to apply. Either 'CPM', 'TFIDF', 'RPKM', 'TPM' or 'feature_size_only'. Note that for all normalization by size (RPKM, TPM, feature_size_only), the features must have defined genomic coordinates.

Value

A SingleCellExperiment object containing normalized counts. (See ?normcounts())
Examples

raw <- create_scDataset_raw()
scExp = create_scExp(raw$mat, raw$annot)
scExp = normalize_scExp(scExp)
head(SingleCellExperiment::normcounts(scExp))

---

num_cell_after_cor_filt_scExp

*Number of cells before & after correlation filtering*

Description

Number of cells before & after correlation filtering

Usage

```
num_cell_after_cor_filt_scExp(scExp, scExp_cf)
```

Arguments

- **scExp**: SingleCellExperiment object before correlation filtering.
- **scExp_cf**: SingleCellExperiment object after correlation filtering.

Value

A colored kable with the number of cells per sample before and after filtering for display

Examples

```
data("scExp")
scExp_cf = correlation_and_hierarchical_clust_scExp(scExp)
scExp_cf = filter_correlated_cell_scExp(scExp Cf,
corr_threshold = 99, percent_correlation = 1)
## Not run: num_cell_after_cor_filt_scExp(scExp,scExp Cf)
```
**num_cell_after_QC_filt_scExp**

Table of cells before / after QC

### Description

Table of cells before / after QC

### Usage

num_cell_after_QC_filt_scExp(scExp, annot, datamatrix)

### Arguments

- **scExp**: A SingleCellExperiment object.
- **annot**: A raw annotation data.frame of cells before filtering.
- **datamatrix**: A matrix of cells per regions before filtering.

### Value

A formatted kable in HTML.

### Examples

```r
raw <- create_scDataset_raw()
scExp = create_scExp(raw$mat, raw$annot)
scExp_filtered = filter_scExp(scExp)
## Not run: num_cell_after_QC_filt_scExp(
  scExp_filtered,SingleCellExperiment::colData(scExp))
## End(Not run)
```

---

**num_cell_before_cor_filt_scExp**

Table of number of cells before correlation filtering

### Description

Table of number of cells before correlation filtering

### Usage

num_cell_before_cor_filt_scExp(scExp)

### Examples

```r
raw <- create_scDataset_raw()
scExp = create_scExp(raw$mat, raw$annot)
scExp_filtered = filter_scExp(scExp)
## Not run: num_cell_after_QC_filt_scExp(
  scExp_filtered,SingleCellExperiment::colData(scExp))
## End(Not run)
```
num_cell_in_cluster_scExp

Arguments

scExp  A SingleCellExperiment Object

Value

A colored kable with the number of cells per sample for display

Examples

data("scExp")
## Not run: num_cell_before_cor_filt_scExp(scExp)

data("scExp")
scExp_cf = correlation_and_hierarchical_clust_scExp(scExp)
scExp_cf = choose_cluster_scExp(scExp_cf,nclust=3,consensus=FALSE)
## Not run: num_cell_in_cluster_scExp(scExp_cf)
num_cell_scExp

Table of cells

Description

Table of cells

Usage

num_cell_scExp(annot, datamatrix)

Arguments

annot An annotation of cells. Can be obtain through 'colData(scExp)'.
datamatrix A matrix of cells per regions before filtering.

Value

A formatted kable in HTML.

Examples

raw <- create_scDataset_raw()
scExp = create_scExp(raw$mat, raw$annot)
## Not run: num_cell_scExp(SingleCellExperiment::colData(scExp))

pca_irlba_for_sparseMatrix

Run sparse PCA using irlba SVD

Description

This function allows to run a PCA using IRLBA Singular Value Decomposition in a fast & memory efficient way. The incremental Lanczos bidagonalisation algorithm allows to keep the matrix sparse as the "loci" centering is implicit. The function then multiplies by the approximate singular values (svd$d) in order to get more importance to the first PCs proportionnally to their singular values. This step is crucial for downstream approaches, e.g. UMAP or T-SNE.

Usage

pca_irlba_for_sparseMatrix(x, n_comp, work = 3 * n_comp)
Arguments

- **x**
  - A sparse normalized matrix (features x cells)

- **n_comp**
  - The number of principal components to keep

- **work**
  - Working subspace dimension, larger values can speed convergence at the cost of more memory use.

Value

The rotated data, e.g. the cells x PC column in case of sc data.

Examples

data("scExp")

plot_cluster_consensus_scExp(scExp)
Description

Plotting correlation of PCs with a variable of interest

Usage

```
plot_correlation_PCA_scExp(
  scExp,
  correlation_var = "total_counts",
  color_by = NULL,
  topPC = 10
)
```

Arguments

- `scExp`: A SingleCellExperiment Object
- `correlation_var`: A string specifying with which numeric variable from colData of scExp to calculate and plot the correlation of each PC with. ('total_counts')
- `color_by`: A string specifying with which categorical variable to color the plot. ('NULL')
- `topPC`: An integer specifying the number of PCs to plot correlation with (10)

Value

A ggplot histogram representing the distribution of count per cell

Examples

```
data("scExp")
plot_correlation_PCA_scExp(scExp, topPC = 25)
plot_correlation_PCA_scExp(scExp, color_by = "cell_cluster")
plot_correlation_PCA_scExp(scExp, color_by = "sample_id")
```
plot_coverage_BigWig  Coverage plot

Description

Coverage plot

Usage

plot_coverage_BigWig(
  coverages,
  label_color_list,
  peaks = NULL,
  chrom,
  start,
  end,
  ref = "hg38"
)

Arguments

coverages  A list containing sample coverage as GenomicRanges
label_color_list  List of colors, list names are labels
peaks  A GRanges object containing peaks location to plot (optional)
chrom  Chromosome
start  Start
end  End
ref  Genomic Reference

Value

A coverage plot annotated with genes

Examples

data(scExp)
plot_differential_summary_scExp

Differential summary barplot

Description

Differential summary barplot

Usage

plot_differential_summary_scExp(
  scExp_cf,
  qval.th = 0.01,
  logFC.th = 1,
  min.percent = 0.01
)

Arguments

scExp_cf  A SingleCellExperiment object
qval.th   Adjusted p-value threshold. (0.01)
logFC.th  Fold change threshold. (1)
min.percent Minimum fraction of cells having the feature active to consider it as significantly differential. (0.01)

Value

A barplot summary of differential analysis

Examples

data("scExp")
plot_differential_summary_scExp(scExp)

plot_differential_volcano_scExp

Volcano plot of differential features

Description

Volcano plot of differential features
Usage

plot_differential_volcano_scExp(
    scExp_cf,
    group = "C1",
    logFC.th = 1,
    qval.th = 0.01,
    min.percent = 0.01
)

Arguments

scExp_cf A SingleCellExperiment object

Arguments

group A character indicating the group for which to plot the differential volcano plot. ("C1")

logFC.th Fold change threshold. (1)

qval.th Adjusted p-value threshold. (0.01)

min.percent Minimum fraction of cells having the feature active to consider it as significantly differential. (0.01)

Value

A volcano plot of differential analysis of a specific cluster

Examples

data("scExp")
plot_differential_volcano_scExp(scExp,"C1")

plot_distribution_scExp

Plotting distribution of signal

Description

Plotting distribution of signal

Usage

plot_distribution_scExp(
    scExp,
    raw = TRUE,
    log10 = FALSE,
    pseudo_counts = 1,
    bins = 150
)
**plot_gain_or_loss_barplots**

Arguments

- **scExp**: A SingleCellExperiment Object
- **raw**: Use raw counts?
- **log10**: Transform using log10?
- **pseudo_counts**: Pseudo-count to add if using log10
- **bins**: Number of bins in the histogram

**Value**

A ggplot histogram representing the distribution of count per cell

**Examples**

```r
data("scExp")
plot_distribution_scExp(scExp)
```

**Description**

Plot Gain or Loss of cytobands of the most variables cytobands

**Usage**

```r
plot_gain_or_loss_barplots(scExp, cells = NULL, top = 20)
```

Arguments

- **scExp**: A SingleCellExperiment with "logRatio_cytoBand" reducedDim slot filled. See `calculate_logRatio_CNA`
- **cells**: Cell IDs of the tumor samples to
- **top**: Number of most variables cytobands to plot

**Value**

Plot the gains/lost in the selected cells of interest as multiple barplots

Plot the gains/lost in the selected cells of interest as multiple barplots
Examples

```r
data("scExp")
scExp = calculate_CNA(scExp, control_samples = unique(scExp$sample_id)[1],
ref_genome="hg38", quantiles_to_define_gol = c(0.05,0.95))
plot_gain_or_loss_barplots(scExp, cells = scExp$cell_id[which(
scExp$sample_id %in% unique(scExp$sample_id)[2])])
```

```r
data("scExp")
scExp = calculate_CNA(scExp, control_samples = unique(scExp$sample_id)[1],
ref_genome="hg38", quantiles_to_define_gol = c(0.05,0.95))
plot_gain_or_loss_barplots(scExp, cells = scExp$cell_id[which(
scExp$sample_id %in% unique(scExp$sample_id)[2])])
```

---

**plot_heatmap_scExp**  
*Plot cell correlation heatmap with annotations*

Description

Plot cell correlation heatmap with annotations

Usage

```r
plot_heatmap_scExp(
  scExp,
  name_hc = "hc_cor",
  corColors = (grDevices::colorRampPalette(c("royalblue", "white", "indianred1")))(256),
  color_by = NULL,
  downsample = 1000,
  hc_linkage = "ward.D"
)
```

Arguments

- `scExp` A SingleCellExperiment Object
- `name_hc` Name of the hclust contained in the SingleCellExperiment object
- `corColors` A palette of colors for the heatmap
- `color_by` Which features to add as additional bands on top of plot
- `downsample` Number of cells to downsample
- `hc_linkage` A linkage method for hierarchical clustering. See `cor` ('ward.D')

Value

A heatmap of cell to cell correlation, grouping cells by hierarchical clustering.
Examples

```r
data("scExp")
plot_heatmap_scExp(scExp)
```

---

**Description**

Violin plot of inter-correlation distribution between one or multiple groups and one reference group

**Usage**

```r
plot_inter_correlation_scExp(
  scExp_cf,
  by = c("sample_id", "cell_cluster")[1],
  jitter_by = NULL,
  reference_group = unique(scExp_cf[[by]])[1],
  other_groups = unique(scExp_cf[[by]]),
  downsample = 5000
)
```

**Arguments**

- `scExp_cf` A SingleCellExperiment
- `by` Color by sample_id or cell_cluster
- `jitter_by` Add jitter points of another layer (cell_cluster or sample_id)
- `reference_group` Character containing the reference group name to calculate correlation from.
- `other_groups` Character vector of the other groups for which to calculate correlation with the reference group.
- `downsample` Downsampling for plotting

**Value**

A violin plot of inter-correlation

**Examples**

```r
data(scExp)
plot_intra_correlation_scExp(scExp)
```
plot_intra_correlation_scExp

Violin plot of intra-correlation distribution

Description

Violin plot of intra-correlation distribution

Usage

plot_intra_correlation_scExp(
    scExp_cf, 
    by = c("sample_id", "cell_cluster")[1], 
    jitter_by = NULL, 
    downsample = 5000
)

Arguments

scExp_cf A SingleCellExperiment
by Color by sample_id or cell_cluster
jitter_by Add jitter points of another layer (cell_cluster or sample_id)
downsample Downsample for plotting

Value

A violin plot of intra-correlation

Examples

data(scExp)
plot_intra_correlation_scExp(scExp)

plot_most_contributing_features

Plot Top/Bottom most contributing features to PCA

Description

Plot Top/Bottom most contributing features to PCA
Usage

plot_most_contributing_features(
    scExp,
    component = "Component_1",
    n_top_bot = 10
)

Arguments

scExp       A SingleCellExperiment containing "PCA" in reducedDims and gene annotation in rowRanges
component   The name of the component of interest
n_top_bot   An integer number of top and bot regions to plot

Details

If a gene TSS is within 10,000bp of the region, the name of the gene(s) will be displayed instead of the region

Value

A barplot of top and bottom features with the largest absolute value in the component of interest

Examples

data(scExp)
plot_most_contributing_features(scExp, component = "Component_1")

plot_percent_active_feature_scExp

Barplot of the % of active cells for a given features

Description

Barplot of the % of active cells for a given features

Usage

plot_percent_active_feature_scExp(
    scExp,
    gene,
    by = c("cell_cluster", "sample_id")[1],
    highlight = NULL,
    downsample = 5000,
    max_distanceToTSS = 1000
)
Arguments

scExp: A SingleCellExperiment

gene: A character specifying the gene to plot

by: Color violin by cell_cluster or sample_id ("cell_cluster")

highlight: A specific group to highlight in a one vs all fashion

downsampling: Downsampling for plotting (5000)

max_distanceToTSS: Numeric. Maximum distance to a gene’s TSS to consider a region linked to a gene. (1000)

Value

A violin plot of intra-correlation

Examples

data(scExp)
plot_percent_active_feature_scExp(scExp, "UBXN10")

Description

Pie chart of top contribution of chromosomes in the 100 most contributing features to PCA #'

Usage

plot_pie_most_contributing_chr(
    scExp,
    component = "Component_1",
    n_top_bot = 100
)

Arguments

scExp: A SingleCellExperiment containing "PCA" in reducedDims and gene annotation in rowRanges

component: The name of the component of interest
	n_top_bot: An integer number of top and bot regions to plot (100)
**Value**

A pie chart showing the distribution of chromosomes in the top features with the largest absolute value in the component of interest.

**Examples**

```r
data(scExp)
plot_pie_most_contributing_chr(scExp, component = "Component_1")
```

---

**plot_reduced_dim_scExp**

*Plot reduced dimensions (PCA, TSNE, UMAP)*

**Description**

Plot reduced dimensions (PCA, TSNE, UMAP)

**Usage**

```r
plot_reduced_dim_scExp(
  scExp,
  color_by = "sample_id",
  reduced_dim = c("PCA", "TSNE", "UMAP"),
  select_x = NULL,
  select_y = NULL,
  downsample = 5000,
  transparency = 0.6,
  size = 1,
  max_distanceToTSS = 1000,
  annotate_clusters = "cell_cluster" %in% colnames(colData(scExp)),
  min_quantile = 0.01,
  max_quantile = 0.99
)
```

**Arguments**

- `scExp` A SingleCellExperiment Object
- `color_by` Character of feature used for coloration. Can be cell metadata ("total_counts", "sample_id", ...) or a gene name.
- `reduced_dim` Reduced Dimension used for plotting
- `select_x` Which variable to select for x axis
- `select_y` Which variable to select for y axis
- `downsample` Number of cells to downsample
- `transparency` Alpha parameter, between 0 and 1
- `size` Size of the points.
max_distanceToTSS
The maximum distance to TSS to consider a gene linked to a region. Used only if "color_by" is a gene name.

annotate_clusters
A logical indicating if clusters should be labelled. The 'cell_cluster' column should be present in metadata.

min_quantile
The lower threshold to remove outlier cells, as quantile of cell embeddings (between 0 and 0.5).

max_quantile
The upper threshold to remove outlier cells, as quantile of cell embeddings (between 0.5 and 1).

Value
A ggplot geom_point plot of reduced dimension 2D representation

Examples

data("scExp")
plot_reduced_dim_scExp(scExp, color_by = "sample_id")
plot_reduced_dim_scExp(scExp, color_by = "total_counts")
plot_reduced_dim_scExp(scExp, reduced_dim = "UMAP")
plot_reduced_dim_scExp(scExp, color_by = "CD52", reduced_dim = "UMAP")

plot_reduced_dim_scExp_CNA
Plot UMAP colored by Gain or Loss of cytobands

Description
Plot UMAP colored by Gain or Loss of cytobands

Usage
plot_reduced_dim_scExp_CNA(scExp, cytoBand)

Arguments
scExp A SingleCellExperiment with "gainOrLoss_cytoBand" reducedDim slot filled. See calculate_gain_or_loss

Value
Which cytoBand to color cells by

Value
Plot the gains/lost of the cytoband overlayed on the epigenetic UMAP.
plot_top_TF_scExp

Examples

data("scExp")
scExp = calculate_CNA(scExp, control_samples = unique(scExp$sample_id)[1],
                     ref_genome="hg38", quantiles_to_define_gol = c(0.05,0.95))
plot_reduced_dim_scExp_CNA(scExp, get_most_variable_cyto(scExp)$cytoBand[1])

plot_top_TF_scExp

Barplot of top TFs from ChEA3 TF enrichment analysis

Description

Barplot of top TFs from ChEA3 TF enrichment analysis

Usage

plot_top_TF_scExp(
  scExp,
  group = unique(scExp$cell_cluster)[1],
  set = c("Differential", "Enriched", "Depleted")[1],
  type = c("Score", "nTargets", "nTargets_over_TF", "nTargets_over_genes")[1],
  n_top = 25
)

Arguments

scExp A SingleCellExperiment

group A character string specifying the differential group to display the top TFs

set A character string specifying the set of genes in which the TF were enriched, either 'Differential', 'Enriched' or 'Depleted'.

type A character string specifying the Y axis of the plot, either the number of differential targets or the ChEA3 integrated mean score. E.g. either "Score", "nTargets", "nTargets_over_TF" for the number of target genes over the total number of genes targeted by the TF or "nTargets_over_genes" for the number of target genes over the number of genes in the gene set.

n_top An integer specifying the number of top TF to display

Value

A bar plot of top TFs from ChEA3 TF enrichment analysis
Examples

```r
data("scExp")

plot_top_TF_scExp(
  scExp,
  group = "C1",
  set = "Differential",
  type = "Score",
  n_top = 10)

plot_top_TF_scExp(
  scExp,
  group = "C1",
  set = "Enriched",
  type = "nTargets_over_genes",
  n_top = 20)
```

Description

Violin plot of features

Usage

```r
plot_violin_feature_scExp(
  scExp,
  gene,
  by = c("cell_cluster", "sample_id")[1],
  downsample = 5000,
  max_distanceToTSS = 1000
)
```

Arguments

- `scExp`: A SingleCellExperiment
- `gene`: A character specifying the gene to plot
- `by`: Color violin by cell_cluster or sample_id ("cell_cluster")
- `downsample`: Downsample for plotting (5000)
- `max_distanceToTSS`: Numeric. Maximum distance to a gene’s TSS to consider a region linked to a gene. (1000)
A violin plot of intra-correlation

Examples

```r
data(scExp)
plot_violin_feature_scExp(scExp, "UBXN10")
```

## Description
Preprocess and filter matrix annotation data project folder to SCE

## Usage

```r
preprocessing_filtering_and_reduction(
  datamatrix, annot_raw, 
  min_reads_per_cell = 1600, 
  max_quantile_read_per_cell = 95, 
  n_top_features = 40000, 
  norm_type = "CPM", 
  n_dims = 10, 
  remove_PC = NULL, 
  subsample_n = NULL, 
  ref_genome = "hg38", 
  exclude_regions = NULL, 
  doBatchCorr = FALSE, 
  batch_sels = NULL
)
```

## Arguments

- **datamatrix**: A sparse count matrix of features x cells.
- **annot_raw**: A data.frame with barcode, cell_id, sample_id, batch_id, total_counts
- **min_reads_per_cell**: Minimum read per cell to keep the cell
- **max_quantile_read_per_cell**: Upper count quantile threshold above which cells are removed
- **n_top_features**: Number of features to keep
- **norm_type**: Normalization type c("CPM", "TFIDF", "RPKM", "TPM", "feature_size_only")
n_dims  An integer specifying the number of dimensions to keep for PCA
remove_PC  A vector of string indicating which principal components to remove before downstream analysis as probably correlated to library size. Should be under the form: 'Component_1', 'Component_2', ... Recommended when using 'TFIDF' normalization method. (NULL)
subsample_n  Number of cells to subsample.
ref Genome  Reference genome ("hg38" or "mm10").
exclude_regions  GenomicRanges with regions to remove from the object.
doBatchCorr  Run batch correction ? TRUE or FALSE
batch_sels  If doBatchCorr is TRUE, List of characters. Names are batch names, characters are sample names.

Value
A SingleCellExperiment object containing feature spaces.

Examples
raw <- create_scDataset_raw()
scExp = preprocessing_filtering_and_reduction(raw$mat, raw$annot)

preprocess_CPM  Preprocess scExp - Counts Per Million (CPM)

Description
Preprocess scExp - Counts Per Million (CPM)

Usage
preprocess_CPM(scExp)

Arguments
scExp  A SingleCellExperiment Object

Value
A SingleCellExperiment object.

Examples
raw <- create_scDataset_raw()
scExp = create_scExp(raw$mat, raw$annot)
scExp = preprocess_CPM(scExp)
head(SingleCellExperiment::normcounts(scExp))
### preprocess_feature_size_only

**Preprocess scExp - size only**

**Description**

Preprocess scExp - size only

**Usage**

preprocess_feature_size_only(scExp)

**Arguments**

scExp  
A SingleCellExperiment Object

**Value**

A SingleCellExperiment object.

**Examples**

```r
raw <- create_scDataset_raw()
scExp = create_scExp(raw$mat, raw$annot)
scExp = preprocess_feature_size_only(scExp)
head(SingleCellExperiment::normcounts(scExp))
```

### preprocess_RPKM

**Preprocess scExp - Read per Kilobase Per Million (RPKM)**

**Description**

Preprocess scExp - Read per Kilobase Per Million (RPKM)

**Usage**

preprocess_RPKM(scExp)

**Arguments**

scExp  
A SingleCellExperiment Object

**Value**

A SingleCellExperiment object.
preprocess_TFIDF

Examples

raw <- create_scDataset_raw()
scExp = create_scExp(raw$mat, raw$annot)
scExp = preprocess_RPKM(scExp)
head(SingleCellExperiment::normcounts(scExp))

---

preprocess_TFIDF  
**Preprocess scExp - TF-IDF**

Description

Preprocess scExp - TF-IDF

Usage

preprocess_TFIDF(scExp, scale = 10000, log = TRUE)

Arguments

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>scExp</td>
<td>A SingleCellExperiment Object</td>
</tr>
<tr>
<td>scale</td>
<td>A numeric to multiply the matrix in order to have human readable numbers. Has no impact on the downstream analysis</td>
</tr>
<tr>
<td>log</td>
<td>Wether to use neperian log on the TF-IDF normalized data or not.</td>
</tr>
</tbody>
</table>

Value

A SingleCellExperiment object.

Examples

raw <- create_scDataset_raw()
scExp = create_scExp(raw$mat, raw$annot)
scExp = preprocess_TFIDF(scExp)
head(SingleCellExperiment::normcounts(scExp))
### preprocess_TPM

**Preprocess scExp - Transcripts per Million (TPM)**

**Description**
Preprocess scExp - Transcripts per Million (TPM)

**Usage**

```r
preprocess_TPM(scExp)
```

**Arguments**

- `scExp` A SingleCellExperiment Object

**Value**
A SingleCellExperiment object.

**Examples**

```r
cat <- create_scDataset_raw()
scExp = create_scExp(cat$mat, cat$annot)
scExp = preprocess_TPM(scExp)
head(SingleCellExperiment::normcounts(scExp))
```

---

### rawfile_ToBigWig

**rawfile_ToBigWig : reads in BAM file and write out BigWig coverage file, normalized and smoothed**

**Description**

rawfile_ToBigWig : reads in BAM file and write out BigWig coverage file, normalized and smoothed

**Usage**

```r
rawfile_ToBigWig(input, 
    BigWig_filename, 
    format = "BAM", 
    bin_width = 150, 
    norm_factor, 
    n_smoothBin = 5, 
    ref = "hg38", 
    read_size = 101,
    
```
raw_counts_to_sparse_matrix

Arguments

input
Either a named list of character vector of path towards single-cell BED files or a sparse raw matrix of small bins (<=500bp). If a named list specifying scBEDn the names MUST correspond to the 'sample_id' column in your SingleCellExperiment object. The single-cell BED files names MUST match the barcode names in your SingleCellExperiment (column 'barcode'). The scBED files can be gzipped or not.

BigWig_filename
Path to write the output BigWig file

format
File format, either "BAM" or "BED"

bin_width
Bin size for coverage

norm_factor
Then number of cells or total number of reads in the given sample, for normalization.

n_smoothBin
Number of bins for smoothing values

ref
Reference genome.

read_size
Length of the reads.

original_bins
Original bins GenomicRanges in case the format is raw matrix.

quantile_for_peak_calling
The quantile to define the threshold above which signal is considered as a peak.

Value

Writes in the output directory a bigwig file displaying the cumulative coverage of cells and a basic set of peaks called by taking all peaks above a given threshold

Writes a BigWig file as output

Description

This function takes three different type of single-cell input: - Single cell BAM files (sorted) - Single cell BED files (gzipped) - A combination of an index file, a peak file and cell barcode file (The index file is composed of three column: index i, index j and value x for the non zeroes entries in the sparse matrix.)
raw_counts_to_sparse_matrix

Usage

raw_counts_to_sparse_matrix(
  files_dir_list,
  file_type = c("scBED", "scBAM", "FragmentFile"),
  use_Signac = TRUE,
  peak_file = NULL,
  n_bins = NULL,
  bin_width = NULL,
  genebody = NULL,
  extendPromoter = 2500,
  verbose = TRUE,
  ref = c("hg38", "mm10")[1],
  progress = NULL,
  BPPARAM = BiocParallel::bpparam()
)

Arguments

files_dir_list  A named character vector of directories containing the files. The names correspond to sample names.

file_type     Input file(s) type(s) (’scBED’,’scBAM’,’FragmentFile’)

use_Signac   Use Signac wrapper function ’FeatureMatrix’ if the Signac package is installed (TRUE).

peak_file    A file containing genomic location of peaks (NULL)

n_bins       The number of bins to tile the genome (NULL)

bin_width    The size of bins to tile the genome (NULL)

genebody     Count on genes (body + promoter) ? (NULL)

extendPromoter If counting on genes, number of base pairs to extend up or downstream of TSS (2500).

verbose      Verbose (TRUE)

ref           reference genome to use (hg38)

progress     Progress object for Shiny

BPPARAM      BPPARAM object for multiprocessing. See bpparam for more informations. Will take the default BPPARAM set in your R session.

Details

This functions re-counts signal on either fixed genomic bins, a set of user-defined peaks or around the TSS of genes.

Value

A sparse matrix of features x cells
References

Stuart el al., Multimodal single-cell chromatin analysis with Signac bioRxiv https://doi.org/10.1101/2020.11.09.373613

read_count_mat_with_separated_chr_start_end

Read a count matrix with three first columns (chr,start,end)

Description

Read a count matrix with three first columns (chr,start,end)

Usage

read_count_mat_with_separated_chr_start_end(path_to_matrix, format_test, separator)

Arguments

path_to_matrix Path to the count matrix
format_test Sample of the read.table
separator Separator character

Value

A sparseMatrix with rownames in the form "chr1:1222-55555"

read_sparse_matrix

Read in one or multiple sparse matrices (10X format)

Description

Given one or multiple directories, look in each directory for a combination of the following files:

- A 'features' file containing unique feature genomic locations - in tab separated format ( *_features.bed / .txt / .tsv / .gz), e.g. chr, start and end
- A 'barcodes' file containing unique barcode names ( _barcode.txt / .tsv / .gz)
- A 'matrix' A file containing indexes of non zero entries (_matrix.mtx / .gz)

Usage

read_sparse_matrix(files_dir_list, ref = c("hg38", "mm10")[1], verbose = TRUE)
**Arguments**

- `files_dir_list` A named character vector containing the full path towards folders. Each folder should contain only the Feature file, the Barcode file and the Matrix file (see description).
- `ref` Reference genome (used to filter non-canonical chromosomes).
- `verbose` Print?

**Value**

Returns a list containing a datamatrix and cell annotation

**Examples**

```r
## Not run:
sample_dirs = c("/path/to/folder1/", "/path/to/folder2/")
names(sample_dirs) = c("sample_1", "sample_2")
out <- read_sparse_matrix(sample_dirs, ref = "hg38")
head(out$datamatrix)
head(out$annot_raw)
## End(Not run)
```

---

**rebin_helper**

Rebin Helper for `rebin_matrix` function

**Description**

Rebin Helper for `rebin_matrix` function

**Usage**

`rebin_helper(mat_df)`

**Arguments**

- `mat_df` A data.frame corresponding to sparse matrix indexes & values.

**Value**

A data.frame grouped mean-summarised by col and new_row
rebin_matrix

Transforms a bins x cells count matrix into a larger bins x cells count matrix.

Description

This function is best used to re-count large number of small bins or peaks (e.g. <= 5000bp) into equal or larger sized bins. The genome is either cut in fixed bins (e.g. 50,000bp) or into an user defined number of bins. Bins are calculated based on the canonical chromosomes. Note that if peaks are larger than bins, or if peaks are overlapping multiple bins, the signal is added to each bin. Users can increase the minimum overlap to consider peaks overlapping bins (by default 150bp, size of a nucleosome) to diminish the number of peaks overlapping multiple regions. Any peak smaller than the minimum overlap threshold will be dismissed. Therefore, library size might be slightly different from peaks to bins if signal was duplicated into multiple bins or omitted due to peaks smaller than minimum overlap.

Usage

rebin_matrix(
  mat,
  bin_width = 50000,
  custom_annotation = NULL,
  minoverlap = 500,
  verbose = TRUE,
  ref = "hg38",
  nthreads = 1,
  rebin_function = rebin_helper
)

Arguments

mat
A matrix of peaks x cells

bin_width
Width of bins to produce in base pairs (minimum 500) (50000)

custom_annotation
A GenomicRanges object specifying the new features to count the matrix on instead of recounting on genomic bins. If not NULL, takes precedence over bin_width.

minoverlap
Minimum overlap between the original bins and the new features to consider the peak as overlapping the bin. We recommend to put this number at exactly half of the original bin size (e.g. 500bp for original bin size of 1000bp) so that no original bins are counted twice. (500)

verbose
Verbose

ref
Reference genome to use (hg38)

nthreads
Number of threads to use for parallel processing
**reduce_dims_scExp**

**Value**

A sparse matrix of larger bins or peaks.

**Examples**

```r
mat = create_scDataset_raw()$mat
binned_mat = rebin_matrix(mat, bin_width = 10e6)
dim(binned_mat)
```

---

**reduce_dims_scExp**  
*Reduce dimensions (PCA, TSNE, UMAP)*

**Description**

Reduce dimensions (PCA, TSNE, UMAP)

**Usage**

```r
reduce_dims_scExp(
  scExp,
  dimension_reductions = c("PCA", "UMAP"),
  n = 10,
  batch_correction = FALSE,
  batch_list = NULL,
  remove_PC = NULL,
  verbose = TRUE
)
```

**Arguments**

- **scExp**  
  A SingleCellExperiment object.

- **dimension_reductions**  
  A character vector of methods to apply. (c("PCA", "UMAP"))

- **n**  
  Numbers of dimensions to keep for PCA. (50)

- **batch_correction**  
  Do batch correction ? (FALSE)

- **batch_list**  
  List of characters. Names are batch names, characters are sample names.

- **remove_PC**  
  A vector of string indicating which principal components to remove before downstream analysis as probably correlated to library size. Should be under the form: 'Component_1', 'Component_2', ... Recommended when using 'TFIDF' normalization method. (NULL)

- **verbose**  
  Print messages ?(TRUE)

**Value**

A SingleCellExperiment object containing feature spaces. See ?reduceDims().
Examples

```r
raw <- create_scDataset_raw()
scExp = create_scExp(raw$mat, raw$annot)
scExp = normalize_scExp(scExp, "CPM")
scExp = reduce_dims_scExp(scExp, dimension_reductions=c("PCA", "UMAP"))
```

**reduce_dim_batch_correction**

*Reduce dimension with batch corrections*

**Description**

Reduce dimension with batch corrections

**Usage**

```r
reduce_dim_batch_correction(scExp, mat, batch_list, n)
```

**Arguments**

- **scExp**: SingleCellExperiment
- **mat**: The normalized count matrix
- **batch_list**: List of batches
- **n**: Number of PCs to keep

**Value**

A list containing the SingleCellExperiment with batch info and the corrected pca

**remove_chr_M_fun**

*Remove chromosome M from scExprownames*

**Description**

Remove chromosome M from scExprownames

**Usage**

```r
remove_chr_M_fun(scExp, verbose)
```

**Arguments**

- **scExp**: A SingleCellExperiment
- **verbose**: Print?
**remove_non_canonical_fun**

---

**Value**

A SingleCellExperiment without chromosome M (mitochondrial chr)

---

**Description**

Remove non canonical chromosomes from scExp

**Usage**

```r
remove_non_canonical_fun(scExp, verbose)
```

**Arguments**

- `scExp`: A SingleCellExperiment
- `verbose`: Print ?

**Value**

A SingleCellExperiment without non canonical chromosomes (random, unknown, contigs etc...)

---

**results_enrichmentTest**

---

**Description**

Run hypergeometric enrichment test and combine significant pathways into a data.frame

**Usage**

```r
results_enrichmentTest(  
  differentialGenes,  
  enrichment_qval,  
  GeneSets,  
  GeneSetsDf,  
  GenePool
)
```
Arguments

differentialGenes  
Genes significantly over / under expressed

enrichment_qval  
Adjusted p-value threshold above which a pathway is considered significative

GeneSets  
List of pathways

GeneSetsDf  
Data.frame of pathways

GenePool  
Pool of possible genes for testing

Value

A data.frame with pathways passing q.value threshold

Description

Retrieve Top and Bot most contributing features of PCA

Usage

retrieve_top_bot_features_pca(  
pca,  
counts,  
component,  
n_top_bot,  
absolute = FALSE  
)

Arguments

pca  
A matrix/data.frame of rotated data

counts  
the normalized counts used for PCA

component  
the component of interest

n_top_bot  
the number of top & bot features to take

absolute  
If TRUE, return the top features in absolute values instead.

Value

a data.frame of top bot contributing features in PCA
run_pairwise_tests

Description

Run pairwise tests

Usage

run_pairwise_tests(
  affection,  
  by, 
  counts, 
  feature, 
  method, 
  progress = NULL, 
  BPPARAM = BiocParallel::bpparam()
)

Arguments

affectation An annotation data.frame with cell_cluster and cell_id columns
by = A character specifying the column of the object containing the groups of cells
to compare.
counts Count matrix
feature Feature data.frame
method DA method, Wilcoxon or edgeR
progress A shiny Progress instance to display progress bar.
BPPARAM BPPARAM object for multiprocessing. See bpparam for more informations. 
Will take the default BPPARAM set in your R session.

Value

A list containing objects for DA function

run_tsne_scExp

Description

Run tsne on single cell experiment
Usage

run_tsne_scExp(scExp, verbose = FALSE)

Arguments

scExp A SingleCellExperiment Object  
verbose Print ?

Value

A colored kable with the number of cells per sample for display

Description

Data from a single-cell ChIP-seq experiment against H3K4me3 active mark from two cell lines, Jurkat B cells and Ramos T cells from Grosselin et al., 2019. The count matrices, on 5kbp bins, were given to ChromSCape and the filtering parameter was set to 3% of cells active in regions and subsampled down to 150 cells per sample. After correlation filtering, the experiment is composed of respectively 51 and 55 cells from Jurkat & Ramos and 5499 5kbp-genomic bins where signal is located.

Usage

data("scExp")

Format

scExp - a SingleCellExperiment with 106 cells and 5499 features (genomic bins) in hg38:

chr A SingleCellExperiment

Details

The scExp is composed of:

- counts and normcounts assays, PCA, UMAP, and Correlation matrix in reducedDims(scExp)
- Assignation of genes to genomic bins in rowRanges(scExp)
- Cluster information in colData(scExp) correlation
- Hierarchical clustering dendogram in metadata$hc_cor
- Consensus clustering raw data in metadata$consclust
- Consensus clustering cluster-consensus and item consensus dataframes in metadata$icl
- Differential analysis in metadata$diff
- Gene Set Analysis in metadata$enr
```
Examples

data("scExp")
plot_reduced_dim_scExp(scExp)
plot_reduced_dim_scExp(scExp,color_by = "cell_cluster")
plot_heatmap_scExp(scExp)
plot_differential_volcano_scExp(scExp, "C1")
plot_differential_summary_scExp(scExp)
```

```
separate_BAM_into_clusters

Separate BAM files into cell cluster BAM files

Description

Separate BAM files into cell cluster BAM files

Usage

separate_BAM_into_clusters(affectation, odir, merged_bam)

Arguments

- affectation: An annotation data.frame containing cell_id and cell_cluster columns
- odir: A valid output directory path
- merged_bam: A list of merged bam file paths

Value

Create one BAM per cluster from one BAM per condition

```
separator_count_mat

Determine Count matrix separator ("tab" or ",")

Description

Determine Count matrix separator ("tab" or ",")

Usage

separator_count_mat(path_to_matrix)

Arguments

- path_to_matrix: A path towards the count matrix to check
```
smoothBin

Smooth a vector of values with nb_bins left and right values

Usage

smoothBin(bin_score, nb_bins = 10)

Arguments

bin_score: A numeric vector of values to be smoothed
nb_bins: Number of values to take left and right

Value

A smooth vector of the same size

subsample_scExp

Subsample scExp

Description

Randomly sample x cells from each sample in a SingleCellExperiment to return a subsampled
SingleCellExperiment with all samples having maximum n cells. If n is higher than the number of
cell in a sample, this sample will not be subsampled.

Usage

subsample_scExp(scExp, n_cell_per_sample = 500, n_cell_total = NULL)

Arguments

scExp: A SingleCellExperiment
n_cell_per_sample: An integer number of cells to subsample for each sample. Exclusive with n_cells_total.
(500)
n_cell_total: An integer number of cells to subsample in total. Exclusive with n_cell_per_sample
(NULL).
subset_bam_call_peaks

Value
A subsampled SingleCellExperiment

Examples

```r
raw <- create_scDataset_raw()
scExp = create_scExp(raw$mat, raw$annot)
scExp_sub = subsample_scExp(scExp,50)
## Not run: num_cell_scExp(scExp_sub)
```

subset_bam_call_peaks  Peak calling on cell clusters

Description
This function does peak calling on each cell population in order to refine gene annotation for large bins. For instance, a 50000bp bins might contain the TSS of several genes, while in reality only one or two of these genes are overlapping the signal (peak). To do so, first in-silico cell sorting is applied based on previously defined clusters contained in the SingleCellExperiment. Taking BAM files of each sample as input, samtools pools then splits reads from each cell barcode into 1 BAM file per cell cluster (pseudo-bulk). Then MACS2 calls peaks on each cluster. The peaks are aggregated and merged if closer to a certain distance defined by user (10000bp). Then, this function takes as input a SingleCellExperiment, that must contain a 'cell_cluster' column in it's colData, an output directory where to store temporary files, the list of BAM files corresponding to each sample and containing the cell barcode information as a tag (for instance tag CB:Z:xxx, XB:Z:xxx or else...) or single-cell BED files containing the raw reads and corresponding to the 'barcode' column metadata, the p.value used by MACS2 to distinguish significant peaks, the reference genome (either hg38 or mm10), the maximal merging distance in bp and a data.frame containing gene TSS genomic coordinates of corresponding genome (if set to NULL, will automatically load geneTSS). The output is a SingleCellExperiment with GRanges object containing ranges of each merged peaks that falls within genomic bins of the SingleCellExperiment, saving the bin range as additional column (window_chr, window_start, window_end), as well as the closest genes and their distance relative to the peak. The peaks may be present in several rows if multiple genes are close / overlap to the peaks.

Note that the user must have MACS2 installed and available in the PATH. Users can open command terminal and type 'which macs2' to verify the availability of these programs. Will only work on unix operating system. Check operating system with 'print(.Platform)'.

Usage

```r
subset_bam_call_peaks(
  scExp,
  odir,
  input,
  format = "BAM",
)```
subset_bam_call_peaks

p.value = 0.05,
ref = "hg38",
peak_distance_to_merge = 10000,
geneTSS_annotation = NULL,
run_coverage = FALSE,
progress = NULL
)

Arguments

scExp: A SingleCellExperiment object
odir: Output directory where to write temporary files and each cluster’s BAM file
input: A character vector of file paths to each sample’s BAM file, containing cell barcode information as tags. BAM files can be paired-end or single-end.
format: Format of the input data, either "BAM" or "scBED".
p.value: a p-value to use for MACS2 to determine significant peaks. (0.05)
ref: A reference genome, either hg38 or mm10. ('hg38')
peak_distance_to_merge: Maximal distance to merge peaks together after peak calling, in bp. (10000)
geneTSS_annotation: A data.frame annotation of genes TSS. If NULL will automatically load GeneCode list of genes fro specified reference genome.
run_coverage: Create coverage tracks (.bw) for each cluster ?
progress: A shiny Progress instance to display progress bar.

Details

The BED files of the peaks called for each clusters, as well as the merged peaks are written in the output directory.

Value

A SingleCellExperiment with refined annotation

Examples

## Not run:
data("scExp")
subset_bam_call_peaks(scExp, "path/to/out/", list("sample1" = "path/to/BAM/sample1.bam", "sample2" = "path/to/BAM/sample2.bam"),
p.value = 0.05, ref = "hg38", peak_distance_to_merge = 10000,
geneTSS_annotation = NULL)

## End(Not run)
**summary_DA**

*Summary of the differential analysis*

**Description**

Summary of the differential analysis

**Usage**

```r
summary_DA(scExp, qval.th = 0.01, logFC.th = 1, min.percent = 0.01)
```

**Arguments**

- `scExp`: A SingleCellExperiment object containing consclust with selected number of cluster.
- `qval.th`: Adjusted p-value threshold. (0.01)
- `logFC.th`: Fold change threshold. (1)
- `min.percent`: Minimum fraction of cells having the feature active to consider it as significantly differential. (0.01)

**Value**

A table summary of the differential analysis

**Examples**

```r
data('scExp')
summary_DA(scExp)
```

---

**swapAltExp_sameColData**

*Swap main & alternative Experiments, with fixed colData*

**Description**

Swap main & alternative Experiments, with fixed colData

**Usage**

```r
swapAltExp_sameColData(scExp, alt)
```

**Arguments**

- `scExp`: A SingleCellExperiment
- `alt`: Name of the alternative experiment
**table_enriched_genes_scExp**

*Creates table of enriched genes sets*

**Description**

Creates table of enriched genes sets

**Usage**

```r
table_enriched_genes_scExp(
  scExp, 
  set = "Both", 
  group = "C1", 
  enr_class_sel = c("c1_positional", "c2_curated", "c3_motif", "c4_computational", "c5_GO", "c6_oncogenic", "c7_immunologic", "hallmark")
)
```

**Arguments**

- `scExp` A SingleCellExperiment object containing list of enriched gene sets.
- `set` A character vector, either 'Both', 'Overexpressed' or 'Underexpressed'. ('Both')
- `group` The "group" name from differential analysis. Can be the cluster name or the custom name in case of a custom differential analysis.
- `enr_class_sel` Which classes of gene sets to show. (c('c1_positional', 'c2_curated', ...))

**Value**

A DT::data.table of enriched gene sets.

**Examples**

```r
data("scExp")
## Not run: table_enriched_genes_scExp(scExp)
```
Description

Warning for differential_analysis_scExp

Usage

warning_DA(scExp, by, de_type, method, block, group, ref)

Arguments

- **scExp**: A SingleCellExperiment object containing consclust with selected number of cluster.
- **by**: A character specifying the column of the object containing the groups of cells to compare. Exclusive with de_type == custom
- **de_type**: Type of comparisons. Either 'one_vs_rest', to compare each cluster against all others, or 'pairwise' to make 1 to 1 comparisons. ('one_vs_rest')
- **method**: Wilcoxon or edgerGLM
- **block**: Use batches as blocking factors?
- **group**: If de_type is custom, the group to compare (data.frame), must be a one-column data.frame with cell_clusters or sample_id as character in rows
- **ref**: If de_type is custom, the reference to compare (data.frame), must be a one-column data.frame with cell_clusters or sample_id as character in rows

Value

Warnings or Errors if the input are not correct
Usage

```r
warning_filter_correlated_cell_scExp(
  scExp,
  random_iter,
  corr_threshold,
  percent_correlation,
  run_tsne,
  downsample,
  verbose
)
```

Arguments

- **scExp**: A SingleCellExperiment object containing 'Cor', a correlation matrix, in reducedDims.
- **random_iter**: Number of random matrices to create to calculate random correlation scores. (50)
- **corr_threshold**: Quantile of random correlation score above which a cell is considered to be 'correlated' with another cell. (99)
- **percent_correlation**: Percentage of the cells that any cell must be 'correlated' to in order to not be filtered. (1)
- **run_tsne**: Re-run tsne? (FALSE)
- **downsample**: Number of cells to calculate correlation filtering threshold? (2500)
- **verbose**: (TRUE)

Value

Warnings or Errors if the input are not correct

Description

A warning helper for plot_reduced_dim_scExp

Usage

```r
warning_plot_reduced_dim_scExp(
  scExp,
  color_by,
  reduced_dim,
  downsample,
  ...)
Arguments

- **scExp**: A SingleCellExperiment Object
- **color_by**: Feature used for coloration
- **reduced_dim**: Reduced Dimension used for plotting
- **downsample**: Number of cells to downsample
- **transparency**: Alpha parameter, between 0 and 1
- **size**: Size of the points.
- **max_distanceToTSS**: Numeric. Maximum distance to a gene’s TSS to consider a region linked to a gene.
- **annotate_clusters**: A logical indicating if clusters should be labelled. The 'cell_cluster' column should be present in metadata.
- **min_quantile**: The lower threshold to remove outlier cells, as quantile of cell embeddings (between 0 and 0.5).
- **max_quantile**: The upper threshold to remove outlier cells, as quantile of cell embeddings (between 0.5 and 1).

Value

Warning or errors if the inputs are not correct

Description

Warning for raw_counts_to_sparse_matrix
Usage

```r
warning_raw_counts_to_sparse_matrix(
  files_dir_list,  # A named character vector of directory containing the raw files
  file_type = c("scBAM", "scBED", "SparseMatrix"),  # Input file(s) type(s) ('scBED', 'scBAM', 'SparseMatrix')
  peak_file = NULL,  # A file containing genomic location of peaks (NULL)
  n_bins = NULL,  # The number of bins to tile the genome (NULL)
  bin_width = NULL,  # The size of bins to tile the genome (NULL)
  genebody = NULL,  # Count on genes (body + promoter) ? (NULL)
  extendPromoter = 2500,  # If counting on genes, number of base pairs to extend up or downstream of TSS (2500).
  verbose = TRUE,  # Verbose (TRUE)
  ref = "hg38"  # reference genome to use (hg38)
)
```

Arguments

- **files_dir_list**: A named character vector of directory containing the raw files
- **file_type**: Input file(s) type(s) ('scBED', 'scBAM', 'SparseMatrix')
- **peak_file**: A file containing genomic location of peaks (NULL)
- **n_bins**: The number of bins to tile the genome (NULL)
- **bin_width**: The size of bins to tile the genome (NULL)
- **genebody**: Count on genes (body + promoter) ? (NULL)
- **extendPromoter**: If counting on genes, number of base pairs to extend up or downstream of TSS (2500).
- **verbose**: Verbose (TRUE)
- **ref**: reference genome to use (hg38)

Value

Error or warnings if the input are not correct

---

**wrapper_Signac_FeatureMatrix**

*Wrapper around ‘FeatureMatrix’ function from Signac Package*

---

**Description**

Wrapper around ‘FeatureMatrix’ function from Signac Package
wrapper_Signac_FeatureMatrix

Usage

```r
wrapper_Signac_FeatureMatrix(
  files_dir_list,
  which,
  ref = "hg38",
  process_n = 2000,
  set_future_plan = TRUE,
  verbose = TRUE,
  progress = NULL
)
```

Arguments

- `files_dir_list`: A named character vector of directories containing the files. The names correspond to sample names.
- `which`: A GenomicRanges containing the features to count on.
- `ref`: Reference genome to use (hg38). Chromosomes that are not present in the canonical chromosomes of the given reference genome will be excluded from the matrix.
- `process_n`: Number of regions to load into memory at a time, per thread. Processing more regions at once can be faster but uses more memory. (2000)
- `set_future_plan`: Set `multisession` plan within the function (TRUE). If TRUE, the previous plan (e.g. `future::plan()`) will be set back on exit.
- `verbose`: Verbose (TRUE).
- `progress`: Progress object for Shiny.

Details

Signac & future are not required packages for ChromSCape as they are required only for the fragment matrix calculations. To use this function, install Signac package first (future will be installed as a dependency). For the simplicity of the application & optimization, the function by default sets `future::plan("multisession")` with `workers = future::availableCores(omit = 1)` in order to allow parallel processing with Signac. On exit the plan is re-set to the previously set future plan. Note that future multisession may have trouble running when VPN is on. To run in parallel, first deactivate your VPN if you encounter long runtimes.

Value

A sparse matrix of features x cells

References

Stuart el al., Multimodal single-cell chromatin analysis with Signac bioRxiv https://doi.org/10.1101/2020.11.09.373613
Examples

## Not run:
gr_bins = define_feature("hg38", bin_width = 50000)
wrapper_Signac_FeatureMatrix("/path/to/dir_containing_fragment_files",
gr_bins, ref = "hg38")

## End(Not run)
Index

* datasets
  CheA3_TF_nTargets, 14
  hg38.chromosomes, 62
  hg38.cytoBand, 62
  hg38.GeneTSS, 63
  mm10.chromosomes, 70
  mm10.cytoBand, 70
  mm10.GeneTSS, 71
  scExp, 106

  annotation_from_merged_peaks, 5
  annotToCol2, 6
  anocol_binary, 7
  anocol_categorical, 7

  bams_to_matrix_indexes, 8
  beds_to_matrix_indexes, 8
  bpparam, 8, 21, 30–32, 36, 42, 45, 64, 97, 105

  calculate_CNA, 9
  calculate_cyto_mat, 9, 10, 12
  calculate_gain_or_loss, 9, 11, 88
  calculate_logRatio_CNA, 9, 11, 12, 81
  call_macs2_merge_peaks, 12
  changeRange, 13
  CheA3_TF_nTargets, 14
  check_correct_datamatrix, 15
  chisq.test, 35
  choose_cluster_scExp, 15, 50
  choose_perplexity, 16
  col2hex, 16
  colors_scExp, 17
  combine_datamatrix, 18
  combine_enrichmentTests, 18
  comparable_variables, 19
  CompareedgeRGLM, 20
  CompareWilcox, 21
  concatenate_scBed_into_clusters, 22
  consensus_clustering_scExp, 23
  ConsensusClusterPlus, 23

  cor, 15, 24, 82
  correlation_and_hierarchical_clust_scExp, 24
  count_coverage, 25
  create_project_folder, 26
  create_sample_name_mat, 27
  create_scDataset_raw, 27
  create_scExp, 28

  DA_custom, 30
  DA_one_vs_rest, 31
  DA_pairwise, 32
  define_feature, 33
  detect_samples, 33
  differential_activation, 34
  differential_analysis_scExp, 35, 35
  distPearson, 37

  enrich_TF_ChEA3_genes, 38
  enrich_TF_ChEA3_scExp, 38
  enrichmentTest, 37
  exclude_features_scExp, 40

  feature_annotation_scExp, 41
  filter_correlated_cell_scExp, 41
  filter_genes_with_refined_peak_annotation, 43
  filter_scExp, 43
  find_clusters_louvain_scExp, 44
  find_top_features, 45

  gene_set_enrichment_analysis_scExp, 52
  generate_analysis, 46
  generate_count_matrix, 49
  generate_coverage_tracks, 49
  generate_feature_names, 51
  generate_report, 51
  get_color_dataframe_from_input, 55
  get_cyto_features, 56
  get_genomic_coordinates, 57
get_most_variable_cyto, 57
get_pathway_mat_scExp, 58
getExperimentNames, 54
getMainExperiment, 55
gg_fill_hue, 59
groupMat, 59
H1proportion, 60
has_genomic_coordinates, 60
hclustAnnotHeatmapPlot, 61
hg38.chromosomes, 62
hg38.cytoBand, 62
hg38.GeneTSS, 63
imageCol, 63
import_count_input_files, 64
import_scExp, 65
index_peaks_barcodes_to_matrix_indexes, 66
inter_correlation_scExp, 66
intra_correlation_scExp, 67
launchApp, 68
load_MSIGdb, 69
merge_MACS2_peaks, 69
mm10.chromosomes, 70
mm10.cytoBand, 70
mm10.GeneTSS, 71
normalize_scExp, 71
num_cell_after_cor_filt_scExp, 72
num_cell_after_QC_filt_scExp, 73
num_cell_before_cor_filt_scExp, 73
num_cell_in_cluster_scExp, 74
num_cell_scExp, 75
pca_irlba_for_sparseMatrix, 75
plot_cluster_consensus_scExp, 76
plot_correlation_PCA_scExp, 77
plot_coverage_BigWig, 78
plot_differential_summary_scExp, 79
plot_differential_volcano_scExp, 79
plot_distribution_scExp, 80
plot_gain_or_loss_barplots, 81
plot_heatmap_scExp, 82
plot_inter_correlation_scExp, 83
plot_intra_correlation_scExp, 84
plot_most_contributing_features, 84
plot_percent_active_feature_scExp, 85
plot_pie_most_contributing_chr, 86
plot_reduced_dim_scExp, 87
plot_reduced_dim_scExp_CNA, 88
plot_top_TF_scExp, 89
plot_violin_feature_scExp, 90
preprocess_CPM, 92
preprocess_feature_size_only, 93
preprocess_RPKM, 93
preprocess_TFIDF, 94
preprocess_TPM, 95
preprocessing_filtering_and_reduction, 91
raw_counts_to_sparse_matrix, 96
rawfile_ToBigWig, 95
read_count_mat_with_separated_chr_start_end, 98
read_sparse_matrix, 98
rebin_helper, 99
rebin_matrix, 100
reduce_dim_batch_correction, 102
reduce_dims_scExp, 101
remove_chr_M_fun, 102
remove_non_canonical_fun, 103
results_enrichmentTest, 103
retrieve_top_bot_features_pca, 104
run_pairwise_tests, 105
run_tsne_scExp, 105
runApp, 68
scExp, 106
separate_BAM_into_clusters, 107
separator_count_mat, 107
smoothBin, 108
subset_bam_call_peaks, 109
summary_DA, 111
swapAltExp_sameColData, 111
table_enriched_genes_scExp, 112
warning_DA, 113
warning_filter_correlated_cell_scExp, 113
warning_plot_reduced_dim_scExp, 114
warning_raw_counts_to_sparse_matrix, 115
wrapper_Signac_FeatureMatrix, 116