Package ‘GladiaTOX’

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Type Package

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Title R Package for Processing High Content Screening data

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Description GladiaTOX R package is an open-source, flexible solution to high-content screening data processing and reporting in biomedical research. GladiaTOX takes advantage of the tcpl core functionalities and provides a number of extensions: it provides a web-service solution to fetch raw data; it computes severity scores and exports ToxPi formatted files; furthermore it contains a suite of functionalities to generate pdf reports for quality control and data processing.

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assignDefaultMthds

Assign default processing methods

Description

Function to assign default processing method to asid in input

Usage

assignDefaultMthds(asid, params = NULL)
Arguments

asid Integer, the asid value(s) to which assign the default methods
params Parameters for level 2, 3, and 5 processing

Details

This function loads all components and endpoints for the given asid(s) in the database, and assigns a default set of processing methods to them.
This function will overwrite any previously assigned methods.
By default, each assay will receive 'none' at level 2. Level 3 data will receive, in order, 'bval.pmi' (39), 'resp.fc' (9), 'resp.log2' (7), and for endpoints with "down" analysis direction, 'resp.multneg1' (6).

Value

None

Examples

```r
## Prepare for analysis before QC + process data
assignDefaultMthds(asid = 1L)

## Process data
gtoxRun(asid = 1L, slvl = 1, elvl = 6, mc.cores = 2)
```

---

### blineShift

#### Shift the baseline to 0

**Description**

blineShift Takes in dose-response data and shifts the baseline to 0 based on the window.

**Usage**

```r
blineShift(resp, logc, wndw)
```

**Arguments**

- `resp` Numeric, the response values
- `logc` Numeric, the log10 concentration values
- `wndw` Numeric, the threshold window

**Value**

A numeric vector containing the shifted response values
buildAssayTab

**Note**

This function is not exported and is not intended to be used by the user.

**See Also**

mc3_mthds, mc3

---

**Description**

This function parses plate annotations and create a mapping between assay endpoints and channels.

**Usage**

```r
buildAssayTab(plate.mtd, chn.map)
```

**Arguments**

- `plate.mtd`: Legacy study annotation file from biobanking.
- `chn.map`: List of endpoints to thermo channels mapping.

**Details**

Function used only when processing historical data.

**Value**

Table with assay information.

**Examples**

```r
## Load sample data
load(system.file("extdata", "data_for_vignette.rda", package="GladiaTOX"))

# Build assay table
assay <- buildAssayTab(plate, chnmap)
```
Configure functions

Functions for configuring the gtox package

Description

These functions are used to configure the gtox settings.
Load the current configuration file

Usage

gtoxConf(drvr = NULL, user = NULL, pass = NULL, host = NULL, db = NULL)
gtoxConfDefault()
gtoxConfList(show.pass = FALSE)
gtoxConfLoad(list.new = TRUE)
gtoxConfReset()
gtoxConfSave()

Arguments

drvr Character of length 1, which database driver to use
user Character of length 1, the database server username
pass Character of length 1, the database server password
host Character of length 1, the database server
db Character of length 1, the name of the gtox database
show.pass Logical, should the password be returned
list.new Logical of length 1, should the new settings be printed?

Details

Currently, the gtox package only supports the "MariaDB" and "SQLite" database drivers.
The settings can be stored in a configuration file to make the using the package more user-friendly.
To create the configuration file, the user must first create a system environment variable ("TCPL_CONF")
that points to to the file. There is more information about system environment variables in Startup
and Sys.getenv. Briefly, the user needs to modify the '.Renviron' file in their home directory. If
the file does not exist, create it, and add the following line:
TCPL_CONF=path/to/confFile.conf
Here 'path/to/confFile.conf' can be any path to a file. One suggestion would be to include gtoxConf
in the home directory, eg. TCPL_CONF=~/gtoxConf. Note, '~' may not indicate the home directory
on every operating system. Once the environment variable is added, the user can change the
Configure functions

settings using gtoxConf, then save the settings to the file given by the TCPL_CONF environment variable running gtoxConfSave().

Value

None

gtoxConf changes options to set the gtox-specific options, most importantly to configure the connection to the gtox databases. gtoxConf will only change non-null values, and can be used to change a single value if needed.

gtoxConfSave modifies the configuration file to reflect the current gtox settings.

gtoxConfList lists the values assigned to the gtox global options.

gtoxConfLoad updates the gtox settings to reflect the current configuration file.

gtoxConfDefault changes the options to reflect the default settings for the example SQLite database, but does not alter the configuration file.

gtoxConfReset is used to generate the initial configuration script, and can be used to reset or regenerate the configuration script by the user.

Examples

    gtoxConfList() # List configuration parameters

    ## Configure database
    sqlite <- file.path(system.file(package="GladiaTOX"),
                        "sql",
                        "gladiatoxdb.sqlite")
    gtoxConf(db=sqlite, user=NA, host=NA, drvr="SQLite")

    ## Configure database with default parameters
    gtoxConfDefault()

    ## List configuration of database parameters
    gtoxConfList()

    ## Set the environment variable pointing to the configuration file
    Sys.setenv(TCPL_CONF=file.path(system.file(package="GladiaTOX"),"gtoxConf"))

    ## Configure database
    gtoxConfLoad()

    ## Set the environment variable pointing to the configuration file
    Sys.setenv(TCPL_CONF=file.path(system.file(package="GladiaTOX"),"gtoxConf"))

    ## Configure database
    gtoxConfReset()
## Set the environment variable pointing to the configuration file
Sys.setenv(TCPL_CONF=file.path(system.file(package="GladiaTOX"),"gtoxConf"))

## Configure database
gtoxConfSave()

---

**deleteStudy**

Completely remove all data for a study

### Description

deleteStudy completely removes all data for a study from the database.

### Usage

deleteStudy(asid, db = NULL)

### Arguments

- **asid**: The assay source/study ID
- **db**: (optional) the database to delete from, defaults to the current database settings

### Details

Cannot be undone. Please use carefully. Not exported, as this is intended for development and should not be used with real data.

### Value

None

### Examples

```r
## Not run:
## Load sample data
load(system.file("extdata", "data_for_vignette.rda", package="GladiaTOX"))

## Build assay table
assay <- buildAssayTab(plate, chnmap)

## Set study parameters
std.nm <- "SampleStudy" # study name
phs.nm <- "PhaseII" # study phase

## Load annotation in gtoxDB
loadAnnot(plate, assay, NULL)
```
exportResultForToxpiGUI

## Delete previously loaded study data
asid = gtoxLoadAsid(fld=c("asnm", "asph"), val=list(std.nm, phs.nm))$asid
if(length(asid)>0){ deleteStudy(asid=asid) }

## End(Not run)

---

exportResultForToxpiGUI

*Create the result table for the asi in input*

### Description

This function export results

### Usage

`exportResultForToxpiGUI(asid, tp, outfile, stat)`

### Arguments

- **asid**: Assay source id
- **tp**: Time point
- **outfile**: Path to the output file
- **stat**: Character vector of statistic to export

### Details

This function is useful to export results in a table format

### Value

None

### Examples

```r
## Export MEC (or AC50) values to be visualized in ToxPiGUI
conf_store <- gtoxConfList()
gtoxConfDefault()

out <- "export_for_toxpiGUI.csv"
exportResultForToxpiGUI(asid=1L, tp="4h", outfile=out, stat=quote(modl_acc))

## Reset configuration
options(conf_store)
```
exportResultTable  
*Create the result table for the asi in input*

**Description**

This function export results

**Usage**

```r
exportResultTable(asid, stats, outfile)
```

**Arguments**

- `asid`  
  Assay source id
- `stats`  
  Statistics to export
- `outfile`  
  Path to the output file

**Details**

This function is useful to export results in a table format

**Value**

None

**Examples**

```r
outfile <- "export_stats.csv"
exportResultTable(asid=1L, stats=c("mod1_acc", "mod1_ga"), outfile=outfile)
```

---

**flareFunc**  
*Calculate the weighted mean of a square to detect plate flares*

**Description**

`flareFunc` calculates the weighted mean of square regions to detect plate flares.

**Usage**

```r
flareFunc(val, coli, rowi, apid, r)
```
**glCheckInput**

Arguments

- **val**: Numeric, the well values
- **coli**: Integer, the well column index
- **rowi**: Integer, the well row index
- **apid**: Character, the assay plate id
- **r**: Integer, the number of wells from the center well (in one direction) to make the square

Value

None

See Also

- `MC6_Methods, Method functions, mc6`

---

**glCheckInput**

*Check validity of input file*

Description

This function checks the structure and content of an input file.

Usage

```
glCheckInput(file)
```

Arguments

- **file**: file URL

Details

This function is useful to check the structure and content of an input file from the GladiaTOX GUI.

Value

List of error messages in JSON format
glComputeToxInd

Create toxicological indicator values for all chemicals in input

Description

This function computes the toxicological indicator value for the assay source id in input.

Usage

```r
glComputeToxInd(asid, tp = NULL, stat = quote(modl_acc))
```

Arguments

- `asid`: assay source id
- `tp`: Time point to report
- `stat`: statistic to plot

Details

This function is useful to compute toxicological indicator values. These values, for each chemical, represent an average impact of the chemical across the list of endpoints tested. The function transform the data to minus log scale. Hence the larger the indicator value, larger is the impact of the chemical.

Value

A data.table with toxicological severity index for each chemical.

Examples

```r
## Store the current config settings, so they can be reloaded at the end
## of the examples
cnf_store <- gtoxConfList()
gtoxConfDefault()

## Compute toxicological severity index
dat <- glComputeToxInd(asid = 1L)
dat[]
```
**glLoadInput**
*Check validity of input file*

**Description**
This function checks the structure and content of an input file.

**Usage**
```r
glLoadInput(file = NULL, studyname = "samplestudy", phasename = "samplephase", tab = NULL)
```

**Arguments**
- **file**: file URL
- **studyname**: Name of the study
- **phasename**: Name of the phase
- **tab**: input table is file URL is not provided

**Details**
This function is useful to load an input file in the GladiaTOX GUI

**Value**
List of error messages in JSON format

**glPlotPie**
*Pie chart for Minimal Effective Concentrations (MEC) and AC50 plot*

**Description**
This function plots MEC values

**Usage**
```r
glPlotPie(asid, chnms = NULL, acids = NULL, aeids = NULL, expos.time.ordr = NULL, stat = quote(modl_acc))
```
**Arguments**

- **asid**  Assay source id
- **chnms** Character vector with list of chemical names
- **acids** Numeric vector with list of acids
- **aeids** Character vector with list of assay endpoints IDs
- **expos.time.ordr** Character vector with sorted list of exposure times
- **stat** Statistic to plot (e.g. MEC:modl_acc or modl_acb, AC50:modl_ga)

**Details**

This function is useful to plot MEC or AC50 values.

**Value**

None

**Examples**

```r
## Create a pie plot of MEC values for all chemicals tested in the study
glPlotPie(asid=1L)
```

---

**glPlotPieLogo**

*plot package logo*

**Description**

This function plots the GladiaTOX logo.

**Usage**

```r
glPlotPieLogo()
```

**Details**

This function is only used to plot the package logo.

**Value**

None

**Examples**

```r
glPlotPieLogo()
```
glPlotPosCtrl

---

**glPlotPosCtrl**

*Box plot for positive control check*

---

**Description**

This function plots positive controls as well as vehicle and treatments normalized values.

**Usage**

```r
glPlotPosCtrl(asid)
```

**Arguments**

- `asid` Assay source id

**Details**

This function is useful to select plates to mask.

**Value**

A list of ggplot objects, one per assay X timepoint.

**Examples**

```r
## Store the current config settings, so they can be reloaded at the end
## of the examples
cnf_store <- gtoxConfList()
gtoxConfDefault()

## Create boxplot for all endpoints and chemicals tested. Useful to save
## plots in a pdf file.
pp <- glPlotPosCtrl(asid = 1L)
pp[[1]]

## Reset configuration
options(conf_store)
```
glPlotPosCtrlMEC

*Box plot for positive control check*

**Description**

This function plots positive controls for study id `asid` as well as boxplot historical positive control MECs.

**Usage**

```r
glPlotPosCtrlMEC(asid, masked=NULL)
```

**Arguments**

- `asid`: Assay source id
- `masked`: Masking color

**Details**

This function is useful to select plates to mask.

**Value**

A list of ggplot objects, one per assay X timepoint.

**Note**

PMI-specific

**Examples**

```r
## Store the current config settings, so they can be reloaded at the end of the examples
conf_store <- gtoxConfList()
gtoxConfDefault()

## Create boxplot for all endpoints and chemicals tested. Useful to save plots in a pdf file.
pp <- glPlotPosCtrlMEC(asid = 1L)
pp[[1]]

## Reset configuration
options(conf_store)
```
glPlotStat

Box plot for Minimal Effective Concentrations (MEC) and AC50 plot

Description

This function plots MEC values

Usage

glPlotStat(asid, ref.chm = NULL, stat = quote(modl_acc))

Arguments

asid          Assay source id
ref.chm       Chemical to adopt as reference
stat          Character vector of statistic to export

Details

This function is useful to show the MEC trend over control chemical

Value

A list of ggplot objects, one per assay X timepoint.

Examples

```r
## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- gtoxConfList()
gtoxConfDefault()

## Create boxplot of MEC
## plots in a pdf file.
pp <- glPlotStat(asid = 1L)
pp[[1]]

## Reset configuration
options(conf_store)
```
glPlotToxInd

Plot toxicological indicator values for all chemicals in input

Description
This function plots the toxicological indicator value for the assay source id in input.

Usage

```r
glPlotToxInd(asid, tp = NULL, stat = quote(mod1_acc))
```

Arguments

- `asid`: assay source id
- `tp`: Time point to report
- `stat`: statistic to plot

Details
This function is useful to plot toxicological indicator values. These values, for each chemical, represent an average impact of the chemical across the list of endpoints tested. The function transform the data to minus log scale. Hence the larger the indicator value, larger is the impact of the chemical.

Value
None

Examples

```r
## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- gtoxConfList()
gtoxConfDefault()

## Compute and plot toxicological severity index
glPlotToxInd(asid=1L)
```
gtoxAddModel

*Draw a gtox Model onto an existing plot*

**Description**

`gtoxAddModel` draws a line for one of the gtox Models (see `Models` for more information) onto an existing plot.

**Usage**

`gtoxAddModel(pars, modl = NULL, adj = NULL, ...)`

**Arguments**

- `pars`: List of parameters from level 4 or 5 output
- `modl`: Character of length 1, the model to plot: 'cnst,' 'hill,' or 'gnls'
- `adj`: Numeric of length 1, an adjustment factor, see details for more information
- `...`: Additional arguments passed to `curve`

**Details**

`gtoxAddModel` draws the model line assuming the x-axis represents log base 10 concentration.

If `modl` is NULL, the function checks `pars$modl` and will return an error if `pars$modl` is also NULL.

`adj` is intended to scale the models, so that models with different response units can be visualized on a single plot. The recommended value for `adj` is \(1/(3*bmad)\) for level 4 data and \(1/\text{coff}\) for level 5 data. If `adj` is NULL the function will check `pars$adj` and set `adj` to 1 if `pars$adj` is also NULL.

**Value**

None

**See Also**

`Models`, `gtoxPlotFits`

**Examples**

```r
# Create some dummy data to plot
logc <- 1:10
r1 <- sapply(logc, gtoxHillVal, ga = 5, tp = 50, gw = 0.5)
r2 <- log2(sapply(logc, gtoxHillVal, ga = 4, tp = 30, gw = 0.5))
p1 <- gtoxFit(logc = logc, resp = r1, bmad = 10)
p2 <- gtoxFit(logc = logc, resp = r2, bmad = log2(1.5))

# In the dummy data above, the two plots are on very different scales
```
plot(r1 ~ logc, pch = 16, ylab = "raw response")
gtoxAddModel(pars = p1, mod1 = "hill")
points(r2 ~ logc)
gtoxAddModel(pars = p2, mod1 = "hill", lty = "dashed")

## To visualize the two curves on the same plot for comparison, we can
## scale the values to the bmad, such that a scaled response of 1 will equal
## the bmad for each curve.
plot(r1/10 ~ logc, pch = 16, ylab = "scaled response")
gtoxAddModel(pars = p1, mod1 = "hill", adj = 1/10)
points(r2/log2(5) ~ logc)
gtoxAddModel(pars = p2, mod1 = "hill", adj = 1/log2(5), lty = "dashed")

---

**gtoxAICProb**

*Calculate the AIC probabilities*

**Description**

gtoxAICProb calculates the probability that the model best represents the data based on the AIC value for each model.

**Usage**

gtoxAICProb(...)

**Arguments**

... Numeric vectors of AIC values

**Details**

The function takes vectors of AIC values. Each vector represents the model AIC values for multiple observation sets. Each vector must contain the same number and order of observation sets. The calculation assumes every possible model is accounted for, and the results should be interpreted accordingly.

**Value**

A vector of probability values for each model given, as a list.

**See Also**

[gtoxFit](#), [AIC](#) for more information about AIC values.
**gtoxAppend**

## Examples

```r
## Returns the probability for each model, given models with AIC values
## ranging from 80 to 100
getxICProb(80, 85, 90, 95, 100)
```

```r
## Also works for vectors
m1 <- c(95, 195, 300) ## model 1 for three different observations
m2 <- c(100, 200, 295) ## model 2 for three different observations
getxICProb(m1, m2)
```

---

**gtoxAppend**

*Append rows to a table*

### Description

`gtoxAppend` takes a data.table (`dat`) and appends the data.table into a database table.

### Usage

```r
gtoxAppend(dat, tbl, db)
```

### Arguments

- `dat` - data.table, the data to append to a table
- `tbl` - Character of length 1, the table to append to
- `db` - Character of length 1, the database containing `tbl`

### Value

None

### Note

This function is not exported and not intended to be used by the user.
gtoxCalcVmad

Calculate and update the assay endpoint cutoff values

Description

gtoxCalcVmad takes the input aeid values and uses them to calculate the assay endpoint cutoff based on the median absolute deviation of vehicle values across the given assay endpoints.

Usage

gtoxCalcVmad(inputs, aeid = NULL, notes = NULL)

Arguments

inputs integer, the aeid(s) used to calculate the cutoff values
aeid integer, the aeid(s) to be updated in the database
notes character of length 1, (optional) comments/justification

Details

If 'aeid' is NULL, the value will be returned with no changes made to the database.

Cutoffs are calculated as the median absolute value of the vehicle values across the assay endpoints given by 'inputs'.

Value

None

Examples

## Store the current config settings, so they can be reloaded at the end of the examples
conf_store <- gtoxConfList()
gtoxConfDefault()

## Prepare for analysis before QC + process data
gtoxCalcVmad(inputs = 10L)

## Reset configuration
options(conf_store)
gtoxCascade  

Do a cascading delete on gtox screening data

Description

gtoxCascade deletes the data for the given id(s) starting at the processing level given. The delete will cascade through all subsequent tables.

Usage

gtoxCascade(lvl, type, id)

Arguments

lvl  
Integer of length 1, the first level to delete from

type  
Character of length 1, the data type, "sc" or "mc"

id  
Integer, the id(s) to delete. See details for more information.

Details

The data type can be either 'mc' for multiple concentration data, or 'sc' for single concentration data. Multiple concentration data will be loaded into the level tables, whereas the single concentration will be loaded into the single tables.

If lvl is less than 3, id is interpreted as acid(s) and if lvl is greater than or equal to 3, id is interpreted as aeid(s).

Value

None

---

gtoxCode2CASN  

Convert chemical code to CAS Registry Number

Description

gtoxCode2CASN takes a code and converts it CAS Registry Number.

Usage

gtoxCode2CASN(code)

Arguments

code  
Character of length 1, a chemical code
Details

The function checks for the validity of the CAS Registry Number. Also, the ToxCast data includes chemicals for which there is no CASRN. The convention for these chemicals is to give them a CASRN as NOCAS_chid; the code for these compounds is CNOCASchid. The function handles the NOCAS compounds as they are stored in the database, as shown in the example below.

Value

A CAS Registry Number.

Examples

```
gtoxCode2CASN("C80057")
gtoxCode2CASN("C09812420") ## Invalid CASRN will give a warning
gtoxCode2CASN("CNOCAS0015") ## The underscore is reinserted for NOCAS codes
```

---

gtoxDelete Delete rows from gtox databases

Description

`gtoxDelete` deletes rows from the given table and database.

Usage

```
gtoxDelete(tbl, fld, val, db)
```

Arguments

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>tbl</td>
<td>Character, length 1, the table to delete from</td>
</tr>
<tr>
<td>fld</td>
<td>Character, the field(s) to query on</td>
</tr>
<tr>
<td>val</td>
<td>List, vectors of values for each field to query on. Must be in the same order as 'fld'.</td>
</tr>
<tr>
<td>db</td>
<td>Character, the database containing the table</td>
</tr>
</tbody>
</table>

Value

None

See Also

`gtoxSendQuery`
gtoxFit

Fit the data with the constant, hill, and gain-loss models

Description

gtoxFit fits the constant, hill, and gain-loss models to the given data and returns some summary statistics and the fit parameters in a list.

Usage

gtoxFit(logc, resp, bmad, force.fit = FALSE, ...)

Arguments

logc    Numeric, log concentration values
resp    Numeric, normalized response values
bmad    Numeric, the baseline median absolute deviation for the entire assay
force.fit Logical, TRUE indicates to attempt fitting every concentration series
...     Any other data to be included in list output.

Details

By default, gtoxFit will only attempt to fit concentration series when at least one median value is greater than 3*bmad.

Value

List of summary values and fit parameters for the given data.

See Also

gtoxObjCnst, gtoxObjHill, gtoxObjGnls, constrOptim

Examples

logc <- 1:10
resp <- sapply(1:10, gtoxHillVal, ga = 5, tp = 50, gw = 0.5)
params <- gtoxFit(logc = logc, resp = resp, bmad = 10)
plot(resp ~ logc)
gtoxAddModel(pars = params, modl = "hill")
Functions to solve the Hill model

Description

These functions solve for Hill model parameters.

Usage

\[
\begin{align*}
gtoxHillACXX(XX, tp, ga, gw, bt = 0) \\
gtoxHillConc(val, tp, ga, gw, bt = 0) \\
gtoxHillVal(logc, tp, ga, gw, bt = 0)
\end{align*}
\]

Arguments

- **XX**: Numeric, the activity level (percentage of the top value)
- **tp**: Numeric, the top value from the Hill model
- **ga**: Numeric, the logAC50 value from the Hill model
- **gw**: Numeric, the Hill coefficient from the Hill model
- **bt**: Numeric, the bottom value from the Hill model
- **val**: Numeric, the activity value
- **logc**: Numeric, the log concentration

Details

- `gtoxHillVal` computes the value of the Hill model for a given log concentration.
- `gtoxHillACXX` computes the activity concentration for a Hill model for a given activity level.
- `gtoxHillConc` computes the Hill model concentration for a given value.

Value

None

Examples

```r
## The following code gives examples for a Hill model with a top of 50,
## bottom of 0, AC50 of 1 and Hill coefficient of 1.
## gtoxHillVal calculates activity value given a concentration. gtoxHillVal
## will return the tp/2 when logc equals ga:
gtoxHillVal(logc = 1, tp = 50, ga = 1, gw = 1, bt = 0)

## Here, gtoxHillConc returns the concentration where the value equals 20
## gtoxHillConc(val = 20, tp = 50, ga = 1, gw = 1, bt = 0)
```
## Note how this differs from gtoxHillACXX:
gtoxHillACXX(XX = 20, tp = 50, ga = 1, gw = 1, bt = 0)

## gtoxHillACXX is based on the top value and allows the user to calculate
specific activity concentrations based on a percentage of the top value

## For example, we can calculate the value for the concentration 0.25, then
use that value to check the other two functions.

value <- gtoxHillVal(logc = 0.25, tp = 50, ga = 1, gw = 1, bt = 0)
c1 <- gtoxHillConc(val = value, tp = 50, ga = 1, gw = 1, bt = 0)
c2 <- gtoxHillACXX(XX = value/50*100, tp = 50, ga = 1, gw = 1, bt = 0)
all.equal(0.25, c1, c2)

## Notice, the value had to be transformed to a percentage of the top value
when using gtoxHillACXX

---

### gtoxImportThermoDB

**Import data from ThermoDB by study ID**

**Description**

This function accesses the ThermoDB webservice and imports data from ThermoDB to the gtox database.

**Usage**

```r
gtoxImportThermoDB(asid, verbose = TRUE, write = FALSE,
store = "STORE", type = "mc",
curlurl = "https://YOUR_THERMODB_SERVER_HOSTNAME/HTTPHCSConnect")
```

**Arguments**

- `asid` Integer, the assay study/source ID to import data for
- `verbose` Logical, should the output from the curl be displayed?
- `write` Logical, should the data be written to the database, or just returned?
- `store` Character, the name of the store on ThermoDB to query
- `type` Character, the data type: 'mc' or 'sc'
- `curlurl` URL of the webservice

**Value**

Data table with content fetched from Thermo DB.
### gtoxListFlds

#### Load the field names for a table

**Description**

`gtoxListFlds` loads the column names for the given table and database.

**Usage**

```r
gtoxListFlds(tbl, db = getOption("TCPL_DB"))
```

**Arguments**

- `tbl` Character of length 1, the gtox database table
- `db` Character of length 1, the gtox database

**Details**

This function can be particularly useful in defining the `fld` param in the `gtoxLoad-` functions.

**Value**

A string of field names for the given table.

**Examples**

```r
## Gives the fields in the mc1 table
getxListFlds("mc1")
```
**gtoxLoadApid**

__Load assay plate information__

**Description**

`gtoxLoadApid` queries the gtox database and returns the assay plate information for the given field and values.

**Usage**

```r

gtoxLoadApid(fld = NULL, val = NULL)
```

**Arguments**

- `fld` Character, the field(s) to query on
- `val` List, vectors of values for each field to query on. Must be in the same order as `fld`.

**Value**

A data.table with the assay plate information for the given parameters

**Examples**

```r

## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- gtoxConfList()
gtoxConfDefault()

## Prepare for analysis before QC + process data
gtoxLoadApid()

## Reset configuration
options(conf_store)
```

---

**gtoxLoadChem**

__Load sample/chemical information__

**Description**

`gtoxLoadChem` queries the gtox database and returns the chemical information for the given field and values.
Usage

```r
gtoxLoadChem(field = NULL, val = NULL, exact = TRUE,
              include.spid = TRUE)
```

Arguments

- **field**: Character of length 1, the field to query on
- **val**: Vector of values to subset on
- **exact**: Logical, should chemical names be considered exact?
- **include.spid**: Logical, should spid be included?

Details

The 'field' parameter is named differently from the 'fld' parameter seen in other functions because it only takes one input.

The functionality of the 'exact' parameter cannot be demonstrated within the SQLite environment. However, in the MariaDB environment the user should be able to give partial chemical name strings, to find chemicals with similar names. For example, setting 'val' to "phenol" when 'field' is "chnm" and 'exact' is FALSE might pull up the chemicals "mercury". More technically, setting 'exact' to FALSE passes the string in 'val' to an RLIKE statement within the MariaDB query.

Value

A data.table with the chemical information for the given parameters

Examples

```r
## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- gtoxConfList()
gtoxConfDefault()

## Passing no parameters gives all of the registered chemicals with their
## sample IDs
gtoxLoadChem()

## Or the user can exclude spid and get a unique list of chemicals
## reset configuration
options(conf_store)
```
**gtoxLoadClib**  
*Load chemical library information*

**Description**

*gtoxLoadClib* queries the gtox databases and returns information about the chemical library.

**Usage**

`gtoxLoadClib(field = NULL, val = NULL)`

**Arguments**

- **field**  
  Character of length 1, 'chid' or 'clib', whether to search by chemical id (chid), or chemical library (clib)

- **val**  
  The values to query on

**Details**

Chemicals are stored in different libraries by chemical ID. Therefore, it is not possible to delineate samples with the same chemical ID into two distinct chemical libraries. However, it is possible for a chemical ID to belong to more than one (or no) chemical libraries.

When chemicals belong to more than one library, the chemical is listed multiple times (one for each distinct library).

**Value**

A data.table with the chemical library information for the given parameters.

**Examples**

```r
## Store the current config settings, so they can be reloaded at the end
## of the examples
cnf_store <- gtoxConfList()
gtoxConfDefault()

## Passing no parameters gives all of the chemical ISs that have a chemical
## library registered
clib <- gtoxLoadClib()

## Reset configuration
options(cnf_store)
```
gtoxLoadData  Load gtox data

Description

gtoxLoadData queries the gtox databases and returns a data.table with data for the given level and data type.

Usage

gtoxLoadData(lvl, fld = NULL, val = NULL, type = "mc")

Arguments

lvl  Integer of length 1, the level of data to load
fld  Character, the field(s) to query on
val  List, vectors of values for each field to query on. Must be in the same order as 'fld'.
type  Character of length 1, the data type, "sc" or "mc"

Details

The data type can be either 'mc' for multiple concentration data, or 'sc' for single concentration data. Multiple concentration data will be loaded into the 'mc' tables, whereas the single concentration will be loaded into the 'sc' tables.

Setting 'lvl' to "agg" will return an aggregate table containing the m4id with the concentration-response data and m3id to map back to well-level information.

Leaving fld NULL will return all data.

Valid fld inputs are based on the data level and type:

```
<table>
<thead>
<tr>
<th>type</th>
<th>lvl</th>
<th>Queried tables</th>
</tr>
</thead>
<tbody>
<tr>
<td>sc</td>
<td>0</td>
<td>sc0</td>
</tr>
<tr>
<td>sc</td>
<td>1</td>
<td>sc0, sc1</td>
</tr>
<tr>
<td>sc</td>
<td>agg</td>
<td>sc1, sc2_agg</td>
</tr>
<tr>
<td>sc</td>
<td>2</td>
<td>sc2</td>
</tr>
<tr>
<td>mc</td>
<td>0</td>
<td>mc0</td>
</tr>
<tr>
<td>mc</td>
<td>1</td>
<td>mc0, mc1</td>
</tr>
<tr>
<td>mc</td>
<td>2</td>
<td>mc0, mc1, mc2</td>
</tr>
<tr>
<td>mc</td>
<td>3</td>
<td>mc0, mc1, mc3</td>
</tr>
<tr>
<td>mc</td>
<td>agg</td>
<td>mc3, mc4_agg</td>
</tr>
<tr>
<td>mc</td>
<td>4</td>
<td>mc4</td>
</tr>
<tr>
<td>mc</td>
<td>5</td>
<td>mc4, mc5</td>
</tr>
<tr>
<td>mc</td>
<td>6</td>
<td>mc4, mc6</td>
</tr>
</tbody>
</table>
```
Value

A data.table containing data for the given fields.

See Also

gtoxQuery, data.table

Examples

## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- gtoxConfList()
gtoxConfDefault()

## Load all of level 0 for multiple-concentration data, note 'mc' is the
## default value for type
gtoxLoadData(lvl = 0)

## Load all of level 1 for single-concentration
gtoxLoadData(lvl = 1, type = "sc")

## List the fields available for level 1, coming from tables mc0 and mc1
gtoxListFlds(tbl = "mc0")
gtoxListFlds(tbl = "mc1")

## Load level 0 data where the well type is "t" and the concentration
## index is 3 or 4
gtoxLoadData(lvl = 1, fld = c("wllt", "cndx"), val = list("t", c(3:4)))

## Reset configuration
options(conf_store)

gtoxLoadUnit(aeid)

Description

gtoxLoadUnit queries the gtox databases and returns a data.table with the response units for the
given assay endpoint ids (aeid).

Usage

gtoxLoadUnit(aeid)

Arguments

aeid Integer, assay endpoint ids
Value
A data.table containing level 3 correction methods for the given aeids.

See Also

gtoxQuery, data.table

Description
gtoxLoadVehicle queries the gtox database and returns the vehicle information for the given field and values.

Usage

gtoxLoadVehicle(field = NULL, val = NULL)

Arguments

field Character of length 1, the field to query on
val Vector of values to subset on

Value
A data.table with the list of vehicles and vehicles ids.

Examples

## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- gtoxConfList()
gtoxConfDefault()

## Prepare for analysis before QC + process data
gtoxLoadVehicle()

## Reset configuration
options(conf_store)
**gtoxLoadVmad**  
*Load cutoff values for assay endpoints*

**Description**

`gtoxLoadVmad` queries the gtox databases and returns a data.table with the cutoff values for the given assay endpoint ids (aeid).

**Usage**

```r
gtoxLoadVmad(aeid = NULL)
```

**Arguments**

- `aeid`: Integer, assay endpoint ids

**Value**

A data.table containing cutoff values for the given aeids.

**Examples**

```r
# Store the current config settings, so they can be reloaded at the end
# of the examples
conf_store <- gtoxConfList()
gtoxConfDefault()

# Prepare for analysis before QC + process data
gtoxLoadVmad()

# Reset configuration
options(conf_store)
```

---

**gtoxLoadWaid**  
*Load well annotation information*

**Description**

`gtoxLoadWaid` queries the gtox database and returns the well annotation information for the given field and values.

**Usage**

```r
gtoxLoadWaid(fld = NULL, val = NULL)
```
gtoxMakeAeidPlts

Arguments

- `fld`: Character, the field(s) to query on
- `val`: List, vectors of values for each field to query on. Must be in the same order as `fld`.

Value

A data.frame with the well annotation information for the given parameters

Examples

```r
## Store the current config settings, so they can be reloaded at the end
## of the examples
cnf_store <- gtoxConfList()
gtoxConfDefault()

## Prepare for analysis before QC + process data
gtoxLoadWaid()

## Reset configuration
options(cnf_store)
```

gtoxMakeAeidPlts  
Create a .pdf with dose-response plots

Description

gtoxMakeAeidPlts creates a .pdf file with the dose-response plots for the given aeid.

Usage

gtoxMakeAeidPlts(aeid, lvl = 4L, fname = NULL, odir = getwd(),
                 ordr.fitc = TRUE, clib = NULL)

Arguments

- `aeid`: Integer of length 1, the assay endpoint id
- `lvl`: Integer of length 1, the data level to use (4-6)
- `fname`: Character, the filename
- `odir`: The directory to save the .pdf file in
- `ordr.fitc`: Logical, should the fits be ordered by fit category?
- `clib`: Character, the chemical library to subset on, see gtoxLoadClib for more information.
Details

gtoxMakeAeidPlts provides a wrapper for gtoxPlotFits, allowing the user to produce PDFs with the curve plots without having to separately load all of the data and establish the PDF device.

If 'fname' is NULL, a default name is given by concatenating together assay information.

Note, the default value for ordr.fitc is TRUE in gtoxMakeAeidPlts, but FALSE in gtoxPlotFits

Value

None

Examples

## Save Aeid plot in a pdf file
gtoxMakeAeidPlts(aeid = 10, lvl = 6, ordr.fitc = FALSE)

---

**gtoxMthdAssign**  
*Functions for managing processing methods*

Description

These functions are used to manage which methods are used to process data. They include methods for assigning, clearing, and loading the assigned methods. Also, gtoxMthdList lists the available methods.

Usage

```r
 gtoxMthdAssign(lvl, id, mthd_id, ordr = NULL, type)
 gtoxMthdClear(lvl, id, mthd_id = NULL, type)
 gtoxMthdList(lvl, type = "mc")
 gtoxMthdLoad(lvl, id = NULL, type = "mc")
```

Arguments

- `lvl` Integer of length 1, the method level
- `id` Integer, the assay component or assay endpoint id(s)
- `mthd_id` Integer, the method id(s)
- `ordr` Integer, the order in which to execute the analysis methods, must be the same length as mthd_id, does not apply to levels 5 or 6
- `type` Character of length 1, the data type, "sc" or "mc"
### gtoxMthdAssign

**Details**

gtoxMthdAssign loads the assigned methods for the given level and ID(s). Similarly, gtoxMthdList displays the available methods for the given level. These two functions do not make any changes to the database.

Unlike the `-Load` and `-List` functions, the `-Assign` and `-Clear` functions alter the database and trigger a delete cascade. gtoxMthdAssign assigns methods to the given ID(s), and gtoxMthdClear removes methods. In addition to the method ID (`mthd_id`), assigning methods at some levels require an order (`ordr`). The `ordr` parameter is necessary to allow progression of methods at level one for single-concentration processing, and levels two and three for multiple-concentration processing. More information about method assignments and the delete cascade are available in the package vignette.

**Value**

None

**Examples**

```r
## Not run:
## Assign level 2 methods (none for all acid values)
gtoxMthdAssign(lvl = 2L, id = 1L, mthd_id = 1, ordr = 1, type = "mc")

## Process data
gtoxRun(asid = 1L, slvl = 1, elvl = 6, mc.cores = 2)

## Not run:
## Clear level 2 methods
gtoxMthdClear(lvl = 2L, id = 1L, mthd_id = NULL, type = "mc")

## Assign level 2 methods (none for all acid values)
gtoxMthdAssign(lvl = 2L, id = 1L, mthd_id = 1, ordr = 1, type = "mc")

## Process data
gtoxRun(asid = 1L, slvl = 1, elvl = 6, mc.cores = 2)

## Not run:
## Store the current config settings, so they can be reloaded at the end of the examples
conf_store <- gtoxConfList()
gtoxConfDefault()

## gtoxListMthd allows the user to display the available methods for
## a given level and data type
head(gtoxMthdList(lvl = 2, type = "mc"))

## gtoxLoadMthd shows which methods are assigned for the given ID, level,
## and data type. Here we will show how to register, load, and clear methods
```
gtoxPlotErrBar

## using an acid not in the example database. Note: There is no check for
## whether an ID exists before assigning/clearing methods.
gtoxMthdLoad(lvl = 2, id = 1, type = "mc")

## Reset configuration
options(conf_store)

gtoxPlotErrBar  Create error bar plots

Description

gtoxPlotErrBar creates the error bar plots.

Usage

gtoxPlotErrBar(c1, c2, aeid, ngrp = NULL)

Arguments

c1  Integer of length 1, the chid value for the first chemical
c2  Integer of length 1, the chid value for the first chemical
aeid  Integer, the aeid value(s) to plot
ngrp  Integer, the number of "slots" to draw; overridden if the number of aeid values
       is greater than 'ngrp'

Value

None

Examples

## Plot error bar plot
gtoxPlotErrBar(c1=1, c2=3, aeid=17:18)
gtoxPlotFitc  
*Plot the fit category tree*

**Description**

gtoxPlotFitc makes a plot showing the level 5 fit categories.

**Usage**

gtoxPlotFitc(fitc = NULL, main = NULL, fitc_sub = NULL)

**Arguments**

- **fitc**  
  Integer, the fit categories

- **main**  
  Character of length 1, the title (optional)

- **fitc_sub**  
  Integer, a subset of fit categories to plot

**Value**

None

**Note**

Suggested device size (inches): width = 10, height = 7.5, pointsize = 9

**Examples**

```r
## Store the current config settings, so they can be reloaded at the end
## of the examples
cconf_store <- gtoxConfList()
gtoxConfDefault()

## Display the fit category tree.
gtoxPlotFitc()
```

gtoxPlotFits  
*Plot summary fits based on fit and dose-response data*

**Description**

gtoxPlotFits takes the dose-response and fit data and produces summary plot figures.

**Usage**

gtoxPlotFits(dat, agg, flg = NULL, ordr.fitc = FALSE, bline = "bmad")
Arguments

dat  data.table, level 4 or level 5 data, see details.
agg  data.table, concentration-response aggregate data, see details.
flg  data.table, level 6 data, see details.
ordr.fitc Logical, should the fits be ordered by fit category?
bline Character of length 1, the value used for drawing the baseline noise

Details

The data for 'dat', 'agg', and 'flg' should be loaded using the \texttt{gtoxLoadData} function with the appropriate 'lvl' parameter. See help page for \texttt{gtoxLoadData} for more information.
Supplying level 4 data for the 'dat' parameter will result in level 4 plots. Similarly, supp
If fits are not ordered by fit category, they will be ordered by chemical ID. Inputs with multiple
assay endpoints will first be ordered by assay endpoint ID.
Any values for 'bline' other than 'coff' will use 3*bmad.

Value

None

Examples

```r
## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- gtoxConfList()
gtoxConfDefault()

## gtoxPlotFits needs data.tables supplying the concentration/response
## data stored in mc4_agg, as well as the fit information from mc4 or mc5.
## Additionally, gtoxPlotFits will take level 6 data from mc6 and add the
## flag information to the plots. The following shows how to make level 6
## plots. Omitting the 'flg' parameter would result in level 5 plots, and
## loading level 4, rather than level 5 data, would result in level 4 plots.

aeid = 2
l5 <- gtoxLoadData(lvl = 5, fld = "aeid", val = aeid)
l4_agg <- gtoxLoadData(lvl = "agg", fld = "aeid", val = aeid)
l6 <- gtoxLoadData(lvl = 6, fld = "aeid", val = aeid)

## Not run:
pdf(file = "gtoxPlotFits.pdf", height = 6, width = 10, pointsize = 10)
gtoxPlotFits(dat = l5, agg = l4_agg, flg = l6)
graphics.off()

## End(Not run)

## While it is most likely the user will want to just save all of the plots
## to view in a PDF, the 'browse' parameter can be used to quickly view
## some plots.
```
## Start by identifying some sample IDs to plot, then call gtoxPlotFits with
## a subset of the data. This browse function is admittedly clunky.

```r
cpa <- gtoxLoadChem(field = "chnm", val = "chromium")[, spid]
l5_sub <- l5[spid %in% cpa]
gtoxPlotFits(dat = l5_sub, agg = l4_agg[m4id %in% l5_sub$m4id])
```

## Reset configuration
```r
options(conf_store)
```

---

### Description

`gtoxPlotM4ID` creates a summary plots for the given m4id(s) by loading the appropriate data from the gtox databases and sending it to `gtoxPlotFits`

### Usage

```r
gtoxPlotM4ID(m4id, lvl = 4L, bline = "bmad")
```

### Arguments

- `m4id` Integer, m4id(s) to plot
- `lvl` Integer, the level of data to plot
- `bline` Character of length 1, the value used for drawing the baseline noise

### Details

A level 4 plot (`l4l` = 4) will plot the concentration series and the applicable curves, without an indication of the activity call or the winning model. Level 4 plots can be created without having done subsequent processing.

Level 5 plots include the level 4 information with the activity call and model selection. The winning model will be highlighted red in the side panel containing the summary statistics. Level 6 plots, in addition the all of the level 4 and 5 information, include the positive flag IDs. If the flag has an associated value, the value will be in parentheses following the flag ID.

Any values for `bline` other than `coff` will use 3*bmad.

### Value

None

### See Also

`gtoxPlotFits, gtoxMakeAeidPlts`
Examples

```r
## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- gtoxConfList()
gtoxConfDefault()

acnm <- "Cytotoxicity (TIER1)_Cytochrome C release_24h"
pltnm <- "S-000049119"
myaid <- gtoxLoadApid()[u_boxtrack == pltnm, aid]
myaid <- myaid[myaid%in%gtoxLoadAid(fld = "asid", val = 1L)$aid]
apid <- gtoxLoadApid()[u_boxtrack == pltnm & aid == myaid, apid]
acid <- gtoxLoadAcid(fld = c("aid", "acnm"), val = list(myaid, acnm))[, acid]
aeid = gtoxLoadAeid(fld = c("acid", "analysis_direction"),
val = list(acid, "up"))[.aeid]
spid = gtoxLoadWaid(fld = c("acid", "wllt"),
val = list(apid, "c"))[.unique(spid)]
m4id = gtoxLoadData(lvl = 4L, fld = c("spid", "aeid"),
val = list(spid, aeid))[. m4id]
gtoxPlotM4ID(m4id = m4id, lvl = 6, bline = "coff") ## Create a level 4 plot
gtoxPlotM4ID(m4id = m4id, lvl = 5) ## Create a level 5 plot
gtoxPlotM4ID(m4id = m4id, lvl = 6) ## Create a level 6 plot

## Reset configuration
options(conf_store)
```

---

**gtoxPlotPie**

Create piechart plots

**Description**

`gtoxPlotPie` creates the piechart plots.

**Usage**

```r
gtoxPlotPie(chid, mrks, aeid, col = NULL, lbl = NULL)
```

**Arguments**

- `chid` : Integer of length 1, the chid value
- `mrks` : Numeric, the values for concentration label rings
- `aeid` : Integer, the aeid values to plot
- `col` : Vector of colors
- `lbl` : Vector with pie labels (optional)
gtoxPlotPieLgnd

Create piechart plot legend

Description

gtoxPlotPieLgnd creates the piechart plots.

Usage

gtoxPlotPieLgnd(aenm, ncol = 2, col = NULL, fit.labels = TRUE)

Arguments

- `aenm`: Character, the assay endpoint names
- `ncol`: Integer, the number of columns for the legend
- `col`: Vector of colors
- `fit.labels`: Boolean, if TRUE, scale the text to fit

Value

None

Examples

## Plot pie legend

gtoxPlotPieLgnd(aenm=c("Endpoint1", "Endpoint2"))
gtoxPlotPlate

Plot plate heatmap

Description

gtoxPlotPlate generates a heatmap of assay plate data

Usage

gtoxPlotPlate(dat, apid, id = NULL, quant = c(0.001, 0.999))

Arguments

dat  data.table containing gtox data
apid  Character of length 1, the apid to plot
id    Integer of length 1, the assay component id (acid) or assay endpoint id (aeid), depending on level. Only need to specify for multiplexed assays when more than one acid/aeid share an apid.
quant Numeric vector, the range of data to include in the legend

Details

The legend represents the range of the data supplied to dat, for the applicable ID. The additional horizontal lines on the legend indicate the range of the plotted plate, to show the relation of the plate to the assay as a whole. A plot with a legend specific for the given apid can be created by only supplying the data for the apid of interest to ‘dat’.

The quant parameter, by default including 99.8 allows for extreme outliers without losing resolution. Outliers in either direction will be highlighted with a dark ring, as seen in the example. A NULL value for ‘quant’ will not restrict the data at all, and will use the full range for the legend.

Wells with a well quality of 0 (only applicable for level 1 plots), will have an "X" through their center.

Value

None

Note

For the optimal output size, use width = 12, height = 8, pointsize = 12, units = "in"
Examples

```r
## Define assay component and extract assay component ID
acnm <- "Cytotoxicity (TIER1)_Cytochrome C release_24h"
acid <- gtoxLoadAcid(fld=c("asid", "acnm"), val=list(1L, acnm))[, acid]
## Extract assay plate ID corresponding to plate name S-000049119
apid <- gtoxLoadApid()[u_boxtrack == "S-000049119", apid]
## Load level 2 data (Raw data before normalization)
l2 <- gtoxLoadData(lvl = 2L, fld = "acid", val = acid)
gtoxPlotPlate(dat = l2, apid = apid, id = acid)
```

---

gtoxPlotWin  
Create winning curve plots

Description

gtoxPlotWin creates best fit plot.

Usage

```r
gtoxPlotWin(chid, aeid, bline = "bmad", collapse = TRUE)
```

Arguments

- `chid`  
  Integer of length 1, the chid value
- `aeid`  
  Integer, the aeid values to plot
- `bline`  
  Character of length 1, the value used for drawing the baseline noise
- `collapse`  
  Logical, collapse the data by spid when true

Details

When 'collapse' is TRUE the plotted points will be the mean of the values based on spid. Any values for 'bline' other than 'coff' will use 3*bmad.

Value

None

Examples

```r
## Not run:
## Load chemical ID
chid <- gtoxLoadChem(field="chnm", val="acrylamide", include.spid=FALSE)$chid

## Load Assay endpoint ID
aeid <- gtoxLoadAeid(fld=c("asid","aenm"),
  val=list(1L, "GSH content_GSH content_4h_dn"), add.fld="asid")$aeid
```
Map assay/chemical ID values to annotation information

Description

gtoxPrepOtpt queries the chemical and assay information from the gtox database, and maps the annotation information to the given data.

Usage

gtoxPrepOtpt(dat, ids = NULL)

Arguments

dat  data.table, output from gtoxLoadData
ids  Character, (optional) a subset of ID fields to map

Details

gtoxPrepOtpt is used to map chemical and assay identifiers to their respective names and annotation information to create a human-readable table that is more suitable for an export/output.

By default the function will map sample ID (spid), assay component id (acid), and assay endpoint ID (aeid) values. However, if 'ids' is not null, the function will only attempt to map the ID fields given by 'ids.'

Value

The given data.table with chemical and assay information mapped

Examples

## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- gtoxConfList()
gtoxConfDefault()

## Load some example data
d1 <- gtoxLoadData(1)

## Check for chemical name in 'dat'
"chnm" %in% names(d1) ## FALSE
## Map chemical annotation only

d2 <- gtoxPrepOtpt(d1, ids = "spid")
"chnm" %in% names(d2) ## TRUE
"acnm" %in% names(d2) ## FALSE

## Map all annotations

d3 <- gtoxPrepOtpt(d1) ## Also works if function is given d2
"chnm" %in% names(d2) ## TRUE
"acnm" %in% names(d2) ## FALSE

## Reset configuration

options(conf_store)

---

gtoxQuery

*Wrappers for sending queries and fetching results*

**Description**

These functions send a query to the given database, and are the access point for all gtox functions that query or update the gtox database.

**Usage**

```r

  gtoxQuery(query, db = getOption("TCPL_DB"),
            drvrv = getOption("TCPL_DVVR"))

  gtoxSendQuery(query, db = getOption("TCPL_DB"),
                 drvrv = getOption("TCPL_DVVR"))

```

**Arguments**

- `query` Character of length 1, the query string
- `db` Character of length 1, the name of the gtox database
- `drvr` Character of length 1, which database driver to use

**Details**

Currently, the gtox package only supports the "MariaDB" and "SQLite" database drivers. 
`gtoxQuery` returns a `data.table` object with the query results. `gtoxSendQuery` sends a query, but does not fetch any results, and returns 'TRUE' or the error message given by the database.

**Value**

None
Examples

```r
## Perform query
gtoxSendQuery(paste0("SELECT * FROM assay_source"))

## Store the current config settings, so they can be reloaded at the end
## of the examples
cnf_store <- gtoxConfList()
gtoxConfDefault()
gtoxQuery("SELECT 'Hello World';")
gtoxQuery("SELECT * FROM assay;")

## Reset configuration
options(cnf_store)
```

---

**gtoxRegister**

*Functions for registering & updating annotation information*

**Description**

These functions are used to register and update the chemical and assay annotation information.

**Usage**

```r
gtoxRegister(what, flds)
gtoxUpdate(what, id, flds)
```

**Arguments**

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td><code>what</code></td>
<td>Character of length 1, the name of the ID to register or update</td>
</tr>
<tr>
<td><code>flds</code></td>
<td>Named list, the other fields and their values</td>
</tr>
<tr>
<td><code>id</code></td>
<td>Integer, the ID value(s) to update</td>
</tr>
</tbody>
</table>

**Details**

These functions are used to populate the gtox database with the necessary annotation information to complete the processing. As shown in the package vignette, the package requires some information about the samples and assays before data can be loaded into the gtox database.

Depending on what is being registered, different information is required. The following table lists the fields that can be registered/updated by these functions, and the minimal fields required for registering a new ID. (The database table affected is in parentheses.)

- `asid` (assay_source): assay_source_name
- `aid` (assay): asid, assay_name, assay_footprint
- acid (assay_component): aid, assay_component_name
- aeid (assay_component_endpoint): acid, assay_component_endpoint_name, normalized_data_type
- spid (sample): spid, chid
- chid (chemical): chid, casn
- clib (chemical_library): chid, clib
- *vehicle (vehicle): vehicle_name
- *waid (assay_plate_well): apid, spid, rowi, coli, wllt, vhid, conc
- *apid (assay_plate): aid

Note: The functions accept the abbreviated forms of the names, i.e. "aenm" rather than the full "assay_component_endpoint_name." More information about the registration process and all of the fields is available in the vignette. * indicate PMI-specific fields.

Value

None

Examples

```r
## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- gtoxConfList()
gtoxConfDefault()

## Load current ASID information
gtoxLoadAsid()

## Register a new assay source
gtoxRegister(what = "asid", flds = list(asnm = "example_asid",
                                           asph = "example_phase"))

## Show the newly registered ASID
gtoxLoadAsid(add.fld = "assay_source_desc")

## Notice that the newly created ASID does not have an assay_source_desc.
## The field could have been defined during the registration process, but
## can also be updated using gtoxUpdate
i1 <- gtoxLoadAsid()[asnm == "example_asid", asid]
gtoxUpdate(what = "asid",
           id = i1,
           flds = list(assay_source_desc = "example asid description"))
gtoxLoadAsid(add.fld = "assay_source_desc")

## Remove the created ASID. Note: Manually deleting primary keys can cause
## serious database problems and should not generally be done.
gtoxSendQuery(paste0("DELETE FROM assay_source WHERE asid = ", i1, ";"))

## Reset configuration
options(conf_store)
```
**gtoxReport**

**Generate a report**

**Description**

gtoxReport generates a report.

**Usage**

gtoxReport(type, asid, params = NULL, odir = getwd(), report_author, 
report_title = "Report", sumfile = NULL, 
keep.intermediates = FALSE)

**Arguments**

- **type**  
The type of report to generate
- **asid**  
The assay source/study ID
- **params**  
Named list containing report type-specific parameters, see details
- **odir**  
The output directory
- **report_author**  
The author for the report
- **report_title**  
The title for the report
- **sumfile**  
Path to a text file that inserts into the report
- **keep.intermediates**  
TRUE/FALSE, keep intermediate files when TRUE

**Details**

'type' can have three values, "all," "compare," or "qc." Each report contains slightly different elements, but in general:

- "all" – summarizes the results for all or some compounds
  - "chids" – (optional) a vector of chid values to report, rather than all available compounds
- "compare" – compares the results for two compounds
  - "c1" – (required) the chid for the first compound to compare
  - "c2" – (required) the chid for the second compound to compare
- "qc" – summarizes low-level data for quality control purposes
  - "aids" – (optional) a vector of aid values to report, rather than all available assays

The required list elements vary depending on the type of report, and are described under the report descriptions above.

'sumfile' allows the user to inject a Tex file into the report. The file contents will be inserted into the Study Overview section, immediately after the autogenerated text. Technically, 'sumfile' is brewed, so 'sumfile' can make use of brew and Sweave syntax, and all data loaded for the report.
gtoxRun

Perform data processing

Description

gtoxRun is the function for performing the data processing, for both single-concentration and multiple-concentration formats.

Usage

gtoxRun(asid = NULL, slvl, elvl, id = NULL, type = "mc", mc.cores = NULL, outfile = NULL, runname = NULL)

Arguments

<table>
<thead>
<tr>
<th>Name</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>asid</td>
<td>Integer, assay source id</td>
</tr>
<tr>
<td>slvl</td>
<td>Integer of length 1, the starting level to process</td>
</tr>
<tr>
<td>elvl</td>
<td>Integer of length 1, the ending level to process</td>
</tr>
<tr>
<td>id</td>
<td>Integer, rather than assay source id, the specific assay component or assay endpoint id(s) (optional)</td>
</tr>
<tr>
<td>type</td>
<td>Character of length 1, the data type, &quot;sc&quot; or &quot;mc&quot;</td>
</tr>
<tr>
<td>mc.cores</td>
<td>Integer of length 1, the number of cores to use, set to 1 when using Windows operating system</td>
</tr>
<tr>
<td>outfile</td>
<td>Character of length 1, the name of the log file (optional)</td>
</tr>
<tr>
<td>runname</td>
<td>Character of length 1, the name of the run to be used in the outfile (optional)</td>
</tr>
</tbody>
</table>

Examples

```rst
## Generate full analysis report

## Not run:
## Generate report
    gtoxReport(type = "all", asid = 1L, report_author = "author", report_title = "Processing report")

## End(Not run)
```
gtoxSetWllq

Details

The gtoxRun function is the core processing function within the package. The function acts as a wrapper for individual processing functions, (ie. mc1, sc1, etc.) that are not exported. If possible, the processing is done in parallel by 'id' by utilizing the mclapply function within the parallel package.

If slvl is less than 4, 'id' is interpreted as acid and if slvl is 4 or greater 'id' is interpreted as aeid. Must give either 'asid' or 'id'. If an id fails no results get loaded into the database, and the id does not get placed into the cue for subsequent level processing.

The 'type' parameter specifies what type of processing to complete: "mc" for multiple-concentration processing, and "sc" for single-concentration processing.

Value

A list containing the results from each level of processing. Each level processed will return a named logical vector, indicating the success of the processing for the id.

Examples

## Process data for asid 1
## Process data

gtoxRun(asid = 1L, slvl = 1, elvl = 6, mc.cores = 2)

---

gtoxSetWllq

*Change the well quality for a vector of lvl 0 IDs*

Description

gtoxSetWllq changes the well quality to either 100 or 0 for a given list of 'm0id' or 's0id' values. Changing the well quality initiates a delete cascade for the affected assay components.

Usage

gtoxSetWllq(ids, wllq, type)

Arguments

ids Integer, the 'm0id' or 's0id' values to change
wllq Integer of length 1, the new well quality value, 0 or 1
type Character of length 1, the data type, "sc" or "mc"

Value

TRUE if successful.
Examples

```r
## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- gtoxConfList()
gtoxConfDefault()

## Set well quality to zero for specific lvl zero ids.
gtoxSetWllq(ids = 1633, wllq = 0, type = "mc")

## Reset configuration
options(conf_store)
```

---

### gtoxSubsetChid

**Subset level 5 data to a single sample per chemical**

**Description**

gtoxSubsetChid subsets level 5 data to a single tested sample per chemical. In other words, if a chemical is tested more than once (a chid has more than one spid) for a given assay endpoint, the function uses a series of logic to select a single "representative" sample.

**Usage**

gtoxSubsetChid(dat, flag = TRUE)

**Arguments**

- **dat**: data.table, a data.table with level 5 data
- **flag**: Integer, the mc6_mthd_id values to go into the flag count, see details for more information

**Details**

gtoxSubsetChid is intended to work with level 5 data that has chemical and assay information mapped with gtoxPrepOtpt.

To select a single sample, first a "consensus hit-call" is made by majority rule, with ties defaulting to active. After the chemical-wise hit call is made, the samples corresponding to to chemical-wise hit call are logically ordered using the fit category, the number of the flags, and the modl_ga, then the first sample for every chemical is selected.

The flag param can be used to specify a subset of flags to be used in the flag count. Leaving flag TRUE utilize all the available flags. Setting flag to FALSE will do the subsetting without considering any flags.

**Value**

A data.table with a single sample for every given chemical-assay pair.
### Examples

```r
## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- gtoxConfList()
gtoxConfDefault()

## Load the example level 5 data
d1 <- gtoxLoadData(lvl = 5, fld = "aeid", val = 2)
d1 <- gtoxPrepOtpt(d1)

## Subset to an example of a duplicated chid
d2 <- d1[chid == 10]
d2[, list(m4id, hitc, fitc, modl_ga)]

## Here the consensus hit-call is 1 (active), and the fit categories are
## all equal. Therefore, if the flags are ignored, the selected sample will
## be the sample with the lowest modl_ga.
gtoxSubsetChid(dat = d2, flag = FALSE)[, list(m4id, modl_ga)]

## Reset configuration
options(conf_store)
```

---

**gtoxWriteData**  
*Write screening data into the gtox databases*

### Description

gtoxWriteData takes a data.table with screening data and writes the data into the given level table in the gtox databases.

### Usage

gtoxWriteData(dat, lvl, type)

### Arguments

- **dat**: data.table, the screening data to load
- **lvl**: Integer of length 1, the data processing level
- **type**: Character of length 1, the data type, "sc" or "mc"
Details

This function appends data onto the existing table. It also deletes all the data for any acids or aeids dat contains from the given and all downstream tables.

The data type can be either 'mc' for multiple concentration data, or 'sc' for single concentration data. Multiple concentration data will be loaded into the level tables, whereas the single concentration will be loaded into the single tables.

Value

None

See Also

gtoxCascade, gtoxAppend

Examples

```r
## Not run:
## Load sample data
go <- system.file("extdata", "data_for_vignette.rda", package="GladiaTOX")

# Build assay table
assay <- buildAssayTab(plate, chnmap)

## Set study parameters
std.nm <- "SampleStudy" # study name
phs.nm <- "PhaseII" # study phase

## Load annotation in gtoxDB
go <- loadAnnot(plate, assay, NULL)

## Get the created study ID
asid = gtoxLoadAsid(fld = c("asnm", "asph"), val = list(std.nm, phs.nm))$asid

## Prepare and load data
dat <- prepareDatForDB(asid, dat)
go <- gtoxWriteData(dat[, list(acid, waid, wllq, rval)], lvl = 0, type = "mc")

## End(Not run)
```
is.odd

Usage

interlaceFunc(val, intq, coli, rowi, apid, r)

Arguments

val Numeric, the well values
intq Numeric, interlace quadrant
coli Integer, the well column index
rowi Integer, the well row index
apid Character, the assay plate id
r Integer, the number of wells from the center well (in one direction) to make the square

Value

None

See Also

MC6_Methods, Method functions, mc6

is.odd Check for odd numbers

Description

is.odd takes an integer vector, x, and returns TRUE for odd integers.

Usage

is.odd(x)

Arguments

x An integer

Value

TRUE for odd integers and FALSE for even integers.

See Also

Other gtox abbreviations: lu, lw, sink_reset
Load assay information

Functions for loading assay information

Description

These functions query the gtox databases and returns a data.table with assay ID and name information. More information about the assay hierarchy is available in the overview vignette.

Usage

```r

gtoxLoadAcid(fld = NULL, val = NULL, add.fld = NULL)

gtoxLoadAeid(fld = NULL, val = NULL, add.fld = NULL)

gtoxLoadAid(fld = NULL, val = NULL, add.fld = NULL)

gtoxLoadAsid(fld = NULL, val = NULL, add.fld = NULL)
```

Arguments

- **fld**: Character, the field(s) to query/subset on
- **val**: List, vectors of values for each field to query/subset on. Must be in the same order as 'fld'.
- **add.fld**: Character, additional field(s) to include, but not query/subset on

Details

Each element in the assay hierarchy has its own function, loading the ID and name for the given assay element. For example, `gtoxLoadAsid` will return the assay source ID (asid) and assay source name (asnm).

Value

A data.table containing the ID, name, and any additional fields.

Examples

```r

## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- gtoxConfList()
gtoxConfDefault()

## The load assay functions can be used without any parameters to list the
## full list of registered assay elements:

## Assay source ID table

gtoxLoadAsid()
```
## Assay ID table

gtoxLoadAid()

## Assay component ID table

gtoxLoadAcid()

## Assay endpoint ID table

gtoxLoadAeid()

## Similarly, the user can add fields without doing any element selection:

gtoxLoadAeid(add.fld = c("asid", "aid", "acid"))

## Or, the user can look only at a subset:

gtoxLoadAeid(fld = "aeid", val = 1, add.fld = "asid")

## The field can be any value in one of the corresponding assay element tables, but the functions also recognize the abbreviated version of the name fields.

gtoxListFlds("assay")
a1 <- gtoxLoadAeid(fld = "anm", val = "Apo Necro (casp37)_4h")
a2 <- gtoxLoadAeid(fld = "assay_name", val = "Apo Necro (casp37)_4h")
identical(a1, a2)

## Reset configuration

options(conf_store)

---

**loadAnnot**

Register the annotations provided by GUI

**Description**

This function parses the output from the GUI and registers the appropriate data within the GladiaTOX database.

**Usage**

loadAnnot(plate, assay, outFile = "out.json")

**Arguments**

- **plate** path to ‘plate’ JSON file produced by the GUI
- **assay** path to ‘assay’ JSON file produced by the GUI
- **outFile** character of length 1, name of the output file

**Details**

If loading legacy data, ‘outFile’ should be set to NULL and no JSON file will be written.
Value

Logical value

Examples

## Not run:
## Load sample data
load(system.file("extdata", "data_for_vignette.rda", package="GladiaTOX"))

## Build assay table
assay <- buildAssayTab(plate, chnmap)

## Set study parameters
std.nm <- "SampleStudy" # study name
phs.nm <- "PhaseII" # study phase

## Delete previously loaded study data
asid = gtoxLoadAsid(fld=c("asnm", "asph"), val=list(std.nm, phs.nm))$asid
if(length(asid)>0){ deleteStudy(asid=asid) }

## Load annotation in gtoxDB
loadAnnot(plate, assay, NULL)

## End(Not run)

---

**lu**

Abbreviation for `length(unique(x))`

Description

`lu` takes a logical vector, `x`, and returns `length(unique(x))`.

Usage

`lu(x)`

Arguments

`x` A logical

Value

The unique of the TRUE values in `x`

See Also

`unique, which`

Other gtox abbreviations: `is.odd, lw, sink_reset`
**lw**

*Abbreviation for length(which(x))*

**Description**

1w takes a logical vector, x, and returns length(which(x)).

**Usage**

1w(x)

**Arguments**

x 
A logical

**Value**

The length of the TRUE values in x

**See Also**

length, which

Other gtox abbreviations: is.odd, lu, sink_reset

**Examples**

1w(c(TRUE, FALSE, TRUE))

---

**mc1**

*Perform level 1 multiple-concentration processing*

**Description**

mc1 loads level 0 data from the gtox database for the given id and performs level 1 multiple-concentration processing. The processed data is then loaded into the mc1 table and all subsequent data is deleted with getxCascade. See details for more information.

The individual processing functions are no longer exported, as it is typically more convenient and suggested to use the getxRun wrapper function.
Usage

mc2(ac, wr = FALSE)

Arguments

ac   Integer of length 1, assay component id (acid) for processing.
wr   Logical, whether the processed data should be written to the gtox database

Details

Level 2 multiple-concentration processing includes defining the corrected value, cval, based on the correction methods listed in the mc2_acid and mc2_methods tables.
Description

`mc2_mthds` returns a list of correction/transformation functions to be used during level 2 multiple-concentration processing.

Usage

`mc2_mthds()`

Details

The functions contained in the list returned by `mc2_mthds` return a list of expressions to be executed in the `mc2` (not exported) function environment. The functions are described here for reference purposes. The `mc2_mthds` function is not exported, nor is it intended for use. All available methods are described in the Available Methods section, listed by the function/method name.

Value

A list functions

Available Methods

More information about the level 2 multiple-concentration processing is available in the package vignette, "Pipeline_Overview."

- **log2** Take the logarithm of `cval` with the base 2.
- **log10** Take the logarithm of `cval` with the base 10.
- **rmneg** Remove entries where `cval` is less than 0.
- **rmzero** Remove entries where `cval` is 0.
- **mult25** Multiply `cval` by 25.
- **mult100** Multiply `cval` by 100.
**mc3**

Perform level 3 multiple-concentration processing

**Description**

mc3 loads level 2 data from the gtox database for the given id and performs level 3 multiple-concentration processing. The processed data is then loaded into the mc3 table and all subsequent data is deleted with gtoxCascade. See details for more information.

The individual processing functions are no longer exported, as it is typically more convenient and suggested to use the gtoxRun wrapper function.

**Usage**

```r
mc3(ac, wr = FALSE)
```

**Arguments**

- `ac` Integer of length 1, assay component id (acid) for processing.
- `wr` Logical, whether the processed data should be written to the gtox database

**Details**

Level 3 multiple-concentration processing includes mapping assay component to assay endpoint, duplicating the data when the assay component has multiple assay endpoints, and any normalization of the data. Data normalization based on methods listed in mc3_aeid and mc3_methods tables.

**Value**

A boolean of length 1, indicating the success of the processing, or when `wr` is FALSE, a list where the first element is a boolean indicating the success of processing and the second element is a data.table containing the processed data.

**Note**

This function is not exported and is not intended to be used by the user.

**See Also**

mc2, Method functions to query what methods get applied to each acid

---

**negshift** Shift cval by subtracting out the minimum of cval and adding 1, such that the new minimum of cval is 1.

**mult25** Multiply cval by 2.5.

**mult3** Multiply cval by 3.

**mult6** Multiply cval by 6.
See Also

Method functions, MC3_Methods
Other multiple-concentration data processing functions: mc1, mc2, mc4, mc5, mc6

Description

mc3_mthds returns a list of normalization methods to be used during level 3 multiple-concentration processing.

Usage

mc3_mthds()

Details

The functions contained in the list returned by mc3_mthds take 'aeids' (a numeric vector of aeid values) and returns a list of expressions to be executed in the mc3 (not exported) function environment. The functions are described here for reference purposes, The mc3_mthds function is not exported, nor is it intended for use.

All available methods are described in the Available Methods section, listed by the type of function and the function/method name.

Value

A list of functions

Available Methods

The methods are broken into three types, based on what fields they define. Different methods are used to define "bval" (the baseline value), "pval" (the positive control value), and "resp" (the final response value).

Although it does not say so specifically in each description, all methods are applied by aeid.

More information about the level 3 multiple-concentration processing is available in the package vignette, "Pipeline_Overview."

bval Methods:

bval.apid.nwlls.med Calculate bval as the median of cval for wells with wllt equal to "n," by apid.

bval.apid.1owconc.med Calculate bval as the median of cval for wells with wllt equal to "t" and cndx equal to 1 or 2, by apid.

bval.apid.twlls.med Calculate bval as the median of cval for wells with wllt equal to "t," by apid.
bval.apid.tn.med  Calculate bval as the median of cval for wells with wllt equal to "t" or "n," by apid.
bval.apid.nwllslowconc.med  Calculate bval as the median of cval for wells with wllt equal to "n" or wells with wllt equal to "t" and cndx equal to 1 or 2, by apid.
bval.spid.lowconc.med  Calculate bval as the median of cval for wells with wllt equal to "t" and cndx equal to 1, 2, or 3, by spid.

pval Methods:
pval.apid.pwlls.med  Calculate pval as the median of cval for wells with wllt equal to "p," by apid.
pval.apid.mwlls.med  Calculate pval as the median of cval for wells with wllt equal to "m," by apid.
pval.apid.medpcbyconc.max  First calculate the median of cval for wells with wllt equal to "p" or "c," by wllt, conc, and apid. Then calculate pval as the maximum of the calculated medians, by apid.
pval.apid.medpcbyconc.min  First calculate the median of cval for wells with wllt equal to "p" or "c," by wllt, conc, and apid. Then calculate pval as the minimum of the calculated medians, by apid.
pval.apid.medncbyconc.min  First calculate the median of cval for wells with wllt equal to "m" or "o," by wllt, conc, and apid. Then calculate pval as the minimum of the calculated medians, by apid.
pval.apid.pmv.min  First calculate the median of cval for wells with wllt equal to "p," "m," or "v," by wllt, conc, and apid. Then calculate pval as the minimum of the calculated medians, by apid.
pval.apid.pmv.max  First calculate the median of cval for wells with wllt equal to "p," "m," or "v," by wllt, conc, and apid. Then calculate pval as the maximum of the calculated medians, by apid.
pval.apid.f.max  First calculate the median of cval for wells with wllt equal to "f," by wllt, conc, and apid. Then calculate pval as the maximum of the calculated medians, by apid.
pval.apid.f.min  First calculate the median of cval for wells with wllt equal to "f," by wllt, conc, and apid. Then calculate pval as the minimum of the calculated medians, by apid.
pval.apid.p.max  First calculate the median of cval for wells with wllt equal to "p," by wllt, conc, and apid. Then calculate pval as the maximum of the calculated medians, by apid.
pval.apid.p.min  First calculate the median of cval for wells with wllt equal to "p," by wllt, conc, and apid. Then calculate pval as the minimum of the calculated medians, by apid.
pval.apid.v.min  First calculate the median of cval for wells with wllt equal to "v," by wllt, conc, and apid. Then calculate pval as the minimum of the calculated medians, by apid.
pval.zero  Define pval as 0.

resp Methods:
resp.pc  Calculate resp as \( \frac{cval - bval}{pval - bval} \times 100. \)
resp.fc  Calculate resp as \( \frac{cval}{bval}. \)
resp.logfc  Calculate resp as \( cval - \log bval. \)
resp.log2  Take the logarithm of resp with base 2.
resp.mult25  Multiply resp by 25.
**resp.scale.mad.log2fc** Multiply resp by the scale factor \( \frac{\log_2(1.2)}{3\text{bmad}} \).

**resp.scale.quant.log2fc** Determine the maximum response \( md \) where \( md = \text{abs}(\text{1st centile} - \text{50th centile}) \) or \( \text{abs}(99\text{th centile} - \text{50th centile}) \), whichever is greater. Scale the response such that 20 percent of \( md \) equals \( \log_2(1.2) \).

**resp.multneg1** Multiply resp by -1.

**resp.shiftneg.3bmad** Shift all resp values less than 3*bmad to 0.

**resp.shiftneg.6bmad** Shift all resp values less than 6*bmad to 0.

**resp.shiftneg.10bmad** Shift all resp values less than 10*bmad to 0.

**resp.blineshift.3bmad.repi** Shift resp values with the blineShift function by repi, where the window (wndw) is 3*bmad.

**resp.blineshift.50.repi** Shift resp values with the blineShift function by repi, where the window (wndw) is 50.

**resp.blineshift.3bmad.spid** Shift resp values with the blineShift function by spid, where the window (wndw) is 3*bmad.

**resp.blineshift.50.spid** Shift resp values with the blineShift function by spid, where the window (wndw) is 50.

**none** Do no normalization; make resp equal to cval.

**Note**

This function is not exported and is not intended to be used by the user.

**See Also**

mc3, gtoxMthdLoad to query what methods get applied to each aeid

---

**mc4**

*Perform level 4 multiple-concentration processing*

**Description**

mc4 loads level 3 data from the gtox database for the given id and performs level 4 multiple-concentration processing. The processed data is then loaded into the mc4 table and all subsequent data is deleted with gtoxCascade. See details for more information.

The individual processing functions are no longer exported, as it is typically more convenient and suggested to use the gtoxRun wrapper function.

**Usage**

mc4(ae, wr = FALSE)

**Arguments**

- **ae** Integer of length 1, assay endpoint id (aeid) for processing.
- **wr** Logical, whether the processed data should be written to the gtox database
Details

Level 4 multiple-concentration modeling takes the dose-response data for chemical-assay pairs, and fits three models to the data: constant, hill, and gain-loss. For more information about the models see Models. When a chemical has more than one sample, the function fits each sample separately.

Value

A boolean of length 1, indicating the success of the processing, or when ‘wr’ is FALSE, a list where the first element is a boolean indicating the success of processing and the second element is a data.table containing the processed data.

See Also

gtoxFit, Models

Other multiple-concentration data processing functions: mc1, mc2, mc3, mc5, mc6

Description

mc5 loads level 4 data from the gtox database for the given id and performs level 5 multiple-concentration processing. The processed data is then loaded into the mc5 table and all subsequent data is deleted with gtoxCascade. See details for more information.

The individual processing functions are no longer exported, as it is typically more convenient and suggested to use the gtoxRun wrapper function.

Usage

mc5(ae, wr = FALSE)

Arguments

ae Integer of length 1, assay endpoint id (aeid) for processing.
wr Logical, whether the processed data should be written to the gtox database.

Details

Level 5 multiple-concentration hit-calling uses the fit parameters and the activity cutoff methods from mc5_aeid and mc5_methods to make an activity call and identify the winning model for each fit.

Value

A boolean of length 1, indicating the success of the processing, or when ‘wr’ is FALSE, a list where the first element is a boolean indicating the success of processing and the second element is a data.table containing the processed data.
**Description**

`mc5_mthds` returns a list of additional activity cutoff methods to be used during level 5 multiple-concentration processing.

**Usage**

```r
mc5_mthds()
```

**Value**

A list of functions

**Available Methods**

More information about the level 5 multiple-concentration processing is available in the package vignette, "Pipeline_Overview."

- **bmad3** Add a cutoff value of 3*bmad.
- **pc20** Add a cutoff value of 20.
- **log2_1.2** Add a cutoff value of log2(1.2).
- **log10_1.2** Add a cutoff value of log10(1.2).
- **bmad5** Add a cutoff value of 5*bmad.
- **bmad6** Add a cutoff value of 6*bmad.
- **bmad10** Add a cutoff value of 10*bmad.
- **log2_2** Add a cutoff value of log2(2).
- **log10_2** Add a cutoff value of log10(2).
- **neglog2_0.88** Add a cutoff value of -1*log2(0.88).
- **vmad3** Add a cutoff value of 3*vmad.
- **vmad5** Add a cutoff value of 5*vmad.
- **vmad10** Add a cutoff value of 10*vmad.

**See Also**

`mc5, Method functions` to query what methods get applied to each aeid.
mc6

Perform level 6 multiple-concentration processing

Description

mc6 loads level 5 data from the gtox database for the given id and performs level 6 multiple-concentration processing. The processed data is then loaded into the mc6 table and all subsequent data is deleted with gtoxCascade. See details for more information.

The individual processing functions are no longer exported, as it is typically more convenient and suggested to use the gtoxRun wrapper function.

Usage

mc6(ae, wr = FALSE)

Arguments

ae   Integer of length 1, assay endpoint id (aeid) for processing.
wr   Logical, whether the processed data should be written to the gtox database

Details

Level 6 multiple-concentration flagging uses both the plate level concentration-response data and the modeled parameters to flag potential false positives and false negative results.

Value

A boolean of length 1, indicating the success of the processing, or when ‘wr’ is FALSE, a list where the first element is a boolean indicating the success of processing and the second element is a data.table containing the processed data

See Also

Method functions, MC6_Methods
Other multiple-concentration data processing functions: mc1, mc2, mc3, mc4, mc5
**Description**

mc6_mthds returns a list of flag methods to be used during level 6 multiple-concentration processing.

**Usage**

mc6_mthds()

**Value**

A list functions

**Available Methods**

More information about the level 6 multiple-concentration processing is available in the package vignette, "Pipeline_Overview."

**row.dev.up** The row.dev.up flag looks at the individual point data, searching for row effects across an apid. To get flagged the point has to be greater than 3 standard deviations above the mean response for the plate, and the row mean must be greater than 3 standard deviations above the row means for the plate.

**row.dev.dn** The row.dev.dn flag is identical to the row.dev.up flag, but identifies points falling in rows with decreased signals.

**col.dev.up** The col.dev.up flag is identical to the row.dev.up flag, but identifies points falling in columns with increased signals.

**col.dev.dn** The col.dev.dn flag is identical to the row.dev.up flag, but identifies points falling in columns with decreased signals.

**plate.flare** The plate.flare flag looks at the individual point data, searching for overly active regions across an apid. Intended for use in fluorometric assays that are read by a plate-reader that measures the plate as a whole, rather than measuring individual wells. For each well the flare value is calculated as a weighted mean a 5 well by 5 well box centered on the well where the weight given to each well in the box is the euclidian distance from the center well. The flag then identifies points with flare values greater than 3 standard deviations above the mean flare values for the plate.

**plate.interlace** The plate.interlace flag is specific to one experimental design that plates chemicals from a 386 well chemical plate to a 1536 well assay plate. The flag looks for any chemical-plate affects, by looking for an increased signal in the wells originating from the same chemical plate.

**rep.mismatch** The rep.mismatch flag is still in development and is not suggested for use at this time.
pintool  Deprecated. The pintool flag uses a complicated algorithm to look for signal potentially caused by residual in the pintool used to deliver the chemical to assay plates in some experimental designs. The gnls.lowconc is a faster and simpler way to identify where this problem may be driving the activity or hit-call.

singlept.hit.high  The singlept.hit.high flag identifies concentration series where the median response was greater than 3*bmad only at the highest tested concentration and the series had an active hit-call.

singlept.hit.mid  The singlept.hit.mid flag identifies concentration series where the median response was greater than 3*bmad at only one concentration (not the highest tested concentration) and the series had an active hit-call.

multipoint.neg  The multipoint.neg flag identifies concentration series with response medians greater than 3*bmad at multiple concentrations and an inactive hit-call.

gnls.lowconc  The gnls.lowconc flag identifies concentration series where the gain-loss model won, the gain AC50 is less than the minimum tested concentration, and the loss AC50 is less than the mean tested concentration.

noise  The noise flag attempts to identify noisy concentration series by flagging series where the root mean square error for the series is greater than the cutoff for the assay endpoint.

border.hit  The border.hit flag identifies active concentration series where the top parameter of the winning model was less than or equal to 1.2*cut-off or the activity probability was less than 0.9.

border.miss  The border.miss flag identifies inactive concentration series where either the Hill or gain-loss top parameter was greater than or equal to 0.8*cut-off and the activity probability was greater than 0.5.

overfit.hit  The overfit.hit flag recalculates the model winner after applying a small sample correction factor to the AIC values. If the hit-call would be changed after applying the small sample correction factor the series is flagged. Series with less than 5 concentrations where the hill model won and series with less than 7 concentrations where the gain-loss model won are automatically flagged.

efficacy.50  The efficacy.50 flag identifies concentration series with efficacy values (either the modeled top parameter for the winning model or the maximum median response) are less than 50. Intended for use with biochemical assays where one might expect at least a 50% change in real responses.

See Also

mc6, Method functions to query what methods get applied to each aid

Models  Model objective functions

Description

These functions take in the dose-response data and the model parameters, and return a likelihood value. They are intended to be optimized using constrOptim in the gtoxFit function.
Usage

\[ gtoxObjCnst(p, \text{resp}) \]
\[ gtoxObjGnls(p, \text{lconc}, \text{resp}) \]
\[ gtoxObjHill(p, \text{lconc}, \text{resp}) \]

Arguments

\( p \)  
Numeric, the parameter values. See details for more information.

\( \text{resp} \)  
Numeric, the response values

\( \text{lconc} \)  
Numeric, the log10 concentration values

Details

These functions produce an estimated value based on the model and given parameters for each observation. Those estimated values are then used with the observed values and a scale term to calculate the log-likelihood.

Let \( t(z, \nu) \) be the Student’s t-distribution with \( \nu \) degrees of freedom, \( y_i \) be the observed response at the \( i^{th} \) observation, and \( \mu_i \) be the estimated response at the \( i^{th} \) observation. We calculate \( z_i \) as:

\[
z_i = \frac{y_i - \mu_i}{e^\sigma}
\]

where \( \sigma \) is the scale term. Then the log-likelihood is:

\[
\sum_{i=1}^{n} \left[ \ln(t(z_i, 4)) - \sigma \right]
\]

Where \( n \) is the number of observations.

Value

The log-likelihood.

Constant Model (cnst)

\[ gtoxObjCnst \] calculates the likelihood for a constant model at 0. The only parameter passed to \[ gtoxObjCnst \] by \( p \) is the scale term \( \sigma \). The constant model value \( \mu_i \) for the \( i^{th} \) observation is given by:

\[
\mu_i = 0
\]

Gain-Loss Model (gnls)

\[ gtoxObjGnls \] calculates the likelihood for a 5 parameter model as the product of two Hill models with the same top and both bottoms equal to 0. The parameters passed to \[ gtoxObjGnls \] by \( p \) are (in order) top \( (t_p) \), gain log AC50 \( (ga) \), gain hill coefficient \( (gw) \), loss log AC50 \( (la) \), loss hill coefficient \( (lw) \), and the scale term \( (\sigma) \). The gain-loss model value \( \mu_i \) for the \( i^{th} \) observation is given by:

\[
g_i = \frac{1}{1 + 10^{(ga-x_i)gw}}
\]
\[ l_i = \frac{1}{1 + 10^{(x_i - l_0)/l_w}} \]
\[ \mu_i = l_i (g_i) (l_i) \]

where \( x_i \) is the log concentration for the \( i^{th} \) observation.

### Hill Model (hill)

\( gtoxObjHill \) calculates the likelihood for a 3 parameter Hill model with the bottom equal to 0. The parameters passed to \( gtoxObjHill \) by \( p \) are (in order) top (\( tp \)), log AC50 (\( ga \)), hill coefficient (\( gw \)), and the scale term (\( \sigma \)). The hill model value \( \mu_i \) for the \( i^{th} \) observation is given by:

\[ \mu_i = \frac{tp}{1 + 10^{(ga-x_i)/gw}} \]

where \( x_i \) is the log concentration for the \( i^{th} \) observation.

### Examples

```r
## Load level 3 data for an assay endpoint ID
dat <- gtoxLoadData(lvl=3L, type="mc", fld="aeid", val=3L)

## Compute fitting log-likelyhood
gtoxObjCnst(1, dat$resp)

## Load level 3 data for an assay endpoint ID
dat <- gtoxLoadData(lvl=3L, type="mc", fld="aeid", val=2L)

## Compute fitting log-likelyhood
gtoxObjGnls(p=c(rep(0.5,5),1e-3), lconc=dat$logc, resp=dat$resp)

## Load level 3 data for an assay endpoint ID
dat <- gtoxLoadData(lvl=3L, type="mc", fld="aeid", val=3L)

## Compute fitting log-likelyhood
gtoxObjHill(c(rep(0,3), 1e-3), dat$logc, dat$resp)
```

### Description

This function is a wrapper to ease the creation of the dataframe containing data and metadata to be loaded in the database.

### Usage

```
prepareDatForDB(asid, dat)
```
registerMthd

Arguments

- `asid` Integer, the asid value(s) to assign the default methods to
- `dat` Data.table containing metadata and data to load in DB

Details

This function formats a dat table to be loaded in DB

Value

Data table with data and metadata to store in database

Examples

```r
## Not run:
## Load sample data
load(system.file("extdata", "data_for_vignette.rda", package="GladiaTOX"))

# Build assay table
assay <- buildAssayTab(plate, chnmap)

# Set study parameters
std.nm <- "SampleStudy" # study name
phs.nm <- "PhaseII" # study phase

# Delete previously loaded study data
asid = gtoxLoadAsid(fld=c("asnm", "asph"), val=list(std.nm, phs.nm))$asid
if(length(asid)>0){ deleteStudy(asid=asid) }

# Load annotation in gtoxDB
loadAnnot(plate, assay, NULL)

# Get the created study ID
asid = gtoxLoadAsid(fld = c("asnm", "asph"), val = list(std.nm, phs.nm))$asid

# Prepare and load data
dat <- prepareDatForDB(asid, dat)

## End(Not run)
```

Description

registerMthd registers a new analysis method to the gtox databases.
Usage

```
registerMthd(lvl, mthd, desc, nddr = 0L, type)
```

Arguments

- **lvl**: Integer of length 1, the level for the analysis method
- **mthd**: Character, the name of the method
- **desc**: Character, same length as mthd, the method description
- **nddr**: Integer, 0 or 1, 1 if the method requires loading the dose-response data
- **type**: Character of length 1, the data type, "sc" or "mc"

Details

'mthd' must match a corresponding function name in the functions that load the methods, ie. `mc2_mthds`. 'nddr' only applies to level 6 methods.

Value

None

---

**sc1**

*Perform level 1 single-concentration processing*

---

Description

`sc1` loads level 0 data from the gtox database for the given id and performs level 1 single-concentration processing. The processed data is then loaded into the sc1 table and all subsequent data is deleted with `gtoxCascade`. See details for more information.

The individual processing functions are no longer exported, as it is typically more convenient and suggested to use the `gtoxRun` wrapper function.

Usage

```
sc1(ac, wr = FALSE)
```

Arguments

- **ac**: Integer of length 1, assay component id (acid) for processing.
- **wr**: Logical, whether the processed data should be written to the gtox database

Details

Level 1 single-concentration processing includes mapping assay component to assay endpoint, duplicating the data when the assay component has multiple assay endpoints, and any normalization of the data. Data normalization based on methods listed in `sc1_aeid` and `sc1_methods` tables.
**Value**

A boolean of length 1, indicating the success of the processing, or when 'wr' is FALSE, a list where the first element is a boolean indicating the success of processing and the second element is a data.table containing the processed data.

**See Also**

Method functions, SC1_Methods

Other single-concentration data processing functions: sc2

---

**Description**

sc1_mthds returns a list of functions to be used during level 1 single-concentration processing.

**Usage**

```r
sc1_mthds()
```

**Details**

The functions contained in the list returned by sc1_mthds return a list of expressions to be executed in the sc2 (not exported) function environment. The functions are described here for reference purposes. The sc1_mthds function is not exported, nor is it intended for use.

All available methods are described in the Available Methods section, listed by the function/method name.

**Value**

A list functions

**Available Methods**

The methods are broken into three types, based on what fields they define. Different methods are used to define "bval" (the baseline value), "pval" (the positive control value), and "resp" (the final response value).

Although it does not say so specifically in each description, all methods are applied by acid.

More information about the level 3 single-concentration processing is available in the package vignette, "Pipeline_Overview."

**bval Methods:**

- **bval.apid.nwlls.med** Calculate bval as the median of rval for wells with wllt equal to "n," by apid.
- **bval.apid.twlls.med** Calculate bval as the median of rval for wells with wllt equal to "t," by apid.
bval.apid.tn.med  Calculate bval as the median of rval for wells with wllt equal to "t" or "n," by apid.

pval Methods:

pval.apid.pwlls.med  Calculate pval as the median of rval for wells with wllt equal to "p," by apid.

pval.apid.mwlls.med  Calculate pval as the median of rval for wells with wllt equal to "m," by apid.

pval.apid.medpcbyconc.max  First calculate the median of rval for wells with wllt equal to "p" or "c," by wllt, conc, and apid. Then calculate pval as the maximum of the calculated medians, by apid.

pval.apid.medpcbyconc.min  First calculate the median of rval for wells with wllt equal to "p" or "c," by wllt, conc, and apid. Then calculate pval as the minimum of the calculated medians, by apid.

pval.apid.medncbyconc.min  First calculate the median of rval for wells with wllt equal to "m" or "o," by wllt, conc, and apid. Then calculate pval as the minimum of the calculated medians, by apid.

pval.zero  Define pval as 0.

resp Methods:

resp.pc  Calculate resp as $\frac{rval - bval}{pval - bval}$ × 100.

resp.fc  Calculate resp as $\frac{rval}{bval}$.

resp.logfc  Calculate resp as $rval - bval$.

resp.log2  Take the logarithm of resp with base 2.

resp.multneg1  Multiply resp by -1.

none  Do no normalization; make resp equal to rval.

Note

This function is not exported and is not intended to be used by the user.

See Also

sc1, Method functions to query what methods get applied to each acid

sc2  Perform level 2 single-concentration processing

Description

sc2 loads level 1 data from the gtox database for the given id and performs level 2 single-concentration processing. The processed data is then loaded into the sc2 table and all subsequent data is deleted with gtoxCascade. See details for more information.

The individual processing functions are no longer exported, as it is typically more convenient and suggested to use the gtoxRun wrapper function.
Usage

```
sc2(ae, wr = FALSE)
```

Arguments

- `ae`: Integer of length 1, assay endpoint id (aeid) for processing.
- `wr`: Logical, whether the processed data should be written to the gtox database

Details

Level 2 single-concentration processing defines the bmad value, and uses the activity cutoff methods from `sc2_aeid` and `sc2_methods` to make an activity call.

Value

A boolean of length 1, indicating the success of the processing, or when `wr` is FALSE, a list where the first element is a boolean indicating the success of processing and the second element is a data.table containing the processed data.

See Also

- Method functions, `SC2_Methods`
- Other single-concentration data processing functions: `sc1`

---

**SC2_Methods**

List of level 2 single-concentration hit-call functions

---

Description

`sc2_mthds` returns a list of functions to be used during level 2 single-concentration processing.

Usage

```
sc2_mthds()
```

Details

The functions contained in the list returned by `sc2_mthds` return a list of expressions to be executed in the `sc2` (not exported) function environment. The functions are described here for reference purposes. The `sc2_mthds` function is not exported, nor is it intended for use.

All available methods are described in the Available Methods section, listed by the function/method name.

Value

A list functions
Available Methods

More information about the level 2 single-concentration processing is available in the package vignette, "Pipeline_Overview."

- **bmad3** Add a cutoff value of 3*bmad.
- **pc20** Add a cutoff value of 20.
- **log2_1.2** Add a cutoff value of log2(1.2).
- **log10_1.2** Add a cutoff value of log10(1.2).
- **bmad5** Add a cutoff value of 5*bmad.
- **bmad6** Add a cutoff value of 6*bmad.
- **bmad10** Add a cutoff value of 10*bmad.
- **pc30orbmad3** Add a cutoff value of either 30 or 3*bmad, whichever is less.

Note

This function is not exported and is not intended to be used by the user.

See Also

- `sc2`, Method functions to query what methods get applied to each acid

| sink_reset | Reset all sinks |

Description

`sink_reset` resets all sinks and returns all output to the console.

Usage

`sink_reset()`

Details

`sink_reset` identifies all sinks with `sink.number` then returns all output and messages back to the console.

Value

None

See Also

- `sink, sink.number`
- Other gtox abbreviations: `is.odd, lu, lw`
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