

# Package ‘MAST’

May 22, 2019

**Type** Package

**Title** Model-based Analysis of Single Cell Transcriptomics

**Version** 1.10.0

**Date** 2019-01-16

**VignetteBuilder** knitr

**Imports** Biobase, BiocGenerics, S4Vectors, data.table, ggplot2, plyr, stringr, abind, methods, parallel, reshape2, stats, stats4, graphics, utils, blme, SummarizedExperiment(>= 1.5.3), progress

**Depends** SingleCellExperiment (>= 1.2.0), R(>= 3.5)

**Suggests** knitr, rmarkdown, testthat, lme4(>= 1.0), roxygen2(> 4.0.0), numDeriv, car, gdata, lattice, GGally, GSEABase, NMF, TxDb.Hsapiens.UCSC.hg19.knownGene, rsvd, limma, RColorBrewer, BiocStyle, scater

**Description** Methods and models for handling zero-inflated single cell assay data.

**License** GPL(>= 2)

**Collate** 'AllGenerics.R' 'AllClasses.R' 'CovFromBoots.R' 'Fluidigm-methods.R' 'GSEA-by-boot.R' 'Hypothesis.R' 'LmWrapper.R' 'MultidimensionalScaling.R' 'RNASeqAssay-methods.R' 'Readers.R' 'SingleCellAssay-methods.R' 'UtilityFunctions.R' 'ZlmFit-bootstrap.R' 'ZlmFit-logFC.R' 'ZlmFit.R' 'bayesglm.R' 'convertMASTClassic.R' 'ebayes-helpers.R' 'filterEval.R' 'helper-methods.R' 'lmWrapper-bayesglm.R' 'lmWrapper-glm.R' 'lmWrapper-glmer.R' 'lmWrapper-ridge.R' 'lrtest.R' 'predict.R' 'stat\_ell.R' 'thresholdSCRNA.R' 'zeroinf.R' 'zlmHooks.R'

**RoxygenNote** 6.1.1

**LazyData** true

**biocViews** GeneExpression, DifferentialExpression, GeneSetEnrichment, RNASeq, Transcriptomics, SingleCell

**BugReports** <https://github.com/RGLab/MAST/issues>

**URL** <https://github.com/RGLab/MAST/>

**git\_url** <https://git.bioconductor.org/packages/MAST>

**git\_branch** RELEASE\_3\_9

**git\_last\_commit** 7223e4f

**git\_last\_commit\_date** 2019-05-02

**Date/Publication** 2019-05-21

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MAST-package

*MAST: Model-based Analysis of Single- cell Transcriptomics***Description**

Methods for analysing single cell assay data using hurdle models.

**Details**

This packages provides data structures and functions for statistical analysis of single-cell assay data such as Fluidigm single cell gene expression assays.

**Author(s)**

**Maintainer:** Andrew McDavid <Andrew\_McDavid@urmc.rochester.edu>

Authors:

- Greg Finak <gfinak@fredhutch.org>
- Masanao Yajima <myajima@fredhutch.org>

**References**

Finak, et al. MAST: a flexible statistical framework for assessing transcriptional changes and characterizing heterogeneity in single-cell RNA sequencing data. *Genome Biology* (2015).

**See Also**

Useful links:

- <https://github.com/RGLab/MAST/>
- Report bugs at <https://github.com/RGLab/MAST/issues>

---

applyFlat

*Apply a vectorized binary operation recycling over last dimension*

---

**Description**

When  $x$  is an array of order  $K$ , and  $y$  is an array of order  $K-1$ , whose dimensions otherwise agree, apply FUN by recycling  $y$  as necessary over dimension  $K$  of  $x$ .

**Usage**

```
applyFlat(x, y, FUN = "-")
```

**Arguments**

$x$	array, order $K$
$y$	array, order $K-1$
FUN	vectorized binary operation

**Value**

array, order  $K$  equal to FUN( $x,y$ )

**Examples**

```
##Dumb example, could be done with scale(...,scale=FALSE)
x0 = matrix(1:10, nrow=2)
y0 = rowMeans(x0)
dim(y0) = c(1, 2)
x1 = MAST:::applyFlat(x0,y0)
stopifnot(rowMeans(x1)==0)
```

---

BayesGLMlike-class	<i>Wrapper for bayesian GLM</i>
--------------------	---------------------------------

---

**Description**

Wrapper for bayesian GLM

**Slots**

prior numeric optional 3d array used to specify prior for coefficients

useContinuousBayes logical should bayesglm be used to fit the continuous component as well?

---

bootVcov1	<i>Bootstrap a zlmfit</i>
-----------	---------------------------

---

**Description**

Sample cells with replacement to find bootstrapped distribution of coefficients

**Usage**

```
bootVcov1(zlmfit, R = 99)
```

**Arguments**

zlmfit class ZlmFit

R number of bootstrap replicates

**Value**

array of bootstrapped coefficients

**Examples**

```
data(vbetaFA)
zlmVbeta <- zlm(~ Stim.Condition, subset(vbetaFA, ncells==1)[1:5,])
#Only run 3 boot straps, which you wouldn't ever want to do in practice...
bootVcov1(zlmVbeta, R=3)
```

---

 calcZ

*Get Z or T statistics and P values after running gseaAfterBoot*


---

## Description

The Z or T statistics may be reported by component (discrete/continuous) when combined='no' or combined by Fisher's or Stouffer's method (combined='fisher' or combined='stouffer'. Fisher's method uses the product of the p-values, while Stouffer's method uses the sum of the Z/T scores. The "Z" score returned by Fisher is the normal quantile that would yield the observed Fisher P-value, whose sign is derived from the sign of the maximum component Z score. The "Z" score returned by Stouffer when testType='normal' is the sum of the Z scores, over sqrt(2). When testType='t' it is a weighted combination of the Z scores, with weights corresponding to the degrees of freedom in each of the t statistics. A t-approximation to this sum of t-variables is derived by matching moments. It seems to be fairly accurate in practice.

## Usage

```
calcZ(gseaObj, testType = "t", combined = "none")
```

## Arguments

gseaObj	output from gseaAfterBoot
testType	either 'normal' or 't'. The 't' test adjusts for excess kurtosis due to the finite number of bootstrap replicates used to estimate the variance of the statistics. This will result in more conservative inference.
combined	character one of 'none', 'fisher' or 'stouffer'

## Value

3D array with dimensions set (modules) comp ('cont'iguous or 'disc'rete) and metric ('Z' stat and two sided 'P' value that  $P(z>|Z|)$ ) if combined='no', otherwise just a matrix.

## See Also

gseaAfterBoot

## Examples

```
## See the examples in gseaAfterBoot
example(gseaAfterBoot)
```

---

`cData`*Deprecated cell/feature data accessors/mutators*

---

### Description

These functions are now all deprecated and will be removed in a future release.

### Usage

```
cData(sc)
```

```
cData(sc) <- value
```

```
## S4 method for signature 'SingleCellAssay'  
cData(sc)
```

```
## S4 replacement method for signature 'SingleCellAssay'  
cData(sc) <- value
```

```
## S4 method for signature 'SingleCellAssay,SingleCellAssay'  
combine(x, y, ...)
```

```
## S4 method for signature 'SingleCellAssay,ANY'  
combine(x, y, ...)
```

### Arguments

<code>sc</code>	An object with <code>cellData</code>
<code>value</code>	replacement value
<code>x</code>	<code>SingleCellAssay</code>
<code>y</code>	<code>SingleCellAssay</code>
<code>...</code>	<code>SingleCellAssay</code>

### Details

`cData(sc)`: Return the `cellData` data.frame.

`cData(sc)<-value`: Replace the `cellData` with `value`, which can be either an `AnnotatedDataFrame` or `data.frame`. The replacement is checked that it has mandatory fields defined by its class.

`combine(x,y,...)`: Concatenate two experiments along rows/columns

### Value

`DataFrame` or modifies the `SingleCellAssay` object in place

### Replacement Functions

You should transition to use the following replacements:

`cData` [colData](#)

`fData` [mcols](#)

exprs [assay](#)  
 combine [cbind2](#) or [rbind2](#)

### See Also

[exprs](#)

### Examples

```
data(vbetaFA)
stopifnot(all.equal(hushWarning(cData(vbetaFA), 'deprecated'), colData(vbetaFA)))
stopifnot(all.equal(hushWarning(fData(vbetaFA), 'deprecated'), mcols(vbetaFA)))
stopifnot(all.equal(hushWarning(exprs(vbetaFA), 'deprecated'), t(assay(vbetaFA))))
```

---

`colData<-`, `SingleCellAssay`, `DataFrame`-method  
*Replace colData*

---

### Description

Replace `colData` with a `DataFrame`. Checks to make sure that `row.names(value)` match `colnames{x}`, in contrast to the parent method Checks for a `wellKey` column, as well.

### Usage

```
## S4 replacement method for signature 'SingleCellAssay,DataFrame'
colData(x) <- value
```

### Arguments

<code>x</code>	<code>SingleCellAssay</code>
<code>value</code>	<code>DataFrame</code>

### Value

modified `SingleCellAssay`

---

`collectResiduals`      *Residual hooks and collection methods*

---

### Description

After each gene is fit, a hook function can optionally be run and the output saved. This allows extended computations to be done using the fitted model, without keeping it in memory. Here this is used to calculate various residuals, though in some cases they can be done using only the information contained in the `ZlmFit`-class.



**Usage**

```

collectResiduals(x, sca, newLayerName = "Residuals")

discrete_residuals_hook(x)

continuous_residuals_hook(x)

combined_residuals_hook(x)

deviance_residuals_hook(x)

fitted_phat(x)

partialScore(x, effectRegex)

```

**Arguments**

x	ZlmFit-class
sca	SingleCellAssay object to which the residuals should be added
newLayerName	character name of the assay layer
effectRegex	a regular expression naming columns of the design corresponding to $Z_0$ . Generally these should be the treatment effects of interest.

**Value**

copy of sca with new layer

**Functions**

- `discrete_residuals_hook`: Hook to get the discrete residuals, ie, difference between expected probability of expression and observed
- `continuous_residuals_hook`: Hook to get the continuous residuals, ie, residuals for conditionally positive observations. If an observation is zero, it's residual is defined to be zero as well.
- `combined_residuals_hook`: Hook to get the combined residuals, ie,  $Y - E(U) * E(V)$
- `deviance_residuals_hook`: Standardized deviance residuals hook. Computes the sum of the standardized deviance residuals for the discrete and continuous models (scaled to have unit variance). If the observation is zero then only the discrete component is used.
- `fitted_phat`: Hook to return `p_hat`, the predicted probability of expression.
- `partialScore`: Compute  $Y_i - E(V_i | X_i, Z_0) E(U | X_i, Z_0)$ , where  $Z_0$  is a treatment effect (being left in) and  $X_i$  is a nuisance effect (being regressed out).

**Total residual types**

Each component of the model contributes several flavors of residual, which can be combined in various fashions. The discrete residual can be on the response scale (thus subtracting the predicted probability of expression from the 0/1 expression value). Or it can be a deviance residual, revealing something about the log-likelihood.

**Partial residuals**

It's also possible to consider partial residuals, in which the contribution of a particular covariate is added back into the model.

**See Also**

zlm

**Examples**

```
data(vbetaFA)
svbeta <- subset(vbetaFA, ncells==1)
svbeta <- svbeta[freq(svbeta)>.4,]
window <- function(x1) lapply(assays(x1), function(x2) x2[1:3, 1:6])
#total residuals of the response
z1 <- zlm(~ Stim.Condition, svbeta, hook=discrete_residuals_hook)
window(collectResiduals(z1, svbeta))
z2 <- zlm(~ Stim.Condition, svbeta, hook=continuous_residuals_hook)
window(collectResiduals(z2, svbeta))
z3 <- zlm(~ Stim.Condition, svbeta, hook=combined_residuals_hook)
window(collectResiduals(z3, svbeta))
#total deviance residuals
z4 <- zlm(~ Stim.Condition, svbeta, hook=deviance_residuals_hook)
window(collectResiduals(z4, svbeta))
#partial residuals
colData(svbeta)$ngeneson <- colMeans(assay(svbeta)>0)
z5 <- zlm(~ Stim.Condition + ngeneson, svbeta)
partialScore(z5, 'Stim.Condition')
```

---

computeEtFromCt

*Compute the Et from the Ct*

---

**Description**

Computes the Et value from the Ct value in an existing data frame and returns a new data frame with the Et column appended

**Usage**

```
computeEtFromCt(df, column = "Ct", Cmax = 40)
```

**Arguments**

df	a data.frame
column	The name of the Ct column. A character. 'Ct' by default.
Cmax	the maximum number of cycles performed. 40 by default.

**Value**

A copy of df with the 'Et' column appended

**Author(s)**

Greg Finak

**Examples**

```
data(vbeta)
vbeta <- computeEtFromCt(vbeta)
```

---

```
convertMASTClassicToSingleCellAssay
      Convert a MASTClassic SingleCellAssay
```

---

**Description**

Convert a `SingleCellAssay` object created with the `MASTClassic` package to an object recognized by the new `MAST` package

**Usage**

```
convertMASTClassicToSingleCellAssay(object = NULL)
```

**Arguments**

`object` of class `SingleCellAssay` created by `MASTClassic`

**Details**

The function will extract the relevant information from the attributes of the old object and construct a new `SingleCellAssay` that is recognized by `MAST`. This function checks that the object is a `MASTClassic SingleCellAssay` object. It will stop if it is not a `SingleCellAssay`, return a converted `SingleCellAssay` if object was created by `MASTClassic`, and return the original object if the object is already compatible.

**Value**

A `MAST SingleCellAssay` object.

**Note**

Type checking for old object is not performed.

**Examples**

```
data(vbetaFA)
convertMASTClassicToSingleCellAssay(vbetaFA)
```

---

CovFromBoots	<i>Extract the intere-gene covariance matrices for continuous and discrete components of a MAST model for a given coefficient from bootstrap replicates</i>
--------------	---

---

### Description

Computes the genewise covariance for a model coefficient from bootstrap replicates from `'MAST::bootVcov1()'`. If coefficients are unestimable (i.e. NA) for a gene, that row/column in the covariance matrix will be NA. Returns a list with components "C" and "D" containing the covariance matrices for the "C"ontinuous and "D"iscrete components of the MAST model.

### Usage

```
CovFromBoots(boots = NULL, coefficient = NULL)
```

### Arguments

boots	a multidimensional array returned by <code>'bootVcov1'</code> or <code>'pbootVcov1'</code> .
coefficient	<code>'character'</code> the name of the model coefficient for which to return the inter-gene covariance matrices.

### Value

list with components "C" and "D" containing covariance matrices for the continuous and discrete components of the mode.

---

defaultPrior	<i>Initialize a prior to be used a prior for BayeGLMlike/BayesGLMlike2</i>
--------------	--

---

### Description

Initialize a prior to be used a prior for BayeGLMlike/BayesGLMlike2

### Usage

```
defaultPrior(names)
```

### Arguments

names	character vector of coefficients. The <code>'(Intercept)'</code> will be ignored.
-------	---

### Value

3d array, with leading dimension giving the prior `'loc'`ation, `'scale'` and degrees of freedom (df), second dimension giving the component (`'C'`ontinuous or `'D'`iscrete) and trailing dimension giving the coefficient to which the prior applies. The location is initialized to be 0, the scale to 2, and degrees of freedom of 1, following the default of `bayesglm`.

**Examples**

```

dp <- defaultPrior('Stim.ConditionUnstim')
## Not run:
data(vbetaFA)
zlmVbeta <- zlm(~ Stim.Condition, vbeta.sc, method='bayesglm', coefPrior=dp)

## End(Not run)

```

---

dof *Degrees of freedom of Zero inflated model*

---

**Description**

Degrees of freedom of Zero inflated model

**Usage**

```
dof(object)
```

**Arguments**

object           LMlike or subclass

**Value**

vector giving the model degrees of freedom for continuous and discrete

---

Drop *Drop specified dimension from an array*

---

**Description**

Like `drop(x)` but only dropping specified dimensions. There is no testing that the specified dimensions are actually singletons.

**Usage**

```
Drop(x, d)
```

**Arguments**

x                   array of at least d dimensions  
d                   dimension(s) to drop

**Value**

array x

**Examples**

```
x = array(1:4, dim=c(1, 2, 1, 2))
dx = MAST:::Drop(x, 1)
stopifnot(all(dim(dx)==c(2,1,2)))
```

---

ebayes	<i>Estimate hyperparameters for hierarchical variance model for continuous component</i>
--------	--

---

**Description**

ebayesControl is a named list with (optional) components 'method' (one of 'MOM' or 'MLE') and 'model' (one of 'H0' or 'H1') method MOM uses a method-of-moments estimator, while MLE using the marginal likelihood. H0 model estimates the precisions using the intercept alone in each gene, while H1 fits the full model specified by formula

**Usage**

```
ebayes(sca, ebayesControl, Formula, truncate = Inf)
```

**Arguments**

sca	SingleCellAssay
ebayesControl	list with (optional) components 'method', 'model'. See details.
Formula	a formula (using variables in colData(sca) used when model='H1').
truncate	Genes with sample precisions exceeding this value are discarded when estimating the hyper parameters

**Value**

numeric of length two, giving the hyperparameters in terms of a variance ( $v$ ) and prior observations ( $df$ ), inside a structure, with component hess, giving the Fisher Information of the hyperparameters.

---

expavg	<i>Exponential average</i>
--------	----------------------------

---

**Description**

Puts log transformed values onto natural scale and takes mean of vector. Calculates  $\text{mean}(2^x - 1)$

**Usage**

```
expavg(x)
```

**Arguments**

x	numeric
---	---------

**Value**

numeric

**Examples**

```
x <- 1:10
logmean(expavg(x))
```

---

<i>fData</i>	<i>fData</i>
--------------	--------------

---

**Description**

Accessor for featureData data.frame

**Arguments**

object            An object with featureData

**Details**

Returns the featureData data.frame.

**Value**

data.frame

---

featureData	<i>Accessor for featureData AnnotatedDataFrame</i>
-------------	--

---

**Description**

Returns the featureData.

**Arguments**

object            An object with featureData

**Value**

AnnotatedDataFrame

---

```
filterLowExpressedGenes
```

*Filter low-expressing genes*

---

### Description

Filter out genes that have less than some percent threshold expression across all libraries

### Usage

```
filterLowExpressedGenes(assay, threshold = 0.1)
```

### Arguments

assay	a SingleCellAssay object
threshold	a numeric between 0, and 1, specifying the threshold frequency below which genes will be filtered out

### Value

SingleCellAssay

### Examples

```
data(vbetaFA)
filterLowExpressedGenes(vbetaFA)
```

---

```
fit
```

*fit a zero-inflated regression*

---

### Description

Given a design and formula, fit the zero inflated regression, storing the fits in slots fitC and fitD

### Usage

```
fit(object, response, ...)

## S4 method for signature 'LMERlike,missing'
fit(object, response, silent = TRUE, ...)
```

### Arguments

object	inheriting from LMlike
response	a vector, same length as the design, or if missing then use the current response
...	currently ignored
silent	mute some warnings emitted from the underlying modeling functions

### Value

LMlike or subclass



---

`freq`*Summary statistics for genes in an experiment*

---

## Description

`freq` returns the frequency of expression, i.e., the proportion of non-zero values in `sc`. NAs can be optionally removed

## Usage

```
freq(sc, na.rm = TRUE)
```

```
condmean(sc)
```

```
condSd(sc)
```

```
numexp(sc)
```

## Arguments

`sc`                    `SingleCellAssay`

`na.rm`                should NAs be removed, or carried through?

## Value

vector of proportions

## Functions

- `condmean`: Report the mean non-zero expression value for each gene. NAs are always removed.
- `condSd`: Report standard deviation of expression, for positive et for each gene. NAs are always removed.
- `numexp`: Report number of expressing cells ( $>0$ ) per gene. NAs are removed.

## Examples

```
data(vbetaFA)
freq(vbetaFA)
condmean(vbetaFA)
```

---

FromFlatDF	<i>Construct a SingleCellAssay (or derived subclass) from a 'flat' (melted) data.frame/data.table</i>
------------	---

---

## Description

SingleCellAssay are a generic container for such data and are simple wrappers around SummarizedExperiment objects. Subclasses exist that imbue the container with additional attributes, eg [FluidigmAssay](#).

## Usage

```
FromFlatDF(dataframe, idvars, primerid, measurement, id = numeric(0),
  cellvars = NULL, featurevars = NULL, phenovars = NULL,
  class = "SingleCellAssay", check_sanity = TRUE, ...)
```

```
FluidigmAssay(...)
```

## Arguments

dataframe	A 'flattened' data.frame or data.table containing columns giving cell and feature identifiers and a measurement column
idvars	character vector naming columns that uniquely identify a cell
primerid	character vector of length 1 that names the column that identifies what feature (i.e. gene) was measured
measurement	character vector of length 1 that names the column containing the measurement
id	An identifier (eg, experiment name) for the resulting object
cellvars	Character vector naming columns containing additional cellular metadata
featurevars	Character vector naming columns containing additional feature metadata
phenovars	Character vector naming columns containing additional phenotype metadata
class	desired subclass of object. Default SingleCellAssay.
check_sanity	(default: TRUE) Set FALSE to override sanity checks that try to ensure that the default assay is log-transformed and has at least one exact zero. See <a href="#">defaultAssay</a> for details on the "default assay" which is assumed to contain log transformed data.
...	additional arguments are ignored

## Value

SingleCellAssay, or derived, object

## Examples

```
data(vbeta)
colnames(vbeta)
vbeta <- computeEtFromCt(vbeta)
vbeta.fa <- FromFlatDF(vbeta, idvars=c("Subject.ID", "Chip.Number", "Well"),
  primerid='Gene', measurement='Et', ncells='Number.of.Cells',
  geneid="Gene", cellvars=c('Number.of.Cells', 'Population'),
```

```
phenovars=c('Stim.Condition','Time'), id='vbeta all', class='FluidigmAssay')
show(vbeta.fa)
nrow(vbeta.fa)
ncol(vbeta.fa)
head(mcols(vbeta.fa)$primerid)
table(colData(vbeta.fa)$Subject.ID)
vbeta.sub <- subset(vbeta.fa, Subject.ID=='Sub01')
show(vbeta.sub)
```

---

FromMatrix

*Construct a SingleCellAssay from a matrix or array of expression*


---

## Description

If the gene expression measurements are already in a rectangular form, then this function allows an easy way to construct a SingleCellAssay object while still doing some sanity checking of inputs.

## Usage

```
FromMatrix(exprsArray, cData, fData, class = "SingleCellAssay",
  check_sanity = TRUE, check_logged = check_sanity)
```

## Arguments

exprsArray	matrix, or a list of matrices, or an array. Columns are cells, rows are genes.
cData	cellData an object that can be coerced to a DataFrame, ie, data.frame, AnnotatedDataFrame. Must have as many rows as ncol(exprsArray)
fData	featureData an object that can be coerced to a DataFrame, ie, data.frame, AnnotatedDataFrame. Must have as many rows as nrow(exprsArray).
class	desired subclass of object. Default SingleCellAssay.
check_sanity	(default: TRUE) Set FALSE to override sanity checks that try to ensure that the default assay is log-transformed and has at least one exact zero. See <a href="#">defaultAssay</a> for details on the "default assay" which is assumed to contain log transformed data.
check_logged	alias for check_sanity

## Value

an object of class class

## See Also

[defaultAssay](#)

**Examples**

```

ncells <- 10
ngenes <- 5
fData <- data.frame(primerid=LETTERS[1:ngenes])
cData <- data.frame(wellKey=seq_len(ncells))
mat <- matrix(rnorm(ncells*ngenes), nrow=ngenes)
sca <- FromMatrix(mat, cData, fData)
stopifnot(inherits(sca, 'SingleCellAssay'))
stopifnot(inherits(sca, 'SummarizedExperiment'))
##If there are mandatory keywords expected by a class, you'll have to manually set them yourself
cData$ncells <- 1
fd <- FromMatrix(mat, cData, fData)
stopifnot(inherits(fd, 'SingleCellAssay'))

```

---

getConcordance

*Get the concordance between two experiments*


---

**Description**

Return the concordance between two assays (i.e. single cell and hundred cell). The "average" of singleCellRef (after adjusting for the number of cells) and singleCellComp are taken per gene, per groups. A data.frame with one row per gene-groups is returned with some additional columns.

**Usage**

```

getConcordance(singleCellRef, singleCellcomp, groups = NULL,
  fun.natural = expavg, fun.cycle = logmean)

```

```

getwss(concord, nexp)

```

```

getss(concord)

```

```

getrc(concord)

```

**Arguments**

```

singleCellRef  "reference" SingleCellAssay
singleCellcomp "comparison" SingleCellAssay
groups         character vector giving variable(s) on which the comparison is conditioned
fun.natural    function to transform the SingleCellAssays to a mRNA proportional level
fun.cycle      inverse function of fun.natural
concord        data.frame returned by getConcordance
nexp           number of expressed cells per row in concord

```

**Value**

concordance between two assays

**Functions**

- getwss: getrc the sum of squares, weighted by nexp
- getss: return the sum of squares
- getrc: Return Lin's (1989) concordance correlation coefficient

**Author(s)**

Andrew McDavid

**See Also**

[plotSCAConcordance](#)

**Examples**

```
data(vbetaFA)
sca1 <- subset(vbetaFA, ncells==1)
sca100 <- subset(vbetaFA, ncells==100)
concord <- getConcordance(sca1, sca100)
getss(concord)
getrc(concord)
```

---

getwellKey

*Accessor for wellKey*

---

**Description**

This returns the wellKey, which is a unique identifier generated by idvars in the mapping

**Usage**

```
getwellKey(sc)
```

**Arguments**

sc                    An object with a wellKey

**Value**

integer giving the unique id generated

**Examples**

```
data(vbetaFA)
getwellKey(vbetaFA)
colData(vbetaFA)$wellKey
```

---

GLMlike-class	<i>Wrapper for regular glm/lm</i>
---------------	-----------------------------------

---

**Description**

Wrapper for regular glm/lm

**Usage**

```
## S4 method for signature 'GLMlike'
vcov(object, which, ...)
```

**Arguments**

object	GLMlike
which	character, one of 'C', 'D'.
...	ignored

**Value**

covariance matrix

**Methods (by generic)**

- `vcov`: return the variance/covariance of component which

**Slots**

`weightFun` function to map expression values to probabilities of expression. Currently unused.

---

gseaAfterBoot	<i>Gene set analysis for hurdle model</i>
---------------	---

---

**Description**

Modules defined in sets are tested for average differences in expression from the "average" gene. By using bootstraps, the between-gene covariance of terms in the hurdle model is found, and is used to adjust for coexpression between genes. We drop genes if the coefficient we are testing was not estimable in original model fit in `zFit` or in any of the bootstrap replicates (evidenced an NA in the bootstrap array). This might yield overly conservative inference. Since bootstrapping is a randomized procedure, the degrees of freedom of a module (and its variance parameters) might differ from run-to-run. You might try setting `var_estimate='modelbased'` to relax this requirement by assuming independence between genes and then using the asymptotic covariance estimates, which are deterministic, but may result in overly-generous inference.

**Usage**

```
gseaAfterBoot(zFit, boots, sets, hypothesis,
  control = gsea_control(n_randomize = Inf, var_estimate = "bootall"))

gsea_control(n_randomize = Inf, var_estimate = "bootall")
```

**Arguments**

zFit	object of class ZlmFit
boots	bootstraps of zFit
sets	list of indices of genes
hypothesis	a Hypothesis to test. Currently only one degree CoefficientHypothesis are supported.
control	parameters as provided by gsea_control. See details.
n_randomize	the number of genes to sample to approximate the non-module average expression. Set to Inf to turn off the approximation (the default).
var_estimate	the method used to estimate the variance of the modules, one of bootall, bootdiag, or modelbased.

**Value**

Object of class GSEATests, containing slots tests, 4D array and bootR, the number of bootstrap replicates.

**Functions**

- `gsea_control`: set control parameters. See Details.

**control**

`control` is a list with elements:

- `n_randomize`, giving the number of genes to sample to approximate the non-module average expression. Set to Inf to turn off the approximation (the default).
- `var_estimate`, giving the method used to estimate the variance of the modules. `bootall` uses the bootstrapped covariance matrices. `bootdiag` uses only the diagonal of the bootstrapped covariance matrix (so assuming independence across genes). `modelbased` assumes independence across genes and uses the variance estimated from the model.

**Return Value**

A 4D array is returned, with dimensions "set" (each module), "comp" ('disc'rete or 'cont'inuous), "metric" ('stat' gives the average of the coefficient, 'var' gives the variance of that average, 'dof' gives the number of genes that were actually tested in the set), "group" ('test' for the genes in test-set, "null" for all genes outside the test-set).

**See Also**

[calcZ](#)

`summary,GSEATests-method`

**Examples**

```
data(vbetaFA)
vb1 = subset(vbetaFA, ncells==1)
vb1 = vb1[,freq(vb1)>.1][1:15,]
zf = zlm(~Stim.Condition, vb1)
boots = bootVcov1(zf, 5)
sets = list(A=1:5, B=3:10, C=15, D=1:5)
```

```

gsea = gseaAfterBoot(zf, boots, sets, CoefficientHypothesis('Stim.ConditionUnstim'))
## Use a model-based estimate of the variance/covariance.
gsea_mb = gseaAfterBoot(zf, boots, sets, CoefficientHypothesis('Stim.ConditionUnstim'),
control = gsea_control(var_estimate = 'modelbased'))
calcZ(gsea)
summary(gsea)

```

---

GSEATests-class      *An S4 class for Gene Set Enrichment output*

---

### Description

This holds output from a call to `gseaAfterBoot`. It primarily provides a summary method.

### Slots

`tests` array: gene sets X discrete,continuous X stat, variance, degrees of freedom, avg correlation  
X test, null  
`bootR` number of bootstrap replicates

### See Also

`gseaAfterBoot`  
`calcZ`  
`summary,GSEATests-method`

---

hushWarning      *Selectively muffle warnings based on output*

---

### Description

Selectively muffle warnings based on output

### Usage

```
hushWarning(expr, regexp)
```

### Arguments

`expr`            an expression  
`regexp`          a regexp to be matched (with `str_detect`)

### Value

the result of `expr`

### Examples

```

hushWarning(warning('Beware the rabbit'), 'rabbit')
hushWarning(warning('Beware the rabbit'), 'hedgehog')

```



---

Hypothesis	<i>Describe a linear model hypothesis to be tested</i>
------------	--

---

**Description**

A Hypothesis can be any linear combination of coefficients, compared to zero. Specify it as a character vector that can be parsed to yield the desired equalities ala `makeContrasts`. A `CoefficientHypothesis` is a hypothesis for which terms are singly or jointly tested to be zero (generally the case in a t-test or F-test), by dropping coefficients from the model.

**Usage**

```
Hypothesis(hypothesis, terms)
```

**Arguments**

<code>hypothesis</code>	a character vector specifying a hypothesis, following <code>makeContrasts</code> , or a character vector naming coefficients to be dropped.
<code>terms</code>	an optional character vector giving the terms (column names from the <code>model.matrix</code> ) out of which the contrasts will be contrasted. If missing then most functions will attempt to fill this in for you at run time.

**Value**

a Hypothesis with a "transformed" component

**See Also**

`zlm` `waldTest` `lrTest`

**Examples**

```
h <- Hypothesis('Stim.ConditionUnstim', c('(Intercept)', 'Stim.ConditionUnstim'))
h@contrastMatrix
```

---

<code>impute</code>	<i>impute missing continuous expression for plotting</i>
---------------------	--

---

**Description**

If there are no positive observations for a contrast, it is generally not estimable. However, for the purposes of testing we can replace it with the least favorable value with respect to the contrasts that are defined.

**Usage**

```
impute(object, groupby)
```

**Arguments**

object	Output of predict
groupby	Variables (column names in predict) to group by for imputation (facets of the plot)

**Value**

data.table

**Examples**

```
##See stat_ell
example(stat_ell)
```

---

influence.bayesglm	<i>Influence bayesglm object</i>
--------------------	----------------------------------

---

**Description**

The influence function

**Usage**

```
## S3 method for class 'bayesglm'
influence(model, do.coef = TRUE, ...)
```

**Arguments**

model	bayesglm
do.coef	see <a href="#">influence.glm</a>
...	ignored

**Value**

see [influence.glm](#)

---

invlogit	<i>Inverse of logistic transformation</i>
----------	---

---

**Description**

Inverse of logistic transformation

**Usage**

```
invlogit(x)
```

**Arguments**

x	numeric
---	---------

**Value**

numeric

**Examples**

```
x <- 1:5
invlogit(log(x/(1-x)))
```

---

LMERlike-class	<i>Wrapper for lmer/glmer</i>
----------------	-------------------------------

---

**Description**

A horrendous hack is employed in order to do arbitrary likelihood ratio tests: the model matrix is built, the names possibly mangled, then fed in as a symbolic formula to glmer/lmer. This is necessary because there is no (easy) way to specify an arbitrary fixed-effect model matrix in glmer.

**Usage**

```
## S4 method for signature 'LMERlike'
update(object, formula., design, ...)

## S4 method for signature 'LMERlike'
vcov(object, which, ...)

## S4 method for signature 'LMERlike'
coef(object, which, singular = TRUE, ...)

## S4 method for signature 'LMERlike'
logLik(object)
```

**Arguments**

object	LMERlike
formula.	formula
design	something coercible to a data.frame
...	In the case of vcov, ignored. In the case of update, passed to model.matrix.
which	character, one of 'C', 'D'.
singular	logical. Should NA coefficients be returned?

**Value**

see the section "Methods (by generic)"

**Methods (by generic)**

- update: update the formula or design matrix
- vcov: return the variance/covariance of component which
- coef: return the coefficients. The horrendous hack is attempted to be undone.
- logLik: return the log-likelihood

**Slots**

pseudoMM part of this horrendous hack.  
 strictConvergence logical (default: TRUE) return results even when the optimizer or \*lmer complains about convergence  
 optimMsg character record warnings from lme. NA\_character\_ means no warnings.

---

LMlike-class	<i>Linear Model-like Class</i>
--------------	--------------------------------

---

**Description**

Wrapper around modeling function to make them behave enough alike that Wald tests and Likelihood ratio are easy to do. To implement a new type of zero-inflated model, extend this class. Depending on how different the method is, you will definitely need to override the fit method, and possibly the model.matrix, model.matrix<-, update, coef, vcov, and logLik methods.

**Usage**

```
## S4 method for signature 'LMlike'
summary(object)

## S4 method for signature 'LMlike'
update(object, formula., design, ...)

## S4 method for signature 'LMlike,CoefficientHypothesis'
waldTest(object, hypothesis)

## S4 method for signature 'LMlike,matrix'
waldTest(object, hypothesis)

## S4 method for signature 'LMlike,character'
lrTest(object, hypothesis)

## S4 method for signature 'LMlike,CoefficientHypothesis'
lrTest(object, hypothesis)

## S4 method for signature 'LMlike,Hypothesis'
lrTest(object, hypothesis)

## S4 method for signature 'LMlike,matrix'
lrTest(object, hypothesis)

## S4 method for signature 'GLMlike'
logLik(object)
```

**Arguments**

object	LMlike
formula.	formula
design	something coercible to a data.frame

... passed to `model.matrix`  
 hypothesis one of a `CoefficientHypothesis`, `Hypothesis` or contrast matrix.

### Value

see section "Methods (by generic)"

### Methods (by generic)

- `summary`: Print a summary of the coefficients in each component.
- `update`: update the formula or design from which the `model.matrix` is constructed
- `waldTest`: Wald test dropping single term specified by `CoefficientHypothesis hypothesis`
- `waldTest`: Wald test of contrast specified by contrast matrix hypothesis
- `lrTest`: Likelihood ratio test dropping entire term specified by character hypothesis naming a term in the symbolic formula.
- `lrTest`: Likelihood ratio test dropping single term specified by `CoefficientHypothesis hypothesis`
- `lrTest`: Likelihood ratio test dropping single term specified by `Hypothesis hypothesis`
- `lrTest`: Likelihood ratio test dropping single term specified by contrast matrix hypothesis
- `logLik`: return the log-likelihood of a fitted model

### Slots

**design** a `data.frame` from which variables are taken for the right hand side of the regression  
**fitC** The continuous fit  
**fitD** The discrete fit  
**response** The left hand side of the regression  
**fitted** A logical with components "C" and "D", TRUE if the respective component has converged  
**formula** A formula for the regression  
**fitArgsC**  
**fitArgsD** Both lists giving arguments that will be passed to the fitter (such as convergence criteria or case weights)

### See Also

`coef`  
`lrTest`  
`waldTest`  
`vcov`  
`logLik`

---

logFC	<i>Calculate log-fold changes from hurdle model components</i>
-------	--

---

### Description

Using the delta method, estimate the log-fold change from a state given by a vector `contrast0` and the state(s) given by `contrast1`.

### Usage

```
logFC(zlmfit, contrast0, contrast1)

getLogFC(zlmfit, contrast0, contrast1)
```

### Arguments

<code>zlmfit</code>	ZlmFit output
<code>contrast0</code>	vector of coefficients giving baseline contrast, or a <a href="#">Hypothesis</a> . If missing, then the '(Intercept)' is used as baseline.
<code>contrast1</code>	matrix of coefficients giving comparison contrasts, or a <a href="#">Hypothesis</a> . If missing, then all non-(Intercept) coefficients are compared.

### Details

The log-fold change is defined as follows. For each gene, let  $u(x)$  be the expected value of the continuous component, given a covariate  $x$  and the estimated coefficients `coefC`, ie,  $u(x) = \text{crossprod}(x, \text{coefC})$ . Likewise, Let  $v(x) = 1/(1+\exp(-\text{crossprod}(\text{coefD}, x)))$  be the expected value of the discrete component. The log fold change from `contrast0` to `contrast1` is defined as

$$u(\text{contrast1})v(\text{contrast1}) - u(\text{contrast0})v(\text{contrast0}).$$

Note that for this to be a log-fold change, then the regression for  $u$  must have been fit on the log scale. This is returned in the matrix `logFC`. An approximation of the variance of `logFC` (applying the delta method to formula defined above) is provided in `varLogFC`.

### Value

list of matrices 'logFC' and 'varLogFC', giving the log-fold-changes for each contrast (columns) and genes (rows) and the estimated sampling variance thereof

### Functions

- `getLogFC`: Return results as a perhaps friendlier `data.table`

### Caveats

1. When `method='bayesglm'` (the default), it's no longer necessarily true that the log fold change from condition A to B will be the inverse of the log fold change from B to A if the models are fit separately. This is due to the shrinkage in `bayesglm`.
2. The log fold change can be small, but the Hurdle p-value small and significant when the sign of the discrete and continuous model components are discordant so that the marginal log fold change

cancels out. The large sample sizes present in many single cell experiments also means that there is substantial power to detect even small changes.

3. When there is no expression in a gene for a coefficient that is non-zero in either condition0 or condition1 we return NA because there is not any information to estimate the continuous component. Technically we might return plus or minus infinity, but there is not a straightforward way to estimate a confidence interval in any case. See <https://support.bioconductor.org/p/99244/> for details

### See Also

[Hypothesis](#)

[summary,ZlmFit-method](#)

### Examples

```
data(vbetaFA)
zz <- zlm(~ Stim.Condition+Population, vbetaFA[1:5,])
##log-fold changes in terms of intercept (which is Stim(SEB) and CD154+VbetaResponsive)
lfcStim <- logFC(zz)
##If we want to compare against unstim, we can try the following
coefnames <- colnames(coef(zz, 'D'))
contrast0 <- setNames(rep(0, length(coefnames)), coefnames)
contrast0[c('(Intercept)', 'Stim.ConditionUnstim')] <- 1
contrast1 <- diag(length(coefnames))
rownames(contrast1)<-colnames(contrast1)<-coefnames
contrast1['(Intercept)',]<-1
lfcUnstim <- logFC(zz, contrast0, contrast1)
##log-fold change with itself is 0
stopifnot(all(lfcUnstim$logFC[,2]==0))
##inverse of log-fold change with Stim as reference
stopifnot(all(lfcStim$logFC[,1]==(-lfcUnstim$logFC[,1])))
##As a data.table:
getLogFC(zz)
```

---

logmean

*Log mean*

---

### Description

Takes mean of natural scaled values and then logarithm Approximately the inverse operation of [expavg](#) Calculates  $\log_2(\text{mean}(x) + 1)$

### Usage

```
logmean(x)
```

### Arguments

x                    numeric

### Value

numeric

**Examples**

```
x <- 1:10
expavg(logmean(x))
```

LRT

*Likelihood Ratio Tests for SingleCellAssays***Description**

Tests for a change in ET binomial proportion or mean of positive ET Likelihood Ratio Test for SingleCellAssay objects

**Usage**

```
LRT(sca, comparison, ...)

## S4 method for signature 'SingleCellAssay,character'
LRT(sca, comparison,
    referent = NULL, groups = NULL, returnall = FALSE)
```

**Arguments**

sca	A SingleCellAssay class object
comparison	A character specifying the factor for comparison
...	ignored
referent	A character specifying the reference level of comparison.
groups	A optional character specifying a variable on which to stratify the test. For each level of groups, there will be a separate likelihood ratio test.
returnall	A logical specifying if additional rows should be returned with information about the different components of the test.

**Details**

Combined Likelihood ratio test (binomial and normal) for SingleCellAssay and derived objects. This function is deprecated, please use [lrTest](#) instead.

**Value**

data.frame

**See Also**

zlm ZlmFit

**Examples**

```
data(vbetaFA)
LRT(vbetaFA, 'Stim.Condition', 'Unstim')
```



---

lrTest	<i>Run a likelihood-ratio test</i>
--------	------------------------------------

---

**Description**

Compares the change in likelihood between the current model and one subject to contrasts tested in hypothesis. hypothesis can be one of a character giving complete factors or terms to be dropped from the model, CoefficientHypothesis giving names of coefficients to be dropped, Hypothesis giving contrasts using the symbolically, or a contrast matrix, with one row for each coefficient in the full model, and one column for each contrast being tested.

**Usage**

```
lrTest(object, hypothesis)
```

**Arguments**

object	LMlike or subclass
hypothesis	the hypothesis to be tested. See details.

**Value**

array giving test statistics

**See Also**

fit  
waldTest  
Hypothesis  
CoefficientHypothesis

**Examples**

```
#see ZlmFit-class for examples
example('ZlmFit-class')
```

---

lrTest, ZlmFit, character-method	<i>Likelihood ratio test</i>
----------------------------------	------------------------------

---

**Description**

A 3D array with first dimension being the genes, next dimension giving information about the test (the degrees of freedom, Chisq statistic, and P value), and final dimension being the value of these quantities on the discrete, continuous and hurdle (combined) levels.

**Usage**

```
## S4 method for signature 'ZlmFit,character'
lrTest(object, hypothesis)
```

**Arguments**

object	ZlmFit
hypothesis	See Details

**Value**

3D array

---

magic_assay_names	<i>Default assay returned</i>
-------------------	-------------------------------

---

**Description**

Methods in this package operate on log-transformed (multiplicative scale) expression. We attempt to check for this at construction, and then over-ride the assay method to return the "layer" containing such log-transformed data.

**Usage**

```
magic_assay_names()

## S4 method for signature 'SingleCellAssay,missing'
assay(x, i, ...)
```

**Arguments**

x	SingleCellAssay
i	must be missing for this method to apply
...	passed to parent method

**Details**

By default we return the assay whose names, as given by `assayNames(x)`, matches the first element in the vector `c('thresh', 'et', 'Et', 'lCount', 'logTPM', 'logCounts', 'logcounts')`.

**Functions**

- `magic_assay_names`: list of names assumed to represent log-transformed data, in order of usage preference

**Examples**

```
data(vbetaFA)
assay(vbetaFA)[1:3, 1:3]
assay(vbetaFA, 'thresh') = assay(vbetaFA)*0 - 9
assay(vbetaFA)[1:3, 1:3]
```

---

maits	<i>MAITs data set, RNASeq</i>
-------	-------------------------------

---

**Description**

MAITs data set, RNASeq

**Format**

a list containing an expression matrix (expressionmat), cell cdat and feature fdat.

**See Also**

[FromMatrix](#)

---

mast_filter	<i>Filter a SingleCellAssay</i>
-------------	---------------------------------

---

**Description**

Remove, or flag wells that are outliers in discrete or continuous space.

**Usage**

```
mast_filter(sc, groups = NULL, filt_control = NULL,
            apply_filter = TRUE)
```

```
burdenOfFiltering(sc, groups, byGroup = FALSE, filt_control = NULL)
```

**Arguments**

sc	The SingleCellAssay object
groups	An optional character naming the grouping variable
filt_control	The list with configuration parameters for the filter.
apply_filter	logical should the filter be applied, or should a matrix of booleans giving if a well would be subject to a filtering criteria be returned?
byGroup	in the case of burdenOfFiltering should the filter be stratified by groups, or only the plotting.

**Details**

The function filters wells that don't pass filtering criteria described in `filt_control`. `filt_control` is a list with named elements `nOutlier` (minimum number of outlier cells for a cell to be filtered [default = 2]) `sigmaContinuous` (the z-score outlier threshold for the continuous part of the signal) [default = 7] and `sigmaProportion` (the z-score outlier threshold for the discrete part of the signal) [default = 7].

If `groups` is provided, the filtering is calculated within each level of the group, then combined again as output.

**Value**

A filtered result

**Functions**

- `burdenOfFiltering`: plot the proportions of wells are filtered due to different criteria

**Author(s)**

Andrew McDavid

**See Also**

`burdenOfFiltering`

**Examples**

```
data(vbetaFA)
## Split by 'ncells', apply to each component, then recombine
vbeta.filtered <- mast_filter(vbetaFA, groups='ncells')
## Returned as boolean matrix
was.filtered <- mast_filter(vbetaFA, apply_filter=FALSE)
## Wells filtered for being discrete outliers
head(subset(was.filtered, pctout))
burdenOfFiltering(vbetaFA, groups='ncells', byGroup=TRUE)
burdenOfFiltering(vbetaFA, groups='ncells')
```

---

`melt.SingleCellAssay` *"Melt" a SingleCellAssay matrix*

---

**Description**

Return a molten (flat) representation, taking the cross-product of the expression values, the `colData` (column meta data), and the feature data (`mcols`).

**Usage**

```
## S3 method for class 'SingleCellAssay'
melt(data, ..., na.rm = FALSE,
      value.name = "value")
```

**Arguments**

<code>data</code>	<code>SingleCellAssay</code>
<code>...</code>	ignored
<code>na.rm</code>	ignored
<code>value.name</code>	name of 'values' column in returned value

**Value**

A `data.table`, with the cartesian product of the row and column attributes and the expression values

**Examples**

```
data(vbetaFA)
melt.SingleCellAssay(vbetaFA[1:10,])
as(vbetaFA[1:10,], 'data.table')
```

---

model.matrix	<i>Model matrix accessor</i>
--------------	------------------------------

---

**Description**

Model matrix accessor

**Usage**

```
model.matrix(object, ...)

## S4 method for signature 'LMlike'
model.matrix(object, ...)
```

**Arguments**

object	LMlike or subclass
...	ignored

**Value**

model.matrix if present

**Methods (by class)**

- LMlike: return the model.matrix

---

model.matrix<-	<i>Replace model matrix</i>
----------------	-----------------------------

---

**Description**

Replace model matrix

**Usage**

```
model.matrix(object) <- value
```

**Arguments**

object	LMlike or subclass
value	matrix

**Value**

modify object

---

myBiplot	<i>Makes a nice BiPlot</i>
----------	----------------------------

---

**Description**

Creates a custom BiPlot for visualizing the results of PCA

**Usage**

```
myBiplot(pc, colorfactor, scaling = 100, nudge = 1.2, N = 10,
  dims = 1:2, ...)
```

**Arguments**

pc	output of prcomp
colorfactor	a factor the same length as nrow(pc\$x) to color the points
scaling	integer to scale the vectors showing loadings
nudge	numeric to offset labels for loadings
N	number of variables with longest dim[1] or dim[2] projections to display
dims	numeric vector of length 2 indicating which PCs to plot
...	passed to plot

**Value**

printed plot

---

pbootVcov1	<i>Bootstrap a zlmfit</i>
------------	---------------------------

---

**Description**

Sample cells with replacement to find bootstrapped distribution of coefficients

**Usage**

```
pbootVcov1(cl, zlmfit, R = 99)
```

**Arguments**

cl	a cluster object created by makeCluster
zlmfit	class ZlmFit
R	number of bootstrap replicates

**Value**

array of bootstrapped coefficients

---

```
plot.thresholdSCRNACountMatrix
```

*Plot cutpoints and densities for thresholding*

---

**Description**

Plot cutpoints and densities for thresholding

**Usage**

```
## S3 method for class 'thresholdSCRNACountMatrix'
plot(x, ask = FALSE, wait.time = 0,
     type = "bin", indices = NULL, ...)
```

**Arguments**

x	output of thresholdSCRNACountMatrix
ask	if TRUE then will prompt before displaying each plot
wait.time	pause (in seconds) between each plot
type	one or more of the following: 'bin' (plot the genes by the binning used for thresholding), or 'gene' (plot thresholding by gene – see next argument)
indices	if type is equal to 'gene', and is a integer of length 1, then a random sample of indices genes is taken. If it is NULL, then 10 genes are sampled. If it is a integer vector of length > 1, then it is interpreted as giving a list of indices of genes to be displayed.
...	further arguments passed to plot

**Value**

displays plots

**Examples**

```
## See thresholdSCRNACountMatrix
example(thresholdSCRNACountMatrix)
```

---

```
plotlrt
```

*Plot a likelihood ratio test object*

---

**Description**

Constructs a forest-like plot of signed log<sub>10</sub> p-values, possibly adjusted for multiple comparisons adjust can be one of "holm", "hochberg", "hommel", "bonferroni", "BH", "BY", "fdr", "none".

**Usage**

```
plotlrt(lr, adjust = "fdr", thres = 0.1, trunc = 1e-06,
       groups = NULL)
```

**Arguments**

lr	output from lrtest, with returnall=FALSE
adjust	character, passed along to p.adjust, see below
thres	numeric genes with adjusted pvalues above this value are not depicted
trunc	numeric p values below this value are truncated at this value
groups	character grouping value. If provided, must match groups argument passed to lrtest. Plots done separately for each group.

**Value**

Constructs a dotplot

**Author(s)**

andrew

---

plotSCAConcordance      *Concordance plots of filtered single vs n-cell assays*

---

**Description**

Plot the average expression value of two subsets of the data. Generally these might be 1 cell and multiple-cell replicates, in which case if the mcols column ncells is set then the averages will be adjusted accordingly. But it could be any grouping.

**Usage**

```
plotSCAConcordance(SCellAssay, NCellAssay, filterCriteria = list(nOutlier
  = 2, sigmaContinuous = 9, sigmaProportion = 9), groups = NULL, ...)
```

**Arguments**

SCellAssay	is a FluidigmAssay for the 1-cell per well assay
NCellAssay	is a FluidigmAssay for the n-cell per well assay
filterCriteria	is a list of filtering criteria to apply to the SCellAssay and NCellAssay
groups	is a character vector naming the group within which to perform filtering. NULL by default.
...	passed to getConcordance

**Value**

printed plot

**See Also**

getConcordance



**Examples**

```
data(vbetaFA)
sca1 <- subset(vbetaFA, ncells==1)
sca100 <- subset(vbetaFA, ncells==100)
plotSCAConcordance(sca1, sca100)
```

---

predict.ZlmFit	<i>Return predictions from a ZlmFit object.</i>
----------------	---

---

**Description**

Return predictions from a ZlmFit object.

**Usage**

```
## S3 method for class 'ZlmFit'
predict(object, newdata = NULL, modelmatrix = NULL,
  ...)
```

**Arguments**

object	A ZlmFit
newdata	The data to predict from. Currently ignored, will use the data in the object.
modelmatrix	The model matrix specifying the linear combination of coefficients.
...	ignored

**Value**

Predictions (on the link scale) and standard errors.

**Examples**

```
##See stat_e11
example(stat_e11)
```

---

predicted_sig	<i>Predicted signatures</i>
---------------	-----------------------------

---

**Description**

Predicted signatures

**Format**

A data frame of predicted gene expression signatures for stimulated and unstimulated cells.

---

primerAverage	<i>Average expression values for duplicated/redundant genes</i>
---------------	---

---

### Description

Takes an average, potentially on a different scale given by `fun.natural` of some genes. The average is then transformed with `fun.cycle`.

### Usage

```
primerAverage(fd, geneGroups, fun.natural = expavg,
             fun.cycle = logshift)
```

### Arguments

<code>fd</code>	SingleCellAssay or subclass
<code>geneGroups</code>	character naming a column in the featureData that keys the duplicates
<code>fun.natural</code>	transformation to be used to collapse the duplicate expression values
<code>fun.cycle</code>	transformation to be used after collapsing

### Value

averaged version of `fd`.

### Note

This code needs to be tested more extensively after a refactoring. Caveat calculator.

---

print.summaryZlmFit	<i>Print summary of a ZlmFit</i>
---------------------	----------------------------------

---

### Description

Shows the top '`n`' genes by z score on '`by`'

### Usage

```
## S3 method for class 'summaryZlmFit'
print(x, n = 2, by = "logFC", ...)
```

### Arguments

<code>x</code>	output from <code>summary(ZlmFit)</code>
<code>n</code>	number of genes to show
<code>by</code>	one of 'C', 'D' or 'logFC' for continuous, discrete and log fold change z-scores for each contrast
<code>...</code>	ignored

**Value**

prints a pretty table and invisibly returns a `data.table` representing the table.

**See Also**

`summary,ZlmFit-method`

---

read.fluidigm	<i>Reads a Fluidigm Biomark (c. 2011) raw data file (or set of files)</i>
---------------	---

---

**Description**

This function reads a raw Fluidigm Biomark data file or set of files and constructs a `SingleCellAssay` (or `FluidigmAssay`) object. This was written c. 2011 and has not been tested lately. The Biomark format may have changed.

**Usage**

```
read.fluidigm(files = NULL, metadata = NULL, header.size = 2,
  skip = 8, cycle.threshold = 40, metadataColClasses = NULL,
  meta.key = NULL, idvars = NULL, splitby = NULL,
  unique.well.id = "Chamber.ID", raw = TRUE, assay = NULL,
  geneid = "Assay.Name", sample = NULL, well = "Well",
  measurement = "X40.Ct", measurement.processed = "Ct",
  ncells = "SampleRConc")
```

**Arguments**

<code>files</code>	A character vector of files to read.
<code>metadata</code>	A character path and filename of a CSV file containing additional metadata about the samples
<code>header.size</code>	A numeric indicating the number of lines in the header (default 2)
<code>skip</code>	numeric how many lines to skip before reading (default 8)
<code>cycle.threshold</code>	The maximum number of PCR cycles performed (default 40) numeric
<code>metadataColClasses</code>	Optional character vector giving the column classes of the metadata file. See <a href="#">read.table</a> .
<code>meta.key</code>	Optional character vector that identifies the key column between the metadata and the fluidigm data
<code>idvars</code>	Optional character vector that defines the set of columns uniquely identifying a well (unique cell, gene, and condition).
<code>splitby</code>	Optional character that defines the column / variable used to split the resulting data into a list of <code>SingleCellAssay</code> , such that unique levels of <code>splitby</code> each fall into their own <code>SingleCellAssay</code> . Usually the experimental unit subjected to different treatments.
<code>unique.well.id</code>	The column that uniquely identifies a sample well in the data. Default is "Chamber.ID".

raw	logical flag indicating this is raw data coming off the instrument. Thus we make some assumptions about the column names that are present.
assay	character name of a column that uniquely identifies an Assay (i.e. gene). Default is NULL
geneid	character names of the column that identifies a gene. Default is "Assay.Name"
sample	character name of a column that uniquely identifies a sample
well	character name of a column that uniquely identifies a well. Default "Well".
measurement	character name of the column that holds the measurement. Default "X40.Ct".
measurement.processed	character one of "Ct", "40-Ct", or "et". If not "Ct", the measurement will be transformed.
ncells	The column with the number of cells in this well.

**Value**

list of SingleCellAssay holding the data.

**Author(s)**

Greg Finak

---

removeResponse	<i>Remove the left hand side (response) from a formula</i>
----------------	--

---

**Description**

The order of terms will be rearrange to suit R's liking for hierarchy but otherwise the function should be idempotent for

**Usage**

```
removeResponse(Formula, warn = TRUE)
```

**Arguments**

Formula	formula
warn	Issue a warning if a response variable is found?

**Value**

formula

**Author(s)**

Andrew

---

rstandard.bayesglm     *rstandard for bayesglm objects.*

---

**Description**

rstandard bayesglm object S3 method

**Usage**

```
## S3 method for class 'bayesglm'
rstandard(model, infl = influence(model, do.coef =
  FALSE), type = c("deviance", "pearson"), ...)
```

**Arguments**

model	bayesglm
infl	see <a href="#">rstandard</a>
type	see <a href="#">rstandard</a>
...	ignored

**Value**

numeric residuals

---

SceToSingleCellAssay     *Coerce a SingleCellExperiment to some class defined in MAST*

---

**Description**

Coerce a SingleCellExperiment to some class defined in MAST

**Usage**

```
SceToSingleCellAssay(sce, class = "SingleCellAssay",
  check_sanity = TRUE)
```

**Arguments**

sce	object inheriting from SingleCellExperiment
class	character naming the class to be coerced to
check_sanity	(default: TRUE) Set FALSE to override sanity checks that try to ensure that the default assay is log-transformed and has at least one exact zero. See <a href="#">defaultAssay</a> for details on the "default assay" which is assumed to contain log transformed data.

**Value**

object of the indicated class.

---

se.coef	<i>Return coefficient standard errors</i>
---------	---

---

**Description**

Given a fitted model, return the standard errors of the coefficient

**Usage**

```
se.coef(object, ...)
```

**Arguments**

object	a model implementing vcov
...	passed to methods

**Value**

vector or matrix

**See Also**

ZlmFit-class

**Examples**

```
#see ZlmFit-class for examples
example('ZlmFit-class')
```

---

show,LMlike-method	<i>show</i>
--------------------	-------------

---

**Description**

Display info

**Usage**

```
## S4 method for signature 'LMlike'
show(object)

## S4 method for signature 'ZlmFit'
show(object)
```

**Arguments**

object	an object of some type
--------	------------------------

**Details**

Prints information on a LMlike object

**Value**

side effect of printing to console

**Methods (by class)**

- ZlmFit: print info on ZlmFit

---

split, SingleCellAssay, character-method  
*Split into list*

---

**Description**

Splits a SingleCellAssay into a list by a factor (or something coercible into a factor) or a character giving a column of colData(x)

**Usage**

```
## S4 method for signature 'SingleCellAssay,character'  
split(x, f, drop = FALSE, ...)
```

**Arguments**

x	SingleCellAssay
f	length-1 character, or atomic of length ncol(x)
drop	drop unused factor levels
...	ignored

**Value**

List

**Examples**

```
data(vbetaFA)  
split(vbetaFA, 'ncells')  
fa <- as.factor(colData(vbetaFA)$ncells)  
split(vbetaFA, fa)
```

stat\_ell

*Plot confidence ellipse in 2D***Description**

The focus of the ellipse will be the point (x, y) and semi-major axes aligned with the coordinate axes and scaled by xse, yse and the level.

**Usage**

```
stat_ell(mapping = NULL, data = NULL, geom = "polygon",
         position = "identity", na.rm = FALSE, show.legend = NA,
         inherit.aes = TRUE, fill = NA, level = 0.95, lty = 2,
         invert = FALSE, alpha = 1, ...)
```

**Arguments**

mapping	Set of aesthetic mappings created by aes or aes_. If specified and inherit.aes = TRUE (the default), it is combined with the default mapping at the top level of the plot. You must supply mapping if there is no plot mapping.
data	The data to be displayed in this layer. There are three options: If NULL, the default, the data is inherited from the plot data as specified in the call to ggplot. A data.frame, or other object, will override the plot data. All objects will be fortified to produce a data frame. See fortify for which variables will be created. A function will be called with a single argument, the plot data. The return value must be a data.frame., and will be used as the layer data.
geom	The geometric object to use display the data
position	Position adjustment, either as a string, or the result of a call to a position adjustment function.
na.rm	If FALSE (the default), removes missing values with a warning. If TRUE silently removes missing values.
show.legend	logical. Should this layer be included in the legends? NA, the default, includes if any aesthetics are mapped. FALSE never includes, and TRUE always includes.
inherit.aes	If FALSE, overrides the default aesthetics, rather than combining with them. This is most useful for helper functions that define both data and aesthetics and shouldn't inherit behaviour from the default plot specification, e.g. borders.
fill	A color or aesthetic mapping to fill color. Defaults to NA for empty ellipses.
level	The confidence level at which to draw an ellipse (default is level=0.95).
lty	The linetype to use. Can map to a variable. Defaults to 2 (dashed line)
invert	vector of length 1 that should either be "x", "y", or TRUE. Specifies whether to plot the estimates from the discrete component on the inverse logit scale. invert specifies which axis to invert.
alpha	transparency
...	other arguments passed on to layer. These are often aesthetics, used to set an aesthetic to a fixed value, like color = "red" or size = 3. They may also be parameters to the paired geom/stat.



**Value**

ggplot layer

**Examples**

```
data(vbetaFA)
library(ggplot2)
zlmCond <- zlm(~Stim.Condition, vbetaFA[1:10,])
MM <- model.matrix(~Stim.Condition,unique(colData(vbetaFA)[,c("Stim.Condition"),drop=FALSE]))
predicted <- predict(zlmCond,modelmatrix=MM)
plt <- ggplot(predicted)+aes(x=invlogit(etaD),y=muC,xse=seD,yse=seC,col=sample)+
  facet_wrap(~primerid,scales="free_y")+theme_linedraw()+
  geom_point(size=0.5)+scale_x_continuous("Proportion expression")+
  scale_y_continuous("Estimated Mean")+
  stat_ell(aes(x=etaD,y=muC),level=0.95, invert='x')
## plot with inverse logit transformed x-axis
print(plt)
# doesn't do anything in this case because there are no inestimable coefficients
predictI <- impute(predicted, groupby='primerid')
```

---

subset,SingleCellAssay-method

*Subset a SingleCellAssay by cells (columns)*

---

**Description**

Evaluates the expression in ... in the context of colData(x) and returns a subsetted version of x

**Usage**

```
## S4 method for signature 'SingleCellAssay'
subset(x, ...)
```

**Arguments**

x	SingleCellAssay
...	expression

**Value**

SingleCellAssay

**Examples**

```
data(vbetaFA)
subset(vbetaFA, ncells==1)
```

---

 summarize

*Return programmatically useful summary of a fit*


---

**Description**

Return programmatically useful summary of a fit

**Usage**

```
summarize(object, ...)
```

**Arguments**

object	LMlike or subclass
...	other arguments

**Value**

list of parameters characterizing fit

---

summary,GSEATests-method

*Summarize gene set enrichment tests*


---

**Description**

Returns a data.table with one row per gene set. This data.table contains columns:

**set** name of gene set

**cond\_Z** Z statistic for continuous component

**cont\_P** wald P value

**cont\_effect** difference in continuous regression coefficients between null and test sets (ie, the numerator of the Z-statistic.)

**disc\_Z** Z statistic for discrete

**disc\_P** wald P value

**disc\_effect** difference in discrete regression coefficients between null and test sets.

**combined\_Z** combined discrete and continuous Z statistic using Stouffer's method

**combined\_P** combined P value

**combined\_adj** FDR adjusted combined P value

**Usage**

```
## S4 method for signature 'GSEATests'
summary(object, ...)
```

**Arguments**

object            A GSEATests object  
 ...                passed to calcZ

**Value**

data.table

**See Also**

gseaAfterBoot

**Examples**

```
## See the examples in gseaAfterBoot
example(gseaAfterBoot)
```

---

summary,ZlmFit-method *Summarize model features from a ZlmFit object*

---

**Description**

Returns a data.table with a special print method that shows the top 2 most significant genes by contrast. This data.table contains columns:

**primerid** the gene

**component** C=continuous, D=discrete, logFC=log fold change, S=combined using Stouffer's method, H=combined using hurdle method

**contrast** the coefficient/contrast of interest

**ci.hi** upper bound of confidence interval

**ci.lo** lower bound of confidence interval

**coef** point estimate

**z** z score (coefficient divided by standard error of coefficient)

**Pr(>Chisq)** likelihood ratio test p-value (only if doLRT=TRUE)

Some of these columns will contain NAs if they are not applicable for a particular component or contrast.

**Usage**

```
## S4 method for signature 'ZlmFit'
summary(object, logFC = TRUE, doLRT = FALSE,
  level = 0.95, ...)
```

**Arguments**

object            A ZlmFit object  
 logFC            If TRUE, calculate log-fold changes, or output from a call to getLogFC.  
 doLRT            if TRUE, calculate lrTests on each coefficient, or a character vector of such coefficients to consider.  
 level            what level of confidence coefficient to return. Defaults to 95 percent.  
 ...                ignored

**Value**

data.table

**See Also**

print.summaryZlmFit

**Examples**

```
data(vbetaFA)
z <- zlm(~Stim.Condition, vbetaFA[1:5,])
zs <- summary(z)
names(zs)
print(zs)
##Select `datatable` component to get normal print method
zs$datatable
```

---

```
summary.thresholdSCRNACountMatrix
```

*Summarize the effect of thresholding*

---

**Description**

Returns the proportion of (putative) expression, the variance of expressed cells, and -log10 shapiro-wilk tests for normality on the expressed cells

**Usage**

```
## S3 method for class 'thresholdSCRNACountMatrix'
summary(object, ...)
```

```
## S3 method for class 'summaryThresholdSCRNA'
print(x, ...)
```

**Arguments**

object	a thresholdSCRNACountMatrix
...	currently ignored
x	a summaryThresholdSCRNA object, ie output from summary.thresholdSCRNACountMatrix

**Value**

a list of statistics on the original data, and thresholded data

**Methods (by generic)**

- print: prints five-number distillation of the statistics and invisibly returns the table used to generate the summary

---

 thresholdSCRNACountMatrix

*Threshold a count matrix using an adaptive threshold.*


---

### Description

An adaptive threshold is calculated from the conditional mean of expression, based on 10 bins of the genes with similar expression levels. Thresholds are chosen by estimating cutpoints in the bimodal density estimates of the binned data. These density estimates currently exclude the zeros due to complications with how the bandwidth is selected. (If the bandwidth is too small, then extra peaks/modes are found and everything goes haywire). If the diagnostic plots don't reveal any bimodal bins, this is probably the reason, and you may not need to threshold since background in the data are exact zeros.

### Usage

```
thresholdSCRNACountMatrix(data_all, conditions = NULL, cutbins = NULL,
  nbins = 10, bin_by = "median", qt = 0.975, min_per_bin = 50,
  absolute_min = 0, data_log = TRUE, adj = 1)
```

### Arguments

data_all	matrix of (possibly log-transformed) counts or TPM. Rows are genes and columns are cells.
conditions	Bins are determined per gene and per condition. Typically contrasts of interest should be specified.
cutbins	vector of cut points.
nbins	integer number of bins when cutbins is not specified.
bin_by	character "median", "proportion", "mean"
qt	when bin_by is "quantile", what quantile should be used to form the bins
min_per_bin	minimum number of genes within a bin
absolute_min	numeric giving a hard threshold below which everything is assumed to be noise
data_log	is data_all log+1 transformed? If so, it will be returned on the (log+1)-scale as well.
adj	bandwidth adjustment, passed to density

### Value

list of thresholded counts (on natural scale), thresholds, bins, densities estimated on each bin, and the original data

### Examples

```
data(maits, package='MAST', envir = environment())
sca <- FromMatrix(t(maits$expressionmat[,1:1000]), maits$cdat, maits$fdat[1:1000,])
tt <- thresholdSCRNACountMatrix(assay(sca))
tt <- thresholdSCRNACountMatrix(2^assay(sca)-1, data_log=FALSE)
opar <- par()
on.exit(par(opar))
par(mfrow=c(4,2))
plot(tt)
```

---

vbeta	<i>Vbeta Data Set</i>
-------	-----------------------

---

**Description**

Vbeta Data Set

**Format**

a data frame with 11 columns. Column Ct contains the cycle threshold, with NA denoting that the threshold was never crossed. So it is inversely proportional to the log2 mRNA, and should be negated (and NAs set to zero) if it is used as a expression measurement for a FluidigmAssay.

---

vbetaFA	<i>Vbeta Data Set, FluidigmAssay</i>
---------	--------------------------------------

---

**Description**

Vbeta Data Set, FluidigmAssay

**Format**

a FluidigmAssay of the vbeta data set.

**See Also**

[vbeta](#), [FromFlatDF](#)

---

waldTest	<i>Run a Wald test</i>
----------	------------------------

---

**Description**

Run a Wald tests on discrete and continuous components hypothesis can be one of a character giving complete factors or terms to be dropped from the model, `CoefficientHypothesis` giving names of coefficients to be dropped, `Hypothesis` giving contrasts using the symbolically, or a contrast matrix, with one row for each coefficient in the full model, and one column for each contrast being tested.

**Usage**

```
waldTest(object, hypothesis)
```

**Arguments**

object	LMlike or subclass
hypothesis	the hypothesis to be tested. See details.

**Value**

array giving test statistics

**See Also**

fit

lrTest

lht

**Examples**

```
#see ZlmFit-class for examples  
example('ZlmFit-class')
```

---

waldTest,ZlmFit,matrix-method  
*Wald test*

---

**Description**

A 3D array with first dimension being the genes, next dimension giving information about the test (the degrees of freedom, Chisq statistic, and P value), and final dimension being the value of these quantities on the discrete, continuous and hurdle (combined) levels.

**Usage**

```
## S4 method for signature 'ZlmFit,matrix'  
waldTest(object, hypothesis)
```

**Arguments**

object	ZlmFit
hypothesis	See Details

**Value**

3D array

---

xform	<i>Make matrix of continuous expression values, orthogonal to discrete</i>
-------	--

---

### Description

This centers each column of `mat` around the mean of its non-zero values.

### Usage

```
xform(mat, scale = FALSE)
```

### Arguments

<code>mat</code>	matrix (such as produced by <code>exprs</code> )
<code>scale</code>	should the columns also be scaled to have unit variance

### Value

matrix

---

<code>zlm</code>	<i>Zero-inflated regression for SingleCellAssay</i>
------------------	---

---

### Description

For each gene in `sca`, fits the hurdle model in `formula` (linear for  $\epsilon > 0$ ), logistic for  $\epsilon = 0$  vs  $\epsilon > 0$ . Return an object of class `ZlmFit` containing slots giving the coefficients, variance-covariance matrices, etc. After each gene, optionally run the function on the fit named by `'hook'`

### Usage

```
zlm(formula, sca, method = "bayesglm", silent = TRUE, ebayes = TRUE,
     ebayesControl = NULL, force = FALSE, hook = NULL,
     parallel = TRUE, LMlike, onlyCoef = FALSE, ...)
```

### Arguments

<code>formula</code>	a formula with the measurement variable on the LHS and predictors present in <code>colData</code> on the RHS
<code>sca</code>	<code>SingleCellAssay</code> object
<code>method</code>	character vector, either <code>'glm'</code> , <code>'glmer'</code> or <code>'bayesglm'</code>
<code>silent</code>	Silence common problems with fitting some genes
<code>ebayes</code>	if TRUE, regularize variance using empirical bayes method
<code>ebayesControl</code>	list with parameters for empirical bayes procedure. See <a href="#">ebayes</a> .
<code>force</code>	Should we continue testing genes even after many errors have occurred?
<code>hook</code>	a function called on the fit after each gene.
<code>parallel</code>	If TRUE and <code>option(mc.cores) &gt; 1</code> then multiple cores will be used in fitting.



LMLike	if provided, then the model defined in this object will be used, rather than following the formulas. This is intended for internal use.
onlyCoef	If TRUE then only an array of model coefficients will be returned (probably only useful for bootstrapping).
...	arguments passed to the S4 model object upon construction. For example, fitArgsC and fitArgsD, or coefPrior.

### Value

a object of class `ZlmFit` with methods to extract coefficients, etc. OR, if data is a `data.frame` just a list of the discrete and continuous fits.

### Empirical Bayes variance regularization

The empirical bayes regularization of the gene variance assumes that the precision (1/variance) is drawn from a gamma distribution with unknown parameters. These parameters are estimated by considering the distribution of sample variances over all genes. The procedure used for this is determined from `ebayesControl`, a named list with components 'method' (one of 'MOM' or 'MLE') and 'model' (one of 'H0' or 'H1') method MOM uses a method-of-moments estimator, while MLE using the marginal likelihood. H0 model estimates the precisions using the intercept alone in each gene, while H1 fits the full model specified by formula

### See Also

`ZlmFit`-class, `ebayes`, `GLMlike`-class, `BayesGLMlike`-class

### Examples

```
data(vbetaFA)
zlmVbeta <- zlm(~ Stim.Condition, subset(vbetaFA, ncells==1)[1:10,])
slotNames(zlmVbeta)
#A matrix of coefficients
coef(zlmVbeta, 'D')['CCL2',]
#An array of covariance matrices
vcov(zlmVbeta, 'D')[, 'CCL2']
waldTest(zlmVbeta, CoefficientHypothesis('Stim.ConditionUnstim'))

## Can also provide just a \code{data.frame} instead
data<- data.frame(x=rnorm(500), z=rbinom(500, 1, .3))
logit.y <- with(data, x*2 + z*2); mu.y <- with(data, 10+10*x+10*z + rnorm(500))
y <- (runif(500)<exp(logit.y)/(1+exp(logit.y)))*1
y[y>0] <- mu.y[y>0]
data$y <- y
fit <- zlm(y ~ x+z, data)
summary.glm(fit$disc)
summary.glm(fit$cont)
```

---

ZlmFit-class

An S4 class to hold the output of a call to `zlm`


---

### Description

This holds output from a call to `zlm`. Many methods are defined to operate on it. See below.

### Usage

```
## S4 method for signature 'ZlmFit,CoefficientHypothesis'
lrTest(object, hypothesis)

## S4 method for signature 'ZlmFit,Hypothesis'
lrTest(object, hypothesis)

## S4 method for signature 'ZlmFit,matrix'
lrTest(object, hypothesis)

## S4 method for signature 'ZlmFit,CoefficientHypothesis'
waldTest(object, hypothesis)

## S4 method for signature 'ZlmFit,Hypothesis'
waldTest(object, hypothesis)

## S4 method for signature 'ZlmFit'
coef(object, which, ...)

## S4 method for signature 'ZlmFit'
vcov(object, which, ...)

## S4 method for signature 'ZlmFit'
se.coef(object, which, ...)
```

### Arguments

<code>object</code>	ZlmFit
<code>hypothesis</code>	call to <a href="#">Hypothesis</a> or <a href="#">CoefficientHypothesis</a> or a matrix giving such contrasts.
<code>which</code>	character vector, one of "C" (continuous) or "D" (discrete) specifying which component should be returned
<code>...</code>	ignored

### Value

see "Methods (by generic)"

### Methods (by generic)

- `lrTest`: Returns an array with likelihood-ratio tests on contrasts defined using `CoefficientHypothesis()`.
- `lrTest`: Returns an array with likelihood-ratio tests specified by `Hypothesis`, which is a [Hypothesis](#).

- `lrTest`: Returns an array with likelihood-ratio tests specified by `Hypothesis`, which is a contrast matrix.
- `waldTest`: Returns an array with Wald Tests on contrasts defined using `CoefficientHypothesis()`.
- `waldTest`: Returns an array with Wald Tests on contrasts defined in `Hypothesis()`
- `coef`: Returns the matrix of coefficients for component which.
- `vcov`: Returns an array of variance/covariance matrices for component which.
- `se.coef`: Returns a matrix of standard error estimates for coefficients on component which.

### Slots

`coefC` matrix of continuous coefficients  
`coefD` matrix of discrete coefficients  
`vcovC` array of variance/covariance matrices for coefficients  
`vcovD` array of variance/covariance matrices for coefficients  
`LModel` the `LmWrapper` object used  
`sca` the `SingleCellAssay` object used  
`deviance` matrix of deviances  
`loglik` matrix of loglikelihoods  
`df.null` matrix of null (intercept only) degrees of freedom  
`df.resid` matrix of residual DOF  
`dispersion` matrix of dispersions (after shrinkage)  
`dispersionNoShrink` matrix of dispersion (before shrinkage)  
`priorDOF` shrinkage weight in terms of number of psuedo-obs  
`priorVar` shrinkage target  
`converged` output that may optionally be set by the underlying modeling function  
`hookOut` a list of length `ngenes` containing output from a hook function, if `zlm` was called with one

### See Also

`zlm` summary, `ZlmFit`-method

### Examples

```

data(vbetaFA)
zlmVbeta <- zlm(~ Stim.Condition+Population, subset(vbetaFA, ncells==1)[1:10,])
#Coefficients and standard errors
coef(zlmVbeta, 'D')
coef(zlmVbeta, 'C')
se.coef(zlmVbeta, 'C')
#Test for a Population effect by dropping the whole term (a 5 degree of freedom test)
lrTest(zlmVbeta, 'Population')
#Test only if the VbetaResponsive cells differ from the baseline group
lrTest(zlmVbeta, CoefficientHypothesis('PopulationVbetaResponsive'))
# Test if there is a difference between CD154+/Unresponsive and CD154-/Unresponsive.
# Note that because we parse the expression
# the columns must be enclosed in backquotes
# to protect the \quote{+} and \quote{-} characters.
lrTest(zlmVbeta, Hypothesis('\`PopulationCD154+VbetaUnresponsive` -
  `PopulationCD154-VbetaUnresponsive`'))
waldTest(zlmVbeta, Hypothesis('\`PopulationCD154+VbetaUnresponsive` -
  `PopulationCD154-VbetaUnresponsive`'))

```

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