Package ‘MEAL’

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Title  Perform methylation analysis
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Description Package to integrate methylation and expression data. It can also perform methylation or expression analysis alone. Several plotting functionalities are included as well as a new region analysis based on redundancy analysis. Effect of SNPs on a region can also be estimated.
Depends  R (>= 3.6.0), Biobase, MultiDataSet
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computeRDAR2

Compute signification of RDA test

Description

Compare R2 obtained in our region of interest with the global R^2 and the R^2 of regions with the same number of probes.

Usage

computeRDAR2(
    fullMat,
    varsmodel,
    covarssmodel = NULL,
    featNum,
    R2,
    num_permutations = 1e+05 - 1
)
correlationMethExprs

Arguments

fullMat      Matrix with the whole genome expression or methylation values
varsmodel   Matrix with the model
covarsmodel Matrix with the covariables model
featNum     Numeric with the number of features of the RDA model
R2          Numeric with the R2 of the RDA model
num_permutations Numeric with the number of permutations.

Value

Numeric vector with the probability of finding a region with the same number of probes with a bigger R2 and the global R2.

correlationMethExprs  Computes the correlation between methylation and expression

Description

Estimates the correlation between methylation and expression. When there are known variables that affect methylation and/or expression, their effect can be subtracted using a linear model and then the residuals are used.

Usage

correlationMethExprs(
  multiset,
  meth_set_name = NULL,
  exprs_set_name = NULL,
  vars_meth = NULL,
  vars_exprs = NULL,
  sel_cpgs = NULL,
  flank = 250000,
  betas = TRUE,
  num_cores = 1,
  verbose = TRUE
)

Arguments

multiset      MultiDataSet containing a methylation and an expression slots.
meth_set_name Character vector with the name of the MultiDataSet’s slot containing methylation data.
exprs_set_name Character vector with the name of the MultiDataSet’s slot containing expression data.
vars_meth  Character vector with the names of the variables that will be used to obtain the methylation residuals. By default, none is used and residuals are not computed.

vars_exprs  Character vector with the names of the variables that will be used to obtain the expression residuals. By default, none is used and residuals are not computed.

sel_cpgs  Character vector with the name of the CpGs used in the analysis. If empty, all the CpGs of the methylation set will be used.

flank  Numeric with the number of pair bases used to define the cpg-expression probe pairs.

betas  If set is a GenomicRatioSet, should beta values be used? (Default: TRUE)

num_cores  Numeric with the number of cores to be used.

verbose  Logical value. If TRUE, it writes out some messages indicating progress. If FALSE nothing should be printed.

Details

For each cpg, a range is defined by the position of the cpg plus the flank parameter (upstream and downstream). Only those expression probes that are entirely in this range will be selected. For these reason, it is required that the ExpressionSet contains a featureData with the chromosome and the starting and ending positions of the probes.

Value

Data.frame with the results of the linear regression:

- cpg: Name of the cpg
- exprs: Name of the expression probe
- beta: coefficient of the methylation change
- se: standard error of the beta
- P.Value: p-value of the beta coefficient
- adj.P.Val: q-value computed using B&H

exportResults  Exports results data.frames to csv files.

Description

Exports results to csv files. If more than one variable is present, subfolders with the name of the variable are created. For each variable, four files will be generated: probeResults.csv, dmrCateResults.csv, bumphunterResults.csv and blockFinderResults.csv
**filterResults**

Filter the data.frame obtained from probe analysis

**Usage**

```r
filterResults(results, range, position = "position", chr = "chromosome")
```

**Arguments**

- `results`: Data.frame with the results of probe analysis
- `range`: GenomicRanges with the desired range.
- `position`: Character with the name of the column containing the positions
- `chr`: Character with the name of the column containing the chromosome

**Value**

Data.frame with the results of the probes of the range
getGeneVals  

Get all probes related to a gene

Description

Given a ResultSet and a gene name returns the results of the analysis of all the probes of the gene.

Usage

getGeneVals(
  object,  
gene,  
  rid = 1,  
genecol = "genes",  
fNames = c("chromosome", "start"),  
...)

Arguments

  object  
    ResultSet  
  gene  
    Character with the name of the gene  
  rid  
    Name of the results: "DiffMean" for mean differences, "DiffVar" for variance differences. (Default: DiffMean)  
  genecol  
    Character with the column of object fData with the gene information  
  fNames  
    Names of the columns of object fData that will be added to the results data.frame.  
  ...  
    Further arguments passed to getProbeResults

Value

data.frame with the results of the analysis of the probes belonging to the gene

Examples

```r
## Not run:
if (require(minfiData)){
  set <- ratioConvert(mapToGenome(MsetEx[1:10,]))
methyOneVar <- runPipeline(set, variable_names = "sex")
geneVals(set, methyOneVar, "TSPY4")
}
## End(Not run)
```
getProbeResults

Obtain probe results from a ResultSet

Description

It computes the statistics from the MArrayLM computed with DiffMeanAnalysis or DiffVarAnalysis. This function allows to specify the contrasts and to get F-statistics for a group of variables.

Usage

getProbeResults(
  object, 
  rid = "DiffMean", 
  coef = 2, 
  contrast = NULL, 
  fName = c("chromosome", "start"), 
  robust = FALSE, 
  ...
)

Arguments

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>object</td>
<td>ResultSet</td>
</tr>
<tr>
<td>rid</td>
<td>Name of the results: &quot;DiffMean&quot; for mean differences, &quot;DiffVar&quot; for variance differences. (Default: DiffMean)</td>
</tr>
<tr>
<td>coef</td>
<td>Number of the coefficient used to compute the statistics. If a vector is supplied, F-statistics evaluating the global effect of the coefficients are computed. (Default: 2).</td>
</tr>
<tr>
<td>contrast</td>
<td>Matrix of contrasts</td>
</tr>
<tr>
<td>fName</td>
<td>Names of the columns of object fData that will be added to the results data.frame.</td>
</tr>
<tr>
<td>...</td>
<td>Further arguments passed to getAssociation.</td>
</tr>
</tbody>
</table>

Value

data.frame with the probe results.
getRDAresults  

*Get a summary of RDA results*

**Description**

Get statistics from RDA result.

**Usage**

```r
getRDAresults(object)
```

**Arguments**

- `object`  
  `ResultSet`

**Value**

Numeric vector with the RDA statistics

---

**MEAL**

*MEAL (Methylation and Expression AnaLizer): Package for analysing methylation and expression data*

**Description**

MEAL is a package designed to facilitate the analysis methylation and expression data. The package can analyze one dataset and can find correlations between methylation and expression data. MEAL has a vignette that explains the main functionalities of the package.

---

**MEAL-defunct**

*Defunct functions*

**Description**

These functions are defunct and no longer available.

**Details**

Defunct functions are: multiCorrMethExprs, DAPipeline, DAProbe, DARegion, RDAs, filterSet, plotBestFeatures, preparePhenotype, createRanges, prepareMethylationSet, calculateRelevantSNPs, correlationMethSNPs, explainedVariance, normalSNP, plotLM

Defunct classes are: analysisRegionResults, analysisResults
### plotFeature

**Plot values of a feature**

#### Description

Plot values of a feature splitted by one or two variables.

#### Usage

```r
plotFeature(set, feat, variables = colnames(pheno)[1], betas = TRUE)
```

#### Arguments

- `set`: ExpressionSet, GenomicRatioSet or SummarizedExperiment.
- `feat`: Numeric with the index of the feature or character with its name.
- `variables`: Character vector with the names of the variables to be used in the splitting. Two variables is the maximum allowed.
- `betas`: If `set` is a GenomicRatioSet, should beta values be used? (Default: TRUE)

#### Value

A plot is generated on the current graphics device.

#### Examples

```r
if (require(minfiData)){
  set <- ratioConvert(mapToGenome(MsetEx[1:10,]))
  plotFeature(set, 1, variables = "Sample_Group")
}
```

---

### plotRDA

**Plot RDA results**

#### Description

Plot RDA results

#### Usage

```r
plotRDA(object, pheno = data.frame(), n_feat = 5, main = "RDA plot", alpha = 1)
```
plotRegion

Description

Plot the results from the different analyses of a ResultSet in a specific genomic region. It can plot all the results from runPipeline.

Usage

plotRegion(
  rset,
  range,
  results = names(rset),
  genome = "hg19",
  rset2,
  tPV = 5,
  fNames = c("chromosome", "start", "end"),
  fNames2 = c("chromosome", "start", "end")
)
runBlockFinder

Arguments

- **rset**: `ResultSet` with the region coordinates
- **range**: `GenomicRanges` with the region coordinates
- **results**: Character with the analyses that will be included in the plot. By default, all analyses available are included.
- **genome**: String with the genome used to retrieve transcripts annotation: `hg19`, `hg38`, `mm10`. (Default: "hg19")
- **rset2**: Additional `ResultSet`
- **tPV**: Threshold for P-Value
- **fNames**: Names from rset fData
- **fNames2**: Names from rset2 fData

Details

This plot allows to have a quick summary of the methylation or gene expression analyses in a given region. If we use a `ResultSet` obtained from methylation data, transcripts annotation is obtained from archive. If we use a `ResultSet` obtained from gene expression data, transcripts annotation is taken from fData.

This plot can be used to plot the results of one dataset (methylation or gene expression) or to represent the association between methylation and gene expression data. If only one dataset is used, the p-values and the coefficients of DiffMean and DiffVar analyses are plotted. If we pass two `ResultSets`, rset should contain methylation results and a rset2 the gene expression results.

Value

Regional plot

---

**runBlockFinder**  
**Run blockFinder**

Description

Run blockFinder

Usage

```r
runBlockFinder(  
  set,  
  model,  
  coefficient = 2,  
  blockfinder_cutoff = 0.1,  
  num_permutations = 0,  
  resultSet = FALSE,  
  verbose = FALSE,  
  ...  
)
```
Arguments

- **set**: GenomicRatioSet, eSet derived object or SummarizedExperiment
- **model**: Model matrix or formula to get model matrix from set.
- **coefficient**: Numeric with the column of model matrix used in the analysis. (Default: 2)
- **blockfinder_cutoff**: Numeric with the minimum cutoff to include a probe in a block. (Default: 0.1)
- **num_permutations**: Numeric with the number of permutations run to compute the blocks p-value. (Default: 0)
- **resultSet**: Should results be encapsulated in a resultSet? (Default: TRUE)
- **verbose**: Logical value. Should the function be verbose? (Default: FALSE)
- **...**: Further arguments passed to blockFinder.

Details

This function has been deprecated and will be defunct in the new version.

Value

data.frame or resultSet with the result of blockFinder

See Also

blockFinder

---

**runBumphunter**

Run bumphunter

Description

Run bumphunter

Usage

```r
runBumphunter(
  set, model,
  coefficient = 2,
  bumphunter_cutoff = 0.1,
  num_permutations = 0,
  bumps_max = 30000,
  betas = TRUE,
  check_perms = FALSE,
  verbose = FALSE,
  resultSet = FALSE,
  ...
)
```
Arguments

- **set**: GenomicRatioSet, eSet derived object or SummarizedExperiment
- **model**: Model matrix or formula to get model matrix from set.
- **coefficient**: Numeric with the column of model matrix used in the analysis. (Default: 2)
- **bumphunter_cutoff**: Numeric with the minimum cutoff to include a probe in a block. (Default: 0.1)
- **num_permutations**: Numeric with the number of permutations run to compute the bumps p-value. (Default: 0)
- **bumps_max**: Numeric with the maximum number of bumps used in the permutation. This parameter only applies when num_permutations is greater than 0. (Default: 30000)
- **betas**: If set is a GenomicRatioSet, should beta values be used? (Default: TRUE)
- **check_perms**: Logical. Should we check that there are less bumps than bumps_max? This parameter only applies when num_permutations is greater than 0. (Default: TRUE)
- **verbose**: Logical value. Should the function be verbose? (Default: FALSE)
- **resultSet**: Should results be encapsulated in a resultSet? (Default: TRUE)
- **...**: Further arguments passed to bumphunter.

Details

This function has been deprecated and will be defunct in the new version.

Value

data.frame or resultSet with the result of bumphunter

See Also

bumphunter

runDiffMeanAnalysis

---

**runDiffMeanAnalysis**

Run differential mean analysis

Description

Run differential mean analysis using t-moderated statistics. This function relies on lmFit from limma package.
runDiffMeanAnalysis

Usage

runDiffMeanAnalysis(
  set,  
  model, 
  weights = NULL, 
  method = "ls", 
  max_iterations = 100, 
  betas = TRUE, 
  resultSet = TRUE, 
  warnings = TRUE
)

Arguments

set Matrix, GenomicRatioSet, SummarizedExperiment or ExpressionSet.
model Model matrix or formula to get model matrix from set.
weights weights used in the lmFit model.
method String indicating the method used in the regression: "ls" or "robust". (Default: "ls")
max_iterations Numeric indicating the maximum number of iterations done in the robust method.
betas If set is a GenomicRatioSet, should beta values be used? (Default: TRUE)
resultSet Should results be encapsulated in a resultSet? (Default: TRUE)
warnings Should warnings be displayed? (Default: TRUE)

Value

MArrayLM or resultSet with the result of the differential mean analysis.

Examples

if (require(minfiData)){
  mvalues <- getM(MsetEx)[1:100, ]
  model <- model.matrix(~ Sample_Group, data = pData(MsetEx))
  res <- runDiffMeanAnalysis(mvalues, model, method = "ls")
  res
}

runDiffVarAnalysis Run differential variance analysis

Description

Run differential variance analysis. This analysis can only be run with categorical variables. This function relies on varFit from missMethyl package.
Usage

runDiffVarAnalysis(
set,
model,
coefficient = NULL,
resultSet = TRUE,
betas = TRUE,
warnings = TRUE,
...
)

Arguments

set Matrix, GenomicRatioSet, SummarizedExperiment or ExpressionSet.
model Model matrix or formula to get model matrix from set.
coefficient Numeric with the coefficients used to make the groups. If NULL, all possible groups will be computed.
resultSet Should results be encapsulated in a resultSet? (Default: TRUE)
betas If set is a GenomicRatioSet, should beta values be used? (Default: TRUE)
warnings Should warnings be displayed? (Default: TRUE)
... Further arguments passed to varFit.

Value

MArrayLM or resultSet with the result of the differential variance analysis.

Examples

if (require(minfiData)){
mvalues <- getM(MsetEx)[1:100, ]
model <- model.matrix(~ Sample_Group, data = pData(MsetEx))
res <- runDiffVarAnalysis(mvalues, model)
res
}

runDMRcate

Run DMRcate

Description

Run DMRcate

Usage

runDMRcate(set, model, coefficient = 2, resultSet = FALSE, ...)
runPipeline

Arguments

- **set**: GenomicRatioSet, eSet derived object or SummarizedExperiment
- **model**: Model matrix or formula to get model matrix from set.
- **coefficient**: Numeric with the column of model matrix used in the analysis. (Default: 2)
- **resultSet**: Should results be encapsulated in a resultSet? (Default: TRUE)
- **...**: Further arguments passed to cpg.annotate or dmrcate.

Details

This function has been deprecated and will be defunct in the new version.

Value

- data.frame or resultSet with the result of bumphunter

See Also

dmrcate, cpg.annotate

---

**runPipeline**

*Perform differential methylation analysis*

Description

Wrapper for analysing differential methylation and expression at region and probe level.

Usage

```r
runPipeline(
  set,
  variable_names,
  covariable_names = NULL,
  model = NULL,
  weights = NULL,
  num_vars,
  sva = FALSE,
  betas = TRUE,
  range,
  analyses = c("DiffMean"),
  verbose = FALSE,
  warnings = TRUE,
  DiffMean_params = NULL,
  DiffVar_params = list(coefficient = 1:2),
  rda_params = NULL,
  method = "ls",
  big = FALSE
)
```
Arguments

- **set**: GenomicRatioSet, eSet derived object or SummarizedExperiment
- **variable_names**: Character vector with the names of the variables that will be returned as result.
- **covariable_names**: Character vector with the names of the variables that will be used to adjust the model.
- **model**: Model matrix or formula to get model matrix from set.
- **weights**: weights used in the lmFit model (default NULL)
- **num_vars**: Numeric with the number of variables in the matrix for which the analysis will be performed. Compulsory if equation is not null.
- **sva**: Logical. Should Surrogate Variable Analysis be applied? (Default: FALSE)
- **betas**: If set is a GenomicRatioSet, should beta values be used? (Default: TRUE)
- **range**: GenomicRanges with the region used for RDA
- **analyses**: Vector with the names of the analysis to be run (DiffMean and/or DiffVar).
- **verbose**: Logical value. If TRUE, it writes out some messages indicating progress. If FALSE nothing should be printed.
- **warnings**: Should warnings be displayed? (Default: TRUE)
- **DiffMean_params**: List with other parameter passed to runBumphunter function.
- **DiffVar_params**: List with other parameter passed to runBumphunter function.
- **rda_params**: List with other parameter passed to runRDA function.
- **method**: String indicating the method used in the regression: "ls" or "robust". (Default: "ls")
- **big**: Logical value indicating whether SmartSVA should be instead of SVA (TRUE recommended for methylation or when having large number of samples). Default is FALSE.

Details

This function is the main wrapper of the package. First, it simplifies the set to only contain the common samples between phenotype and features. In addition, it allows to change the class of the variables and to apply genomic models (more information on preparePhenotype). Afterwards, analysis per probe and per region are done merging the results in an AnalysisResults object.

Default linear model will contain a sum of the variables and covariables. If interactions are desired, a costum formula can be specified. In that case, variables and covariables must also be specified in order to assure the proper work of the resulting AnalysisResult. In addition, the number of variables of the model for which the calculation will be done must be specified.

Value

ResultSet object
Examples

```r
if (require(minfiData)){
set <- ratioConvert(mapToGenome(MsetEx[1:10,]))
res <- runPipeline(set, variable_names = "Sample_Group")
res
}
```

runRDA  

**Calculate RDA for a set**

Description

Perform RDA calculation for a `AnalysisRegionResults`. Feature values will be considered the matrix X and phenotypes the matrix Y. Adjusting for covariates is done using a model matrix passed in `covarsmodel`.

Usage

```r
runRDA(
  set, model, num_vars = ncol(model), range, betas = FALSE, resultSet = TRUE,
  num_permutations = 10000, ...
)
```

Arguments

- `set`  
  MethylationSet, ExpressionSet or matrix
- `model`  
  Model matrix or formula to get model matrix from set.
- `num_vars`  
  Numeric with the number of variables in the matrix for which the analysis will be performed. Compulsory if equation is not null.
- `range`  
  GenomicRanges with the region used for RDA
- `betas`  
  If set is a GenomicRatioSet, should beta values be used? (Default: TRUE)
- `resultSet`  
  Should results be encapsulated in a `resultSet`? (Default: TRUE)
- `num_permutations`  
  Numeric with the number of permutations run to compute the p-value. (Default: 1e4)
- `...`  
  Further arguments passed to `rda`.

Value

Object of class `rda` or `resultSet`
runRegionAnalysis

See Also
rda

Examples
if (require(minfiData)) {
  set <- ratioConvert(mapToGenome(MsetEx[1:10,]))
  model <- model.matrix(~set$age)
  rda <- runRDA(set, model)
  rda
}

runRegionAnalysis Run different DMR detection methods

Description
Run different DMR detection methods

Usage
runRegionAnalysis(
  set, model,
  methods = c("blockFinder", "bumphunter", "DMRcate"),
  coefficient = 2,
  bumphunter_params = NULL,
  blockFinder_params = NULL,
  dmrcate_params = NULL,
  verbose = FALSE,
  resultSet = TRUE
)

Arguments
set GenomicRatioSet, eSet derived object or SummarizedExperiment
model Model matrix representing a linear model.
methods Character vector with the names of the methods used to estimate the regions. Valid names are: "blockFinder", "bumphunter" and "DMRcate".
coefficient Numeric with the index of the model matrix used to perform the analysis.
bumphunter_params List with other parameter passed to runBumphunter function.
blockFinder_params List with other parameter passed to runBlockFinder function.
dmrcate_params List with other parameter passed to runDMRcate function.
verbose Logical value. Should the function be verbose? (Default: FALSE)
resultSet Should results be encapsulated in a resultSet? (Default: TRUE)
Details

This function has been deprecated and will be defunct in the new version.

Value

List or resultSet with the result of the DMR detection methods.

See Also

bumphunter, blockFinder, dmrcate

Examples

```r
if (require(minfiData)){
  set <- ratioConvert(mapToGenome(MsetEx[1:10,]))
  model <- model.matrix(~Sample_Group, data = pData(MsetEx))
  res <- runRegionAnalysis(set, model)
  res
}
```

topRDAhits

_Get the top features associated with the RDA_

Description

Get a list of the features significantly associated to the first two RDA components

Usage

`topRDAhits(object, tPV = 0.05)`

Arguments

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>object</td>
<td>ResultSet</td>
</tr>
<tr>
<td>tPV</td>
<td>numeric with the p-value threshold. Only features with a p-values below this threshold will be shown.</td>
</tr>
</tbody>
</table>

Value

data.frame with the features, the component, the correlation and the p-value

Examples

```r
if (require(minfiData) & require(GenomicRanges)){
  set <- ratioConvert(mapToGenome(MsetEx[1:10,]))
  model <- model.matrix(~set$sex)
  rda <- runRDA(set, model)
  topRDAhits(rda)
}
```
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