Package ‘MSnID’

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Description Extracts MS/MS ID data from mzIdentML (leveraging mzID package) or text files. After collating the search results from multiple datasets it assesses their identification quality and optimize filtering criteria to achieve the maximum number of identifications while not exceeding a specified false discovery rate. Also contains a number of utilities to explore the MS/MS results and assess missed and irregular enzymatic cleavages, mass measurement accuracy, etc.
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MSnID-package MSnID: Utilities for Handling MS/MS Identifications

Description

Extracts MS/MS ID data from mzIdentML (leveraging mzID package) or text files. After collating
the search results from multiple datasets it assesses their identification quality and optimize filtering
criteria to achieve the maximal identifications at a user specified false discovery rate. Additional
utilities include:

1. post-experimental recalibration of mass measurement accuracy
2. assessment of irregular and missed cleavages given the enzyme cleavage pattern
3. assessment of false discovery rates at peptide-to-spectrum match, unique peptide and protein
   levels
4. leverages brute-force and sophisticated optimization routines (Nelder-Mead and simulated
   annealing) for finding the filtering criteria that provide the maximum spectrum, peptide or pro-
  tein identifications while not exceeding a corresponding preset threshold of false discovery
   rate
5. converts the results into MSnSet class object as spectral counting data
accessions

Details

Package: MSnID
Type: Package
Version: 0.1.0
Date: 2014-04-02
License: Artistic-2.0

Author(s)

Vladislav A. Petyuk (<vladislav.petyuk@pnnl.gov>)

---

accessions | Non-redundant list of accession (protein) identifiers

Description

Returns the non-redundant list of accession (protein) identifiers from the MSnID object. Most of the times accessions and proteins have the same meaning. However, there are cases, for example use of 6-frame stop-to-stop translation as FASTA file, where the entries are called with general term accessions rather then proteins. Currently, accessions and proteins have the same meaning in MSnID.

Usage

accessions(object, ...)
proteins(object, ...)

Arguments

object | An instance of class "MSnID".
... | ignored parameters

Value

Non-redundant list of accession (protein) identifiers.

Author(s)

Vladislav A Petyuk <vladislav.petyuk@pnnl.gov>

See Also

peptides
add_mod_symbol

Examples

data(c_elegans)
head(accessions(msnidObj))
head(proteins(msnidObj))

add_mod_symbol
Annotates peptide sequences with modification symbols

Description

Given the provided modification mass, annotates its position within the peptide sequence with provided symbol.

Usage

add_mod_symbol(object, mod_mass, symbol)

Arguments

object
An instance of class "MSnID".

mod_mass
(string) modification mass. Must match exactly one of the masses in report_mods.

symbol
(string) character that annotates the position of the modification

Value

Returns MSnID object with new or modified peptide_mod column. The column contains peptide sequence with amino acid modifications annotated with symbols.

Note

In current implementation the method can not distinguish modifications with the same mass, but different amino acid specificity as different modifications.

Author(s)

Vladislav A Petyuk <vladislav.petyuk@pnnl.gov>

See Also

report_mods map_mod_sites
Examples

```r
m <- MSnID(".")
mzids <- system.file("extdata","phospho.mzid.gz",package="MSnID")
m <- read_mzIDs(m, mzids)

# to know the present mod masses
report_mods(m)

# TMT modification
m <- add_mod_symbol(m, mod_mass="229.1629", symbol="#")
# alkylation
m <- add_mod_symbol(m, mod_mass="57.021463735", symbol="^")
# phosphorylation
m <- add_mod_symbol(m, mod_mass="79.966330925", symbol="*")

# show the mapping
head(unique(subset(psms(m), select=c("modification", "peptide_mod"))))

# clean-up cache
unlink(".Rcache", recursive=TRUE)
```

apply_filter

**Filters the MS/MS identifications**

Description

Filter out peptide-to-spectrum MS/MS identifications.

Usage

```r
apply_filter(msnidObj, filterObj)
```

Arguments

- `msnidObj`: An instance of class "MSnID".
- `filterObj`: Either an instance of `MSnIDFilter` class or a "character".

Details

`filterObj` argument evaluated to a "logical" for each entry of the MS/MS results table.

Value

Returns an instance of "MSnID" class with with peptide-to-spectrum matches that pass criteria defined in `filterObj` argument.

Author(s)

Vladislav A Petyuk <vladislav.petyuk@pnnl.gov>
assess_missed_cleavages

Counts the missing cleavage sites within the peptides sequence

Description

Bottom-up proteomics approaches utilize endoproteases or chemical agents to digest proteins into smaller fragments called peptides. The enzymes recognize short amino acid motifs and cleave along the peptide bonds. Chemical agents such as CNBr also possess amino acid cleavage specificity. In real-world not every cleavage site get cleaved during the sample processing. Therefore settings of MS/MS search engines quite often explicitly allow up to a certain number missed cleavage sites per peptide sequence.

This function counts the number of missed cleavages in peptide sequence given the endoprotease cleavage motif in the form of regular expression. The default value for missedCleavagePattern is \[KR](?=[^P])\), which corresponds to trypsin.

Usage

assess_missed_cleavages(object, missedCleavagePattern="\[KR](?=[^P])")

See Also

MSnID evaluate_filter
assess_termini

Arguments

object An instance of class "MSnID".
missedCleavagePattern Cleavage pattern in the form of regular expression.

Value

Returns an instance of "MSnID" class with additional column "numMissCleavages"

Warning

If the "MSnID" instance does not contain "peptide" column in MS/MS results table then there will
be an error. E.g. you can check this by
"peptide" %in% names(msnid) where msnid is your "MSnID" instance.

Author(s)

Vladislav A Petyuk <vladislav.petyuk@pnnl.gov>

See Also

assess_termini

Examples

data(c_elegans)
  # adding column numMissCleavages containing count of missed cleavages
  msnidObj <- assess_missed_cleavages(msnidObj,
            missedCleavagePattern="[KR](?=[^P$])")
  # check the distribution
  table(msnidObj$numMissCleavages)

assess_termini Checks if the peptide termini conforms with cleavage specificity

Description

Bottom-up proteomics approaches utilize endoproteases or chemical agents to digest proteins into
smaller fragments called peptides. The enzymes recognize short amino acid motifs and cleave along
the peptide bonds. Chemical agents such as CNBr also possesses amino acid cleavage specificity.

This function checks if peptide termini are as expected given the enzymatic/chemical cleavage
specificity. The default value for validCleavagePattern is [KR]\.[^P], which corresponds to
trypsin.

Usage

assess_termini(object, validCleavagePattern="[KR]\.[^P]")
Arguments

object  An instance of class "MSnID".
validCleavagePattern  Cleavage pattern in the form of regular expression.

Details

N- or C- protein termini are not considered as irregular cleavages sites.

Value

Returns an instance of "MSnID" class with additional column "numIrregCleavages". If both termini conforms with cleavage specificity, then value is 0, if one or two termini are irregular then the values are 1 and 2, correspondingly.

Warning

If the "MSnID" instance does not contain "peptide" column in MS/MS results table then there will be an error. E.g. you can check this by "peptide" %in% names(msnid) where msnid is your "MSnID" instance.

Author(s)

Vladislav A Petyuk <vladislav.petyuk@pnnl.gov>

See Also

assess_missed_cleavages

Examples

data(c_elegans)
# adding column numIrregCleavages
# containing count of irregularly cleaved termini
msnidObj <- assess_termini(msnidObj, validCleavagePattern="[KR]\\\.[^P]")
# check the distribution
table(msnidObj$numIrregCleavages)

correct_peak_selection

Corrects wrong selection of monoisotopic peak
**correct_peak_selection**

**Description**

In a typical setting instruments select ions for fragmentation primarily based on ion intensity. For low molecular weight peptides the most intense peak usually corresponds to monoisotopic peak (that is only C12 carbon isotopes). With increase of molecular weight, intensity of monoisotopic peak becomes smaller relatively to heavier peptide isotopes (that is containing one or a few C13 isotopes).

The function subtracts or adds the mass difference between C13 and C12 isotopes (1.0033548378 Da) if that reduces the mass error. Such a mass error arises from the fact that instrument may peak non-monoisotopic peak for fragmentation and thus report the mass that is different by ~ 1 Da.

**Usage**

correct_peak_selection(object)

**Arguments**

object  
An instance of class "MSnID".

**Value**

Returns an instance of "MSnID" class with updated experimentalMassToCharge value.

**Author(s)**

Vladislav A Petyuk <vladislav.petyuk@pnnl.gov>

**See Also**

MSnID recalibrate mass_measurement_error

**Examples**

data(c_elegans)
# plot original mass error
massErr <- (msnidObj$experimentalMassToCharge -
    msnidObj$calculatedMassToCharge) *
    msnidObj$chargeState
hist(massErr,xlim=c(-1,+1), breaks=seq(-1.5,+1.5,0.01))
# fixing the problem of picking wrong monoisotopic peak
msnidObj <- correct_peak_selection(msnidObj)
# plot fixed mass error
massErr <- (msnidObj$experimentalMassToCharge -
    msnidObj$calculatedMassToCharge) *
    msnidObj$chargeState
hist(massErr,xlim=c(-1,+1), breaks=seq(-1.5,+1.5,0.01))
**evaluate_filter**

**Filters the MS/MS identifications**

**Description**

Filter out peptide-to-spectrum MS/MS identifications.

**Usage**

```r
evaluate_filter(object, filter, level=c("PSM", "peptide", "accession"))
```

**Arguments**

- **object**: An instance of class "MSnID".
- **filter**: Either an instance of MSnIDFilter class or a "character".
- **level**: Level at which the filter will be evaluated. Possible values are "PSM", "peptide" and "accession". Multiple are OK. Default is all of them.

**Value**

Returns a matrix with column names "fdr" and "n". Column "n" contains the number of features (spectra, peptides or proteins/accessions) passing the filter. Column "fdr" is the false discovery rate (i.e. identification confidence) for the corresponding features. Row names correspond to the provided levels.

**Author(s)**

Vladislav A Petyuk <vladislav.petyuk@pnnl.gov>

**See Also**

MSnID id_quality

**Examples**

```r
data(c_elegans)

## Filtering using string:
msnidObj <- assess_termini(msnidObj, validCleavagePattern="[KR]\.[^P]")
table(msnidObj$msIdObj$numIrregCleavages)
evaluate_filter(msnidObj, "numIrregCleavages == 0")

## Filtering using filter object:
# first adding columns that will be used as filters
msnidObj$msmsScore <- -log10(msnidObj$MS-GF:SpecEValue)
msnidObj$mzError <- abs(msnidObj$experimentalMassToCharge -
msnidObj$calculatedMassToCharge)

# setting up filter object
filtObj <- MSnIDFilter(msnidObj)
filtObj msmScore <- list(comparison=">", threshold=10.0)
filtObj mzError <- list(comparison="<", threshold=0.1) # 0.1 Thomson
show(filtObj)
evaluate_filter(msnidObj, filtObj)

---

**fetch_conversion_table**

*Fetches conversion table form one type of identifiers to another*

**Description**

A wrapper function over **AnnationHub** that helps to convert from one protein identifiers to another.

**Usage**

```r
fetch_conversion_table(organism_name, from, to,
                      backend="AnnationHub",
                      snapshot_date=NULL)
```

**Arguments**

organism_name  (string) official organism name. E.g. "Homo sapiens", "Mus musculus" or "Rattus norvegicus".

from, to        (string) identifier names. Recommended names are "SYMBOL", "UNIPROT", "REFSEQ" and "ENSEMBLPROT". Other identifiers are possible, but use them at your own risk.

backend        (string) currently only **AnnationHub**

snapshot_date  (string) snapshot date for **AnnationHub**. Default is NULL meaning latest date.

**Value**

data.frame with first column name of from identifier, the second name of to identifier

**Author(s)**

Vladislav A Petyuk <vladislav.petyuk@pnnl.gov>

**Examples**

```r
conv_tbl <- fetch_conversion_table("Rattus norvegicus", "REFSEQ", "SYMBOL")
head(conv_tbl)

conv_tab <- fetch_conversion_table("Homo sapiens", "UNIPROT", "SYMBOL")
head(conv_tbl)
```
**id_quality**

**Identification quality**

**Description**

Reports quality for a given level of identification (spectra, peptide or protein).

**Usage**

```r
id_quality(object, filter=NULL, level=c("PSM", "peptide", "accession"))
```

**Arguments**

- **object**: An instance of class "MSnID".
- **filter**: Optional argument. Either an instance of MSnIDFilter class or a "character".
- **level**: Level at which the filter will be evaluated. Possible values are "PSM", "peptide" and "accession". Multiple are OK. Default is all of them.

**Value**

Returns a matrix with column names "fdr" and "n". Column "n" contains the number of features (spectra, peptides or proteins/accessions) passing the filter. Column "fdr" is the false discovery rate (i.e. identification confidence) for the corresponding features. Row names correspond to the provided levels.

**Author(s)**

Vladislav A Petyuk <vladislav.petyuk@pnnl.gov>

**See Also**

- [MSnID evaluate_filter](#)

**Examples**

```r
data(c_elegans)
id_quality(msnidObj, level="peptide")
id_quality(msnidObj, filter="MS-GF:PepQValue < 0.01", level="peptide")
```
infer_parsimonious_accessions

Eliminates Redundancy in Peptide-to-Protein Mapping

Description

Infer parsimonious set of accessions (e.g. proteins) that explains all the peptide sequences. The algorithm is a simple loop that looks for the accession explaining most peptides, records the peptide-to-accession mapping for this accession, removes those peptides, and then looks for next best accession. The loop continues until no peptides left. The method does not accept any arguments at this point (except the MSnID object itself).

Usage

infer_parsimonious_accessions(object, unique_only=FALSE, prior=character(0))

Arguments

<table>
<thead>
<tr>
<th>object</th>
<th>An instance of class &quot;MSnID&quot;.</th>
</tr>
</thead>
<tbody>
<tr>
<td>unique_only</td>
<td>If TRUE, peptides mapping to multiple accessions are dropped and only unique are retained. If FALSE, then shared peptides assigned according to Occam’s razor rule. That is a shared peptide is assigned to a protein with larger number of unique peptides. If the number of unique peptides is the same, then to the first accession. Default is FALSE.</td>
</tr>
<tr>
<td>prior</td>
<td>(character) character vector with prior justified proteins/accessions. If unique_only == TRUE, then prior argument is ignored. Essentially evidence by presence of unique peptide supercedes any prior. Default is character(0), that is none.</td>
</tr>
</tbody>
</table>

Details

Although the algorithm is rather simple it is THE algorithm used for inferring maximal matching in bipartate graphs and is used in the IDPicker software.

Value

Returns an instance of "MSnID" with minimal set of proteins necessary to explain all the peptide sequences.

Author(s)

Vladislav A Petyuk <vladislav.petyuk@pnnl.gov>

See Also

MSnID
Examples

```r
data(c_elegans)

# explicitely adding parameters that will be used for data filtering
msnidObj$msmsScore <- -log10(msnidObj$`MS-GF:SpecEValue`)
msnidObj$absParentMassErrorPPM <- abs(mass_measurement_error(msnidObj))

# quick-and-dirty filter. The filter is too strong for the sake of saving time
# at the minimal set of proteins inference step.
msnidObj <- apply_filter(msnidObj, 'msmsScore > 12 & absParentMassErrorPPM < 2')

show(msnidObj)
msnidObj2 <- infer_parsimonious_accessions(msnidObj)
show(msnidObj2)
```

---

**map_mod_sites**  
Maps the modifications to protein sequence

**Description**

Given the peptide sequence with modification `X.XXXX*XXXX.X` and provided protein sequence FASTA, the method maps the location of the modification resulting in `{protein ID}-{aa}{aa position}`.

**Usage**

```r
map_mod_sites(object, 
               fasta, 
               accession_col = "accession", 
               peptide_mod_col = "peptide_mod", 
               mod_char = "*", 
               site_delimiter = "lower")
```

**Arguments**

- **object**: An instance of class MSnID.
- **fasta**: (AAStringSet object) Protein sequences read from a FASTA file. Names must match protein/accession IDs in the accession column of the MSnID object.
- **accession_col**: (string) Name of the column with accession/protein IDs in the MSnID object. Default is "accession".
- **peptide_mod_col**: (string) Name of the column with modified peptide sequences in the MSnID object. Default is "peptide_mod".
- **mod_char**: (string) character that annotates the position of the modification. Default is "*".
- **site_delimiter**: (string) either a single character or "lower" (default) meaning it will be the same amino acid symbol, but in lower case
**Value**

MSnID object with extra columns regarding the modification mapping. Most likely, what you need is SiteID.

- **PepLoc** (list of ints) position of the starting amino acid within protein sequence. It is a list, because there may be multiple occurrences of the same sequence matching the peptide’s sequence.
- **PepLocFirst** (int) position of the first occurrence of the matching sequence
- **ProtLength** (int) protein length
- **ModShift** (vector of ints) positions of modified amino acids within peptide
- **ModAAAs** (vector of characters) single-letter amino acid codes of the modified residues
- **SiteLoc** (list of vectors of ints) positions of the modified amino acids within protein for each occurrence of the peptide
- **Site** (list of vectors of characters) modified sites encoded as amino acid symbol followed by position for each occurrence of the peptide
- **SiteCollapsed** (list of characters) same as Site, but modified sites for each peptide occurrence collapsed into one string
- **SiteCollapsedFirst** (character) first element of the SiteCollapsed list
- **SiteID** (character) accession ID concatenated with SiteCollapsedFirst

**Author(s)**

Vladislav A Petyuk <vladislav.petyuk@pnnl.gov>

**Examples**

```r
m <- MSnID(".")
mzids <- system.file("extdata","phospho.mzid.gz",package="MSnID")
m <- read_mzIDs(m, mzids)

# to know the present mod masses
report_mzs(m)

# TMT modification
m <- add_mod_symbol(m, mod_mass="229.1629", symbol="#")
# alkylation
m <- add_mod_symbol(m, mod_mass="57.021463735", symbol="^")
# phosphorylation
m <- add_mod_symbol(m, mod_mass="79.966330925", symbol="*")

# show the mapping
head(unique(subset(psms(m), select=c("modification", "peptide_mod"))))

# read fasta for mapping modifications
fst_path <- system.file("extdata","for_phospho.fasta.gz",package="MSnID")
library(Biostrings)
fst <- readAAStringSet(fst_path)
```
mass_measurement_error

Computes error of the parent ion mass to charge measurement

Description
Computes error of the parent ion mass to charge measurement from experimentalMassToCharge and calculatedMassToCharge. The returned value is in points per million (ppm).

Usage
mass_measurement_error(object)

Arguments
object
An instance of class "MSnID".

Details
It may be more common to compute "mass measurement error". However, the practical difference in "mass measurement error" and "mass to charge measurement error" is negligible. Moreover, the instruments measure mass/charge ratio, not mass per se.

Value
Returns mass to charge measurement error in "ppm" units.

Author(s)
Vladislav A Petyuk <vladislav.petyuk@pnnl.gov>

See Also
MSnID recalibrate correct_peak_selection

Examples
data(c_elegans)
ppm <- mass_measurement_error(msnidObj)
hist(ppm, 100)
**Description**

The MSnID is a convenience class for manipulating the MS/MS search results.

**Objects from the Class**

The way to create objects is to call MSnID constructor function that takes as an input the project working directory `workDir` and the second argument if the cache from previous analysis should be cleaned `cleanCache`.

**Slots**

- `workDir`: Object of class "character". containing working directory for the project. The `.Rcache` subdirectory stores the cached results from the previous analyses. The mechanism of caching relies on `R.cache` package.

- `psms`: Object of class `data.table` that contains all the MS/MS identification results in the form of peptide(or protein)-spectrum-matches.

**Methods**

- `read_mzIDs` signature(object, mzids):
  Reads mzIdentML files into `psms` data.table slot of object MSnID instance. The functionality leverage `mzID` package facility. Note, the calls are memoised using `R.cache` facility. So if the call with the same list of files issues again, the results will be read from cache instead of re-parsing the mzIdentML files. See `read_mzIDs`

- `psms(object), psms(object)<-value`: Gets and sets MS/MS search results as data.frame. See `psms`

- `dim` signature(x = "MSnID"):
  Returns the dimensions of the table with MS/MS identification data.

- `peptides` signature(object = "MSnID"):
  Returns unique peptide list. See `peptides`

- `accessions` signature(object = "MSnID"):
  Returns unique accessions (typically proteins) list. See `accessions`

- `proteins` signature(object = "MSnID"):
  Returns unique proteins list. See `proteins`

- `assess_termini` Checks the agreement of peptide termini with enzymes cleavage specificity. The return value is the MSnID object with extra variable `numIrregCleavages`. See `assess_termini`

- `assess_missed_cleavages` Checks if the peptide sequence contains the sites that were not cleaved by the enzyme. For details see `assess_missed_cleavages`
mass_measurement_error  Returns parent ion mass measurement error in parts per million (ppm) units. Note, it requires experimentalMassToCharge and calculatedMassToCharge variables to be set. See mass_measurement_error

recalibrate  Recalibrates, that is removes systematic error from experimentalMassToCharge measurements. See recalibrate

correct_peak_selection  Subtracts or adds the mass difference between C13 and C12 isotopes (1.0033548378 Da) if that reduces the mass error. Such a mass error arises from the fact that instrument may peak non-monoisotopic peak for fragmentation and thus report the mass that is different by ~ 1 Da. See correct_peak_selection

apply_filter  signature(msnidObj="MSnID", filterObj="character")
signature(msnidObj="MSnID", filterObj="MSnIDFilter")
The filterObj argument is a "character" or converted to a "character" text string that is evaluated to a "logical" for each entry of the MS/MS results table. Return value is a filtered MSnID object with entries that pass the applied filter. See apply_filter

evaluate_filter  evaluate_filter(object, filter, level = c("PSM","peptide","accession"))
Returns a list with fdr and n elements. Argument filter is either "character" or "MSnIDFilter" object. Argument level can take one of the values c("PSM","peptide","accession") and controls the level filter is evaluated. See evaluate_filter

id_quality  signature(object="MSnID", ...)
Other optional... arguments are filter is an "MSnIDFilter" instance and level. The level values are one of "PSM", "peptide", "accession". The method returns FDR for given level depending of type of identifications. See id_quality

as("MSnSet")  signature(x = "MSnID") Coerce object from MSnID to MSnSet.

names  signature(x="MSnID")
Returns the column names in the MS/MS results table.

object$name, object$name<-value  Access and set name column in MS/MS search results table.

object[[i]], object[[i]]<-value  Access and set column i (character or numeric index) in MS/MS search results table.

as("MSnSet")  signature(from = "MSnID") Coerce object from MSnID to MSnSet.

as("data.table")  signature(from = "MSnID") Coerce object from MSnID to data.table.

Author(s)
Vladislav A Petyuk <vladislav.petyuk@pnnl.gov>

See Also
MSnSet, mzID.

Examples
## Not run:
msnidObj <- MSnID(".")
mzids <- system.file("extdata", "c_elegans.mzid.gz", package="MSnID")
msnidObj <- read_mzIDs(msnidObj, mzids)
# clean up the cache directory
unlink(".Rcache", recursive=TRUE)
## End(Not run)

### MSnIDFilter-class

The "MSnIDFilter" Class for Handling MS/MS Criteria, Relationships and Thresholds for Data Filtration.

#### Description

The `MSnIDFilter` is a convenience class for manipulating the MS/MS filter for MS/MS results.

#### Objects from the Class

The way to create objects is to call `MSnIDFilter` constructor function that takes as input the `MSnID` class instance and (optionally) `filterList`.

#### Slots

- **MSnIDObj**: An instance of class "MSnID".
- **filterList**: An optional argument. A list with element names corresponding to column names available in `MSnID` instance. Each element contains sub-elements "comparison" and "threshold". "Comparison" is one of the relationship operators (e.g. ">") see `Comparison` for details. "Threshold" is the corresponding parameter value the identification has to be more or less (depending on comparison) to pass the filter.

#### Methods

- **show** signature(object="MSnIDFilter"): Prints `MSnIDFilter` object.
- **object$name**, **object$name<-value** Access and set `filterList` elements.
- **names** signature(x="MSnIDFilter") Returns the names of the criteria.
- **as.numeric** signature(x="MSnIDFilter") Converts `filterList` into "numeric" vector. Vector names are the list element names. Vector values are threshold values. Comparison operators are lost.
- **length** signature(x="MSnIDFilter") Returns the number of criteria set in the "MSnIDFilter" object.
- **update** signature(object="MSnIDFilter", ...) The additional ...argument is numeric vector of the same length as the number of criteria in `MSnIDFilter` object. The method update the corresponding thresholds to new provided values.

#### Author(s)

Vladislav A Petyuk <vladislav.petyuk@pnnl.gov>
See Also

MSnSet evaluate_filter apply_filter optimize_filter

Examples

data(c_elegans)

## Filtering using filter object:
# first adding columns that will be used as filters
msnidObj$msmsScore <- -log10(msnidObj$"MS-GF:SpecEValue")
msnidObj$mzError <- abs(msnidObj$experimentalMassToCharge -
msnidObj$calculatedMassToCharge)

# setting up filter object
filtObj <- MSnIDFilter(msnidObj)
filtObj$msmsScore <- list(comparison=">", threshold=10.0)
filtObj$mzError <- list(comparison="<", threshold=0.1) # 0.1 Thomson
show(filtObj)

# applying filter and comparing MSnID object before and after
show(msnidObj)
msnidObj <- apply_filter(msnidObj, filtObj)
show(msnidObj)

Example mzIdenML File and MSnID Object

Description

MSnID object from c_elegans_A_3_1_21Apr10_Draco_10-03-04_msgfplus.mzid.gz dataset from PeptideAtlas repository id PASS00308.

Usage

data(c_elegans)

Examples

data(c_elegans)
msnidObj

## Not run:
## How to download the example mzID file from PeptideAtlas:
try(setInternet2(FALSE),silent=TRUE)
ftp.loc <- "ftp://PASS00308:PJ5348t@ftp.peptideatlas.org/MSGFPlus_Results/MZID_Files/c_elegans_A_3_1_21Apr10_Draco_10-03-04_msgfplus.mzid.gz"
download.file(ftp.loc, "c_elegans.mzid.gz")

## End(Not run)

## Not run:
## Script for generation of the C. elegans example data:
```r
msnidObj <- MSnID(".")
mzids <- system.file("extdata","c_elegans.mzid.gz",package="MSnID")
msnidObj <- read_mzIDs(msnidObj, mzids)
save(msnidObj, file='c_elegans.RData', compress='xz', compression_level=9)
# MD5 sum for the file is: a7c511a6502a6419127f1e46db48ed92
digest::digest(msnidObj)
# clean up the cache directory
unlink(".Rcache", recursive=TRUE)
```

`optimize_filter`  
*Filter criteria optimization to maximize the number of identifications given the FDR upper threshold*

### Description

Adjusts parameters in the "MSnIDFilter" instance to achieve the most number of spectra, peptides or proteins/accessions within pre-set FDR upper limit.

### Usage

```r
optimize_filter(filterObj, msnidObj, fdr.max, method, level, n.iter, mc.cores=NULL)
```

### Arguments

- `filterObj`: An instance of class "MSnIDFilter".
- `msnidObj`: An instance of class "MSnID".
- `fdr.max`: Upper limit on acceptable false discovery rate (FDR).
- `method`: Optimization method. Possible values are "Grid" or same values as for the method argument of the `optim` function. Tested and recommended arguments (besides "Grid") of `method` are "Nelder-Mead" or "SANN".
- `level`: Determines at what level to perform optimization. Possible values are "PSM", "peptides" or "accession".
- `n.iter`: For method "Grid" is approximate number of evaluation point. For "Nelder-Mead" and "SANN" methods see `optim`.
- `mc.cores`: Controls the number of enabled cores for parallel processing. Make sense only for "Grid" optimization. Default is `getOption("mc.cores", 2L)`. See `mclapply` for details.
Details

The "Grid" method is brute-force optimization through evaluation of approximately \( n.\text{iter} \) combinations of the parameters set in the "MSnIDFilter" object. The enumeration of the parameter combinations proceeds as follows. The \( n.\text{iter} \) number is getting split given the dimensionality of the problem (that is the number of filter parameters in the "MSnIDFilter" object. For each parameter the evaluation points are equally spaced according to quantiles of the parameter distribution. This way we enumerate the grid that has more evaluation points in relatively more dense areas.

Note, optimization is computationally expensive. Thus, the optimize_filter call is memoised using facilities from the \texttt{R.cache} package. Once the same call of optimize_filter function issued second time the results will be retrieved from cache.

Value

Returns an instance of "MSnIDFilter" that is maximized to provide the most number of identifications while maintaining a pre-set confidence (FDR).

Author(s)

Vladislav A Petyuk <vladislav.petyuk@pnnl.gov>

See Also

\texttt{MSnID evaluate_filter apply_filter}

Examples

data(c_elegans)

# explictely adding parameters that will be used for data filtering
msnidObj$msmsScore <- -log10(msnidObj$'MS-GF:SpecEValue')
msnidObj$absParentMassErrorPPM <- abs(mass_measurement_error(msnidObj))

# Setting up filter object
filtObj <- MSnIDFilter(msnidObj)
filtObj$absParentMassErrorPPM <- list(comparison="<", threshold=10.0)
filtObj$msmsScore <- list(comparison=">", threshold=10.0)

system.time(
  filtObj.grid <- optimize_filter(filtObj, msnidObj, fdr.max=0.01,
                               method="Grid", level="peptide", n.iter=50))
show(filtObj.grid)

# Fine tuning. Nelder-Mead optimization.
system.time(
  filtObj.nm <- optimize_filter(filtObj.grid, msnidObj, fdr.max=0.01,
                             method="Nelder-Mead", level="peptide",
                             n.iter=50))
show(filtObj.nm)

# Fine tuning. Simulated Annealing optimization.
system.time({
peptides

filtObj.sann <- optimize_filter(filtObj.grid, msnidObj, fdr.max=0.01,  
method="SANN", level="peptide", n.iter=50))
show(filtObj.sann)

peptides  Non-redundant list of peptides

Description

Returns the non-redundant list of peptides from the MSnID object

Usage

peptides(object)

Arguments

object  An instance of class "MSnID".

Value

Non-redundant list of peptides.

Author(s)

Vladislav A Petyuk <vladislav.petyuk@pnnl.gov>

See Also

accessions proteins

Examples

data(c_elegans)
head(peptides(msnidObj))
psms

Peptide-to-spectrum matches

Description

Returns results of MS/MS search (peptide-to-spectrum) matches in the form of data.frame.

Usage

```
psms(object, ...)  
psms(object) <- value
```

Arguments

- **object**: An instance of class "MSnID".
- **value**: Value is a data.frame with MS/MS search results
- **...**: ignored for now

Value

data.frame

Author(s)

Vladislav A Petyuk <vladislav.petyuk@pnnl.gov>

See Also

- MSnID

Examples

```
data(c_elegans)  
msnidDF <- psms(msnidObj)  
head(msnidDF)
```
Description

Reads mzIdentML files into `psms` data.table slot of object MSnID instance. The functionality leverage `mzID` package facility. Note, the calls are memoised using `R.cache` facility. So if the call with the same list of files issues again, the results will be read from cache instead of re-parsing the mzIdentML files.

Usage

```r
read_mzIDs(object, mzids, backend)
```

Arguments

- `object`: An instance of class "MSnID"
- `mzids`: paths to mzIdentML (mzid) files
- `backend`: Package that is leveraged for parsing. Either 'mzID' or 'mzR' corresponding to `mzID-package` and `mzR-class` respectively. The 'mzR' parser is much faster, since it is based on C++ code. 'mzID' will be kept in the package for legacy reasons. Note, the default is 'mzID'.

Details

mzIdentML files can be either as is or in gzip compressed form (*.mzid.gz).

Value

Returns an instance of "MSnID" class with `@psms` data.table slot populated with MS/MS identifications.

Author(s)

Vladislav A Petyuk <vladislav.petyuk@pnnl.gov>

See Also

`flatten mzID`

Examples

```r
## Not run:
msnidObj <- MSnID(".")
mzids <- system.file("extdata","c_elegans.mzid.gz",package="MSnID")
msnidObj <- read_mzIDs(msnidObj, mzids)
# clean up the cache directory
unlink(".Rcache", recursive=TRUE)
```
recalibrate

Post-experimental recalibration of observed mass to charge ratios

Description

Mass spectrometry measurements like any other real-world measurements are prone to systematic errors. Typically they are minimized by instrument calibration prior the analysis. Nonetheless, the calibration may drift over time or be affected by some adverse factors (temperature or space charge fluctuations).

This function estimates and removes the systematic error from the datasets. The side effect is the recalibrated experimentalMassToCharge values.

Usage

recalibrate(object)

Arguments

object

An instance of class "MSnID".

Details

Currently it employs a very simple method of zero-centering the histogram of mass measurement errors. In the future it will contain more sophisticated recalibration routines.

Value

"MSnID" class instance with updated experimentalMassToCharge.

Author(s)

Vladislav A Petyuk <vladislav.petyuk@pnnl.gov>

See Also

MSnID mass_measurement_error correct_peak_selection

Examples

data(c_elegans)

# first let's fix the error of picking wrong monoisotopic peak
# otherwise the mass error range will be very large
msnidObj <- correct_peak_selection(msnidObj)

# original mass error in ppm
remap_accessions <- mass_measurement_error(msnidObj)
hist(remap_accessions, 200, xlim=c(-20,+20))

# The dataset is well calibrated. So let's introduce
# some mass measurement error.
msnidObj$experimentalMassToCharge <-
  msnidObj$experimentalMassToCharge * (1+0.00001)

# mass error (in ppm) after artificial de-calibration
remap_accessions <- mass_measurement_error(msnidObj)
hist(remap_accessions, 200, xlim=c(-20,+20))

# recalibration
msnidObj <- recalibrate(msnidObj)
remap_accessions <- mass_measurement_error(msnidObj)
hist(remap_accessions, 200, xlim=c(-20,+20))

---

remap_accessions

Changes accessions from one protein id to another

Description

Changes accessions from one protein id to another.

Usage

```r
remap_accessions(object, 
  conversion_table, 
  extraction_pttrn=c("\|([^|\-]+)(-\d+)?\|", 
    "([A-Z]P_\d+)", 
    "(ENS[A-Z0-9]+)"), 
  path_to_FASTA=NULL)
```

Arguments

- **object** An instance of class "MSnID".
- **conversion_table** (data.frame) first column in the data frame corresponds to identifiers in the FASTA file. Second column is the new identifier.
- **extraction_pttrn** (string) regex pattern that extract protein identifier from FASTA entry name as first group (that is "\|\|\)). The most common patterns are the one corresponding to UniProt "\|([^|\-]+)\(-\d+)?\|\|", RefSeq "\^([A-Z]P_\d+)" and ENSEMBL "\^ENS[A-Z0-9]+". Other regex patterns can be accepted as well. Default is UniProt pattern.
- **path_to_FASTA** (string) path to FASTA file. If provided only accessions present in the given FASTA file will be retained.
remap_fasta_entry_names

Value

Returns an instance of "MSnID" with updated accessions.

Author(s)

Vladislav A Petyuk <vladislav.petyuk@pnnl.gov>

See Also

remap_fasta_entry_names

Examples

```r
m <- MSnID(".")
mzids <- system.file("extdata", "phospho.mzid.gz", package="MSnID")
m <- read_mzIDs(m, mzids)
head(m$accessions)
conv_tab <- fetch_conversion_table("Homo sapiens", "UNIPROT", "SYMBOL")
m2 <- remap_accessions(m, conv_tab, "\\{[^\|\-]+\}(-\d+)?\|")
head(m2$accessions)
unlink(".Rcache", recursive=TRUE)
```

Description

Remaps entries in the FASTA file from one protein identifier to another according to provided conversion table. Input is a path to FASTA file. Output is also a path to a new FASTA file with updated entry names.

Usage

```r
remap_fasta_entry_names(path_to_FASTA, conversion_table, extraction_pttrn=c("\\{[^\|\-]+\}(-\d+)?\|", "([A-Z]P_\d+)", "(ENS[A-Z0-9]+)"))
```

Arguments

- `path_to_FASTA` (string) path to FASTA file
- `conversion_table` (data.frame) first column in the data frame corresponds to identifiers in the FASTA file. Second column is the new identifier.
(string) regex pattern that extract protein identifier from FASTA entry name as first group (that is "\1"). The most common patterns are the one corresponding to UniProt "\|([\^\-]+)(-\d+)\|", RefSeq "^[A-Z]P_\d+" and ENSEMBL "^\(ENS[A-Z0-9]+\)". Other regex patterns can be accepted as well. Default is UniProt pattern.

Value

path to new FASTA file

Author(s)

Vladislav A Petyuk <vladislav.petyuk@pnnl.gov>

Examples

library(Biostrings)
fst_path <- system.file("extdata", "for_phospho.fasta.gz", package="MSnID")
readAAStringSet(fst_path)
conv_tab <- fetch_conversion_table("Homo sapiens", "UNIPROT", "SYMBOL")
fst_path_2 <- remap_fasta_entry_names(fst_path, conv_tab, "\|([\^\-]+)(-\d+)\|")
readAAStringSet(fst_path_2)
file.remove(fst_path_2)

report_mods

Lists modification masses in the MSnID object

Description

Parses out masses from the modification column and return them as table with counts.

Usage

report_mods(object, ...)

Arguments

object An instance of class "MSnID".

Value

Counts of each modification mass listed in modification column.
See Also

add_mod_symbol map_mod_sites

Examples

msnidObj <- MSnID(".")
mzids <- system.file("extdata","phospho.mzid.gz",package="MSnID")
msnidObj <- read_mzIDs(msnidObj, mzids)
# reports the masses and number of their occurrences
report_mods(msnidObj)
# clean-up cache
unlink(".Rcache", recursive=TRUE)
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