Package ‘PROPER’

April 11, 2024

Type Package
Title PROspective Power Evaluation for RNAseq
Version 1.34.0
Imports edgeR
Depends R (>= 3.3)
Suggests BiocStyle,DESeq2,DSS,knitr
Date 2020-11-25
Author Hao Wu
Maintainer Hao Wu <hao.wu@emory.edu>
Description This package provide simulation based methods for evaluating the statistical power in differential expression analysis from RNA-seq data.
License GPL
biocViews ImmunoOncology, Sequencing, RNASeq, DifferentialExpression
VignetteBuilder knitr
git_url https://git.bioconductor.org/packages/PROPER
git_branch RELEASE_3_18
git_last_commit c294c33
git_last_commit_date 2023-10-24
Repository Bioconductor 3.18
Date/Publication 2024-04-10

R topics documented:

comparePower ................................................................. 2
estParam ................................................................. 4
param ................................................................. 6
plotPowerHist ............................................................ 7
plots ................................................................. 8
power.seqDepth .............................................................. 9
ComparePower

Compute the power-related quantities from simulation results

Description

This function takes the simulation output from "runSims" function and compute a variety of power-related quantities.

Usage

comparePower(simOutput, alpha.type = c("fdr", "pval"), alpha.nominal=0.1,
stratify.by = c("expr", "dispersion"), strata,
filter.by = c("none","expr"), strata.filtered = 1,
target.by=c("lfc", "effectsize"), delta=0.5)

Arguments

simOutput The result from "runSims" function.
alpha.type A string to represent the way to call DE genes. Available options are "fdr" and "pval", for calling DE genes based on FDR or p-values. Default is "fdr".
alpha.nominal The nominal value for call DE genes. Recommend values are 0.1 when using FDR, and 0.01 for using p-values.
stratify.by A string to represent the way to stratify genes. Available options are "expr" and "dispersion", for stratifying gene by average expression levels or over-dispersion.
strata The strata used for stratification.
filter.by A string to represent the way to filter genes. This is used in conjunction with strata.filtered for gene filtering. Available options are "none" and "expr". "none" stands for no filtering, thus all genes will be considered. "expr" stands for filtering based on average expression levels.
strata.filtered The strata to be filtered out in computing power-related quantities. Genes fall into these strata will be excluded when computing power-related quantities. See "Details" for more description of gene filtering.
target.by A string to specify the method to define "biologically important" DE genes. Available options are (1) "lfc": interesting genes are defined by absolute log fold changes. (2) "effectsize": interesting genes are defined by absolute log fold changes divided by the square root of dispersion.
delta A threshold used for defining "biologically important" genes. Genes with absolute log fold changes (when target.by is "lfc") or effect sizes (when target.by is "effectsize") greater than this value are deemed DE in power calculations. See "Details" for more description.
Details

This is the main function to compute various power-related quantities, under stratification and filtering of all genes.

Gene stratification: we advocate to compute and visualize the powers at different stratification of genes. Because of the characteristics of RNA-seq data such as average expression level (counts), powers for calling DE genes are different even when the magnitude of changes are the same. The stratified results will provide a more comprehensive power assessment and better guide the investigators in experimental designs and analysis strategies.

Gene filtering: sometimes it is advisable to filter out some genes (such as the ones with very low count) before DE detection. The filtering option here provides an opportunity to compare the powers before and after filtering.

Define biologically interesting genes: we advocate to compute powers for "biologically interesting genes", because there might be many true DE genes with very low changes which are difficult to detect. In this sense, we are only interested in genes with adequate changes between groups (defined as "biologically interesting"). We provide two options to define biologically interesting genes: by absolute values of log fold changes or effect sizes (absolute values of log fold changes divided by the square root of dispersions). Genes with these quantities over a threshold are deemed interesting, and the power calculation are based on these genes.

Value

A list with following fields:

- TD, FD, alpha, FDR, power
  3D array representing the number of true discoveries, false discoveries, type I error rate, FDR, and power for all simulation settings. The dimension of the arrays are nstrata * N * nsims. Here nstrata is number of specified strata. N is number of different sample sizes settings, and nsims is number of simulations.

- alpha.marginal, FDR.marginal, power.marginal
  Matrix representing the marginal type I error rate, FDR, and power for all simulation settings. The dimension of the matrices are N * nsims. These are the marginalized (over all strata) values of the stratified quantities.

- stratify.by
  The input stratify.by.

- strata
  The input strata.

- Nreps
  Sample sizes one wants to perform simulation on. This is taken from the simulation options.

- target.by
  The input method to define "biologically important" DE genes.

- delta
  The input delta for biologically important genes.

Author(s)

Hao Wu <hao.wu@emory.edu>

See Also

- RNAseq.SimOptions.2grp, simRNAseq, runSims
Examples

```r
## Not run:
simOptions = RNAseq.SimOptions.2grp()
## run a few simulations
simRes = runSims(Nreps=c(3,5,7), sim.opts=simOptions, nsims=5,
    DEmethod="edgeR")

## using FDR 0.1 to call DE, then look at power curves and summary
powers = comparePower(simRes)
summaryPower(powers)
par(mfrow=c(2,2))
plotPower(powers)
plotPowerTD(powers)
plotFDR(powers)
plotFDcost(powers)

## filter out the genes with low counts (<10) and redo power calculation
## Marginal powers are significantly higher.
powers = comparePower(simRes, filter.by="expr", strata.filtered=1)
summaryPower(powers)
par(mfrow=c(2,2))
plotPower(powers)
plotPowerTD(powers)
plotFDR(powers)
plotFDcost(powers)

## Provide higher threshold for log fold change to define true DE.  
## This will result in higher power.
powers2 = comparePower(simRes, delta=2)
summaryPower(powers2)
par(mfrow=c(2,2))
plotPower(powers2)
plotPowerTD(powers2)
plotFDR(powers2)
plotFDcost(powers2)

## use effect size to define biologically interesting genes
powers3 = comparePower(simRes, filter.by="expr", strata.filtered=1,
    target.by="effectsize", delta=1)
summaryPower(powers3)
par(mfrow=c(2,2))
plotPower(powers3)
plotPowerTD(powers3)
plotFDR(powers3)
plotFDcost(powers3)

## End(Not run)
```

---

**estParam**  
estimate simulation parameters from a count table
estParam

Description

This function estimate simulation parameters from a count table, which can be used to set simulation scenarios. The parameters estimated include baseline expression, biological variation in the form of dispersion parameter.

Usage

estParam(X,type = c(1,2))

Arguments

X An ExpressionSet or a matrix, with entries being RNA sequencing counts

type type=1 estimates with minimal assumption based on sample means and variances for each genes. type=2 uses edgeR package.

Details

This is a function that allows a user to establish simulation basis from his/her own data set. Type 1 uses simple average count and sample variance to estimate over dispersion, after normalizing by library size for each sample. Estimated dispersion is bounded at minimum 0.001. Type 2 uses edgeR’s AveLogCPM as the estimate for baseline expression and tagwiseDispersion as the estimate for dispersion.

Value

A list with following fields:

seqDepth Estimated sequencing depth for each sample

lmean A vector of baseline expression rate per gene in log scale, with length nrow(X)

lOD A vector of log dispersion for each gene, with length nrow(X)

Author(s)

Jean Wu <zhijin_wu@brown.edu>

Examples

## Not run:
lmu0=rnorm(20000,5,2)
lOD0=rnorm(20000,-4,1)
lmu=exp(rnorm(20000*10,lmu0,exp(lOD0)))
X=matrix(rpois(20000*10,lmu),20000,10)
param=estParam(X)

## End(Not run)
Some existing RNA-seq and gene expression microarray data

Description

These are several datasets distributed with the package. They are simulation parameters estimated from existing RNA-seq data. The original raw count data were obtained from reCount website.

Usage

data(cheung)
data(gilad)
data(bottomly)
data(maqc)
data(GE.human)
data(pbmc)

Details

The data include:

- cheung: parameters from Cheung data. They are measurements from unrelated individuals, so the dispersions are large.
- gilad: parameters from Gilad data. They are for Human liver sample comparisons between male and female. This dataset has moderate dispersions.
- bottomly: parameters from Bottomly data. They compare two strains of inbred mice and the within group dispersions are small.
- maqc: MAQC data which are technical replicates. There are no biological variation from the replicates, so the dispersions are close to 0.
- GE.human: a vector of aggregated counts from all samples in Cheung data. This is used for generating marginal expression distribution when provide sequencing depth.
- pbmc: data from gene expression microarray data. This is for an example of using historical data to estimate effect sizes.

Author(s)

Hao Wu <hao.wu@emory.edu>
plotPowerHist

Plot the histogram of power

Description
Using histogram-alike graph to visualize the distribution of all genes and DE genes in all user specified strata.

Usage
plotPowerHist(powerOutput, simResult, main = "Histogram of power", return = FALSE)

Arguments
- powerOutput: Result object from "comparePower" function.
- simResult: Result object from "runSims" function.
- main: Figure caption.
- return: Return a matrix for average number of genes and true DE genes in each strata.

Value
A matrix of two rows. First row is for number of genes, and the second row is for number of DE genes. Columns are strata.

Author(s)
Hao Wu <hao.wu@emory.edu>

Examples
## Not run:
simOptions = RNAseq.SimOptions.2grp()
## run a few simulations
simRes = runSims(Nreps=c(3,5,7), sim.opts=simOptions, nsims=5,
                DEmethod="edgeR")

## using FDR 0.1 to call DE, then look at power curves and summary
powers = comparePower(simRes)
plotPowerHist(powers, simRes)

## End(Not run)
plots

Plot the stratified curves for power-related quantities

Description

These are a group of functions to generate plots to visualize the stratified power metrics, including stratified power, FDR, true discoveries, false discoveries, false discovery cost, and type I error. There will be a line for each sample size. X-axis are strata and y-axis are the metrics. Error bars will be plotted if requested.

Usage

plotPower(powerOutput, cols = 1:ncol(powerOutput$FD), lty = 1:ncol(powerOutput$power), main = "", ylab = "Power", leg = TRUE, error.bar=TRUE)

plotFDR(powerOutput, cols = 1:ncol(powerOutput$FDR), lty=1:ncol(powerOutput$FDR), main = "", ylab="FDR", leg = TRUE, error.bar=TRUE)

plotPowerTD(powerOutput, cols = 1:ncol(powerOutput$TD), lty=1:ncol(powerOutput$TD), main="", ylab = "# True Discoveries", leg = TRUE, error.bar=TRUE)

plotPowerFD(powerOutput, cols = 1:ncol(powerOutput$FD), lty=1:ncol(powerOutput$FD), main="", ylab = "# False Discoveries", leg = TRUE, error.bar=TRUE)

plotFDcost(powerOutput, cols = 1:ncol(powerOutput$FD), lty=1:ncol(powerOutput$FD), main="", ylab = "False discovery cost", leg = TRUE, error.bar=TRUE)

plotPowerAlpha(powerOutput, cols = 1:ncol(powerOutput$alpha), lty=1:ncol(powerOutput$alpha), main="", ylab = "False positive rate", leg = TRUE, error.bar=TRUE)

plotAll(powerOutput)

Arguments

- `powerOutput`: Result object from "comparePower" function.
- `cols`, `lty`: Colors and line types of the TD curves.
- `main`: Figure title.
- `ylab`: Y-axis label.
- `leg`: Indicator for having figure legend or not.
- `error.bar`: Indicator for whether plotting error bars in the figures.

Author(s)

Hao Wu <hao.wu@emory.edu>
power.seqDepth

See Also

comparePower

Examples

## Not run:
simOptions = RNAseq.SimOptions.2grp()
## run a few simulations
simRes = runSims(Nreps=c(3,5,7), sim.opts=simOptions, nsims=5, 
        DEmethod="edgeR")

## using FDR 0.1 to call DE, then look at power curves and summary
powers = comparePower(simRes)
## plot
par(mfrow=c(2,3))
plotPower(powers)
plotPowerTD(powers)
plotFDR(powers)
plotFDcost(powers)
plotPowerAlpha(powers)

## in one figure
par(mfrow=c(2,3))
plotAll(powers)

## End(Not run)

power.seqDepth  Estimate the marginal power under different sequencing depth

Description

The function helps study the power under different sequencing depth and sample sizes. It estimates
the marginal powers based on existing simulation results, given new sequencing depth.

Usage

power.seqDepth(simResult, powerOutput, depth.factor = c(0.2, 0.5, 1, 2, 5, 10))

Arguments

  simResult  Result object from "runSims" function.
  powerOutput Result object from "comparePower" function.
  depth.factor A vector of numbers specifying the *relative* sequencing depth, comparing to
               the depth used in the simulation. 1 means using the same number of total reads as the simulation.
Details
The powers under different sequencing depth and sample sizes provides important guidance in
experimental design. Under the same total number of sequence reads, investigator can choose to
use more replicates and shallower coverage for each, or less replicates and deeper coverage.
This function provides estimated marginal power holding all experimental variables fixed (biological
variation, effect sizes, sample sizes, etc.) except the sequencing depth. Changing sequencing
depth will only alter the marginal distribution of average counts. Since the stratified power (by av-
erage counts) won’t change, those numbers are used in estimating the powers under different depth.
This approaches allows skipping new simulations, which saves computation.

Value
A matrix for marginal powers. Each row is for a sequencing depth, each columns is for a sample
size.

Author(s)
Hao Wu <hao.wu@emory.edu>

See Also
comparePower, summary.power

Examples
## Not run:
simOptions = RNAseq.SimOptions.2grp()
simRes = runSims(Nreps=c(3,5,7), sim.opts=simOptions, nsims=5,
               DEmethod="edgeR")
powers = comparePower(simRes)
power.seqDepth(simRes, powers)
## End(Not run)
Arguments

ngenes Number of genes in the simulation.

lBaselineExpr log baseline expression for each gene. This can be: (1) a constant; (2) a vector with length ngenes; (3) a function that takes an integer n, and generate a vector of length n; (4) a string specifying the name of existing datasets, from which the mean expressions will be sampled. Details for this option is provide in "Details" section.

lOD log over-dispersion for each gene. Available options are the same as for lBaselineExpr.

seqDepth Sequencing depth, in terms of total read counts. This will be ignored if lBaselineExpr is specified.

p.DE Percentage of genes being differentially expressed (DE). By default it’s 5%.

lfc log-fold change for DE genes. This can be: (1) a constant; (2) a vector with length being number of DE genes; (3) a function that takes an integer n, and generate a vector of length n. If the input is a vector and the length is not the number of DE genes, it will be sampled with replacement to generate log-fold change.

sim.seed Simulation seed.

Details

The simulation of RNA-seq data requires a lot of parameters. This function provides users an interface to specify the simulation parameters. The result from this function will be used for simulating RNA-seq count data. By default, the simulation parameters are similar to that from Cheung data (for unrelated individuals, with large biological variance).

The baseline expression levels and log over-dispersions can be sampled from real data. There are parameters estimated from several real datasets distributed with the package. Available string options for "lBaselineExpr" and "lOD" include: (1) "cheung": parameters from Cheung data, which measures unrelated individuals, so the dispersions are large; (2) "gilad": from Gilad data which are for Human liver sample comparisons between male and female. This dataset has moderate dispersions; (3) "bottomly": from Bottmly data which are from comparing two strains of inbred mice. The dispersions are small. (4) "maqc": from MAQC data which are technical replicates. There are no biological variation from the replicates because the data are technical replicates. The dispersions from this dataset is very small.

The effect sizes (log fold changes of the DE genes) are arbitrarily specified. It is possible to estimate those from real data. We provide a simple example in the package vignette for doing so.

Value

A list with following fields:

ngenes An integer for number of genes.

p.DE Percentage of DE genes.

lBaselineExpr A vector of length ngenes for log baseline expression.

lOD A vector of length ngenes for log over-dispersion.
runSims

Description

This is the “wrapper” function for running RNA-seq DE detection simulation. It runs simulations under different sample sizes (replicates in each group) for a certain numbers. In each simulation, the RNA-seq data are generated, and then DE detection (using user specified method/software) is performed. The return object contains DE test results (test statistics, p-values, FDRs, etc.) from all simulations.

Usage

runSims(Nreps = c(3, 5, 7, 10), Nreps2, nsims = 100, sim.opts, 
DEmethod = c("edgeR", "DSS", "DESeq2"), verbose =TRUE)
runSims

Arguments

Nreps  Sample sizes one wants to perform simulation on. This is a vector, each element specifies the number of biological replicates in each group. Default value is c(3, 5, 7, 10), which means we want to perform simulation when there are 3, 5, 7, or 10 replicates in each group.

Nreps2  Sample sizes for the second treatment group. If this is missing, it'll take the same value as Nreps and then two groups will be assumed to have the same sample size. This parameter allows two treatment groups have different sample sizes. When specified, it must be a vector of the same length as Nreps.

nsims  Number of simulations.

sim.opts  An object for simulation option. This should be the return object from "RNAseq.SimOptions.2grp" function.

DEmethod  String to specify the DE detection method to be used. Available options are "edgeR", "DSS", and "DESeq2".

verbose  Logical value to indicate whether to output some messages (progress report).

Details

This is the main simulation function in the package. After specifying the simulation parameters (from "RNAseq.SimOptions.2grp" function), one wants to evaluate the power vs different sample sizes under that simulation setting. This function simulates the count data and performs statistical tests for DE detection. It only stores and returns the DE test results (test statistics, p-values, FDRs, etc.) but doesn’t make inferences. The inferences will be conducted in the "comparePower" function. The advantage is that for one simulation setting, the simulation only need to be run once. The inferences using different critical values and type I error controls can then be drawn from the same results. This greatly save the computation because the simulation part is the most computationally intensive.

This function can be slow, depends on the setting (number of genes, replicates, simulations, etc). For the default (50000 genes, 100 simulations, for 3, 5, 7, or 10 replicates), it takes about an hour to run on a single core of i7 2.7GHz CPU. But again, this only need to be run once for a particular simulation setting.

Value

A list with following fields:

pvalue, fdrs  3D array for p-values and FDR from each simulation. The dimension of the array is ngenes * N * nsims. Here N is length(Nreps), of the number of different sample sizes settings.

xbar  3D array for average read counts for genes. Dimension is the same as pvalue/fdr. This will be used in "comparePower" function for stratified power quantities.

DEid  A list of length nsims. Each contains the id of DE genes.

lfcs  A list of length nsims. Each contains the log fold changes of DE genes.

Nreps  The input Nreps.

sim.opts  The input sim.opts.
simRNAseq

Simulate RNA-seq count data

Description

This function generate simulated RNA-seq count data for a two-group comparison design. It takes an object for simulation option (return object from 'RNAseq.SimOptions.2grp' function, and sample sizes (replicates in each condition, then generate a matrix of counts.

Usage

simRNAseq(simOptions, n1, n2)

Arguments

simOptions: An object for simulation option. This should be the return object from 'RNAseq.SimOptions.2grp' function.
n1, n2: Sample size in two treatment groups.
**Value**

A list with following fields:

- **counts**: A matrix of dimension ngenes x (n1+n2). Each row is for a gene, each column is for a sample. Columns 1 to n1 are for the first condition. The rest columns are for the other condition.

- **designs**: A vector/matrix representing the experimental designs. In a two-group comparison, it's a simple 0/1 vector.

- **DEid**: The ID (row index) for true DE genes.

- **simOptions**: The simulation option object. This is exactly the same as the input simOptions.

**Author(s)**

Hao Wu <hao.wu@emory.edu>

**See Also**

RNAseq.SimOptions.2grp, runSims

**Examples**

```r
simOptions = RNAseq.SimOptions.2grp()
data = simRNAseq(simOptions, n1=3, n2=3)
names(data)
```

---

**summaryPower**

*Generate a summary table for power analysis result*

**Description**

Takes a result object from "comparePower" function and print out a table to summarize important power-related quantities. The results are marginalized, meaning that they are averaged quantities over all strata and simulations. This provides a quick view of the marginal results for power-related quantities under all sample sizes.

**Usage**

`summaryPower(powerOutput)`

**Arguments**

- **powerOutput**: The result object from "comparePower" function.
Value

A matrix with multiple rows, each for a different sample size. Columns include sample size, specified nominal type I control value (for FDR or p-values), actual FDR, marginal power, averaged number of true and false discoveries, and false discovery costs.

Author(s)

Hao Wu <hao.wu@emory.edu>

See Also

runSims, comparePower

Examples

```r
## Not run:
simOptions = RNAseq.SimOptions.2grp()
simRes = runSims(Nreps=c(3,5,7), sim.opts=simOptions, nsims=5, DEmethod="edgeR")
powers = comparePower(simRes)
summaryPower(powers)
## End(Not run)
```
Index

bottomly (param), 6
cheung (param), 6
comparePower, 2
estParam, 4
GE.human (param), 6
gilad (param), 6
maqc (param), 6
param, 6
pbmc (param), 6
plotAll (plots), 8
plotFDCost (plots), 8
plotFDR (plots), 8
plotPower (plots), 8
plotPowerAlpha (plots), 8
plotPowerFD (plots), 8
plotPowerHist, 7
plotPowerTD (plots), 8
plots, 8
power.seqDepth, 9
RNAseq.SimOptions.2grp, 10
runSims, 12
simRNAseq, 14
summaryPower, 15