Package ‘clipper’

May 29, 2024

Version 1.44.0
Date 2019-04-09
Title Gene Set Analysis Exploiting Pathway Topology
Author Paolo Martini <paolo.cavei@gmail.com>, Gabriele Sales <gabriele.sales@unipd.it>, Chiara Romualdi <chiara.romualdi@unipd.it>
Maintainer Paolo Martini <paolo.cavei@gmail.com>
Description Implements topological gene set analysis using a two-step empirical approach. It exploits graph decomposition theory to create a junction tree and reconstruct the most relevant signal path. In the first step clipper selects significant pathways according to statistical tests on the means and the concentration matrices of the graphs derived from pathway topologies. Then, it “clips” the whole pathway identifying the signal paths having the greatest association with a specific phenotype.
Depends R (>= 2.15.0), Matrix, graph
Imports methods, Biobase, Rcpp, igraph, gRbase (>= 1.6.6), qpgraph, KEGGgraph, corpcor
Suggests RUnit, BiocGenerics, graphite, ALL, hgu95av2.db, MASS, BiocStyle
Enhances RCy3
License AGPL-3

git_url https://git.bioconductor.org/packages/clipper
git_branch RELEASE_3_19
git_last_commit 0ebdc8d
git_last_commit_date 2024-04-30
Repository Bioconductor 3.19
Date/Publication 2024-05-29

Contents

clipper ................................................................. 2
**Description**

Basing on either variance or mean clique test, this function identifies the paths that are mostly related with the phenotype under study.

**Usage**

```r
clipper(expr, classes, graph, method=c("variance","mean", "both", "paired"), nperm=100, alphaV=0.05, b=100, root=NULL, trZero=0.001, signThr=0.05, maxGap=1, permute=TRUE, alwaysShrink=FALSE)
```

**Arguments**

- `expr` an expression matrix or ExpressionSet with colnames for samples and row name for genes.
- `classes` vector of 1, 2 indicating the classes of samples (columns).
- `graph` a graphNEL object.
- `method` the kind of test to perform on the cliques. It could be mean, variance, mixed (the best between variance and mean) or paired mean.
- `nperm` number of permutations. Default = 100.
- `alphaV` pvalue threshold for variance test to be used during mean test. Default = 0.05.
- `b` number of permutations for mean analysis. Default = 100.
- `root` nodes by which rip ordering is performed (as far as possible) on the variables using the maximum cardinality search algorithm.
- `trZero` lowest pvalue detectable. This threshold avoids that -log(p) goes infinite.
**clipper**

- **signThr**: significance threshold for clique pvalues.
- **maxGap**: allow up to maxGap gaps in the best path computation. Default = 1.
- **permute**: always performs permutations in the concentration matrix test. If FALSE, the test is made using the asymptotic distribution of the log-likelihood ratio. This option should be used only if samples size is >=40 per class.
- **alwaysShrink**: always perform the shrinkage estimates of variance.

**Details**

The both method combines the results obtained from the mean and variance test. In particular it assign to the cliques the minimum of mean and variance p-values.

**Value**

A matrix with a row for each paths. Columns are organized as follows:

1. Index of the starting clique
2. Index of the ending clique
3. Index of the clique where the maximum value is reached
4. Length of the path
5. Maximum score of the path
6. Average score along the path
7. Percentage of path activation
8. Impact of the path on the entire pathway
9. Cliques involved and significant
10. Cliques forming the path
11. Genes forming the significant cliques
12. Genes forming the path

**References**


Massa MS, Chiogna M, Romualdi C. Gene set analysis exploiting the topology of a pathway. BMC System Biol. 2010 Sep 1;4:121.

**See Also**

cliqueVarianceTest, cliqueMeanTest, getJunctionTreePaths
clipperAllRoots

Examples

```r
if (require(graphite) & require(ALL)){
  kegg <- pathways("hsapiens", "kegg")
  graph <- pathwayGraph(convertIdentifiers(kegg$'Chronic myeloid leukemia', "entrez"))
  genes <- nodes(graph)
  data(ALL)
  all <- ALL[1:length(genes),1:20]
  classes <- c(rep(1,10), rep(2,10))
  featureNames(all@assayData)<- genes
  graph <- subGraph(genes, graph)
  clipped <- clipper(all, classes, graph, "var", trZero=0.01, permute=FALSE)
  clipped[,1:5]
}
```

clipperAllRoots

Dissect the pathway to find the path with the greatest association with phenotype.

Description

Basing on either variance or mean clique test, this function identifies the paths that are mostly related with the phenotype under study.

Usage

```r
clipperAllRoots(expr, classes, graph, method=c("variance","mean", "both", "paired"), nperm=100, alphaV=0.05, b=100, trZero=0.001, signThr=0.05, maxGap=1, permute=TRUE, alwaysShrink=FALSE)
```

Arguments

- `expr`: an expression matrix or ExpressionSet with colnames for samples and row name for genes.
- `classes`: vector of 1,2 indicating the classes of samples (columns).
- `graph`: a graphNEL object.
- `method`: the kind of test to perform on the cliques. It could be mean, variance, mixed (the best between variance and mean) or paired mean.
- `nperm`: number of permutations. Default = 100.
- `alphaV`: pvalue threshold for variance test to be used during mean test. Default = 0.05.
- `b`: number of permutations for mean analysis. Default = 100.
- `trZero`: lowest pvalue detectable. This threshold avoids that -log(p) goes infinite.
- `signThr`: significance threshold for clique pvalues.
- `maxGap`: allow up to maxGap gaps in the best path computation. Default = 1.
- `permute`: always performs permutations in the concentration matrix test. If FALSE, the test is made using the asymptotic distribution of the log-likelihood ratio. This option should be use only if samples size is >=40 per class.
- `alwaysShrink`: always perform the shrinkage estimates of variance.
Details

The both method combines the results obtained from the mean and variance test. In particular it assign to the cliques the minimum of mean and variance p-values.

Value

A matrix with a row for each paths. Rownames have the form: roots-paths.

Columns are organized as follows:

1. Index of the starting clique
2. Index of the ending clique
3. Index of the clique where the maximum value is reached
4. Length of the path
5. Maximum score of the path
6. Average score along the path
7. Percentage of path activation
8. Impact of the path on the entire pathway
9. Cliques involved and significant
10. Cliques forming the path
11. Genes forming the significant cliques
12. Genes forming the path

References


Massa MS, Chiogna M, Romualdi C. Gene set analysis exploiting the topology of a pathway. BMC System Biol. 2010 Sep 1;4:121.

See Also

cliqueVarianceTest, cliqueMeanTest, getJunctionTreePaths

Examples

if (require(graphite) & require(ALL)){
  kegg <- pathways("hsapiens", "kegg")
  graph <- pathwayGraph(convertIdentifiers(kegg$'Chronic myeloid leukemia', "entrez"))
  genes <- nodes(graph)
  data(ALL)
  all <- ALL[1:length(genes),1:20]
  classes <- c(rep(1,10), rep(2,10))
  featureNames(all@assayData)<- genes
  graph <- subGraph(genes, graph)
  clipped <- clipperAllRoots(all, classes, graph, "var", trZero=0.01, permute=FALSE)
cliqueMeanTest  

Mean test for cliques.

Description

It decomposes the graph in cliques and performs the mean test in every one.

Usage

cliqueMeanTest(expr, classes, graph, nperm, alphaV=0.05, b=100, root=NULL, permute=TRUE, alwaysShrink=FALSE)

Arguments

expr an expression matrix or ExpressionSet with colnames for samples and row name for genes.
classes vector of 1,2 indicating the classes of samples (columns).
graph a graphNEL object.
nperm number of permutations.
alphaV pvalue threshold for variance test to be used during mean test.
b number of permutations for mean analysis.
root nodes by which rip ordering is performed (as far as possible) on the variables using the maximum cardinality search algorithm.
permute always performs permutations in the concentration matrix test. If FALSE, the test is made using the asymptotic distribution of the log-likelihood ratio. This option should be use only if samples size is >=40 per class.
alwaysShrink always perform the shrinkage estimates of variance.

Value

a list with alphas (vector of cliques pvalues based on the mean test) and cliques (list of the cliques and related elements).

References


Massa MS, Chiogna M, Romualdi C. Gene set analysis exploiting the topology of a pathway. BMC System Biol. 2010 Sep 1;4:121.

See Also

cliqueVarianceTest.


Examples

```r
if (require(graphite) & require(ALL)){
  kegg <- pathways("hsapiens", "kegg")
  graph <- pathwayGraph(convertIdentifiers(kegg$'Chronic myeloid leukemia', "entrez"))
  genes <- nodes(graph)
  data(ALL)
  all <- ALL[1:length(genes),1:20]
  classes <- c(rep(1,10), rep(2,10))
  featureNames(all@assayData)<- genes
  graph <- subGraph(genes, graph)
  cliqueMeanTest(all, classes, graph, nperm=100, permute=FALSE)$alpha
}
```

---

cliqueMixedTest

Mean test for cliques.

Description

It decomposes the graph in cliques and performs the combination of mean e variance test in every one.

Usage

```r
cliqueMixedTest(expr, classes, graph, nperm, alphaV=0.05, b=100, 
root=NULL, permute=TRUE, alwaysShrink=FALSE)
```

Arguments

- `expr`: an expression matrix or ExpressionSet with colnames for samples and row name for genes.
- `classes`: vector of 1,2 indicating the classes of samples (columns).
- `graph`: a graphNEL object.
- `nperm`: number of permutations.
- `alphaV`: pvalue threshold for variance test to be used during mean test.
- `b`: number of permutations for mean analysis.
- `root`: nodes by which rip ordering is performed (as far as possible) on the variables using the maximum cardinality search algorithm.
- `permute`: always performs permutations in the concentration matrix test. If FALSE, the test is made using the asymptotic distribution of the log-likelihood ratio. This option should be use only if samples size is >=40 per class.
- `alwaysShrink`: always perform the shrinkage estimates of variance.

Details

The method combines the results obtained from the mean and variance test. In particular it assign to the cliques the minimum of mean and variance p-values.
cliquePairedTest

Value

a list with alphas (vector of cliques pvalues based on the variance test) and cliques (list of the cliques and related elements).

References

Massa MS, Chiogna M, Romualdi C. Gene set analysis exploiting the topology of a pathway. BMC System Biol. 2010 Sep 1;4:121.

See Also

cliqueVarianceTest.

Examples

if (require(graphite) & require(ALL)){
  kegg <- pathways("hsapiens", "kegg")
  graph <- pathwayGraph(convertIdentifiers(kegg$'Chronic myeloid leukemia', "entrez"))
  genes <- nodes(graph)
  data(ALL)
  all <- ALL[1:length(genes),1:20]
  classes <- c(rep(1,10), rep(2,10))
  featureNames(all@assayData)<- genes
  graph <- subGraph(genes, graph)
  cliqueMeanTest(all, classes, graph, nperm=100, permute=FALSE)$alpha
}

cliquePairedTest             Paired mean test for cliques.

Description

It decomposes the graph in cliques and performs the paired mean test in every one.

Usage

cliquePairedTest(expr, classes, graph, nperm, alphaV=0.05, b=100, root=NULL, permute=TRUE, alwaysShrink=FALSE)

Arguments

expr                  an expression matrix or ExpressionSet with colnames for samples and row name for genes.
classes               vector of 1,2 indicating the classes of samples (columns). It is assumed that class labels are ordered so that the first occurrence of class 2 is paired with the first occurrence of class 1 and so on.
cliquePairedTest

graph a graphNEL object.
nperm number of permutations.
alphaV pvalue threshold for variance test to be used during mean test.
b number of permutations for mean analysis.
root nodes by which rip ordering is performed (as far as possible) on the variables using the maximum cardinality search algorithm.
permute always performs permutations in the concentration matrix test. If FALSE, the test is made using the asymptotic distribution of the log-likelihood ratio. This option should be use only if samples size is >=40 per class.
alwaysShrink always perform the shrinkage estimates of variance.

Value

a list with alphas (vector of cliques pvalues based on the variance test) and cliques (list of the cliques and related elements).

References


Massa MS, Chiogna M, Romualdi C. Gene set analysis exploiting the topology of a pathway. BMC System Biol. 2010 Sep 1;4:121.

See Also

cliqueVarianceTest.

Examples

```
if (require(graphite) & require(ALL)){
  kegg <- pathways("hsapiens", "kegg")
  graph <- pathwayGraph(convertIdentifiers(kegg$'Chronic myeloid leukemia', "entrez"))
  genes <- nodes(graph)
  data(ALL)
  all <- ALL[1:length(genes),1:20]
  classes <- c(rep(1,10), rep(2,10))
  featureNames(all@assayData)<- genes
  graph <- subGraph(genes, graph)
  cliquePairedTest(all, classes, graph, nperm=100, permute=FALSE)$alpha
}
```
cliqueVarianceTest  Variance test for cliques.

Description

It decomposes the graph in cliques and performs the variance test in every one.

Usage

cliqueVarianceTest(expr, classes, graph, nperm, alphaV=0.05, b=100, root=NULL, permute=TRUE, alwaysShrink=FALSE)

Arguments

expr  an expression matrix or ExpressionSet with colnames for samples and row name for genes.

classes  vector of 1,2 indicating the classes of samples (columns).

graph  a graphNEL object.

nperm  number of permutations.

alphaV  pvalue threshold for variance test to be used during mean test.

b  number of permutations for mean analysis.

root  nodes by which rip ordering is performed (as far as possible) on the variables using the maximum cardinality search algorithm.

permute  always performs permutations in the concentration matrix test. If FALSE, the test is made using the asymptotic distribution of the log-likelihood ratio. This option should be use only if samples size is >=40 per class.

alwaysShrink  always perform the shrinkage estimates of variance.

Value

a list with alphas (vector of cliques pvalues based on the variance test) and cliques (list of the cliques and related elements).

References


Massa MS, Chiogna M, Romualdi C. Gene set analysis exploiting the topology of a pathway. BMC System Biol. 2010 Sep 1;4:121.

See Also

cliqueMeanTest.
Examples

```r
if (require(graphite) & require(ALL)) {
  kegg <- pathways("hsapiens", "kegg")
  graph <- pathwayGraph(convertIdentifiers(kegg$'Chronic myeloid leukemia', "entrez"))
  genes <- nodes(graph)
  data(ALL)
  all <- ALL[1:length(genes),1:20]
  classes <- c(rep(1,10), rep(2,10))
  featureNames(all@assayData) <- genes
  graph <- subGraph(genes, graph)
  cliqueVarianceTest(all, classes, graph, nperm=100, permute=FALSE)$alpha
}
```

---

**deleteEdge**

*Remove an edge from graphNEL object.*

**Description**

Remove from a graphNEL object the edge specified.

**Usage**

```r
deleteEdge(graph, from, to)
```

**Arguments**

- `graph`: a graphNEL object.
- `from`: a string with the name of the node where the edge start.
- `to`: a string with the name of the node where the edge end.

**Value**

a graphNEL object.

**Examples**

```r
if (require(graphite)) {
  kegg <- pathways("hsapiens", "kegg")
  graph <- pathwayGraph(convertIdentifiers(kegg$'Chronic myeloid leukemia', "entrez"))
  head(edges(graph))
  ## We are going to remove the edge 1026-1019
  head(edges(deleteEdge(graph, "ENTREZID:1026", "ENTREZID:1019")))
}
```
easyClip

**Easy clip analysis.**

**Description**

Easy clip function allows the full exploitation of Clipper Package features in a unique and easy to use function. Starting from an expression matrix and a pathway, these function extract the most transcriptionally altered portions of the graph.

**Usage**

```r
easyClip(expr, classes, graph, method=c("variance","mean"), pathThr=0.05, pruneLevel=0.2, nperm=100, alphaV=0.05, b=100, root=NULL, trZero=0.001, signThr=0.05, maxGap=1, permute=TRUE)
```

**Arguments**

- `expr`: an expression matrix or ExpressionSet with colnames for samples and row name for genes.
- `classes`: vector of 1,2 indicating the classes of samples (columns).
- `graph`: a graphNEL object.
- `method`: the kind of test to perform on the cliques. It could be either mean or variance.
- `pathThr`: The significance threshold of the whole pathway test. Default = 0.05
- `pruneLevel`: a dissimilarity threshold. NULL means no pruning.
- `nperm`: number of permutations. Default = 100.
- `alphaV`: pvalue threshold for variance test to be used during mean test. Default = 0.05.
- `b`: number of permutations for mean analysis. Default = 100.
- `root`: nodes by which rip ordering is performed (as far as possible) on the variables using the maximum cardinality search algorithm.
- `trZero`: lowest pvalue detectable. This threshold avoids that -log(p) goes infinite.
- `signThr`: significance threshold for clique pvalues.
- `maxGap`: allow up to maxGap gaps in the best path computation. Default = 1.
- `permute`: always performs permutations in the concentration matrix test. If FALSE, the test is made using the asymptotic distribution of the log-likelihood ratio. This option should be use only if samples size is >=40 per class.

**Value**

A matrix with row as the different paths. Columns are organized as follows: 1 - Index of the starting clique 2 - Index of the ending clique 3 - Index of the clique where the maximum value is reached 4 - length of the path 5 - maximum score of the path 6 - average score along the path 7 - percentage of path activation 8 - impact of the path on the entire pathway 9 - clique involved and significant 10 - clique forming the path 11 - genes forming the significant cliques 12 - genes forming the path.
easyLook

References
Massa MS, Chiogna M, Romualdi C. Gene set analysis exploiting the topology of a pathway. BMC System Biol. 2010 Sep 1;4:121.

See Also
cliqueVarianceTest, cliqueMeanTest, getJunctionTreePaths

Examples
if (require(graphite) & require(ALL)){
  kegg <- pathways("hsapiens", "kegg")
  graph <- pathwayGraph(convertIdentifiers(kegg$'Chronic myeloid leukemia', "entrez"))
  genes <- nodes(graph)
  data(ALL)
  all <- ALL[1:length(genes),1:24]
  classes <- c(rep(1,12), rep(2,12))
  featureNames(all@assayData)<- genes
  graph <- subGraph(genes, graph)
  easyClip(all, classes, graph, nperm=10)
}

easyLook(clipped)  Summarize clipper output.

Description
Summarization of the result for a quick look of clipper function.

Usage
easyLook(clipped)

Arguments
clipped  the output of either clipper o easyClip.

Value
Nice formatted output of clipper function.

References
Massa MS, Chiogna M, Romualdi C. Gene set analysis exploiting the topology of a pathway. BMC System Biol. 2010 Sep 1;4:121.
getGraphEntryGenes  
Extract all the possible entry point (genes with no entering edges) from graph.

Description

It extracts the possible entry point of the graph. Entry points are defined as nodes with no entering edges.

Usage

getGraphEntryGenes(graph, byCliques=FALSE, root=NULL)

Arguments

graph  
a graphNEL object.
byCliques  
when TRUE it returns a list where entry point are organized by cliques.
root  
nodes by which rip ordering is performed (as far as possible) on the variables using the maximum cardinality search algorithm.

Value

a vector of gene names representing the entry point of graph.

References


Massa MS, Chiogna M, Romualdi C. Gene set analysis exploiting the topology of a pathway. BMC System Biol. 2010 Sep 1;4:121.

Examples

if (require(graphite)) {
  kegg <- pathways("hsapiens", "kegg")
  graph <- pathwayGraph(convertIdentifiers(kegg$'Chronic myeloid leukemia', "entrez"))
  getGraphEntryGenes(graph)
}
getJunctionTreePaths  

Extract the shortest paths along the junction tree of the graph.

Description

Find the shortest paths in the Junction tree designed with the cliques of the graph.

Usage

getJunctionTreePaths(graph, root=NULL)

Arguments

graph  
a graphNEL object.

root  
nodes by which rip ordering is performed (as far as possible) on the variables using the maximum cardinality search algorithm.

Value

list of clique indices representing the shortest paths of the graph.

References


Massa MS, Chiogna M, Romualdi C. Gene set analysis exploiting the topology of a pathway. BMC System Biol. 2010 Sep 1;4:121.

Examples

if (require(graphite)) {
  kegg <- pathways("hsapiens", "kegg")
  graph <- pathwayGraph(convertIdentifiers(kegg$'Chronic myeloid leukemia', "entrez"))
  getJunctionTreePaths(graph)
}

nameCliques  
Generate clique names from their own elements.

Description

Starting from the sorted elements of each clique of the list, this function generates names fusing in a string the element names.
Usage

name Cliques(cliques)

Arguments

cliques a list where each element is a clique.

Value

vector of strings

Examples

toy Cliques <- list(c(45,36,90), c(36,1000,35))
name Cliques(toy Cliques)

pathwayTest Whole pathway test using qpipf.

Description

Performs variance and mean test using qpipf on the whole pathway.

Usage

pathQ(expr, classes, graph, nperm=100, alphaV=0.05, b=100,
permute=TRUE, paired=FALSE, alwaysShrink=FALSE)

Arguments

expr an expression matrix or ExpressionSet with colnames for samples and rownames
for expression features.
classes vector of 1,2 indicating the classes of the samples (columns).
graph a graphNEL object.
nperm number of permutations. Default = 100.
alphaV pvalue significance threshold for variance test to be used during mean test. De-
default = 0.05.
b number of permutations for mean analysis. Default = 100.
permute always performs permutations in the concentration matrix test. If FALSE, the
test is made using the asymptotic distribution of the log-likelihood ratio. This
option should be use only if samples size is >=40 per class.
paired perform the test for paired sample. It assumes that class labels are ordered so
that the first occurrence of class 2 is paired with the first occurrence of class 1
and so on.
alwaysShrink always perform the shrinkage estimates of variance.
plotInCytoscape

Value

a list with alphaVar (pvalue for the variance test) and alphaMean (pvalue for mean test).

Note

This function is based on the Gaussian Graphical Models and to use it in a proper way it is necessary that the graph is an Direct Acyclic Graph. Please check any graph in input using isAcyclic from ggm package.

References


Massa MS, Chiogna M, Romualdi C. Gene set analysis exploiting the topology of a pathway. BMC System Biol. 2010 Sep 1;4:121.

Examples

```r
if (require(graphite) & require(ALL)){
  kegg <- pathways("hsapiens", "kegg")
  graph <- pathwayGraph(convertIdentifiers(kegg$'Chronic myeloid leukemia', "entrez"))
  genes <- nodes(graph)
  data(ALL)
  all <- ALL[1:length(genes),1:24]
  classes <- c(rep(1,12), rep(2,12))
  featureNames(all@assayData)<- genes
  graph <- subGraph(genes, graph)
  pathQ(all, classes, graph, nperm=100, permute=FALSE)
}
```

Description

Plot a pathway graph in Cytoscape highlighting the relevant path.

Usage

```r
plotInCytoscape(graph, path, color="#6699FF", main="graph")
```

Arguments

- `graph`: a graphNEL object.
- `path`: vector summarizing a path (a rows of clipper output matrix).
- `color`: color code string: genes of the most involved fragment will be colored using color. Default = "#6699FF"
- `main`: a graph name to be used in Cytoscape. Default = 'graph'
Details

Requires the RCy3 package.

See Also

clipper

Examples

```r
## Not run: if (require(graphite)) {
  if (requireNamespace("RCy3")){
    kegg <- pathways("hsapiens", "kegg")
    graph <- pathwayGraph(convertIdentifiers(kegg$'Chronic myeloid leukemia', "entrez"))
  }
  path <- c(3,17,5,9,13.04,2.60,0.209,0.321,"6,7,8,9,10", "3,5,6,7,8,9,10,14,17", "ENTREZID:1029;ENTREZID:4193;ENTREZID:7157", "ENTREZID:1019;ENTREZID:1021;ENTREZID:1026;ENTREZID:1029;ENTREZID:595")
  plotInCytoscape(graph,path)
}
## End(Not run)
```

---

**prunePaths**

Summarize the paths obtained by clipper according to their similarity.

Description

This function allows the user to chose only one representant of those paths that have more than 1-thr similarity. The best scoring path is choosen.

Usage

```r
prunePaths(pathSummary, thr=NULL, clust=NULL, sep=";")
```

Arguments

- `pathSummary`: a matrix resulting from clipper function.
- `thr`: a dissimilarity threshold. NULL means no pruning.
- `clust`: filename where path-cluster is saved. NULL means no cluster saved.
- `sep`: the separator to split genes for similarity computation. Default = ;

Value

a matrix

See Also

clipper
Examples

\[
\text{toyEx} \leftarrow \text{matrix(c(1,1,5,3,5,2,5,3,8,2,3,2,1,0.3,0.1,1,2,1,"1;2;3;4;5","1;2;3","1;2;3;4;5","1;2;3","1;2;3;4;5","1;2;3"),2,12)}
\]

\[
\text{row.names(toyEx) } \leftarrow \text{c("1;5","1;3")}
\]

\text{toyEx}

\text{prunePaths(toyEx, thr=0.1)
Index

clipper, 2, 18
clipperAllRoots, 4
cliqueMeanTest, 3, 5, 6, 10, 13
cliqueMixedTest, 7
cliquePairedTest, 8
cliqueVarianceTest, 3, 5, 6, 8, 9, 10, 13
deleteEdge, 11
easyClip, 12
easyLook, 13
getGraphEntryGenes, 14
getJunctionTreePaths, 3, 5, 13, 15

nameCliques, 15

pathQ(pathwayTest), 16
pathwayTest, 16
plotInCytoscape, 17
prunePaths, 18