Package ‘crisprBase’

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Description Provides S4 classes for general nucleases, CRISPR nucleases, CRISPR nickases, and base editors. Several CRISPR-specific genome arithmetic functions are implemented to help extract genomic coordinates of spacer and protospacer sequences. Commonly-used CRISPR nuclease objects are provided that can be readily used in other packages. Both DNA- and RNA-targeting nucleases are supported.
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annotateMismatches

Annotate mismatches between spacer and protospacer sequences

Description

Annotate mismatches between spacer and protospacer sequences.

Usage

annotateMismatches(spacers, protospacers, rnase = FALSE)
**Arguments**

- **spacers**: A character vector specifying spacer sequences (gRNA).
- **protospacers**: A character vector specifying protospacer sequences (target DNA).
- **rnase**: Is it for an RNAse? FALSE by default. If TRUE, spacers and protospacers are expected to be the reverse complement of each other.

**Value**

A data.frame storing spacer and protospacer columns, as well as number of mismatches, and positions for the different mismatches, if any. Positions are relative to the 5’ end of the spacer sequences. For RNAses (e.g. CasRx), this means that a mismatch at position 1 corresponds to the last nucleotide of the protospacer sequence.

**Author(s)**

Jean-Philippe Fortin

**Examples**

```r
spacers <- c("CCGGAGCGAGTTGCAGTAAGCAG",
              "GCCGGAGCGAGTTGCAGTAAGCA",
              "GGCCGGAGCGAGTTGCAGTAAGC")
protospacers=c("CTGCTTACTGCAACTCGCTCTGG",
               "TGCTTAATGCAACCCGCTCCGGC",
               "GCTTACTGCAACTCGCTCCGGCC")
ann <- annotateMismatches(spacers, protospacers, rnase=TRUE)
```

**Description**

CrisprNuclease object for the Wildtype Acidaminococcus Cas12a (AsCas12a) nuclease.

**Usage**

data(AsCas12a, package="crisprBase")

**Format**

CrisprNuclease object.
Details

The AsCas12a nuclease recognizes TTTV PAM sequences. Spacer sequences must be located downstream of PAM sequences.

An S4 class to represent a base editor

Usage

baseEditorName(object)

baseEditorName(object) <- value

editingWeights(object, ...)

editingWeights(object) <- value

editingStrand(object, ...)

editingStrand(object) <- value

BaseEditor(
  CrisprNuclease,
  baseEditorName = NA_character_,
  editingStrand = c("original", "opposite"),
  editingWeights = NULL
)

## S4 method for signature 'BaseEditor'
show(object)

## S4 method for signature 'BaseEditor'
baseEditorName(object)

## S4 replacement method for signature 'BaseEditor'
baseEditorName(object) <- value

## S4 method for signature 'BaseEditor'
editingWeights(object, substitutions = NULL)

## S4 replacement method for signature 'BaseEditor'
editingWeights(object) <- value
## S4 method for signature 'BaseEditor'
editingStrand(object)

## S4 replacement method for signature 'BaseEditor'
editingStrand(object) <- value

### Arguments
- **object**: BaseEditor object.
- **value**: Value to replaced with.
- **...**: Additional arguments for class-specific methods
- **CrisprNuclease**: A CrisprNuclease object.
- **baseEditorName**: String specifying base editor name.
- **editingStrand**: String indicating which strand with respect to the target protospacer sequence will be edited. Must be either "original" or "opposite". "original" by default.
- **editingWeights**: Numeric matrix of editing weights. Column names must be indicating relative position to the PAM site. Row names must be of the form "X2Y" where "X" represents the origin base, and "Y" represents the subtituted base. For instance, "C2T" indicates the row corresponding to C to T editing.
- **substitutions**: Character vector indicating which substitutions should be returned.

### Value
A BaseEditor object

### Functions
- **BaseEditor()**: Create a BaseEditor object

### Slots
- **baseEditorName**: Name of the base editor.
- **editingWeights**: Matrix of editing weights.
- **editingStrand**: String indicating which strand with respect to the target protospacer sequence will be edited. Must be either "original" or "opposite". "original" by default.

### Constructors
Use the constructor link{BaseEditor} to create a BaseEditor object.

### Accessors
- **baseEditorName**: To get the name of the base editor.
- **editingWeights**: To return the matrix of editing weights.
- **editingStrand**: To return the editing strand.
Setters

baseEditorName<-: To change the name of the base editor.
editingWeights<-: To change the matrix of editing weights.
editingStrand<-: To change the editing strand.

Examples

# Creating an object for BE4max (C to T editor)
# based on experimental weights

ws <- c(0.7, 0.7, 0.8, 1.8, 1, 2, 1.4, 1.2, 2.3, 1.3, 2.4, 2.2, 3.4,
  2.2, 2.1, 3.5, 5.8, 16.2, 31.8, 63.2, 90.3, 100, 87, 62, 31.4,
  16.3, 10, 5.6, 3.3, 1.9, 1.8, 2.4, 1.7, 0.5, 0.2, 0.1)
ws <- matrix(ws, nrow=1, ncol=length(ws))
rownames(ws) <- "C2T"
colnames(ws) <- -36:-1
data(SpCas9, package="crisprBase")
BE4max <- BaseEditor(SpCas9,
  baseEditorName="BE4max",
  editingStrand="original",
  editingWeights=ws)
metadata(BE4max)$description_base_editor <- "BE4max cytosine base editor."

BE4max

BE4max BaseEditor object

Description

BaseEditor for the cytosine base editor CRISPR/Cas9 system BE4max. Editing weights were obtained from https://doi.org/10.1016/j.cell.2020.05.037

Usage

data(BE4max, package="crisprBase")

Format

BaseEditor object.

Details

BaseEditor for the cytosine base editor CRISPR/Cas9 system BE4max. Editing weights were obtained from https://doi.org/10.1016/j.cell.2020.05.037.
**CrisprNuclease-class**

An S4 class to represent a CRISPR nuclease.

**Description**

CrisprNuclease object for the Cas13d-NLS from Ruminococcus flavefaciens strain XPD3002 nuclease (RNase).

**Usage**

```r
data(CasRx, package = "crisprBase")
```

**Format**

CrisprNuclease object.

**Details**

The CasRx nuclease was derived from Cas13d Ruminococcus flavefaciens string XPD3002. See [10.1016/j.cell.2018.02.033](https://doi.org/10.1016/j.cell.2018.02.033).

**CrisprNickase-class**

An S4 class to represent a CRISPR nickase.

**Description**

An S4 class to represent a CRISPR nickase.

**Usage**

```r
CrisprNickase(
    nickaseName,
    nickingStrand = c("original", "opposite"),
    pams = NA_character_,
    weights = rep(1, length(pams)),
    metadata = list(),
    pam_side = NA_character_,
    spacer_gap = 0L,
    spacer_length = NA_integer_
)
```

```
## S4 method for signature 'CrisprNickase'
show(object)
```

```
## S4 method for signature 'CrisprNickase'
pamLength(object)
```
## S4 method for signature 'CrisprNickase'
spacelength(object)

## S4 replacement method for signature 'CrisprNickase'
spacelength(object) <- value

## S4 method for signature 'CrisprNickase'
pamSide(object)

## S4 replacement method for signature 'CrisprNickase'
pamSide(object) <- value

## S4 method for signature 'CrisprNickase'
spacergap(object)

## S4 replacement method for signature 'CrisprNickase'
spacergap(object) <- value

## S4 method for signature 'CrisprNickase'
hasSpacergap(object)

## S4 method for signature 'CrisprNickase'
targetlength(object)

## S4 method for signature 'CrisprNickase'
pams(object, primary = TRUE, ignore_pam = FALSE, as.character = FALSE)

## S4 method for signature 'CrisprNickase'
pamIndices(object)

## S4 method for signature 'CrisprNickase'
spacerIndices(object)

## S4 method for signature 'CrisprNickase'
prototypeSequence(object, primary = TRUE)

### Arguments

**nickaseName** Name of the CRISPR nickase.

**nickingStrand** String specifying with strand with respect to the motif sequence (5' to 3') is nicked. Must be either "original" (default) or "opposite".

**pams** Character vector of PAM sequence motifs written from 5' to 3. If the point of cleavage has been determined, the precise site is marked with ^. Only letters in the IUPAC code are accepted. For nickases that cleave away from their recognition sequence, the cleavage sites are indicated in parentheses. See details for more information.
weights  Optional numeric vector specifying relative weights of the PAM sequences to specify cleavage probabilities.
metadata  Optional list providing global metadata information.
pam_side  String specifying the side of the PAM sequence sequence with respect to the protospacer sequence. Must be either '3prime' (e.g. Cas9) or '5prime' (e.g. Cas12a)
spacer_gap  Integer specifying the length (in nucleotides) between the spacer sequence and the PAM sequence (e.g. 0 for Cas9 and Cas12a).
spacer_length  Integer specifying the length of the spacer sequence
object  CrisprNickase object.
value  For spacerLength<- and gapLength<-, must be a non-negative integer. For pamSide, must be either '5prime' or '3prime'.
primary  Should only the PAM sequence with the heighest weight be returned? If no cleavage weights are stored in the CrisprNickase object, all sequences are returned. TRUE by default.
ignore_pam  Should all possible k-mer sequences for a given PAM length be returned, irrespectively of the PAM sequence motifs stored in the CrisprNickase object? FALSE by default.
as.character  Should the PAM sequences be returned as a character vector? FALSE by default.

Value
A CrisprNickase object

Functions
- CrisprNickase(): Create a CrisprNickase object

Slots
- pam_side  String specifying the side of the PAM sequence with respect to the protospacer sequence. Must be either '3prime' (e.g. SpCas9) or '5prime' (e.g. AsCas12a)
- spacer_length  Integer specifying the length of the spacer sequence
- spacer_gap  Integer specifying the length (in nucleotides) between the spacer sequence and the PAM sequence (e.g. 0 for SpCas9 and AsCas12a).

Constructors
Use the constructor link(CrisprNickase) to create a CrisprNickase object.

Accessors
- nickaseName: To get the name of the CRISPR nickase.
- spacerLength: To return the length of the spacer sequence.
- targetLength: To return the length of the target sequence (protospacer + pam).
pamLength: To return the length of the PAM sequence.
pamSide: To return the side of the PAM sequence with respect to the spacer sequence.
spacerGap: To return the length of the gap between the PAM and spacer sequences.
pams: To return the list of PAM sequences.

Setters

spacerGap<-: To change the length of the gap between the PAM and spacer sequences.
pamSide<-: To change the side of the PAM sequence with respect to the protospacer sequence.
spacerLength<-: To change the length of the spacer sequence.

Utility functions for genomic arithmetics

pamIndices: To return the relative coordinates of the PAM sequence within the protospacer sequence.
spacerIndices: To return the relative coordinates of the spacer sequence within the protospacer sequence.

Examples

Cas9D10A <- CrisprNickase("Cas9D10A",
    nickingStrand="opposite",
    pams=c("(3)NGG", "(3)NAG", "(3)NGA"),
    weights=c(1, 0.2593, 0.0694),
    metadata=list(description="D10A-mutated Streptococcus pyogenes Cas9 (SpCas9) nickase"),
    pam_side="3prime",
    spacer_length=20)

Cas9H840A <- CrisprNickase("Cas9H840A",
    nickingStrand="original",
    pams=c("(3)NGG", "(3)NAG", "(3)NGA"),
    weights=c(1, 0.2593, 0.0694),
    metadata=list(description="H840A-mutated Streptococcus pyogenes Cas9 (SpCas9) nickase"),
    pam_side="3prime",
    spacer_length=20)

enAsCas12a

CrisprNuclease object for the Enhanced Acidaminococcus Cas12a (AsCas12a) nuclease.
Usage

data(enAsCas12a, package="crisprBase")

Format

CrisprNuclease object.

Details

The enAsCas12a nuclease recognizes an extended set of PAM sequences beyond the canonical TTTV sequence for AsCas12a. Spacer sequences must be located downstream of PAM sequences.

Description

Extract PAM sequences from target sequences (protospacer + PAM) using information stored in a CrisprNuclease object.

Usage

extractPamFromTarget(targets, object)

Arguments

targets Character vector of target sequences.
oxject CrisprNuclease corresponding to the target sequences.

Value

Character vector of PAM sequences of length equal to that of the targets character vector.

Author(s)

Jean-Philippe Fortin

Examples

data(SpCas9, AsCas12a, package="crisprBase")

# Extracting PAM sequences from Cas9 protospacers:
targets <- c("AGGTGCTGATTGTAGTGCTGCGG", "AGGTGCTGATTGTAGTGCTGAGG")
exttractPamFromTarget(targets, SpCas9)

# Extracting PAM sequences from Cas12a targets:
targets <- c("TTTAAGGTGCTGATTGTAGTGCTGTGT", "TTTCAAGGTGCTGATTGTAGTGCTGAAA")
extectPamFromTarget(targets, AsCas12a)
extractProtospacerFromTarget

Extract protospacer sequences from target sequences

Description

Extract protospacer sequences from target sequences (protospacer + PAM) using information stored in a CrisprNuclease object.

Usage

extractProtospacerFromTarget(targets, object)

Arguments

targets Character vector of targets sequences.
object CrisprNuclease corresponding to the targets sequences.

Value

Character vector of protospacer sequences of length equal to that of the targets character vector.

Author(s)

Jean-Philippe Fortin

Examples

data(SpCas9, AsCas12a, package="crisprBase")
# Extracting protospacer sequences from Cas9 targets:
targets <- c("AGGTGCTGATTGTAGTGCTGCGG",
             "AGGTGCTGATTGTAGTGCTGAGG")
exttractProtospacerFromTarget(targets, SpCas9)
# Extracting protospacer sequences from Cas12a targets:
targets <- c("TTTAAGGTGCTGATTGTAGTGCTGTGT",
             "TTTCAGGTGCTGATTGTAGTGCTGAAA")
exttractProtospacerFromTarget(targets, AsCas12a)
getAvailableCrisprNucleases

Return list of available CrisprNuclease objects in crisprBase

Description
Return list of available CrisprNuclease objects in crisprBase.

Usage
getAvailableCrisprNucleases()

Value
Character vector of available CrisprNuclease objects found in crisprBase.

Author(s)
Jean-Philippe Fortin

Examples
getAvailableCrisprNucleases()

getCutSiteFromPamSite
Return cut site coordinates from PAM site coordinates

Description
Return cut site coordinates from PAM site coordinates.

Usage
getCutSiteFromPamSite(pam_site, strand, nuclease = NULL)

Arguments

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>pam_site</td>
<td>Coordinate of the first nucleotide of the PAM sequence.</td>
</tr>
<tr>
<td>strand</td>
<td>Either &quot;+&quot; or &quot;.&quot;.</td>
</tr>
<tr>
<td>nuclease</td>
<td>A CrisprNuclease object.</td>
</tr>
</tbody>
</table>

Value
numeric vector of cut sites
Author(s)

Jean-Philippe Fortin

Examples

data(SpCas9, package="crisprBase")
getCutSiteFromPamSite(pam_site=100, strand="+", nuclease=SpCas9)
gETCHSiteFromPamSite(pam_site=100, strand="-", nuclease=SpCas9)

getCutSiteRanges

Construct a cut site GRanges from a list of PAM sites

Description

Construct a cut site GRanges from a list of PAM sites using information stored in a CrisprNuclease object.

Usage

getCutSiteRanges(
  gr = NULL,
  seqnames = NULL,
  pam_site = NULL,
  strand = NULL,
  nuclease = NULL
)

Arguments

gr
  GRanges object of width 1 specifying the coordinates of the first nucleotide of the PAM sequences.

seqnames
  Character vector of genomic sequence names. Ignored if gr is not NULL.

pam_site
  Numeric vector specifying the coordinates of the first nucleotide of the PAM sequences corresponding to the PAM sequences. Ignored if gr is not NULL.

strand
  Character vector specifying the strand of the PAM. Ignored if gr is not NULL.

nuclease
  CrisprNuclease object.

Value

GRanges object representing genomic coordinates of the cut sites.

Author(s)

Jean-Philippe Fortin
getEditingSiteFromPamSite

Return optimal editing site coordinates from PAM site coordinates

Description

Return optimal editing site coordinates from PAM site coordinates.

Usage

getEditingSiteFromPamSite(
  pam_site,
  strand,
  baseEditor = NULL,
  substitution = NULL
)

Arguments

pam_site Coordinate of the first nucleotide of the PAM sequence.
strand Either "+" or ".".
baseEditor A BaseEditor object.
substitution String indicating which substitution should be used to estimate the optimal editing position. E.g. "C2T" will return the optimal editing position for C to T editing.

Value

numeric vector of editing sites.

Author(s)

Jean-Philippe Fortin

Examples

data(BE4max, package="crisprBase")
getEditingSiteFromPamSite(pam_site=100, strand="+", baseEditor=BE4max, "C2T")
getPamRanges

Construct a PAM GRanges from a list of PAM sites

Description

Construct a PAM GRanges from a list of PAM sites using information stored in a CrisprNuclease object.

Usage

getPamRanges(
  gr = NULL,
  seqnames = NULL,
  pam_site = NULL,
  strand = NULL,
  nuclease = NULL
)

Arguments

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>gr</td>
<td>GRanges object of width 1 specifying the coordinates of the first nucleotide of the PAM sequences.</td>
</tr>
<tr>
<td>seqnames</td>
<td>Character vector of genomic sequence names. Ignored if gr is not NULL.</td>
</tr>
<tr>
<td>pam_site</td>
<td>Numeric vector specifying the coordinates of the first nucleotide of the PAM sequences corresponding to the PAM sequences. Ignored if gr is not NULL.</td>
</tr>
<tr>
<td>strand</td>
<td>Character vector specifying the strand of the PAM. Ignored if gr is not NULL.</td>
</tr>
<tr>
<td>nuclease</td>
<td>CrisprNuclease object.</td>
</tr>
</tbody>
</table>

Value

GRanges object representing genomic coordinates of PAM sequences.

Author(s)

Jean-Philippe Fortin

Examples

data(SpCas9, AsCas12a, package="crisprBase")
library(GenomicRanges)
gr <- GRanges("chr10",
  IRanges(start=c(100,120), width=1),
  strand=c("+","-"))
getPamRanges(gr, nuclease=SpCas9)
getPamRanges(gr, nuclease=AsCas12a)
**getProtospacerRanges**

*Construct a protospacer GRanges from a list of PAM sites*

**Description**

Construct a protospacer GRanges from a list of PAM sites using information stored in a CrisprNuclease object.

**Usage**

```r
getProtospacerRanges(
  gr = NULL,
  seqnames = NULL,
  pam_site = NULL,
  strand = NULL,
  nuclease = NULL,
  spacer_len = NULL
)
```

**Arguments**

- `gr` : GRanges object of width 1 specifying the coordinates of the first nucleotide of the PAM sequences.
- `seqnames` : Character vector of genomic sequence names. Ignored if `gr` is not NULL.
- `pam_site` : Numeric vector specifying the coordinates of the first nucleotide of the PAM sequences corresponding to the protospacers. Ignored if `gr` is not NULL.
- `strand` : Character vector specifying the strand of the protospacer. Ignored if `gr` is not NULL.
- `nuclease` : CrisprNuclease object.
- `spacer_len` : Non-negative integer to overwrite the default spacer length stored in the CrisprNuclease object.

**Value**

GRanges object representing genomic coordinates of protospacer sequences.

**Author(s)**

Jean-Philippe Fortin

**Examples**

```r
data(SpCas9, AsCas12a, package="crisprBase")
library(GenomicRanges)
gr <- GRanges("chr10",
  IRanges(start=c(100,120), width=1),
```

---

**getProtospacerRanges**

*Construct a protospacer GRanges from a list of PAM sites*

**Description**

Construct a protospacer GRanges from a list of PAM sites using information stored in a CrisprNuclease object.

**Usage**

```r
getProtospacerRanges(
  gr = NULL,
  seqnames = NULL,
  pam_site = NULL,
  strand = NULL,
  nuclease = NULL,
  spacer_len = NULL
)
```

**Arguments**

- `gr` : GRanges object of width 1 specifying the coordinates of the first nucleotide of the PAM sequences.
- `seqnames` : Character vector of genomic sequence names. Ignored if `gr` is not NULL.
- `pam_site` : Numeric vector specifying the coordinates of the first nucleotide of the PAM sequences corresponding to the protospacers. Ignored if `gr` is not NULL.
- `strand` : Character vector specifying the strand of the protospacer. Ignored if `gr` is not NULL.
- `nuclease` : CrisprNuclease object.
- `spacer_len` : Non-negative integer to overwrite the default spacer length stored in the CrisprNuclease object.

**Value**

GRanges object representing genomic coordinates of protospacer sequences.

**Author(s)**

Jean-Philippe Fortin

**Examples**

```r
data(SpCas9, AsCas12a, package="crisprBase")
library(GenomicRanges)
gr <- GRanges("chr10",
  IRanges(start=c(100,120), width=1),
```
getTargetRanges

\[
\text{strand}=c("+","-")
\]
getProtospercerRanges(gr, nuclease=SpCas9)
getProtospercerRanges(gr, nuclease=AsCas12a)

### getTargetRanges

**Construct a target GRanges from a list of PAM sites**

**Description**

Construct a target (protospacer + PAM) GRanges from a list of PAM sites using information stored in a CrisprNuclease object.

**Usage**

```r
getTargetRanges(
  gr = NULL,
  seqnames = NULL,
  pam_site = NULL,
  strand = NULL,
  nuclease = NULL,
  spacer_len = NULL
)
```

**Arguments**

- `gr` - GRanges object of width 1 specifying the coordinates of the first nucleotide of the PAM sequences.
- `seqnames` - Character vector of genomic sequence names. Ignored if `gr` is not NULL.
- `pam_site` - Numeric vector specifying the coordinates of the first nucleotide of the PAM sequences corresponding to the targets. Ignored if `gr` is not NULL.
- `strand` - Character vector specifying the strand of the target. Ignored if `gr` is not NULL.
- `nuclease` - CrisprNuclease object.
- `spacer_len` - Non-negative integer to overwrite the default spacer length stored in the CrisprNuclease object.

**Value**

GRanges object representing genomic coordinates of the target sequences.

**Author(s)**

Jean-Philippe Fortin
MAD7

Examples

data(SpCas9, AsCas12a, package="crisprBase")
library(GenomicRanges)
gr <- GRanges("chr10",
  IRanges(start=c(100,120), width=1),
  strand=c("+","-"))
getTargetRanges(gr, nuclease=SpCas9)
getTargetRanges(gr, nuclease=AsCas12a)

MAD7

MAD7 CrisprNuclease object

Description

CrisprNuclease object for the MAD7 nuclease (Cas12a-like nuclease)

Usage

data(MAD7, package="crisprBase")

Format

CrisprNuclease object.

Details

The MAD7 nuclease recognizes YTTV PAM sequences. Spacer sequences must be located down-stream of PAM sequences.

motifs

An S4 class to represent a nuclease.

Description

Return motif string representations of recognition sites.

Return length of the recognition sites sequences.
Usage

motifs(object, ...)

motifLength(object, ...)

nucleaseName(object)

targetType(object)

weights(object, ...)

nucleaseName(object) <- value

targetType(object) <- value

weights(object) <- value

cutSites(object, ...)

isCutting(object)

isRnase(object)

isDnase(object)

Nuclease(
  nucleaseName,
  targetType = c("DNA", "RNA"),
  motifs = NULL,
  cutSites = NULL,
  weights = rep(1, length(motifs)),
  metadata = list()
)

## S4 method for signature 'Nuclease'
show(object)

## S4 method for signature 'Nuclease'
nucleaseName(object)

## S4 replacement method for signature 'Nuclease'
nucleaseName(object) <- value

## S4 method for signature 'Nuclease'
targetType(object)

## S4 replacement method for signature 'Nuclease'
targetType(object) <- value
## S4 method for signature 'Nuclease'
weights(object, expand = FALSE)

## S4 replacement method for signature 'Nuclease'
weights(object) <- value

## S4 method for signature 'Nuclease'
isCutting(object)

## S4 method for signature 'Nuclease'
isRnase(object)

## S4 method for signature 'Nuclease'
isDnase(object)

## S4 method for signature 'Nuclease'
motifs(
  object,
  primary = FALSE,
  strand = c("+", "-"),
  expand = FALSE,
  as.character = FALSE
)

## S4 method for signature 'Nuclease'
motifLength(object)

## S4 method for signature 'Nuclease'
cutSites(object, strand = c("+", "-", "both"), combine = TRUE, middle = FALSE)

### Arguments

- **object**: Nuclease object.
- **...**: Additional arguments for class-specific methods.
- **value**: New value to pass to the setter functions.
- **nucleaseName**: Name of the nuclease.
- **targetType**: String specifying target type ("DNA" or "RNA").
- **motifs**: Character vector of recognition sequence motifs written from 5’ to 3’ written in Rebase convention. If the point of cleavage has been determined, the precise site is marked with ^\(^\). Only letters in the IUPAC code are accepted. For nuclease that cleave away from their recognition sequence, the cleavage sites are indicated in parentheses. See details for more information.
- **cutSites**: Matrix with 2 rows (+ and - strand, respectively) specifying the cleavage coordinates relative to the first nucleotide of the motif sequence. Each column corresponds to a motif specified in the motifs slot.
weights | Optional numeric vector specifying relative weights for the recognition motifs to specify cleavage probabilities.
metadata | Optional list providing global metadata information.
expand | Should sequences be expanded to only contain ATCG nucleotides? FALSE by default.
primary | Should only the motif with the highest weight be returned? FALSE by default. Only relevant if weights are stored in the Nuclease object.
strand | Strand to allow reverse complementation of the motif. "+" by default.
as.character | Should the motif sequences be returned as a character vector? FALSE by default.
combine | Should only unique values be considered? TRUE by default.
middle | For staggered cuts, should the middle point between the cut on the forward strand and the cut on the reverse strand be considered as the cut site? FALSE by default.

Value

A Nuclease object

Functions

- Nuclease(): Create a Nuclease object

Slots

nucleaseName | Name of the nuclease.
targetType | Character string indicating target type ("DNA" or "RNA").
motifs | DNASTringSet of recognition sequence motifs written from 5’ to 3’.
cutSites | Matrix with 2 rows (+ and - strand, respectively) specifying the cleavage coordinates relative to the first nucleotide of the motif sequence. Each column corresponds to a motif specified in the motifs slot.
weights | Optional numeric vector specifying relative weights for the motifs corresponding to cleavage probabilities.
metadata | Optional string providing a description of the nuclease.

Constructors

Use the constructor link{Nuclease} to create a Nuclease object.

Accessors

nucleaseName: To get the name of the nuclease.
targetType: To get the target type ("DNA" or "RNA").
metadata: To get the metadata list of the nuclease.
motifs: To get the recognition motif nucleotide sequences.
weights: To get nuclease weights.
cutSites: To get nuclease cut sites.
See Also

See the CrisprNuclease for CRISPR-specific nucleases.

Examples

    EcoRI <- Nuclease("EcoRI",
                      motifs=c("G^AATTC"),
                      metadata=list(description="EcoRI restriction enzyme"))

nickaseName

An S4 class to represent a nickase

Description

An S4 class to represent a nickase

Usage

    nickaseName(object)
    nickaseName(object) <- value
    nickingStrand(object)
    nickingStrand(object) <- value

    Nickase(
        nickaseName, 
        nickingStrand = c("original", "opposite"), 
        motifs = NULL, 
        cutSites = NULL, 
        weights = rep(1, length(motifs)), 
        metadata = list()
    )

    ## S4 method for signature 'Nickase'
    show(object)

    ## S4 method for signature 'Nickase'
    nickaseName(object)

    ## S4 replacement method for signature 'Nickase'
    nickaseName(object) <- value

    ## S4 method for signature 'Nickase'
    nickingStrand(object)
## S4 replacement method for signature 'Nickase'

```r
nickingStrand(object) <- value
```

## S4 method for signature 'Nickase'

```r
weights(object, expand = FALSE)
```

## S4 replacement method for signature 'Nickase'

```r
weights(object) <- value
```

## S4 method for signature 'Nickase'

```r
isCutting(object)
```

## S4 method for signature 'Nickase'

```r
motifs(
    object,
    primary = FALSE,
    strand = c("+", "-"),
    expand = FALSE,
    as.character = FALSE
)
```

## S4 method for signature 'Nickase'

```r
motifLength(object)
```

## S4 method for signature 'Nickase'

```r
cutSites(object, combine = TRUE)
```

### Arguments

- **object**  
  Nickase object.

- **value**  
  New value to pass to the setter functions.

- **nickaseName**  
  Name of the nickase.

- **nickingStrand**  
  String specifying with strand with respect to the motif sequence (5’ to 3’) is nicked. Must be either "original" (default) or "opposite".

- **motifs**  
  Character vector of recognition sequence motifs written from 5’ to 3’ written in Rebase convention. If the point of cleavage has been determined, the precise site is marked with ^. Only letters in the IUPAC code are accepted. For nickases that cleave away from their recognition sequence, the cleavage sites are indicated in parentheses. See details for more information.

- **cutSites**  
  Vector specifying the cleavage coordinates relative to the first nucleotide of the motif sequence. Each column corresponds to a motif specified in the motifs slot.

- **weights**  
  Optional numeric vector specifying relative weights for the recognition motifs to specify cleavage probabilities.

- **metadata**  
  Optional list providing global metadata information.
**nickaseName**

expand Should sequences be expanded to only contain ATCG nucleotides? FALSE by default.

primary Should only the motif with the highest weight be returned? FALSE by default. Only relevant if weights are stored in the Nickase object.

strand Strand to allow reverse complementation of the motif. "+" by default.

as.character Should the motif sequences be returned as a character vector? FALSE by default.

combine Should only unique values be considered? TRUE by default.

**Value**

A Nickase object

**Functions**

- Nickase(): Create a Nickase object

**Slots**

- nickaseName: Name of the nickase
- motifs: DNAStringSet of recognition sequence motifs written from 5’ to 3’.
- nickingStrand: String specifying with strand with respect to the motif sequence (5’ to 3’) is nicked. Must be either “original” (default) or “opposite”.
- cutSites: Vector specifying the cleavage coordinates relative to the first nucleotide of the motif sequence. Each column corresponds to a motif specified in the motifs slot.
- weights: Optional numeric vector specifying relative weights for the motifs corresponding to cleavage probabilities.
- metadata: Optional string providing a description of the nickase.

**Constructors**

Use the constructor link(Nickase) to create a Nickase object.

**Accessors**

- nickaseName: To get the name of the nickase.
- nickingStrand: To get the nicking strand.
- metadata: To get the metadata list of the nickase
- motifs: To get the recognition motif nucleotide sequences.
- weights: To get nickase weights.
- cutSites: To get nickase cut sites.

**See Also**

See the CrisprNickase for CRISPR-specific nickases.
plotEditingWeights

Examples

```r
Nb.BsmI <- Nickase("Nb.BsmI",
  motifs=c("GAATG^C"),
  nickingStrand="opposite",
  metadata=list(description="Nb.BsmI nicking enzyme."))
```

---

### Description

Quick plot to visualize editing weights from a BaseEditor object.

### Usage

```r
plotEditingWeights(
  baseEditor,
  discardEmptyRows = TRUE,
  substitutions = NULL,
  ...
)
```

### Arguments

- **baseEditor**: A `BaseEditor` object.
- **discardEmptyRows**: Should rows that have all weight equal to 0 be discarded? TRUE by default.
- **substitutions**: Character vector specifying substitutions to be plotted. If NULL (default), all substitutions are shown.
- **...**: Additional arguments to be passed to `plot`

### Value

Nothing. A plot is generated as a side effect.

### Examples

```r
if (interactive()){  
data(BE4max, package="crisprBase")  
plotEditingWeights(BE4max)  
}
```
**reexports**

*Objects exported from other packages*

### S4Vectors

**metadata, metadata<-**

### restrictionEnzymes

*List of Nuclease objects representing common restriction enzymes*

#### Description

List of Nuclease objects representing common restriction enzymes from REBASE database.

#### Usage

```r
data(restrictionEnzymes, package="crisprBase")
```

#### Format

List of Nuclease objects.

#### Details

List of Nuclease objects representing common restriction enzymes from REBASE database.

### SaCas9

*SaCas9 CrisprNuclease object*

#### Description

CrisprNuclease object for the wildtype Staphylococcus aureus Cas9 (SaCas9) nuclease.

#### Usage

```r
data(SaCas9, package="crisprBase")
```

#### Format

CrisprNuclease object.

#### Details

The AsCas9 nuclease recognizes NNGRRT PAM sequences. Spacer sequences must be located upstream of PAM sequences. Editing weights were obtained from doi:10.1038/nature14299.
An S4 class to represent a CRISPR nuclease.

Usage

- `spacerLength(object, ...)`
- `targetLength(object, ...)`
- `pamLength(object, ...)`
- `spacerGap(object)`
- `hasSpacerGap(object)`
- `spacerGap(object) <- value`
- `spacerLength(object) <- value`
- `pamSide(object, ...)`
- `pamSide(object) <- value`
- `pams(object, ...)`
- `pamIndices(object, ...)`
- `spacerIndices(object, ...)`
- `prototypeSequence(object, ...)`

```r
CrisprNuclease(
  nucleaseName, 
  targetType = c("DNA", "RNA"),
  pams = NA_character_,
  weights = rep(1, length(pams)),
  metadata = list(),
  pam_side = NA_character_,
  spacer_gap = 0L,
  spacer_length = NA_integer_
)
```

## S4 method for signature 'CrisprNuclease'
show(object)
## S4 method for signature 'CrisprNuclease'
pamLength(object)
## S4 method for signature 'CrisprNuclease'
spacerLength(object)
## S4 replacement method for signature 'CrisprNuclease'
spacerLength(object) <- value
## S4 method for signature 'CrisprNuclease'
pamSide(object)
## S4 replacement method for signature 'CrisprNuclease'
pamSide(object) <- value
## S4 method for signature 'CrisprNuclease'
spacerGap(object)
## S4 replacement method for signature 'CrisprNuclease'
spacerGap(object) <- value
## S4 method for signature 'CrisprNuclease'
hasSpacerGap(object)
## S4 method for signature 'CrisprNuclease'
targetLength(object)
## S4 method for signature 'CrisprNuclease'
pams(object, primary = TRUE, ignore_pam = FALSE, as.character = FALSE)
## S4 method for signature 'CrisprNuclease'
pamIndices(object)
## S4 method for signature 'CrisprNuclease'
spacerIndices(object)
## S4 method for signature 'CrisprNuclease'
prototypeSequence(object, primary = TRUE)

Arguments

<table>
<thead>
<tr>
<th>Name</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>object</td>
<td>CrisprNuclease object.</td>
</tr>
<tr>
<td>...</td>
<td>Additional arguments for class-specific methods</td>
</tr>
<tr>
<td>value</td>
<td>For spacerLength&lt;- and gapLength&lt;-, must be a non-negative integer. For pamSide, must be either '5prime' or '3prime'.</td>
</tr>
<tr>
<td>nucleaseName</td>
<td>Name of the CRISPR nuclease.</td>
</tr>
</tbody>
</table>
targetType    String specifying target type ("DNA" or "RNA").
pams          Character vector of PAM sequence motifs written from 5’ to 3. If the point of cleavage has been determined, the precise site is marked with ^. Only letters in the IUPAC code are accepted. For nucleases that cleave away from their recognition sequence, the cleavage sites are indicated in parentheses. See details for more information.
weights       Optional numeric vector specifying relative weights of the PAM sequences to specify cleavage probabilities.
metadata      Optional list providing global metadata information.
pam_side      String specifying the side of the PAM sequence with respect to the protospacer sequence. Must be either '3prime' (e.g. SpCas9) or '5prime' (e.g. AsCas12a).
spacer_gap    Integer specifying the length (in nucleotides) between the spacer sequence and the PAM sequence (e.g. 0 for SpCas9 and AsCas12a).
spacer_length Integer specifying the length of the spacer sequence
primary       Should only the PAM sequence with the highest weight be returned? If no cleavage weights are stored in the CrisprNuclease object, all sequences are returned. TRUE by default.
ignore_pam    Should all possible k-mer sequences for a given PAM length be returned, irrespectively of the PAM sequence motifs stored in the CrisprNuclease object? FALSE by default.

as.character  Should the PAM sequences be returned as a character vector? FALSE by default.

Value
A CrisprNuclease object

Functions
- CrisprNuclease(): Create a CrisprNuclease object

Slots
- pam_side    String specifying the side of the PAM sequence with respect to the protospacer sequence. Must be either '3prime' (e.g. SpCas9) or '5prime' (e.g. AsCas12a)
- spacer_length Integer specifying the length of the spacer sequence
- spacer_gap   Integer specifying the length (in nucleotides) between the spacer sequence and the PAM sequence (e.g. 0 for SpCas9 and AsCas12a).

Constructors
Use the constructor link(CrisprNuclease) to create a CrisprNuclease object.
SpCas9

Accessors

nucleaseName: To get the name of the CRISPR nuclease.
spacerLength: To return the length of the spacer sequence.
targetLength: To return the length of the target sequence (protospacer + pam).
pamLength: To return the length of the PAM sequence.
pamSide: To return the side of the PAM sequence with respect to the spacer sequence.
spacerGap: To return the length of the gap between the PAM and spacer sequences.
pams: To return the list of PAM sequences.

Setters

spacerGap<-: To change the length of the gap between the PAM and spacer sequences.
pamSide<-: To change the side of the PAM sequence with respect to the protospacer sequence.
spacerLength<-: To change the length of the spacer sequence.

Utility functions for genomic arithmetics

pamIndices: To return the relative coordinates of the PAM sequence within the target sequence.
spacerIndices: To return the relative coordinates of the spacer sequence within the target sequence.

Examples

SpCas9 <- CrisprNuclease("SpCas9",
  pams=c("(3/3)NGG", "(3/3)NAG", "(3/3)NGA"),
  weights=c(1, 0.2593, 0.0694),
  metadata=list(description="Wildtype Streptococcus pyogenes Cas9 (SpCas9) nuclease"),
  pam_side="3prime",
  spacer_length=20)

SpCas9

SpCas9 CrisprNuclease object

Description

CrisprNuclease object for the wildtype Streptococcus pyogenes Cas9 (SpCas9) nuclease.

Usage

data(SpCas9, package="crisprBase")

Format

CrisprNuclease object.
Details

The SpCas9 nuclease recognizes NGG PAM sequences. Spacer sequences must be located upstream of PAM sequences.

Description

CrisprNuclease object for the engineered Streptococcus pyogenes Cas9 SpG nuclease.

Usage

data(SpGCas9, package="crisprBase")

Format

CrisprNuclease object.

Details

The SpGCas9 nuclease recognizes NGG PAM sequences. Spacer sequences must be located upstream of PAM sequences.
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