Package ‘epimutacions’

April 3, 2024

Title Robust outlier identification for DNA methylation data

Version 1.6.1

Description The package includes some statistical outlier detection methods for epimutations detection in DNA methylation data.

The methods included in the package are MANOVA, Multivariate linear models, isolation forest, robust mahalanobis distance, quantile and beta.

The methods compare a case sample with a suspected disease against a reference panel (composed of healthy individuals) to identify epimutations in the given case sample.

It also contains functions to annotate and visualize the identified epimutations.

biocViews DNA Methylation, Biological Question, Preprocessing, Statistical Method, Normalization

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Depends R (>= 4.3.0), epimutacionsData


Suggests testthat, knitr, markdown, BiocStyle, a4Base, kableExtra, methods, grDevices

VignetteBuilder knitr

Encoding UTF-8

Roxygen list(markdown = TRUE)

URL https://github.com/isglobal-brge/epimutacions

BugReports https://github.com/isglobal-brge/epimutacions/issues
RoxygenNote 7.2.3

git_url https://git.bioconductor.org/packages/epimutations

git_branch RELEASE_3_18

git_last_commit ba2e314

git_last_commit_date 2023-11-03

Repository Bioconductor 3.18

Date/Publication 2024-04-03

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add_ensemble_regulatory

Add ENSEMBL regulatory regions to epimutations

Description
Add ENSEMBL regulatory regions to epimutations

Usage
add_ensemble_regulatory(epimutations, build = "37")

Arguments
epimutations a data frame object containing the result from epimutations or epimutations_one_leave_out functions.
build the build used to define epimutations coordinates. By default, it is '37', corresponding to Illumina annotation.

Value
The function returns a data frame object containing the results of epimutations or epimutations_one_leave_out with some additional variables describing regulatory elements from ENSEMBL.

Note that a single epimutation might overlap with more than one regulatory region. In that case, the different regulatory regions are separated by ///.

• ensembl_reg_id Region identifier from ENSEMBL
• ensembl_reg_coordinates Coordinates for the ENSEMBL regulatory regions
• ensembl_reg_type Type of regulatory region
• ensembl_reg_tissues Activity of the regulatory region per tissue. The different activation states are separated by /
annotate_cpg

Annotate the DMR resulting from epimutacions package

Description

This function annotates a differentially methylated region

Usage

annotate_cpg(
  data,
  db,
  split = ",",
  epi_col = "cpg_ids",
  gene_col = "GencodeBasicV12_NAME",
  feat_col = "Regulatory_Feature_Group",
  relat_col = "Relation_to_Island",
  build = "37",
  omim = TRUE
)

Arguments

data: DataFrame-like object.
db: a character string specifying the Database to use for annotation. E.g: 'IlluminaHumanMethylationEPICanno.ilm10b2.hg19'.
split: a character string containing the separator for CpG ids. Default ",".
epi_col: CpG ids, should be row names in the database.
gene_col: column name from where to extract gene names. Default: 'GencodeBasicV12_NAME'.
feat_col: column name from where to extract CpG feature groups. Default: 'Regulatory_Feature_Group'.
relat_col: column name from where to extract relation to island info. Default: 'Relation_to_Island'.
build: The build for bioMart. Default '37'.
omim: a boolean, if TRUE will annotate OMIMs as well. Takes a bit longer. Default TRUE.

Value

The function returns a DataFrame-like object annotated.
annotate_epimutations

Annotate the results of epimutations or epimutations_one_leave_out functions

Description

Information about close genes and regulatory elements for epimutations.

Usage

annotate_epimutations(
  epi_results,
  db = "IlluminaHumanMethylationEPICanno.ilm10b2.hg19",
  build = "37",
  ...
)

Arguments

epi_results  a data frame object containing the output from epimutations or epimutations_one_leave_out functions.
db  a character string containing the Illumina annotation package used to annotate the CpGs.
build  a character string containing the genomic build where the epimutations are mapped. The default is GRCh37 (build = "37"). To use GRCh38 set build to NULL.
...  Further arguments passed to annotate_cpg.

Value

The function returns the input object epi_results with additional columns containing the information about the genes or overlapping regulatory features.

See annotate_cpg and add_ensemble_regulatory for an in-depth description of these variables.

Examples

data(res.epi.manova)
#Annotate the epimutations

#anno_results <- annotate_epimutations(res.epi.manova)
Description

Distribution of a positive linear combination of $\chi^2$ random variables.

Usage

\[
\text{AS204}(c, \lambda, \text{mult} = \text{rep}(1, \text{length}(\lambda)), \delta = \text{rep}(0, \text{length}(\lambda)), \text{maxit} = 1e+05, \text{eps} = 1e-14, \text{mode} = 1)
\]

Arguments

- **c**: value point at which distribution is to be evaluated.
- **lambda**: the weights $\lambda_j$.
- **mult**: the multiplicities $m_j$.
- **delta**: the non-centrality parameters $\delta_j^2$.
- **maxit**: the maximum number of terms $K$ (see Details).
- **eps**: the desired level of accuracy.
- **mode**: if "mode" > 0 then $\beta = \text{mode} \lambda_{\text{min}}$, otherwise $\beta = 2/(1/\lambda_{\text{min}} + 1/\lambda_{\text{max}})$.

Details

Algorithm AS 204 evaluates the expression

\[
P[X < c] = P[\sum_{j=1}^{n} \lambda_j \chi^2(m_j, \delta_j^2) < c]
\]

where $\lambda_j$ and $c$ are positive constants and $\chi^2(m_j, \delta_j^2)$ represents an independent $\chi^2$ random variable with $m_j$ degrees of freedom and non-centrality parameter $\delta_j^2$. This can be approximated by the truncated series

\[
\sum_{k=0}^{K-1} a_k P[\chi^2(m + 2k) < c/\beta]
\]

where $m = \sum_{j=1}^{n} m_j$ and $\beta$ is an arbitrary constant (as given by argument "mode").

The C++ implementation of algorithm AS 204 used here is identical to the one employed by the farebrother method in the CompQuadForm package, with minor modifications.
**Value**

The function returns the probability \( P[X > c] = 1 - P[X < c] \) if the AS 204 fault indicator is 0 (see Note below), and NULL if the fault indicator is 4, 5 or 9, as the corresponding faults can be corrected by increasing "eps". Other faults raise an error.

**Note**

The algorithm AS 204 defines the following fault indicators: 7) one or more of the constraints \( \lambda_j > 0, m_j > 0 \) and \( \delta_j^2 \geq 0 \) is not satisfied. 1) non-fatal underflow of \( a_0 \). 2) one or more of the constraints \( n > 0, c > 0, \text{maxit} > 0 \) and \( \text{eps} > 0 \) is not satisfied. 3) the current estimate of the probability is < -1. 4) the required accuracy could not be obtained in \( \text{maxit} \) iterations. 5) the value returned by the procedure does not satisfy \( 0 \leq P[X < c] \leq 1 \). 6) the density of the linear form is negative. 9) faults 4 and 5. 10) faults 4 and 6. 0) otherwise.

**Author(s)**

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**References**


**See Also**

farebrother

---

**betas_from_bump**

Obtains bumps beta values

**Description**

The function obtains beta values corresponding to the CpGs into DMRs.

**Usage**

```
betas_from_bump(bump, fd, betas)
```

**Arguments**

- **bump**: the result from `bumphunter`
- **fd**: a data frame containing the genomic ranges for each CpGs.
- **betas**: a matrix containing the beta values for all CpGs in each sample.
The function returns a list containing the melted beta values, the population mean and 1, 1.5, and 2 standard deviations from the mean of the distribution.

**cols_names**

Sets common column names in a data frame

Description

Sets common column names in a given data frame containing the CpGs genomic ranges or a DMR (result of epimutations or epimutations_one_leave_out function).

Usage

cols_names(x, cpg_ids_col = FALSE)

Arguments

- **x** a data frame containing the genomic ranges or a DMR (a row of the results of epimutations or epimutations_one_leave_out function).
- **cpg_ids_col** a boolean, if TRUE the input data frame contains the CpGs names column.

Value

The function returns a data frame containing the column names to carry out the analysis without any error.
create_GRanges_class

Generates a GRanges object

Description
This function makes a GRanges object from a GenomicRatioSet.

Usage
create_GRanges_class(methy, cpg_ids)

Arguments
methy a GenomicRatioSet object containing the control and case samples used in epimutations or epimutations_one_leave_out function.
cpg_ids a character string specifying the name of the CpGs in the DMR of interest.

Value
The function returns a GRanges object containing the beta values and the genomic ranges of the CpGs of interest.

epimutations Epimutations analysis based on outlier detection methods

Description
The function identifies differentially methylated regions in a case sample by comparing it against a control panel.

Usage
epimutations(
case_samples,
control_panel,
method = "manova",
chr = NULL,
start = NULL,
end = NULL,
epi_params = epi_parameters(),
maxGap = 1000,
bump_cutoff = 0.1,
min_cpg = 3,
verbose = TRUE
)


**Arguments**

- **case_samples** a GenomicRatioSet object containing the case samples. See the constructor function `GenomicRatioSet, makeGenomicRatioSetFromMatrix`.
- **control_panel** a GenomicRatioSet object containing the control panel (control panel).
- **method** a character string naming the outlier detection method to be used. This can be set as: "manova", "mlm", "iForest", "mahdist", "quantile" and "beta". The default is "manova". For more information see **Details**.
- **chr** a character string containing the sequence names to be analysed. The default value is `NULL`.
- **start** an integer specifying the start position. The default value is `NULL`.
- **end** an integer specifying the end position. The default value is `NULL`.
- **epi_params** the parameters for each method. See the function `epi_parameters`.
- **maxGap** the maximum location gap used in `bumphunter` method.
- **bump_cutoff** a numeric value of the estimate of the genomic profile above the cutoff or below the negative of the cutoff will be used as candidate regions.
- **min_cpg** an integer specifying the minimum CpGs number in a DMR.
- **verbose** logical. If TRUE additional details about the procedure will provide to the user. The default is TRUE.

**Details**

The function compares a case sample against a control panel to identify epimutations in the given sample. First, the DMRs are identified using the `bumphunter` approach. After that, CpGs in those DMRs are tested in order to detect regions with CpGs being outliers. For that, different outlier detection methods can be selected:

- Multivariate Analysis of Variance ("manova"). manova
- Multivariate Linear Model ("mlm")
- Isolation Forest ("iForest") isolation.forest
- Robust Mahalanobis Distance ("mahdist") covMcd
- Quantile distribution ("quantile")
- Beta ("beta")

We defined candidate epimutation regions (found in candRegsGR) based on the 450K array design. As CpGs are not equally distributed along the genome, only CpGs closer to other CpGs can form an epimutation. More information can be found in candRegsGR documentation.

**Value**

The function returns an object of class tibble containing the outliers regions. The results are composed by the following columns:

- **epi_id**: systematic name for each epimutation identified. It provides the name of the used anomaly detection method.
epimutations

- **sample**: the name of the sample containing the epimutation.
- **chromosome, start and end**: indicate the location of the epimutation.
- **sz**: the window’s size of the event.
- **cpg_n**: the number of CpGs in the epimutation.
- **cpg_ids**: the names of CpGs in the epimutation.
- **outlier_score**: 
  - For method *manova* it provides the approximation to F-test and the Pillai score, separated by `/`.
  - For method *mlm* it provides the approximation to F-test and the R2 of the model, separated by `/`.
  - For method *iforest* it provides the magnitude of the outlier score.
  - For method *beta* it provides the mean outlier p-value.
  - For methods *quantile* and *mahdist* it is filled with NA.
- **outlier_direction**: indicates the direction of the outlier with "hypomethylation" and "hypermethylation"
  - For *manova*, *mlm*, *iforest*, and *mahdist* it is computed from the values obtained from bumhunter.
  - For *quantile* it is computed from the location of the sample in the reference distribution (left vs. right outlier).
  - For method *beta* it return a NA.
- **pvalue**: 
  - For methods *manova*, *mlm*, and *iforest* it provides the p-value obtained from the model.
  - For method *quantile*, *mahdist* and *beta* is filled with NA.
- **adj_pvalue**: for methods with p-value (manova and mlm adjusted p-value with Benjamini-Hochberg based on the total number of regions detected by Bumphunter.
- **epi_region_id**: Name of the epimutation region as defined in candRegsGR.
- **CRE**: cREs (cis-Regulatory Elements) as defined by ENCODE overlapping the epimutation region. Different cREs are separated by `;`.
- **CRE_type**: Type of cREs (cis-Regulatory Elements) as defined by ENCODE. Different type are separated by `;`.

**Examples**

```r
data(GRset)

#Find epimutations in GSM2562701 sample of GRset dataset

case_samples <- GRset[,11]
control_panel <- GRset[,1:10]
epimutations(case_samples, control_panel, method = "manova")```
epimutations_one_leave_out

Epimutations analysis based on outlier detection methods

Description

This function is similar to epimutations with the particularity that when is more than one case sample, the remaining case samples are included as controls.

Usage

epimutations_one_leave_out(
  methy,
  method = "manova",
  epi_params = epi_parameters(),
  BPPARAM = BiocParallel::SerialParam(),
  verbose = TRUE,
  ...
)

Arguments

methy a GenomicRatioSet object containing the samples for the analysis. See the constructor function GenomicRatioSet, makeGenomicRatioSetFromMatrix.
method a character string naming the outlier detection method to be used. This can be set as: "manova", "mlm", "iForest", "mahdist", "barbosa" and beta. The default is "manova". For more information see Details.
epi_params the parameters for each method. See the function epi_parameters.
BPPARAM ("BiocParallelParam") BiocParallelParam object to configure parallelization execution. By default, execution is non-parallel.
verbose logical. If TRUE additional details about the procedure will provide to the user. The default is TRUE.
... Further parameters passed to epimutations

Details

The function compares a case sample against a control panel to identify epimutations in the given sample. First, the DMRs are identified using the bumphunter approach. After that, CpGs in those DMRs are tested in order to detect regions with CpGs being outliers. For that, different anomaly detection methods can be selected:

- Multivariate Analysis of Variance ("manova"). manova
- Multivariate Linear Model ("mlm")
- Isolation Forest ("iForest") isolation.forest
- Robust Mahalanobis Distance ("mahdist") covMcd
- Barbosa ("barbosa")
The function returns an object of class tibble containing the outliers regions. The results are composed by the following columns:

- **epi_id**: the name of the anomaly detection method that has been used to detect the epimutation.
- **sample**: the name of the sample where the epimutation was found.
- **chromosome, start and end**: indicate the location of the epimutation.
- **sz**: the number of base pairs in the region.
- **cpg_n**: number of CpGs in the region.
- **cpg_ids**: differentially methylated CpGs names.
- **outlier_score**:
  - For method manova it provides the approximation to F-test and the Pillai score, separated by `/`.
  - For method mlm it provides the approximation to F-test and the R2 of the model, separated by `/`.
  - For method iforest it provides the magnitude of the outlier score.
  - For methods barbosa and mahdist is filled with NA.
- **outlier_significance**:
  - For methods manova, mlm, and iforest it provides the p-value obtained from the model.
  - For method barbosa and mahdist is filled with NA.
- **outlier_direction**: indicates the direction of the outlier with "hypomethylation" and "hypermethylation"
  - For manova, mlm, iforest, and mahdist it is computed from the values obtained from bumphunter.
  - For barbosa it is computed from the location of the sample in the reference distribution (left vs. right outlier).

Examples

```r
data(GRset)
manova_result <- epimutations_one_leave_out(GRset,
                                          method = "manova")
```

---

**Description**

*epi_beta* method models the DNA methylation data using a beta distribution. First, the beta distribution parameters of the reference population are precomputed and passed to the method. Then, we compute the probability of observing the methylation values of the case from the reference beta distribution. CpGs with p-values smaller than a threshold `pvalue_threshold` and with a methylation difference with the mean reference methylation higher than `diff_threshold` are defined as outlier CpGs. Finally, epimutations are defined as a group of contiguous outlier CpGs.
**Usage**

```r
epi_beta(
  beta_params,
  beta_mean,
  betas_case,
  case,
  controls,
  betas,
  annot,
  pvalue_threshold,
  diff_threshold,
  min_cpgs = 3,
  maxGap
)
```

**Arguments**

- `beta_params`: matrix with the parameters of the reference beta distributions for each CpG in the dataset.
- `beta_mean`: beta values mean.
- `betas_case`: matrix with the methylation values for a case.
- `case`: case sample name.
- `controls`: control samples names.
- `betas`: a matrix containing the beta values for all samples.
- `annot`: annotation of the CpGs.
- `pvalue_threshold`: minimum p-value to consider a CpG an outlier.
- `diff_threshold`: minimum methylation difference between the CpG and the mean methylation to consider a position an outlier.
- `min_cpgs`: minimum number of CpGs to consider an epimutation.
- `maxGap`: maximum distance between two contiguous CpGs to combine them into an epimutation.

**Value**

The function returns a data frame with the candidate regions to be epimutations.

---

**epi_iForest**

Identifies epimutations using Isolation Forest

**Description**

This function identifies regions with CpGs being outliers using `isolation_forest` approach.
epi_mahdist

Usage

epi_iForest(mixture, case_id, ntrees)

Arguments

mixture  beta values matrix. Samples in columns and CpGs in rows.
case_id  a character string specifying the name of the case sample.
ntrees  number of binary trees to build for the model. Default is 100.

Value

The function returns the outlier score for the given case sample.

epi_mahdist  Identifies epimutations using Robust Mahalanobis distance

Description

This function identifies regions with CpGs being outliers using the Minimum Covariance Determinant (MCD) estimator (covMcd) to compute the Mahalanobis distance.

Usage

epi_mahdist(mixture, nsamp = c("best", "exact", "deterministic"))

Arguments

mixture  beta values matrix. Samples in columns and CpGs in rows.
nsamp  the number of subsets used for initial estimates in the MCD. It can be set as: "best", "exact", or "deterministic".

Details

The implementation of the method here is based on the discussion in this thread of Cross Validated

Value

The function returns the computed Robust Mahalanobis distance.
epi_manova

Identifies epimutations using MANOVA

Description
This function identifies regions with CpGs being outliers using manova approach.

Usage
epi_manova(mixture, model, case_id)

Arguments
- mixture: beta values matrix. Samples in columns and CpGs in rows.
- model: design (or model) matrix.
- case_id: a character string specifying the name of the case sample.

Value
The function returns the F statistic, Pillai and P value.

epi_mlm

Detects epimutations using Multivariate Linear Model (MLM)

Description
Identifies CpGs with outlier methylation values using methylated Multivariate Linear Model

Usage
epi_mlm(mixture, model)

Arguments
- mixture: beta values matrix. Samples in columns and CpGs in rows.
- model: design (or model) matrix.

Value
The function returns the F statistic, R2 test statistic and Pillai.
Description

Allow the user to set the values of the parameters to compute the functions epimutations and epimutations_one_leave_out.

Usage

```r
epi_parameters(
  manova = list(pvalue_cutoff = 0.05),
  mlm = list(pvalue_cutoff = 0.05),
  iForest = list(outlier_score_cutoff = 0.7, ntrees = 100),
  mahdist = list(nsamp = "deterministic"),
  quantile = list(window_sz = 1000, offset_abs = 0.15, qsup = 0.995, qinf = 0.005),
  beta = list(pvalue_cutoff = 1e-06, diff_threshold = 0.1)
)
```

Arguments

- `manova`, `mlm`, `iForest`, `mahdist`, `quantile`, `beta`
  - Method selected in the function epimutations.
- `pvalue_cutoff` - the threshold p value to select which CpG regions are outliers in manova, mlm and beta methods.
- `outlier_score_cutoff` - The outlier score threshold to identify outliers CpGs in isolation forest (iForest) method. Default is 0.5.
- `ntrees` - number of binary trees to build for the model build by isolation forest (iForest) method. Default is 100.
- `nsamp` - the number of subsets used for initial estimates in the Minimum Covariance Determinant which is used to compute the Robust Mahalanobis distance (mahdist). It can be set as: "best", "exact", or "deterministic". For nsamp = "best" exhaustive enumeration is done, as long as the number of trials does not exceed 100,000. For nsamp = "exact" exhaustive enumeration will be attempted however many samples are needed. In this case, a warning message may be displayed saying that the computation can take a very long time. For nsamp = "deterministic". For more information see covMcd. Default is "deterministic".
- `window_sz` - the maximum distance between CpGs to be considered in the same DMR. This parameter is used in quantile (default: 1000).
- `qsup`, `qinf`, `offset_abs`
  - The upper and lower quantiles (threshold) to consider a CpG an outlier when using quantile method, as well as the offset to consider (defaults: 0.005, 0.995, 0.15).
- `diff_threshold` - Minimum methylation difference between the CpG and the mean methylation to consider a position an outlier.
### Details

Invoking `epi_parameters()` with no arguments returns a list with the default values.

### Value

The function returns a list of all set parameters for each method used in `epimutations` and `epimutations_one_leave_out` functions.

### Examples

```r
# Default set of parameters
epi_parameters()

# Change p-value for manova method
epi_parameters(manova = list("pvalue_cutoff" = 0.01))
```

---

### epi_preprocess

**Preprocess methylation array**

### Description

The `epi_preprocess` function reads Illumina methylation sample sheet for case samples and it merges them with `RGChannelSet` reference panel. The final dataset is normalized using minfi package preprocess methods.

### Usage

```r
epi_preprocess(
  cases_dir,  
  reference_panel,  
  pattern = "csv$",  
  normalize = "raw",  
  norm_param = norm_parameters(),  
  verbose = FALSE
)
```

### Arguments

- `cases_dir`: the base directory from which the search is started.
- `reference_panel`: an `RGChannelSet` object containing the reference panel (controls) samples.
- `pattern`: What pattern is used to identify a sample sheet file.
- `normalize`: a character string specifying the selected preprocess method. For more information see Details or minfi package user's Guide. It can be set as: "raw", "illumina", "swan", "quantile", "noob" or "funnorm".
- `norm_param`: the parameters for each preprocessing method. See the function `norm_parameters`.
- `verbose`: logical. If TRUE additional details about the procedure will provide to the user. The default is FALSE.
Details

The epi_preprocess function reads Illumina methylation sample sheet for case samples and it merges them with RGChannelSet reference panel. The final dataset is normalized using different minfi package preprocess methods:

- "raw": preprocessRaw
- "illumina": preprocessIllumina
- "swan": preprocessSWAN
- "quantile": preprocessQuantile
- "noob": preprocessNoob
- "funnorm": preprocessFunnorm

Value

epi_preprocess function returns a GenomicRatioSet object containing case and control (reference panel) samples.

Examples

```r
# The reference panel for this example is available in
# epimutacionsData (ExperimentHub) package

library(ExperimentHub)
eh <- ExperimentHub()
query(eh, c("epimutacionsData"))
reference_panel <- eh[["EH6691"]]
cases_dir <- system.file("extdata", package = "epimutacionsData")
# Preprocessing

epi_preprocess( cases_dir,
               reference_panel,
               pattern = "SampleSheet.csv")
```

---

**epi_quantile**

*Identifies epimutations using quantile distribution*

Description

Identifies CpGs with outlier methylation values using a sliding window approach to compare individual methylation profiles of a single case sample against all other samples from reference panel (controls)
Usage

epi_quantile(
    case,
    fd,
    bctr_pmin,
    bctr_pmax,
    controls,
    betas,
    window_sz = 1000,
    N = 3,
    offset_abs = 0.15
)

Arguments

case  beta values for a single case (data.frame). The samples as single column and
       CpGs in rows (named).
fd    feature description as data.frame having at least chromosome and position as
       columns and and CpGs in rows (named).
bctr_pmin Beta value observed at 0.01 quantile in controls. A beta values has to be lower
           or equal to this value to be considered an epimutation.
bctr_pmax Beta value observed at 0.99 quantile in controls. A beta values has to be higher
           or equal to this value to be considered an epimutation.
controls control samples names.
betas  a matrix containing the beta values for all samples.
window_sz Maximum distance between a pair of CpGs to defined an region of CpGs as
           epimutation (default: 1000).
N     Minimum number of CpGs, separated in a maximum of window_sz bass, to
     defined an epimutation (default: 3).
offset_abs Extra enforcement defining an epimutation based on beta values at 0.005 and
          0.995 quantiles (default: 0.15).

Value

The function returns a data frame with the regions candidates to be epimutations.

getBetaParams  Model methylation as a beta distribution

Description

Model methylation as a beta distribution
**Usage**
```
getBetaParams(x)
```

**Arguments**
- **x**: Matrix of methylation expressed as a beta. CpGs are in columns and samples in rows.

**Value**
Beta distribution.

---

**get_candRegsGR**

*Candidate regions to be epimutations*

**Description**
Load candidate regions to be epimutations from epimutacionsData package in ExperimentHub.

**Usage**
```
get_candRegsGR()
```

**Value**
The function returns a GRanges object containing the candidate regions.

---

**get_ENSEMBL_data**

*Get ENSEMBL regulatory features overlapping a genomic region*

**Description**
This function queries for ENSEMBL regulatory features and collapse them to return a single record.

**Usage**
```
get_ENSEMBL_data(chromosome, start, end, mart)
```

**Arguments**
- **chromosome**: Chromosome of the region
- **start**: Start of the region
- **end**: End of the region
- **mart**: Mart object to perform the ENSEMBL query
Description

A small GenomicRatioSet object to use in the functions examples containing 10 control samples and a case sample.

Usage

data(GRset)

Format

A GenomicRatioSet object with 4243 CpGs and 11 variables

Value

A GenomicRatioSet object with 4243 CpGs and 11 variables

Examples

data(GRset)

merge_records

Merge records for the same ENSEMBL regulatory element

Description

This function collapses the activity status of a given an ENSEMBL regulatory element in different tissues. Notice that tissues identified as inactive will not be reported.

Usage

merge_records(tab)

Arguments

<table>
<thead>
<tr>
<th>arg</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>tab</td>
<td>Results from biomaRt::getBM for the same regulatory element</td>
</tr>
</tbody>
</table>

Value

data.frame of one row after collapsing the
**mlm**

*Non-parametric, Asymptotic P-values for Multivariate Linear Models*

**Description**

Fits a multivariate linear model and computes test statistics and asymptotic P-values for predictors in a non-parametric manner.

**Usage**

```r
mlm(
  formula,
  data,
  transform = "none",
  contrasts = NULL,
  subset = NULL,
  fit = FALSE
)
```

**Arguments**

- `formula`: object of class "formula" (or one that can be coerced to that class): a symbolic description of the model to be fitted.
- `data`: an optional data frame, list or environment (or object coercible by `as.data.frame`) containing the variables in the model. If not found in `data`, the variables are taken from `environment(formula)`, typically the environment from which `mlm` is called.
- `transform`: transformation of the response variables: "none", "sqrt" or "log". Default is "none".
- `contrasts`: an optional list. See `contrasts.arg` in `model.matrix.default`. Default is "contr.sum" for ordered factors and "contr.poly" for unordered factors. Note that this is different from the default setting in `options("contrasts")`.
- `subset`: subset of predictors for which summary statistics will be reported. Note that this is different from the "subset" argument in `lm`.
- `fit`: logical. If TRUE the multivariate fit on transformed and centered responses is returned.

**Details**

A Y matrix is obtained after transforming (optionally) and centering the original response variables. Then, the multivariate fit obtained by `lm` can be used to compute sums of squares, pseudo-F statistics and asymptotic P-values for the terms specified by the `formula` in a non-parametric manner.
mlmtst

Sums of Squares and Pseudo-F Statistics from a Multivariate Fit

Description
Computes the sum of squares, degrees of freedom, pseudo-F statistics and partial R-squared for each predictor from a multivariate fit. It also returns the eigenvalues of the residual covariance matrix.

Usage
mlmtst(fit, X, subset = NULL, tol = 0.001)

Arguments
- **fit**: multivariate fit obtained by `lm`.
- **X**: design matrix obtained by `model.matrix`.
- **subset**: subset of predictors for which summary statistics will be reported. Note that this is different from the "subset" argument in `lm`.
- **tol**: `e[e/sum(e) > tol]`, where `e` is the vector of eigenvalues of the residual covariance matrix. Required to prevent long running times of algorithm AS 204. Default is 0.001 to ensure minimal loss of accuracy.

Details
Different types of sums of squares (i.e. "I", "II" and "III") are available.
**norm_parameters**

**Value**

A list containing:

- **SS**: sums of squares for all predictors (and residuals).
- **df**: degrees of freedom for all predictors (and residuals).
- **f.tilde**: pseudo-F statistics for all predictors.
- **r2**: partial R-squared for all predictors.
- **e**: eigenvalues of the residual covariance matrix.

**Author(s)**

Diego Garrido-Martín

**See Also**

AS204

---

**Description**

`norm_parameters` function allows the user to set the values of the parameters to compute the functions `epi_preprocess`.

**Usage**

```r
norm_parameters(
  illumina = list(bg.correct = TRUE, normalize = c("controls", "no"), reference = 1),
  quantile = list(fixOutliers = TRUE, removeBadSamples = FALSE, badSampleCutoff = 10.5,
                  quantileNormalize = TRUE, stratified = TRUE, mergeManifest = FALSE, sex = NULL),
  noob = list(offset = 15, dyeCorr = TRUE, dyeMethod = c("single", "reference")),
  funnorm = list(nPCs = 2, sex = NULL, bgCorr = TRUE, dyeCorr = TRUE, keepCN = FALSE)
)
```

**Arguments**

- **illumina**: preprocess method selected in the function `epi_preprocess`.
- **bg.correct**: logical. If TRUE background correction will be performed in "illumina" method. Default is TRUE.
- **normalize**: logical. If TRUE control normalization will be performed in "illumina" method.
- **reference**: numeric. The reference array for control normalization in "illumina" method.
- **fixOutliers**: logical. If TRUE low outlier Meth and Unmeth signals will be fixed in "quantile" method. Default is TRUE.
removeBadSamples
logical. If TRUE bad samples will be removed.

badSampleCutoff
a numeric specifying the cutoff to label samples as 'bad' in "quantile" method. Default is 10.5.

quantileNormalize
logical. If TRUE quantile normalization will be performed in "quantile" method. Default is TRUE.

stratified
logical. If TRUE quantile normalization will be performed within region strata in "quantile" method. Default is TRUE.

mergeManifest
logical. If TRUE the information in the associated manifest package will be merged into the output object in "quantile" method. Default is FALSE.

offset
a numeric specifying an offset for the normexp background correction in "noob" method. Default is 15.

dyeCorr
logical. Dye correction will be done in "noob" and "funnorm" methods. Default is TRUE.

dyeMethod
specify the dye bias correction to be done, single sample approach or a reference array in "noob" method.

nPCs
numeric specifying the number of principal components from the control probes PCA in "funnorm" method. Default is 2.

sex
an optional numeric vector containing the sex of the samples in "quantile" and "funnorm" methods.

bgCorr
logical. If TRUE NOOB background correction will be done prior to functional normalization in "funnorm" method. Default is TRUE.

keepCN
logical. If TRUE copy number estimates will be kept in "funnorm" method. Default is FALSE.

Details
Invoking epi_parameters() with no arguments returns a list with the default values for each normalization parameter.

Value
the function returns a list of all set parameters for each normalization method used in epi_peprocess.

Examples
#Default set of parameters
norm_parameters()
#change p value for manova method
norm_parameters(illumina = list("bg.correct" = FALSE))
**p.asympt**

Asymptotic P-values

Description

Computes asymptotic P-values given the numerator of the pseudo-F statistic, its degrees of freedom and the eigenvalues of the residual covariance matrix.

Usage

```r
p.asympt(ss, df, lambda, eps = 1e-14, eps.updt = 2, eps.stop = 1e-10)
```

Arguments

- **ss**: numerator of the pseudo-F statistic.
- **df**: degrees of freedom of the numerator of the pseudo-F statistic.
- **lambda**: eigenvalues of the residual covariance matrix.
- **eps**: the desired level of accuracy.
- **eps.updt**: factor by which eps is updated to retry execution of algorithm AS 204 when it fails with fault indicator 4, 5 or 9.
- **eps.stop**: if eps > eps.stop, execution of algorithm AS 204 is not retried and the function raises an error. Default is 1e-10.

Value

A vector containing the P-value and the level of accuracy.

Author(s)

Diego Garrido-Martín

See Also

AS204
plot_epimutations  

*Plot a given epimutation and locate it along the genome*

**Description**

This function plots a given epimutation and UCSC annotations for the specified genomic region.

**Usage**

```r
plot_epimutations(
  dmr,
  methy,
  genome = "hg19",
  genes_annot = FALSE,
  regulation = FALSE,
  from = NULL,
  to = NULL
)
```

**Arguments**

- `dmr`: epimutation obtained as a result of `epimutations` function.
- `methy`: a GenomicRatioSet object containing the information of control and case samples used for the analysis in the `epimutations` function. See the constructor function `GenomicRatioSet, makeGenomicRatioSetFromMatrix`.
- `genome`: a character string specifying the genome of reference. It can be set as "hg38", "hg19" and "hg18". The default is "hg19".
- `genes_annot`: a boolean. If TRUE gene annotations are plotted. Default is FALSE.
- `regulation`: a boolean. If TRUE UCSC annotations for CpG Islands, H3K27Ac, H3K4Me3 and H3K27Me3 are plotted. The default is FALSE. The running process when regulation is TRUE can take several minutes.
- `from`, `to`: scalar, specifying the range of genomic coordinates for the plot of gene annotation region. If NULL the plotting ranges are derived from the individual track. Note that from cannot be larger than to.

**Details**

The tracks are plotted vertically. Each track is separated by different background colour and a section title. The colours and titles are preset and cannot be set by the user.

Note that if you want to see the UCSC annotations maybe you need to take a bigger genomic region.
The function returns a plot divided in two parts:

- ggplot graph including the individual with the epimutation in red, the control samples in dashed black lines and population mean in blue. Grey shaded regions indicate 1, 1.5 and 2 standard deviations from the mean of the distribution.
- UCSC gene annotations for the specified genomic region (if genes == TRUE)
- UCSC annotations for CpG Islands, H3K27Ac, H3K4Me3 and H3K27Me3 (if regulation == TRUE)

Examples

```r
data(GRset)
data(res.epi.manova)
plot_epimutations(res.epi.manova[1,], GRset)
```

**Description**

Process data from ENSEMBL to combine results from the same regulatory elements in a unique record.

**Usage**

```r
process_ENSEMBL_results(ensembl_res)
```

**Arguments**

- `ensembl_res` Results from `biomaRt::getBM`

**Value**

data.frame of one row after collapsing the input ENSEMBL regulatory regions
Description

A data frame containing the results of epimutations function using "manova" methods for GRset dataset. For more information see the example of epimutations function.

Usage

data(res.epi.manova)

Format

A data frame with 16 variables and 6 epimutations.

Value

A data frame with 16 variables and 6 epimutations.

Examples

data(res.epi.manova)

Description

Creates a data frame containing the genomic regions, statistics and direction for the DMRs.

Usage

res_iForest(bump, sts, outlier_score_cutoff)

Arguments

bump a DMR obtained from bumphunter (i.e. a row from bumphunter method result).
sts the outlier score from epi_iForest function results.
outlier_score_cutoff numeric specifying the outlier score cut off
The function returns a data frame containing the following information for each DMR:

- genomic ranges
- DMR base pairs
- number and name of CpGs in DMR
- statistics:
  - Outlier score
  - Outlier significance
  - Outlier direction
- Sample name

For more information about the output see epimutations.

---

**Description**

Creates a data frame containing the genomic regions, statistics and direction for the DMRs.

**Usage**

`res_mahdist(case, bump, outliers)`

**Arguments**

- `case` a character string specifying the case sample name.
- `bump` a DMR obtained from `bumphunter` (i.e. a row from `bumphunter` method result).
- `outliers` the robust distance computed by `epi_mahdist` function results.

**Value**

The function returns a data frame containing the following information for each DMR:

- genomic ranges
- DMR base pairs
- number and name of CpGs in DMR
- statistics:
  - Outlier score
  - Outlier significance
  - Outlier direction
- Sample name

For more information about the output see epimutations.
res_manova

Description

Creates a data frame containing the results obtained from MANOVA.

Usage

res_manova(bump, sts)

Arguments

bump a DMR obtained from bumphunter (i.e. a row from bumphunter method result).
sts F statistic, Pillai and P value from epi_manova function results.

Value

The function returns a data frame containing the following information for each DMR:

- genomic ranges
- DMR base pairs
- number and name of CpGs in DMR
- statistics:
  - Outlier score
  - Outlier significance
  - Outlier direction
- Sample name

For more information about the output see epimutations.

res_mlm

Description

Creates a data frame containing the results obtained from MLM.

Usage

res_mlm(bump, sts)
 Arguments

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>bump</td>
<td>a DMR obtained from bumphunter (i.e. a row from bumphunter method result).</td>
</tr>
<tr>
<td>sts</td>
<td>the F statistic, R2 test statistic and Pillai obtained as a result of epi_mlm function.</td>
</tr>
</tbody>
</table>

 Value

The function returns a data frame containing the following information for each DMR:

- genomic ranges
- DMR base pairs
- number and name of CpGs in DMR
- statistics:
  - Outlier score
  - Outlier significance
  - Outlier direction
- Sample name

For more information about the output see epimutations.

---

### UCSC_annotation

UCSC gene annotations for a given genome assembly.

#### Usage

```
UCSC_annotation(genome = "hg19")
```

#### Arguments

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>genome</td>
<td>genome assembly. Can be set as: 'hg38', 'hg19' and 'hg18'.</td>
</tr>
</tbody>
</table>

#### Value

The function returns gene annotations for the specified genome assembly.
Description

UCSC annotations for CpG Islands, H3K27Ac and H3K4Me3 for a given genome assembly and genomic coordinates.

Usage

UCSC_regulation(genome, chr, from, to)

Arguments

- **genome**: genome assembly. Can be set as: 'hg38', 'hg19' and 'hg18'.
- **chr**: a character string containing the sequence names to be analysed.
- **from, to**: scalar, specifying the range of genomic coordinates. Note that from cannot be larger than to.

Value

UCSC_regulation returns a list containing CpG Islands, H3K27Ac and H3K4Me3 tracks.
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