Package ‘epiregulon.extra’

May 29, 2024

Title Companion package to epiregulon with additional plotting, differential and graph functions

Version 1.0.0

Description Gene regulatory networks model the underlying gene regulation hierarchies that drive gene expression and observed phenotypes. Epiregulon infers TF activity in single cells by constructing a gene regulatory network (regulons). This is achieved through integration of scATAC-seq and scRNA-seq data and incorporation of public bulk TF ChIP-seq data. Links between regulatory elements and their target genes are established by computing correlations between chromatin accessibility and gene expressions.

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Encoding UTF-8

Roxygen list(markdown = TRUE)

RoxygenNote 7.3.1

Imports scran, ComplexHeatmap, Matrix, SummarizedExperiment, checkmate, circlize, clusterProfiler, ggplot2, ggraph, igraph, lifecycle, patchwork, reshape2, scales, scater, stats

Depends R (>= 4.4), SingleCellExperiment

Suggests epiregulon, knitr, rmarkdown, parallel, BiocStyle, testthat (>= 3.0.0), EnrichmentBrowser, msigdb, dorothea, scMultiome, S4Vectors, scuttle, vdiffr, ggastrix, ggrepel

VignetteBuilder knitr

URL https://github.com/xiaosaiyao/epiregulon.extra/

biocViews GeneRegulation, Network, GeneExpression, Transcription, ChipOnChip, DifferentialExpression, GeneTarget, Normalization, GraphAndNetwork

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BugReports https://github.com/xiaosaiyao/epiregulon.extra/issues

git_url https://git.bioconductor.org/packages/epiregulon.extra

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buildGraph

Creating graphs and related operations

Description

The function enable to create graph objects using as input regulon objects returned by pruneRegulon or addWeights. Both weighted and unweighted graphs can be created that can further be visualized using dedicated functions.
Usage

```r
buildGraph(
  regulon,
  mode = c("tg", "tripartite", "re", "pairs"),
  weights = "weights",
  cluster = "all",
  aggregation_function = function(x) x[which.max(abs(x))],
  na_replace = TRUE,
  keep_original_names = TRUE,
  filter_edges = NULL
)
```

```r
buildDiffGraph(graph_obj_1, graph_obj_2, weighted = TRUE, abs_diff = TRUE)
```

```r
addCentrality(graph)
```

```r
normalizeCentrality(graph, FUN = sqrt, weighted = TRUE)
```

```r
rankTfs(graph, type_attr = "type")
```

Arguments

- **regulon**: an object returned by the getRegulon or addWeights function.
- **mode**: a character specifying which type of graph will be built. In 'tg' mode a bipartite graph is built by connecting transcription factors directly to the target genes and ignoring information about mediating regulatory elements; in 'pairs' mode transcription factors are connected to unique target gene-regulatory element pairs; in 'tripartite' mode the network is made up of three types of vertices (nodes): transcription factors, regulatory elements and target genes; here the path from target gene to regulatory element always contains a regulatory element; in 're' mode data in the target genes is dropped and only connections are between transcription factors and regulatory elements.
- **weights**: a character specifying which variable should be used to assign weights to edges.
- **cluster**: a character specifying the name of the cluster column which should be used to retrieve weight values from regulon object. Using this argument makes sense only with combination with weights parameter when it points to the regulon column that is a matrix.
- **aggregation_function**: a function used to aggregate weights of duplicated edges, which might appear due to the many transcription factor converging at the same regulatory element; starting from this point each transcription factor is supposed to have a separate connection to the target gene, perhaps the same one across several connections. In tripartite mode this might result in many edges in the same node pair, however weights might differ since they are inherited from different tf-re-tg triplets (rows) in the regulon object. Similarly, duplicated edges are generated by one transcription factor using a regulatory element multiple times to reach different target genes. In tg mode the edges became duplicated if one transcription factor reaches the same target genes through many regulatory elements.
na_replace  a logical indicating whether NA values for weights should be replaced with zeros.

keep_original_names  A logical indicating whether gene names should be used as node names in the output graph. Note that this might lead to the duplicated node names if the same gene is present in two layers (transcription factors and target genes).

filter_edges  A numeric defining the cutoff weight used for filtering out edges which have weights equal or greater than cutoff. The isolated vertices are removed then from the graph. Defaults to NULL in which case no filtering is applied.

weighted  a logical indicating whether weighted graphs are used; in tripartite mode tf-re-tg triplet is decomposed into two edges corresponding to tf-re and re-tg pairs, and both edges inherit the same weight, which was originally assigned to the parent triplet.

abs_diff  a logical indicating whether absolute difference in the number edges or their weights will be calculated.

graph, graph_obj_1, graph_obj_2  an igraph object.

FUN  a function used for normalization. The input to this function is be the number of edges connected with each node (incident edges).

type_attr  a character corresponding to the name of the vertex attribute which indicate the type of vertex.

Details

buildGraph function creates a directed graph based on the output of the getRegulon function. Four modes are available: (1) tg in which connections are made directly between transcription factor and target genes. Even if the same tf-tg pair is connected in the original regulon object through many regulatory elements then only one edge is created. In such a case, when weighted graph is created, weights are summarized by the aggregating function (by default the maximum absolute value with the sign of the original value). Similarly, aggregation is made in the re mode leaving only unique transcription factor-regulatory element pairs. In tripartite mode edges connect transcription factors with regulatory elements and regulatory elements with target genes. The same weights are used for both edges that correspond to the single row in the regulon data frame (tf-re and re-tg). Note that the original regulon structure is not fully preserved because each row is now represented by two edges which are independent from each other. Thus they can be coupled with different edges connected to the same regulatory element building the path from transcription factor to the target gene of another transcription factor through the shared regulatory element.

buildDiffGraph a graph difference by subtracting the edges of graph_obj_2 from those of the graph_obj_1. If weighted is set to TRUE then for each ordered pair of vertices (nodes) the difference in number of edges between graph_obj_1 and graph_obj_1 is calculated. The result is used to set the number of corresponding edges in output graph. Note that unless abs_diff is set to TRUE any non-positive difference will translate into lack of the edges for a corresponding ordered pair of vertices in the output graph (equivalent to 0 value in the respective position in adjacency matrix). In case of weighted graphs, the weight of the output graph is calculated as a difference of the corresponding weights between input graphs.

addCentrality calculates degree centrality for each vertex using igraph::strength.
With normalizeCentrality function the normalized values of centrality are calculated from the original ones divided by \( \text{FUN}( \text{total number of non-zero edges associated with each node}) \).

rankTfs assign ranks to transcription factors according to degree centrality of their vertices.

Value

an igraph object. rankTfs returns a data.frame with transcription factors sorted according to the value of the centrality attribute.

Examples

```r
# create an artificial getRegulon output
set.seed(1234)
tf_set <- apply(expand.grid(LETTERS[1:10], LETTERS[1:10]), 1, paste, collapse = '')
regulon <- DataFrame(tf = sample(tf_set, 5e3, replace = TRUE))
gene_set <- expand.grid(LETTERS[1:10], LETTERS[1:10], LETTERS[1:10])
gene_set <- apply(gene_set, 1, function(x) paste0(x, collapse = ''))
regulon$target <- sample(gene_set, 5e3, replace = TRUE)
regulon$idxATAC <- 1:5e3
regulon$corr <- runif(5e3)*0.5+0.5
regulon$weights <- matrix(runif(15000), nrow=5000, ncol=3)
colnames(regulon$weights) <- c('all', 'cluster1', 'cluster2')
graph_tripartite <- buildGraph(regulon, cluster = 'all', mode = 'tripartite')

# build bipartite graph using regulatory element-target gene pairs
graph_pairs_1 <- buildGraph/regulon, cluster = 'cluster1', mode = 'pairs')
graph_pairs_2 <- buildGraph(regulon, cluster = 'cluster2', mode = 'pairs')
graph_diff <- buildDiffGraph(graph_pairs_1, graph_pairs_2)
graph_diff <- addCentrality(graph_diff)
graph_diff <- normalizeCentrality(graph_diff)
tf_ranking <- rankTfs(graph_diff)
```

---

**calculateJaccardSimilarity**

*Calculate Jaccard Similarity between regulons of all transcription factors*

**Description**

Calculate Jaccard Similarity between regulons of all transcription factors

**Usage**

`calculateJaccardSimilarity(graph)`

**Arguments**

- `graph` a igraph object from buildGraph or buildDiffGraph
**Value**

A matrix with Jaccard similarity between all pairs of transcription factors.

**Examples**

```r
regulon <- data.frame(tf = sample(letters[1:4], 100, replace = TRUE), idxATAC = 1:100,
target = sample(letters[5:14], 100, replace = TRUE))
regulon$weights <- runif(100)
GRN_graph <- buildGraph(regulon)
similarity <- calculateJaccardSimilarity(GRN_graph)
```

**Description**

Plot results of regulonEnrich

**Usage**

```r
enrichPlot(results, top = 15, ncol = 3, title = NULL, combine = TRUE)
```

**Arguments**

- `results`: Output from regulonEnrich
- `top`: An integer to indicate the number of pathways to plot ranked by significance. Default is 15.
- `ncol`: An integer to indicate the number of columns in the combined plot, if `combine == TRUE`. Default is 3.
- `title`: String indicating the title of the combined plot
- `combine`: logical to indicate whether to combine and visualize the plots in one panel

**Value**

A combined ggplot object or a list of ggplots if `combine == FALSE`

**Author(s)**

Xiaosai Yao
findDifferentialActivity

**Examples**

```r
# retrieve genesets
H <- EnrichmentBrowser::getGenesets(org = 'hsa', db = 'msigdb',
cat = 'H', gene.id.type = 'SYMBOL')
C6 <- EnrichmentBrowser::getGenesets(org = 'hsa', db = 'msigdb',
cat = 'C6', gene.id.type = 'SYMBOL')

# combine genesets and convert genesets to be compatible with enricher
gs <- c(H, C6)
gs.list <- do.call(rbind, lapply(names(gs), function(x) {
data.frame(gs=x, genes=gs[[x]]))))

# get regulon
library(dorothea)
data(dorothea_hs, package = 'dorothea')
regulon <- dorothea_hs
enrichment_results <- regulonEnrich(c('ESR1', 'AR'), regulon = regulon, weight = 'mor',
genesets = gs.list)

# plot graph
enrichPlot(results = enrichment_results)
```

**Description**

Test for differential TF activity between pairs of single cell clusters/groups

**Usage**

```r
findDifferentialActivity(
  activity_matrix,
  clusters,
  test.type = "t",
pval.type = "some",
direction = c("any", "up", "down"),
groups = deprecated(),
...)
```
findPartners

Find interaction partners of a transcription factor of interest

Description
Find interaction partners of a transcription factor of interest

Usage
findPartners(graph, focal_tf)
getSigGenes

Arguments

- **graph**: a igraph object from buildGraph or buildDiffGraph
- **focal_tf**: character string indicating the name of the transcription factors to find interaction partners of

Value

A list with elements corresponding to each transcription factor apart from the focal one. Each list element is represented as a data frame with columns containing names of all target genes shared with focal transcription factor, weights of edges connecting transcription factor with target genes, equivalent weights for focal transcription factor and the element wise product of both weight columns.

Examples

```r
regulon <- data.frame(tf = sample(letters[1:4], 100, replace = TRUE), idxATAC= 1:100, target = sample(letters[5:14], 100, replace = TRUE))
regulon$weights <- runif(100)
GRN_graph <- buildGraph(regulon)
partners <- findPartners(GRN_graph, 'a')
```

getSigGenes

Compile and summarize the output from findDifferentialActivity function

Usage

```r
getSigGenes(
    da_list, 
    fdr_cutoff = 0.05, 
    logFC_cutoff = NULL, 
    topgenes = NULL, 
    direction = c("any", "up", "down")
)
```

Arguments

- **da_list**: List of dataframes from running findDifferentialActivity
- **fdr_cutoff**: A numeric scalar to specify the cutoff for FDR value. Default is 0.05
- **logFC_cutoff**: A numeric scalar to specify the cutoff for log fold change.
- **topgenes**: A integer scalar to indicate the number of top ordered genes to include in output
- **direction**: A string specifying direction for which differential TF activity was calculated, can be "any", "up" or "down"
Value

A compiled dataframe of TFs with differential activities across clusters/groups

Author(s)

Xiaosai Yao, Shang-yang Chen

Examples

```r
set.seed(1)
score.combine <- cbind(matrix(runif(2000,0,2), 20,100), matrix(runif(2000,0,10), 20,100))
rownames(score.combine) <- paste0("TF",1:20)
colnames(score.combine) <- paste0("cell",1:200)
cluster <- c(rep(1,100),rep(2,100))
markers <- findDifferentialActivity(score.combine, cluster, pval.type = "some", direction = "up", test.type = "t")
sig.genes <- getSigGenes(markers, fdr_cutoff = 1, logFC_cutoff = 0.1)
utils::head(sig.genes)
```

permuteGraph

Calculate similarity score from permuted graphs to estimate background similarity

Description

Calculate similarity score from permuted graphs to estimate background similarity

Usage

```r
permuteGraph(graph, focal_tf, n = 100, p = 1)
```

Arguments

- `graph`: an igraph object from `buildGraph` or `buildDiffGraph`
- `focal_tf`: character string indicating the name of the transcription factors to calculate similarity score
- `n`: an integer indicating the number of permutations
- `p`: a scalar indicating the probability of rewiring the graphs

Value

A matrix with Jaccard similarity between the focal transcription factor and all pairs of transcription factors for n permuted graphs
**Examples**

```r
regulon <- data.frame(tf = sample(letters[1:4], 100, replace = TRUE), idxATAC= 1:100, target = sample(letters[5:14], 100, replace = TRUE))
regulon$weights <- runif(100)
GRN_graph <- buildGraph(regulon)
permuted_graph <- permuteGraph(GRN_graph, focal_tf = "a")
```

**plotActivityDim**

Plot cell-level reduced dimension results stored in a SingleCellExperiment object, colored by activities for a list of TFs

**Description**

Plot cell-level reduced dimension results stored in a SingleCellExperiment object, colored by activities for a list of TFs

**Usage**

```r
plotActivityDim(
  sce = NULL,
  activity_matrix,
  tf,
  dimtype = "UMAP",
  label = NULL,
  ncol = NULL,
  nrow = NULL,
  title = NULL,
  combine = TRUE,
  legend.label = "activity",
  colors = c("blue", "yellow"),
  limit = NULL,
  ...
)
```

**Arguments**

- **sce**: A SingleCellExperiment object containing dimensionality reduction coordinates
- **activity_matrix**: A matrix of TF activities inferred from calculateActivity
- **tf**: A character vector indicating the names of the transcription factors to be plotted
- **dimtype**: String indicating the name of dimensionality reduction matrix to be extracted from the SingleCellExperiment
- **label**: String corresponding to the field in the colData of sce for annotation on plot
- **ncol**: A integer to specify the number of columns in the combined plot, if combine == TRUE
plotActivityViolin

*nrow* A integer to specify the number of rows in the combined plot, if combine == TRUE

title A string to specify the name of the combined plot

combine logical to indicate whether to combine and visualize the plots in one panel

legend.label String indicating the name of variable to be plotted on the legend

colors A vector of 2 colors for the intensity, with the first element referring to the lower value and the second element referring to the higher value. Default is c(’blue’,’yellow’).

limit A vector of lower and upper bounds for the color scale. The default option is NULL and will adjust to minimal and maximal values

... Additional arguments from scater::plotReducedDim

Value

A combined ggplot object or a list of ggplots if combine == FALSE

Author(s)

Xiaosai Yao, Shang-yang Chen

Examples

# create a mock singleCellExperiment object for gene expression matrix
example_sce <- scuttle::mockSCE()
example_sce <- scuttle::logNormCounts(example_sce)
example_sce <- scater::runPCA(example_sce)
example_sce <- scater::runUMAP(example_sce)
example_sce$cluster <- sample(LETTERS[1:5], ncol(example_sce), replace = TRUE)
plotActivityDim(sce = example_sce, activity = logcounts(example_sce),
   tf = c(’Gene_0001’,’Gene_0002’), label =’cluster’)

plotActivityViolin

*Generate violin plots of inferred activities for a list of TFs grouped by cluster/group labels*

Description

Generate violin plots of inferred activities for a list of TFs grouped by cluster/group labels

Usage

plotActivityViolin(
   activity_matrix,  
   tf,  
   clusters,  
   ncol = NULL,  
   nrow = NULL,  
   title = NULL,  
   combine = FALSE,  
   legend.label = NULL,  
   colors = c(’blue’,’yellow’),  
   limit = NULL,  
   ...  
)
plotActivityViolin

```r
nrow = NULL,
combine = TRUE,
legend.label = "activity",
colors = NULL,
title = NULL,
text_size = 10,
facet_grid_variable = NULL,
boxplot = FALSE
```

**Arguments**

- **activity_matrix**
  A matrix of TF activities inferred from calculateActivity
- **tf**
  A character vector indicating the names of the transcription factors to be plotted
- **clusters**
  A vector of cluster or group labels for single cells
- **ncol**
  A integer to indicate the number of columns in the combined plot, if combine = TRUE
- **nrow**
  A integer to indicate the number of rows in the combined plot, if combine = TRUE
- **combine**
  logical to indicate whether to combine and visualize the plots in one panel
- **legend.label**
  String indicating the name of variable to be plotted on the legend
- **colors**
  A character vector representing the names of colors
- **title**
  String indicating the title of the plot if combine = TRUE
- **text_size**
  Scalar indicating the font size of the title
- **facet_grid_variable**
  A character vector of a secondary label to split the plots by facet_grid
- **boxplot**
  logical indicating whether to add boxplot on top of violin plot

**Value**

A combined ggplot object or a list of ggplots if combine = FALSE

**Author(s)**

Xiaosai Yao, Shang-yang Chen

**Examples**

```r
# create a mock singleCellExperiment object for gene expression matrix
dataset <- scuttle::mockSCE()
dataset <- scuttle::logNormCounts(dataset)
dataset$cluster <- sample(LETTERS[1:5], ncol(dataset), replace = TRUE)
plotActivityViolin(activity_matrix = logcounts(dataset),
tf = c('Gene_0001','Gene_0002'), clusters = dataset$cluster)
```
plotBubble

Generate bubble plots of relative activities across cluster/group labels for a list of TFs

Description

Generate bubble plots of relative activities across cluster/group labels for a list of TFs

Usage

plotBubble(
  activity_matrix,
  tf,
  clusters,
  bubblesize = c("FDR", "summary.logFC"),
  color.theme = "viridis",
  legend.label = "relative_activity",
  x.label = "clusters",
  y.label = "transcription factors",
  title = "TF activity",
  ...    
)

Arguments

activity_matrix
  A matrix of TF activities inferred from calculateActivity

tf
  A character vector indicating the names of the transcription factors to be plotted

clusters
  A character or integer vector of cluster or group labels for single cells

bubblesize
  String indicating the variable from findDifferentialActivity output to scale size of bubbles by either FDR or summary.logFC. Default is FDR.

color.theme
  String indicating the color theme used for the bubble plot and corresponding to the color options in scale_color_viridis_c

legend.label
  String indicating the name of legend corresponding to the color scale

x.label
  String indicating the x axis label

y.label
  String indicating the y axis label

title
  String indicating the title of the plot

...
  Additional arguments to pass to findDifferentialActivity

Value

A ggplot object

Author(s)

Shang-yang Chen
Examples

```r
example_sce <- scuttle::mockSCE()
example_sce <- scuttle::logNormCounts(example_sce)
example_sce$cluster <- sample(LETTERS[1:5], ncol(example_sce), replace = TRUE)
plotBubble(activity_matrix = logcounts(example_sce),
            tf = c('Gene_0001','Gene_0002'), clusters = example_sce$cluster)
```

plotDiffNetwork

Plot graph according to grouping factor

Description

Plot graph with separate weights for different levels of the grouping factor

Usage

```r
plotDiffNetwork(
  regulon,
  cutoff = 0.01,
  tf = NULL,
  weight = "weight",
  clusters,
  layout = "stress"
)
```

Arguments

- **regulon**: an object returned by the getRegulon or addWeights function
- **cutoff**: a numeric used to select values of the variables passed in clusters parameter. Values greater than cutoff are retained and used as graph edge weights.
- **tf**: a character vector storing the names of transcription factors to be included in the graph
- **weight**: a string indicating the name of the column in the regulon to be used as the weight of the edges
- **clusters**: a character vector indicating the clusters to be plotted
- **layout**: a layout specification. Any values that are valid for ggraph or create_layout will work.

Value

- a ggraph object

Author(s)

Xiaosai Yao, Tomasz Wlodarczyk
plotEpiregulonNetwork

Plot a graph build based on getRegulon output

Description
This function takes an input an igraph object created by any of the following: buildGraph, addCentrality, igraph::strength, normalizeCentrality. It makes a force-directed layout plot to visualize it at a high level.

Usage

plotEpiregulonNetwork(
  graph,
  layout = "stress",
  label_size = 3,
  tfs_to_highlight = NULL,
  edge_alpha = 0.02,
  point_size = 1,
  point_border_size = 0.5,
  label_alpha = 0.8,
  label_nudge_x = 0.2,
  label_nudge_y = 0.2,
  ...
)

Arguments

graph an igraph object
layout a layout specification. Any values that are valid for ggraph or create_layout will work. Defaults to 'stress'. Consider also trying 'mds', 'nicely', and 'fr' while you experiment.
label_size an integer indicating how large the labels of highlighted transcription factors should be

Examples

#' # create an artificial getRegulon output
set.seed(1234)
tf_set <- apply(expand.grid(LETTERS[1:10], LETTERS[1:10]),1, paste, collapse = '')
regulon <- S4Vectors::DataFrame(tf = sample(tf_set, 5e3, replace = TRUE))
gene_set <- expand.grid(LETTERS[1:10], LETTERS[1:10], LETTERS[1:10])
gene_set <- apply(gene_set,1,function(x) paste0(x,collapse=''))
regulon$target <- sample(gene_set, 5e3, replace = TRUE)
regulon$idxATAC <- 1:5e3
regulon$weight <- cbind(data.frame(C1 = runif(5e3), C2 = runif(5e3), C3 = runif(5e3)))
plotDiffNetwork(regulon, tf = unique(tf_set)[1:3],
clusters = c('C1', 'C2', 'C3'), cutoff = 0.2)
tfs_to_highlight

A character vector specifying which TFs in the plot should be highlighted. Defaults to NULL (no labels).

edge_alpha

A numeric value between 0 and 1 indicating the level of transparency to use for the edge links in the force-directed layout. Defaults to 0.02.

point_size

A numeric value indicating the size of nodes in the force-directed layout.

point_border_size

A numeric value indicating the size of point borders for nodes in the force-directed layout.

label_alpha

A numeric value between 0 and 1 indicating the level of transparency to use for the labels of highlighted nodes.

label_nudge_x

A numeric value indicating the shift of the labels along the x axis that should be used in the force-directed layout.

label_nudge_y

A numeric value indicating the shift of the labels along the y axis that should be used in the force-directed layout.

... optional additional arguments to pass to create_layout

Value

A ggraph object.

Author(s)

Timothy Keyes, Tomasz Wlodarczyk

Examples

# create an artificial getRegulon output
set.seed(1234)
tf_set <- apply(expand.grid(LETTERS[seq_len(5)], LETTERS[seq_len(5)]),1, paste, collapse = '')
regulon <- data.frame(tf = sample(tf_set, 5e2, replace = TRUE))
gene_set <- expand.grid(LETTERS[seq_len(5)], LETTERS[seq_len(5)], LETTERS[seq_len(5)])
gene_set <- apply(gene_set,1,function(x) paste0(x,collapse='')
regulon$target <- sample(gene_set, 5e2, replace = TRUE)
regulon$idxATAC <- seq_len(5e2)
regulon$corr <- runif(5e2)*0.5+0.5
regulon$weights <- runif(500)
# create igraph object
graph_tripartite <- buildGraph(regulon, mode = 'tripartite')
plotEpiregulonNetwork(graph_tripartite, tfs_to_highlight = sample(unique(tf_set),3),
edge_alpha = 0.2)
plotGseaNetwork  

*Plot networks graph of significant genesets from regulonEnrich results*

**Description**

Plot networks graph of significant genesets from regulonEnrich results

**Usage**

```r
plotGseaNetwork(
  tf,  
enrichresults,  
ntop_pathways = 10,  
p.adj_cutoff = 0.05,  
layout = "sugiyama",  
  tf_label = "tf",  
gset_label = "ID",  
    tf_color = "tomato",  
gset_color = "grey"
)
```

**Arguments**

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td><code>tf</code></td>
<td>A vector of gene names to be plotted. They should be present in enrichresults</td>
</tr>
<tr>
<td><code>enrichresults</code></td>
<td>Output from regulonEnrich that computes enriched genesets from user-specified regulons of interest</td>
</tr>
<tr>
<td><code>ntop_pathways</code></td>
<td>An integer indicating the number of top pathways to be included in the graph</td>
</tr>
<tr>
<td><code>p.adj_cutoff</code></td>
<td>A scalar indicating the p.adjusted cutoff for pathways to be included in the graph. Default value is 0.05</td>
</tr>
<tr>
<td><code>layout</code></td>
<td>String indicating layout option from igraph</td>
</tr>
<tr>
<td><code>tf_label</code></td>
<td>String indicating the name of the tf label</td>
</tr>
<tr>
<td><code>gset_label</code></td>
<td>String indicating the name of the geneset label</td>
</tr>
<tr>
<td><code>tf_color</code></td>
<td>String indicating the color of the tf label</td>
</tr>
<tr>
<td><code>gset_color</code></td>
<td>String indicating the color of the geneset label</td>
</tr>
</tbody>
</table>

**Value**

an igraph plot of interconnected pathways through TFs

**Author(s)**

Phoebe Guo, Xiaosai Yao
Examples

```r
AR <- data.frame(ID = c('ANDROGEN RESPONSE', 'PROLIFERATION', 'MAPK'),
p.adjust = c(0.001, 0.01, 0.04))
GATA6 <- data.frame(ID = c('STK33', 'PROLIFERATION', 'MAPK'),
p.adjust = c(0.001, 0.01, 0.04))
enrichresults <- list(AR = AR, GATA6 = GATA6)
plotGseaNetwork(tf = names(enrichresults), enrichresults = enrichresults)
```

Description

Plot transcription factor activity

Usage

```r
plotHeatmapActivity(
    activity_matrix,
    sce,
    tfs,
    downsample = 1000,
    scale = TRUE,
    center = TRUE,
    color_breaks = c(-2, 0, 2),
    colors = c("blue", "white", "red"),
    cell_attributes = NULL,
    col_gap = NULL,
    use_raster = TRUE,
    raster_quality = 10,
    cluster_rows = TRUE,
    cluster_columns = FALSE,
    border = TRUE,
    show_column_names = FALSE,
    ...
)
```

Arguments

- `activity_matrix`: A SingleCellExperiment object containing information of cell attributes
- `sce`: A SingleCellExperiment object containing information of cell attributes
- `tfs`: A character vector indicating the names of the transcription factors to be plotted
- `downsample`: Integer indicating the number of cells to sample from the matrix
- `scale`: Logical indicating whether to scale the heatmap
- `center`: Logical indicating whether to center the heatmap
plotHeatmapRegulon

Plot targets genes of transcription factors in regulons

Description

Plot targets genes of transcription factors in regulons
Usage

`plotHeatmapRegulon(
  sce,
  tfs,
  regulon,
  regulon_column = "weight",
  regulon_cutoff = 0.1,
  downsample = 1000,
  scale = TRUE,
  center = TRUE,
  color_breaks = c(-2, 0, 2),
  colors = c("blue", "white", "red"),
  cell_attributes,
  col_gap = NULL,
  exprs_values = "logcounts",
  use_raster = TRUE,
  raster_quality = 10,
  cluster_rows = FALSE,
  cluster_columns = FALSE,
  border = TRUE,
  show_column_names = FALSE,
  column_col = NULL,
  row_col = NULL,
  ...
)`

Arguments

- **sce**: A `SingleCellExperiment` object containing information of cell attributes
- **tfs**: A character vector indicating the names of the transcription factors to be plotted
- **regulon**: A dataframe of regulons containing `tf`, `targets` and a column for filtering the regulons
- **regulon_column**: String indicating the column names to be used for filtering regulons
- **regulon_cutoff**: A scalar indicating the minimal value to retain the regulons for plotting
- **downsample**: Integer indicating the number of cells to sample from the matrix
- **scale**: Logical indicating whether to scale the heatmap
- **center**: Logical indicating whether to center the heatmap
- **color_breaks**: A vector indicating numeric breaks as input to `circlize::colorRamp2`
- **colors**: A vector of colors corresponding to values in `breaks` as input to `circlize::colorRamp2`
- **cell_attributes**: A character vector matching the column names of `colData(sce)` to be used for plotting
- **col_gap**: String indicating the cell attribute to split the columns of the heatmap by
- **exprs_values**: A string specifying which assay in `assays(object)` to obtain expression values from
use_raster Logical indicating whether to use rasterization to reduce image size
raster_quality Integer indicating the raster quality. The higher the value, the better the resolution
cluster_rows Logical indicating whether to cluster rows
cluster_columns Logical indicating whether to cluster columns
border Logical indicating whether to add border around heatmap
show_column_names Logical indicating whether to show column names
column_col A list specifying the colors in the columns. See here
row_col A list specifying the colors in the rows. See here
...
other arguments for ComplexHeatmap::Heatmap

Value
A Heatmap-class object.

Author(s)
Xiaosai Yao

Examples
example_sce <- scuttle::mockSCE()
example_sce <- scuttle::logNormCounts(example_sce)
example_sce$cluster <- sample(LETTERS[1:5], ncol(example_sce), replace = TRUE)
regulon <- data.frame(tf = c(rep('Gene_0001', 10), rep('Gene_0002', 20)),
target = sample(rownames(example_sce), 30), weight = rnorm(30))
# plot heatmap and rotate labels
plotHeatmapRegulon(example_sce, tfs = c('Gene_0001', 'Gene_0002'), regulon = regulon,
cell_attributes = 'cluster', col_gap = 'cluster', column_title_rot = 90)

regulon regulon created using epiregulon package from reprogram-seq data

Description
regulon created using epiregulon package from reprogram-seq data

Usage
data(regulon)

Format
a DFrame.
regulonEnrich

Value

a DFrame.

Examples

data(regulon)

regulonEnrich

Perform geneset enrichment of user-defined regulons

Usage

regulonEnrich(TF, regulon, weight = "weight", weight_cutoff = 0.5, genesets)

Arguments

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>TF</td>
<td>A character vector of TF names</td>
</tr>
<tr>
<td>regulon</td>
<td>A matrix of weighted regulon consisting of tf, targets, corr and weight</td>
</tr>
<tr>
<td>weight</td>
<td>String indicating the column name that should be used to filter target genes for geneset enrichment. Default is 'weight'.</td>
</tr>
<tr>
<td>weight_cutoff</td>
<td>A numeric scalar to indicate the cutoff to filter on the column specified by corr. Default is 0.5.</td>
</tr>
<tr>
<td>genesets</td>
<td>A dataframe with the first column being the name of the geneset and the second column being the name of the genes</td>
</tr>
</tbody>
</table>

Value

A dataframe showing the significantly enriched pathways

Author(s)

Xiaosai Yao

Examples

#retrieve genesets
H <- EnrichmentBrowser::getGenesets(org = 'hsa', db = 'msigdb',
cat = 'H', gene.id.type = 'SYMBOL')
C6 <- EnrichmentBrowser::getGenesets(org = 'hsa', db = 'msigdb',
cat = 'C6', gene.id.type = 'SYMBOL')

#combine genesets and convert genesets to be compatible with enricher
gs <- c(H,C6)
gs.list <- do.call(rbind,lapply(names(gs), function(x) {
  data.frame(gs=x, genes=gs[[x]])))

head(gs.list)

# get regulon
library(dorothea)
data(dorothea_hs, package = 'dorothea')
regulon <- dorothea_hs
enrichment_results <- regulonEnrich(c('ESR1', 'AR'), regulon = regulon, weight = 'mor',
genesisets = gs.list)
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