Package ‘ggbio’

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Title Visualization tools for genomic data

Description The ggbio package extends and specializes the grammar of graphics for biological data. The graphics are designed to answer common scientific questions, in particular those often asked of high throughput genomics data. All core Bioconductor data structures are supported, where appropriate. The package supports detailed views of particular genomic regions, as well as genome-wide overviews. Supported overviews include ideograms and grand linear views. High-level plots include sequence fragment length, edge-linked interval to data view, mismatch pileup, and several splicing summaries.

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### arrangeGrobByParsingLegend

Arrange grobs by parse their legend.

**Description**

Arrange grobs and parse their legend, then put it together on the right.

**Usage**

```r
arrangeGrobByParsingLegend(..., nrow = NULL, ncol = NULL, widths = c(4, 1), legend.idx = NULL)
```
Arguments

... ggplot graphics.

nrow number of row for layout.

ncol number of columns for layout

widths width ratio for plot group and legend group.

legend.idx legend index you want to keep.

Value

a

Author(s)

Tengfei Yin

Examples

library(ggplot2)
p1 <- qplot(x = mpg, y = cyl, data = mtcars, color = carb)
p2 <- qplot(x = mpg, y = cyl, data = mtcars, color = wt)
p3 <- qplot(x = mpg, y = cyl, data = mtcars, color = qsec)
p4 <- qplot(x = mpg, y = cyl, data = mtcars, color = gear)
arrangeGrobByParsingLegend(p1, p2, p3, p4)
arrangeGrobByParsingLegend(p1, p2, p3, p4, ncol = 1)
arrangeGrobByParsingLegend(p1, p2, p3, p4, legend.idx = 2)

autoplot  

Generic autoplot function

Description

autoplot is a generic function to visualize various data object, it tries to give better default graphics and customized choices for each data type, quick and convenient to explore your genomic data compare to low level ggplot method, it is much simpler and easy to produce fairly complicate graphics, though you may lose some flexibility for each layer.

Usage

## S4 method for signature 'GRanges'
autoplot(object, ..., chr, xlab, ylab, main, truncate.gaps = FALSE,
         truncate.fun = NULL, ratio = 0.0025, space.skip = 0.1,
         legend = TRUE, geom = NULL, stat = NULL,
         chr.weight = NULL,
         coord = c("default", "genome", "truncate_gaps"),
         layout = c("linear", "karyogram", "circle"))
## S4 method for signature 'GRangesList'
autoplot(object, ..., xlab, ylab, main, indName = "grl_name",
         geom = NULL, stat = NULL, coverage.col = "gray50",
         coverage.fill = coverage.col, group.selfish = FALSE)

## S4 method for signature 'IRanges'
autoplot(object, ..., xlab, ylab, main)

## S4 method for signature 'Seqinfo'
autoplot(object, ideogram = FALSE, ...)

## S4 method for signature 'GAlignments'
autoplot(object, ..., xlab, ylab, main, which,
         geom = NULL, stat = NULL)

## S4 method for signature 'BamFile'
autoplot(object, ..., which, xlab, ylab, main, bsgenome, geom = "line",
         stat = "coverage", method = c("raw", "estimate"),
         coord = c("linear", "genome"),
         resize.extra = 10, space.skip = 0.1, show.coverage = TRUE)

## S4 method for signature 'character'
autoplot(object, ..., xlab, ylab, main, which)

## S4 method for signature 'TxDbOREnsDb'
autoplot(object, which, ..., xlab, ylab, main, truncate.gaps = FALSE,
         truncate.fun = NULL, ratio = 0.0025,
         mode = c("full", "reduce"), geom = c("alignment"),
         stat = c("identity", "reduce"),
         names.expr = "tx_name", label = TRUE)

## S4 method for signature 'BSgenome'
autoplot(object, which, ..., xlab, ylab, main, geom = NULL)

## S4 method for signature 'Rle'
autoplot(object, ..., xlab, ylab, main, binwidth, nbin = 30,
         geom = NULL, stat = c("bin", "identity", "slice"),
         type = c("viewSums", "viewMins", "viewMaxs", "viewMeans"))

## S4 method for signature 'RleList'
autoplot(object, ..., xlab, ylab, main, nbin = 30, binwidth,
facetByRow = TRUE, stat = c("bin", "identity", "slice"),
geom = NULL, type = c("viewSums", "viewMins", "viewMaxs", "viewMeans"))

## S4 method for signature 'matrix'
autoplot(object, ..., xlab, ylab, main,
        geom = c("tile", "raster"), axis.text.angle = NULL,
hjust = 0.5, na.value = NULL,
        rownames.label = TRUE, colnames.label = TRUE,
        axis.text.x = TRUE, axis.text.y = TRUE)

## S4 method for signature 'ExpressionSet'
autoplot(object, ..., type = c("heatmap", "none",
        "scatterplot.matrix", "pcp", "MA", "boxplot",
        "mean-sd"), test.method =
        "t", rotate = FALSE, pheno.plot = FALSE, main_to_pheno
        = 4.5, padding = 0.2)

## S4 method for signature 'RangedSummarizedExperiment'
autoplot(object, ..., type = c("heatmap", "link", "pcp", "boxplot", "scatterplot.matrix"), pheno.plot =
        main_to_pheno = 4.5, padding = 0.2, assay.id = 1)

## S4 method for signature 'VCF'
autoplot(object, ..., xlab, ylab, main,
        assay.id,
        type = c("default", "geno", "info", "fixed"),
        full.string = FALSE,
        ref.show = TRUE,
        genome.axis = TRUE,
        transpose = TRUE)

## S4 method for signature 'OrganismDb'
autoplot(object, which, ..., xlab, ylab, main,
        truncate.gaps = FALSE,
        truncate.fun = NULL,
        ratio = 0.0025,
        geom = c("alignment"),
        stat = c("identity", "reduce"),
        columns = c("TXNAME", "SYMBOL", "TXID", "GENEID"),
        names.expr = "SYMBOL",
        label = TRUE,
        label.color = "gray40")

## S4 method for signature 'VRanges'
autoplot(object, ..., which = NULL,
       arrow = TRUE, indel.col = "gray30",
       geom = NULL,
       xlab, ylab, main)

## S4 method for signature 'TabixFile'
 autoplot(object, which, ...)

Arguments

- **object**: object to be plot.
- **columns**: columns passed to method works for TxDb, EnsDb and OrganismDb.
- **label.color**: when label turned on for gene model, this parameter controls label color.
- **arrow**: arrow passed to geome_alignment function to control intron arrow attributes.
- **indel.col**: indel colors.
- **ideogram**: Weather to call plotIdeogram or not, default is FALSE, if TRUE, layout_karyogram will be called.
- **transpose**: logical value, default TRUE, always make features from VCF as x, so we can use it to map to genomic position.
- **axis.text.angle**: axis text angle.
- **axis.text.x**: logical value indicates whether to show x axis and labels or not.
- **axis.text.y**: logical value indicates whether to show y axis and labels or not.
- **hjust**: horizontal just for axis text.
- **rownames.label**: logical value indicates whether to show rownames of matrix as y label or not.
- **colnames.label**: logical value indicates whether to show colnames of matrix as y label or not.
- **na.value**: color for NA value.
- **rotate**: show pheno plot or not.
- **main_to_pheno**: main matrix plot width to pheno plot width ratio.
- **padding**: padding between plots.
- **assay.id**: index for assay you are going to use.
- **geom**: Geom to use (Single character for now). Please see section Geometry for details.
- **truncate.gaps**: logical value indicate to truncate gaps or not.
- **truncate.fun**: shrinkage function. Please see shrinkagefun in package biovizBase.
- **ratio**: used in maxGap.
- **mode**: Display mode for genomic features.
- **space.skip**: space ratio between chromosome spaces in coordate genome.
- **coord**: Cooridinate system.
chr.weight numeric vectors which sum to <1, the names of vectors has to be matched with seqnames in seqinfo, and you can only specify part of the seqnames, other lengths of chromosomes will be assined proportionally to their seqlengths, for example, you could specify chr1 to be 0.5, so the chr1 will take half of the space and other chromosomes squeezed to take left of the space.

legend A logical value indicates whether to show legend or not. Default is TRUE

which A GRanges object to subset the result, usually passed to the ScanBamParam function. For autoplot,EnsDb, which can in addition also be an object extending AnnotationFilter, an AnnotationFilterList combining such objects or a formula representing a filter expression. See examples below or documentation of AnnotationFilter for more details.

show.coverage A logical value indicates whether to show coverage or not. This is used for geom "mismatch.summary".

resize.extra A numeric value used to add buffer to intervals to compute stepping levels on.

bsgenome A BSgenome object. Only need for geom "mismatch.summary".

xlab x label.

ylab y label.

label logic value, default TRUE. To show label by the side of features.

facetByRow A logical value, default is TRUE ,facet RleList by row. If FALSE, facet by column.

type For Rle/RleList, "raw" plot everything, so be careful, that would be pretty slow if you have too much data. For "viewMins", "viewMaxs", "viewMeans", "viewSums", require extra arguments to slice the object. so users need to at least provide lower, more details and control please refer the the manual of slice function in IRanges. For "viewMins", "viewMaxs", we use viewWhichMin and viewWhichMax to get x scale, for "viewMeans", "viewSums", we use window midpoint as x.

For ExpreesionSet, plotting types.

layout Layout including linear, circular and karyogram. for GenomicRangesList, it only supports circular layout.

method method used for parsing coverage from bam files. 'estimate' use fast esitmated method and 'raw' use relatively slow parsing method.

test.method test method

... Extra parameters. Usually are those parameters used in autoplot to control aesthetics or geometries.

main title.

stat statistical transformation.

indName When coerce GRangesList to GRanges, names created for each group.

coverage.col coverage stroke color.

coverage.fill coverage fill color.

group.selfish Passed to addStepping, control whether to show each group as unique level or not. If set to FALSE, if two groups are not overlapped with each other, they will probably be layout in the same level to save space.
names.expr  names expression used for creating labels. For EnsDb objects either "tx_id", "gene_name" or "gene_id".

binwidth  width of the bins.

nbin  number of bins.

genome.axis  logical value, if TRUE, whenever possible, try to parse genomic position for each column (e.g. RangedSummarizedExperiment), show column as exactly the genomic position instead of showing them side by side and indexed from 1.

full.string  logical value. If TRUE, show full string of indels in plot for VCF.

ref.show  logical value. If TRUE, show REF in VCF at bottom track.

chr  characters indicates the seqnames to be subseted.

Value

A ggplot object, so you can use common features from ggplot2 package to manipulate the plot.

Introduction

autoplot is redefined as generic s4 method inside this package, user could use autoplot in the way they are familiar with, and we are also setting limitation inside this package, like

- scales X scales is always genomic coordinates in most cases, x could be specified as start/end/midpoint when it's special geoms for interval data like point/line
- colors Try to use default color scheme defined in biovizBase package as possible as it can

Geometry

We have developed new geom for different objects, some of them may require extra parameters you need to provide. Some of the geom are more like geom + stat in ggplot2 package. e.g. "coverage.line" and "coverage.polygon". We simply combine them together, but in the future, we plan to make it more general.

This package is designed for only biological data, especially genomic data if users want to explore the data in a more flexible way, you could simply coerce the GRanges to a data.frame, then just use formal autoplot function in ggplot2, or autoplot generic for data.frame.

Some objects share the same geom so we introduce all the geom together in this section

Showing all the intervals as stepped rectangle, colored by strand automatically.
For TxDb or EnsDb objects, showing full model.

segment  Showing all the intervals as stepped segments, colored by strand automatically.
For object BSgenome, show nucleotides as colored segment.
For Rle/RleList, show histogram-like segments.

line  Showing interval as line, the interval data could also be just single position when start = end, x is one of start/end/midpoint, y value is unquoted name in elementMetadata column names. y value is required.

point  Showing interval as point, the interval data could also be just single position when start = end, x is one of start/end/midpoint, y value is unquoted name in elementMetadata column names. y value is required.
For object BSgenome, show nucleotides as colored point.
coverage.line Coverage showing as lines for interval data.

coverage.polygon Coverage showing as polygon for interval data.

splice Splicing summary. The size and width of the line and rectangle should represent the counts in each model. Need to provide model.

single For TxDb or EnsDb objects, showing single(reduced) model only.

tx For TxDb or EnsDb objects, showing transcripts isoforms.

mismatch.summary Showing color coded mismatched stacked bar to indicate the proportion of mismatching at each position, the reference is set to gray.

text For object BSgenome, show nucleotides as colored text.

rectangle For object BSgenome, show nucleotides as colored rectangle.

Faceting

Faceting in ggbio package is a little different from ggplot2 in several ways

- The faceted column could only be seqnames or regions on the genome. So we limited the formula passing to facet argument, e.g something ~ seqnames, is accepted formula, you can change "something" to variable name in the elementMetadata. But you can not change the second part.

- Sometime, we need to view different regions, so we also have a facet_gr argument which accept a GRanges. If this is provided, it will override the default seqnames and use provided region to facet the graphics, this might be useful for different gene centric views.

Author(s)

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Examples

```r
set.seed(1)
N <- 1000
library(GenomicRanges)
gr <- GRanges(seqnames =
sample(c("chr1", "chr2", "chr3"),
size = N, replace = TRUE),
IRanges(
  start = sample(1:300, size = N, replace = TRUE),
  width = sample(70:75, size = N,replace = TRUE)),
strand = sample(c("+", "-", "*"), size = N,
  replace = TRUE),
value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
sample = sample(c("Normal", "Tumor"),
  size = N, replace = TRUE),
pair = sample(letters, size = N,
  replace = TRUE))

idx <- sample(1:length(gr), size = 50)
```
### code chunk number 3: default
```r
autoplot(gr[idx])
```

### code chunk number 4: bar-default-pre
```r
set.seed(123)
gr.b <- GRanges(seqnames = "chr1", IRanges(start = seq(1, 100, by = 10),
width = sample(4:9, size = 10, replace = TRUE)),
score = rnorm(10, 10, 3), value = runif(10, 1, 100))
gr.b2 <- GRanges(seqnames = "chr2", IRanges(start = seq(1, 100, by = 10),
width = sample(4:9, size = 10, replace = TRUE)),
score = rnorm(10, 10, 3), value = runif(10, 1, 100))
gr.b <- c(gr.b, gr.b2)
head(gr.b)
```

### code chunk number 5: bar-default
```r
p1 <- autoplot(gr.b, geom = "bar")
## use value to fill the bar
p2 <- autoplot(gr.b, geom = "bar", aes(fill = value))
tracks(default = p1, fill = p2)
```

### code chunk number 6: autoplot.Rnw:236-237
```r
autoplot(gr[idx], geom = "arch", aes(color = value), facets = sample ~ seqnames)
```

### code chunk number 7: gr-group
```r
gra <- GRanges("chr1", IRanges(c(1,7,20), end = c(4,9,30)), group = c("a", "a", "b"))
## if you don't specify group, then group based on stepping levels, and gaps are computed without
## considering extra group method
p1 <- autoplot(gra, aes(fill = group), geom = "alignment")
## when use group method, gaps only computed for grouped intervals.
## default is group.selfish = TRUE, each group keep one row.
## in this way, group labels could be shown as y axis.
p2 <- autoplot(gra, aes(fill = group, group = group), geom = "alignment")
## group.selfish = FALSE, save space
p3 <- autoplot(gra, aes(fill = group, group = group), geom = "alignment", group.selfish = FALSE)
tracks(’non-group’ = p1, ’group.selfish = TRUE’ = p2 , ’group.selfish = FALSE’ = p3)
```

### code chunk number 8: gr-facet-strand
autoplot(gr, stat = "coverage", geom = "area",
          facets = strand ~ seqnames, aes(fill = strand))

### code chunk number 9: gr-autoplot-circle

autoplot(gr[idx], layout = 'circle')

### code chunk number 10: gr-circle

seqlengths(gr) <- c(400, 500, 700)
values(gr)$to.gr <- gr[sample(1:length(gr), size = length(gr))]
idx <- sample(1:length(gr), size = 50)
gr <- gr[idx]

ggplot() + layout_circle(gr, geom = "ideo", fill = "gray70", radius = 7, trackWidth = 3) +
          layout_circle(gr, geom = "bar", radius = 10, trackWidth = 4,
                         aes(fill = score, y = score)) +
          layout_circle(gr, geom = "point", color = "red", radius = 14,
                         trackWidth = 3, grid = TRUE, aes(y = score)) +
          layout_circle(gr, geom = "link", linked.to = "to.gr", radius = 6, trackWidth = 1)

### code chunk number 11: seqinfo-src

data(hg19Ideogram, package = "biovizBase")
sq <- seqinfo(hg19Ideogram)
sq

### code chunk number 12: seqinfo

autoplot(sq[paste0("chr", c(1:22, "X"))])

### code chunk number 13: ir-load

set.seed(1)
N <- 100
ir <- IRanges(start = sample(1:300, size = N, replace = TRUE),
              width = sample(70:75, size = N, replace = TRUE))

# add meta data
df <- DataFrame(value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
                 sample = sample(c("Normal", "Tumor"),
                                size = N, replace = TRUE),
                 pair = sample(letters, size = N,
                               replace = TRUE))
```r
values(ir) <- df
ir

### code chunk number 14: ir-exp
p1 <- autoplot(ir)
p2 <- autoplot(ir, aes(fill = pair)) + theme(legend.position = "none")
p3 <- autoplot(ir, stat = "coverage", geom = "line", facets = sample ~ .)
p4 <- autoplot(ir, stat = "reduce")
tracks(p1, p2, p3, p4)

### code chunk number 15: grl-simul
set.seed(1)
N <- 100
gr <- GRanges(seqnames = sample(c("chr1", "chr2", "chr3"), size = N, replace = TRUE),
  IRanges(start = sample(1:300, size = N, replace = TRUE),
          width = sample(30:40, size = N, replace = TRUE),
          strand = sample(c("+", "-", "*"), size = N, replace = TRUE),
          value = rnorm(N, 10, 3),
          score = rnorm(N, 100, 30),
          sample = sample(c("Normal", "Tumor"), size = N, replace = TRUE),
          pair = sample(letters, size = N, replace = TRUE))
grl <- split(gr, values(gr)$pair)

### code chunk number 16: grl-exp
p1 <- autoplot(grl, group.selfish = TRUE)
p2 <- autoplot(grl, group.selfish = TRUE, main.geom = "arrowrect", gap.geom = "segment")
tracks(p1, p2)

### code chunk number 17: grl-name
autoplot(grl, aes(fill = ..grl_name..))
```
# equal to
# autoplot(grl, aes(fill = grl_name))

```r
### code chunk number 18: rle-simul
library(IRanges)
set.seed(1)
lambda <- c(rep(0.001, 4500), seq(0.001, 10, length = 500),
            seq(10, 0.001, length = 500))
## @knitr create
xVector <- rpois(1e4, lambda)
xRle <- Rle(xVector)
xRle
```

```r
### code chunk number 19: rle-bin
p1 <- autoplot(xRle)
p2 <- autoplot(xRle, nbin = 80)
p3 <- autoplot(xRle, geom = "heatmap", nbin = 200)
tracks("nbin = 30" = p1, "nbin = 80" = p2, "nbin = 200(heatmap)" = p3)
```

```r
### code chunk number 20: rle-id
p1 <- autoplot(xRle, stat = "identity")
p2 <- autoplot(xRle, stat = "identity", geom = "point", color = "red")
tracks("line" = p1, "point" = p2)
```

```r
### code chunk number 21: rle-slice
p1 <- autoplot(xRle, type = "viewMaxs", stat = "slice", lower = 5)
p2 <- autoplot(xRle, type = "viewMaxs", stat = "slice", lower = 5, geom = "heatmap")
tracks("bar" = p1, "heatmap" = p2)
```

```r
### code chunk number 22: rlel-simul
xRleList <- RleList(xRle, 2L * xRle)
xRleList
```

```r
### code chunk number 23: rlel-bin
```
p1 <- autoplot(xRleList)
p2 <- autoplot(xRleList, nbin = 80)
p3 <- autoplot(xRleList, geom = "heatmap", nbin = 200)
tracks('nbin = 30' = p1, "nbin = 80" = p2, "nbin = 200(heatmap)" = p3)

###################################################
### code chunk number 24: rlel-id
###################################################
p1 <- autoplot(xRleList, stat = "identity")
p2 <- autoplot(xRleList, stat = "identity", geom = "point", color = "red")
tracks('line' = p1, "point" = p2)

###################################################
### code chunk number 25: rlel-slice
###################################################
p1 <- autoplot(xRleList, type = "viewMaxs", stat = "slice", lower = 5)
p2 <- autoplot(xRleList, type = "viewMaxs", stat = "slice", lower = 5, geom = "heatmap")
tracks('bar' = p1, "heatmap" = p2)

###################################################
### code chunk number 26: txdb
###################################################
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
data(genesymbol, package = "biovizBase")
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene

###################################################
### code chunk number 27: txdb-visual
###################################################
p1 <- autoplot(txdb, which = genesymbol["ALDOA"], names.expr = "tx_name:::gene_id")
p2 <- autoplot(txdb, which = genesymbol["ALDOA"], stat = "reduce", color = "brown", fill = "brown")
tracks(full = p1, reduce = p2, heights = c(5, 1)) + ylab("")

###################################################
### EnsDb
###################################################
## Fetching gene models from an EnsDb object.
library(EnsDb.Hsapiens.v75)
ensdb <- EnsDb.Hsapiens.v75
## We use a GeneNameFilter to specifically retrieve all transcripts for that gene.
p1 <- autoplot(ensdb, which = GeneNameFilter("ALDOA"), names.expr = "gene_name")
## Instead of providing the GeneNameFilter, we can also use filter expressions
p2 <- autoplot(ensdb, which = ~ genename == "ALDOA", stat = "reduce", color = "brown", fill = "brown")
tracks(full = p1, reduce = p2, heights = c(5, 1)) + ylab("")
## Alternatively, we can specify a GRangesFilter and display all genes
## that are (partially) overlapping with that genomic region:
gr <- GRanges(seqnames=16, IRanges(30768000, 30770000), strand="+")
autoplot(ensdb, GRangesFilter(gr, "any"), names.expr="gene_name")

## Just submitting the GRanges object also works.
autoplot(ensdb, gr, names.expr="gene_name")

## Or genes encoded on both strands.
gr <- GRanges(seqnames = 16, IRanges(30768000, 30770000), strand = "*")
autoplot(ensdb, GRangesFilter(gr), names.expr="gene_name")

## Also, we can specify directly the gene ids and plot all transcripts of these
## genes (not only those overlapping with the region)
autoplot(ensdb, GeneIdFilter(c("ENSG00000196118", "ENSG00000156873")))

### code chunk number 28: ga-load
library(GenomicAlignments)
data("genesymbol", package = "biovizBase")

bamfile <- system.file("extdata", "SRR027894subRBM17.bam", package="biovizBase")
which <- keepStandardChromosomes(genesymbol["RBM17"])
# need to set use.names = TRUE
ga <- readGAlignments(bamfile,
                      param = ScanBamParam(which = which),
                      use.names = TRUE)

### code chunk number 29: ga-exp
p1 <- autoplot(ga)
p2 <- autoplot(ga, geom = "rect")
p3 <- autoplot(ga, geom = "line", stat = "coverage")
tracks(default = p1, rect = p2, coverage = p3)

### code chunk number 30: bf-load (eval = FALSE)
library(Rsamtools)
# bamfile <- "./wgEncodeCaltechRnaSeqK562R1x75dAlignsRep1V2.bam"
# bf <- BamFile(bamfile)

### code chunk number 31: bf-est-cov (eval = FALSE)
# autoplot(bamfile)
# autoplot(bamfile, which = c("chr1", "chr2"))
# autoplot(bf)
# autoplot(bf, which = c("chr1", "chr2"))
#
```r
## data(genesymbol, package = "biovizBase")
## autoplot(bamfile, method = "raw", which = genesymbol["ALDOA"])
##
## library(BSgenome.Hsapiens.UCSC.hg19)
## autoplot(bf, stat = "mismatch", which = genesymbol["ALDOA"], bsgenome = Hsapiens)

###################################################
### code chunk number 32: char-bam (eval = FALSE)
###################################################
## bamfile <- "./wgEncodeCaltechRnaSeqK562R1x75dAlignsRep1V2.bam"
## autoplot(bamfile)

###################################################
### code chunk number 33: char-gr
###################################################
library(rtracklayer)

 test_path <- system.file("tests", package = "rtracklayer")
test_bed <- file.path(test_path, "test.bed")

 autoplot(test_bed, aes(fill = name))

###################################################
### matrix
###################################################
volcano <- volcano[20:70, 20:60] - 150

 autoplot(volcano)

test_path <- system.file("tests", package = "rtracklayer")
test_bed <- file.path(test_path, "test.bed")

test_bed <- file.path(test_path, "test.bed")

 autoplot(test_bed, aes(fill = name))

 autoplot(volcano, xlab = "xlab", main = "main", ylab = "ylab")

# special scale theme for 0-centered values
autoplot(volcano, geom = "raster") + scale_fill_fold_change()

# when a matrix has colnames and rownames label them by default
colnames(volcano) <- sort(sample(1:300, size = ncol(volcano), replace = FALSE))

 autoplot(volcano) + scale_fill_fold_change()

 rownames(volcano) <- letters[sample(1:24, size = nrow(volcano), replace = TRUE)]

 autoplot(volcano)

# even with row/col names, you could also disable it and just use numeric index

 autoplot(volcano) + scale_fill_fold_change()

 rownames(volcano) <- letters[sample(1:24, size = nrow(volcano), replace = TRUE)]

 autoplot(volcano)

# don't want the axis has label??

 autoplot(volcano, axis.text.x = FALSE)

 autoplot(volcano, axis.text.y = FALSE)

# or totally remove axis

colnames(volcano) <- lapply(letters[sample(1:24, size = ncol(volcano), replace = TRUE)],
 function(x){
  paste(rep(x, 7), collapse = "")
})```
autoplot(volcano)

## Oops, overlapped
autoplot(volcano, axis.text.angle = -45, hjust = 0)

## when character is the value
x <- sample(c(letters[1:3], NA), size = 100, replace = TRUE)
mx <- matrix(x, nrow = 5)
autoplot(mx)

## tile gives you a white margin
rownames(mx) <- LETTERS[1:5]
autoplot(mx, color = "white")
colnames(mx) <- LETTERS[1:20]
autoplot(mx, color = "white")
autoplot(mx, color = "white", size = 2)

## weird in aes(), though works
## default tile is flexible
autoplot(mx, aes(width = 0.6, height = 0.6))
autoplot(mx, aes(width = 0.6, height = 0.6), na.value = "white")
autoplot(mx, aes(width = 0.6, height = 0.6)) + theme_clear()

###################################################
### Views
###################################################
lambda <- c(rep(0.001, 4500), seq(0.001, 10, length = 500),
seq(10, 0.001, length = 500))
xVector <- dnorm(1:5e3, mean = 1e3, sd = 200)
xRle <- Rle(xVector)
v1 <- Views(xRle, start = sample(.4e3:.6e3, size = 50, replace = FALSE), width =1000)
autoplot(v1)
names(v1) <- letters[sample(1:24, size = length(v1), replace = TRUE)]
autoplot(v1)
autoplot(v1, geom = "tile", aes(width = 0.5, height = 0.5))
autoplot(v1, geom = "line")
 autoplot(v1, geom = "line", aes(color = row)) + theme(legend.position = "none")
autoplot(v1, geom = "line", facets = NULL)
autoplot(v1, geom = "line", facets = NULL, alpha = 0.1)

###################################################
### ExpressionSet
###################################################
library(Biobase)
data(sample.ExpressionSet)
sample.ExpressionSet
set.seed(1)
## select 50 features
idx <- sample(seq_len(dim(sample.ExpressionSet)[1]), size = 50)
eset <- sample.ExpressionSet[idx,]
eset
autoplot(as.matrix(pData(eset)))
## default heatmap

```r
p1 <- autoplot(eset)
p2 <- p1 + scale_fill_fold_change()
p2
autoplot(eset)
autoplot(eset, geom = "tile", color = "white", size = 2)
autoplot(eset, geom = "tile", aes(width = 0.6, height = 0.6))

autoplot(eset, pheno.plot = TRUE)
idx <- order(pData(eset)[,1])
eset2 <- eset[,idx]
autoplot(eset2, pheno.plot = TRUE)
```

## parallel coordinate plot

```r
autoplot(eset, type = "pcp")
```

## boxplot

```r
autoplot(eset, type = "boxplot")
```

## scatterplot.matrix

```r
## slow, be careful
##autoplot(eset[, 1:7], type = "scatterplot.matrix")
```

## mean-sd

```r
autoplot(eset, type = "mean-sd"
```

### RangedSummarizedExperiment

```r
library(SummarizedExperiment)
nrows <- 200; ncols <- 6
counts <- matrix(runif(nrows * ncols, 1, 1e4), nrows)
counts2 <- matrix(runif(nrows * ncols, 1, 1e4), nrows)
rowRanges <- GRanges(rep(c("chr1", "chr2"), c(50, 150)),
    IRanges(floor(runif(200, 1e5, 1e6)), width=100),
    strand=sample(c("+", "-"), 200, TRUE))
colData <- DataFrame(Treatment=rep(c("ChIP", "Input"), 3),
    row.names=LETTERS[1:6])
sset <- SummarizedExperiment(assays=SimpleList(counts=counts,
    counts2 = counts2),
    rowRanges=rowRanges, colData=colData)
autoplot(sset) + scale_fill_fold_change()
autoplot(sset, pheno.plot = TRUE)
```

### pcp
autoplot(sset, type = "pcp")

###################################################
### boxplot
###################################################
autoplot(sset, type = "boxplot")

###################################################
### scatterplot matrix
###################################################
## autoplot(sset, type = "scatterplot.matrix")

###################################################
### vcf
###################################################
## Not run:
library(VariantAnnotation)
vcffile <- system.file("extdata", "chr22.vcf.gz", package="VariantAnnotation")
vcf <- readVcf(vcffile, "hg19")
## default use type 'geno'
## default use genome position
autoplot(vcf)
## or disable it
autoplot(vcf, genome.axis = FALSE)
## not transpose
autoplot(vcf, genome.axis = FALSE, transpose = FALSE, rownames.label = FALSE)
autoplot(vcf)
## use
autoplot(vcf, assay.id = "DS")
## equivalent to
autoplot(vcf, assay.id = 2)
## doesn't work when assay.id cannot find
autoplot(vcf, assay.id = "NO")
## use AF or first
autoplot(vcf, type = "info")
## geom bar
autoplot(vcf, type = "info", aes(y = THETA))
autoplot(vcf, type = "info", aes(y = THETA, fill = VT, color = VT))
autoplot(vcf, type = "fixed")
autoplot(vcf, type = "fixed", size = 10) + xlim(c(50310860, 50310890)) + ylim(0.75, 1.25)
p1 <- autoplot(vcf, type = "fixed") + xlim(50310860, 50310890)
p2 <- autoplot(vcf, type = "fixed", full.string = TRUE) + xlim(50310860, 50310890)
tracks("full.string = FALSE" = p1, "full.string = TRUE" = p2)+
scale_y_continuous(breaks = NULL, limits = c(0, 3))
p3 <- autoplot(vcf, type = "fixed", ref.show = FALSE) + xlim(50310860, 50310890) +
scale_y_continuous(breaks = NULL, limits = c(0, 2))
p3
## End(Not run)

```r
library(BSgenome.Hsapiens.UCSC.hg19)
data(genesymbol, package = "biovizBase")
p1 <- autoplot(Hsapiens, which = resize(genesymbol["ALDOA"], width = 50))
p2 <- autoplot(Hsapiens, which = resize(genesymbol["ALDOA"], width = 50), geom = "rect")
tracks(text = p1, rect = p2)
```

```r
sessionInfo()
```

---

### geom_alignment

**Alignment geoms for GRanges object**

#### Description

Show interval data as alignment.

#### Usage

```r
## S4 method for signature 'GRanges'
geom_alignment(data, ..., xlab, ylab, main, facets = NULL, stat = c("stepping", "identity"), range.geom = c("rect", "arrowrect"), gap.geom = c("chevron", "arrow", "segment"), rect.height = NULL, group.selfish = TRUE, label = TRUE)

## S4 method for signature 'TxDbOREnsDb'
geom_alignment(data, ..., which, columns = c("tx_id", "tx_name", "gene_id"), names.expr = "tx_name", facets = NULL, truncate.gaps = FALSE, truncate.fun = NULL, ratio = 0.0025)

## S4 method for signature 'GRangesList'
geom_alignment(data, ..., which = NULL, cds.rect.h = 0.25, exon.rect.h = cds.rect.h, utr.rect.h = cds.rect.h/2, xlab, ylab, main, facets = NULL, geom = "alignment",
```

```r
```
```r
stat = c("identity", "reduce"),
range.geom = "rect",
gap.geom = "arrow",
utr.geom = "rect",
names.expr = NULL,
label = TRUE,
label.color = "gray40",
label.size = 3,
arrow.rate = 0.015,
length = unit(0.1, "cm")
```

```r
## S4 method for signature 'OrganismDb'
geom_alignment(data, ..., which, columns = c("TXNAME", "SYMBOL", "TXID", "GENEID"), names.expr = "SYMBOL", facets = NULL, truncate.gaps = FALSE, truncate.fun = NULL, ratio = 0.0025)
```

### Arguments
- **data**: A GRanges, data.frame, TxDb or EnsDb object.
- **...**: Extra parameters such as aes() passed.
- **which**: GRanges object to subset the TxDb or EnsDb object. For EnsDb: can also be a single object extending AnnotationFilter, an AnnotationFilterList combining such objects or a filter expression in form of a formula.
- **cds.rect.h**: cds heights.
- **exon.rect.h**: exon heights.
- **utr.rect.h**: utr heights.
- **label.color**: label color.
- **label.size**: label size.
- **arrow.rate**: arrow rate.
- **length**: arrow length.
- **columns**: columns to get from object.
- **xlab**: Label for x
- **ylab**: Label for y
- **main**: Title for plot.
- **facets**: Faceting formula to use.
- **stat**: For GRanges: Character vector specifying statistics to use. "stepping" with randomly assigned stepping levels as y varialbe. "identity" allow users to specify y value in aes.
  - For TxDb: defualt "identity" give full gene model and "reduce" for reduced model.
geom_alignment

- `gap.geom`: Geom for 'gap' computed from the data you passed based on the group information.
- `rect.height`: Half height of the arrow body.
- `group.selfish`: Passed to addStepping, control whether to show each group as unique level or not. If set to `FALSE`, if two groups are not overlapped with each other, they will probably be layout in the same level to save space.
- `truncate.gaps`: logical value indicate to truncate gaps or not.
- `truncate.fun`: shrinkage function. Please see `shrinkagefun` in package biovizBase.
- `ratio`: used in `maxGap`.
- `geom`: geometric object. only support "gene" now.
- `range.geom`: geom for main intervals or exons.
- `utr.geom`: geom for utr region.
- `names.expr`: expression for showing y label.
- `label`: logical value. Whether to label the intervals with names specified by argument `names.expr`.

**Value**

A 'Layer'.

**Author(s)**

Tengfei Yin

**Examples**

```r
set.seed(1)
N <- 100
require(GenomicRanges)
## simmulated GRanges
gr <- GRanges(seqnames = sample(c("chr1", "chr2", "chr3"), size = N, replace = TRUE),
              IRanges(
                start = sample(1:300, size = N, replace = TRUE),
                width = sample(70:75, size = N,replace = TRUE)),
              strand = sample(c("+", "-", "x"), size = N, replace = TRUE),
              value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
              sample = sample(c("Normal", "Tumor"), size = N, replace = TRUE),
              pair = sample(letters, size = N, replace = TRUE))
```

## simmulated GRanges
## geom_arch

Arch geoms for GRanges object

### Description

Show interval data as arches.
Usage

```r
## S4 method for signature 'data.frame'
geom_arch(data, ..., n = 25, max.height = 10)
```

```r
## S4 method for signature 'GRanges'
geom_arch(data, ..., xlab, ylab, main, facets = NULL,
          rect.height = 0, n = 25, max.height = 10)
```

Arguments

- `data`: A GRanges or data.frame object.
- `...`: Extra parameters passed to autoplot function, aes mapping support `height`, `x`, `xend`.
  - `xstart` of the arches
  - `xend` end of the arches
  - `height` height of arches
- `xlab`: Label for `x`
- `ylab`: Label for `y`
- `main`: Title for plot.
- `n`: Integer values at which interpolation takes place to create `n` equally spaced points spanning the interval `[min(x)` , `max(x)]`

- `facets`: Faceting formula to use.
- `rect.height`: When data is GRanges, this padding the arches from original y value to allow users putting arches `around` the interval rectangles.
- `max.height`: Max height of all arches.

Details

To draw a interval data as arches, we need to provide a special geom for this purpose. Arches is popular in gene viewer or genomoe browser, when they try to show isoforms or gene model. `geom_arch`, just like any other geom_* function in ggplot2, you can pass `aes()` to it to map variable to height of arches.

Value

A 'Layer'.

Author(s)

Tengfei Yin
Examples

```r
set.seed(1)
N <- 100
library(GenomicRanges)

## simmulated GRanges
gr <- GRanges(seqnames = sample(c("chr1", "chr2", "chr3"), size = N, replace = TRUE),
              IRanges(start = sample(1:300, size = N, replace = TRUE),
                      width = sample(70:75, size = N, replace = TRUE),
                      strand = sample(c("+", "-", "*"), size = N, replace = TRUE),
                      value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
                      sample = sample(c("Normal", "Tumor"), size = N, replace = TRUE),
                      pair = sample(letters, size = N, replace = TRUE))

## default
ggplot(gr) + geom_arch()
# or
ggplot() + geom_arch(gr)

## facetting and aesthetics
ggplot(gr) + geom_arch(aes(color = value, height = value, size = value),
                        alpha = 0.2, facets = sample ~ seqnames)
```

---

**geom_arrow**

Arrow geoms for GRanges object

**Description**

Show interval data as arrows.

**Usage**

```r
## S4 method for signature 'GRanges'
geom_arrow(data, ..., xlab, ylab, main,
           angle = 30, length = unit(0.12, "cm"), type = "open",
```
stat = c("stepping", "identity"), facets = NULL,
arrow.rate = 0.03, group.selfish = TRUE)

Arguments

data A GRanges object.

... Extra parameters such as aes() passed.
xlab Label for x
ylab Label for y
main Title for plot.
angle The angle of the arrow head in degrees (smaller numbers produce narrower,
pointier arrows). Essentially describes the width of the arrow head.
length A unit specifying the length of the arrow head (from tip to base).
type One of "open" or "closed" indicating whether the arrow head should be a closed
triangle.
stat Character vector specifying statistics to use. "stepping" with randomly assigned
stepping levels as y varialbe. "identity" allow users to specify y value in aes.
facets Faceting formula to use.
arow.rate Arrow density of the arrow body.
group.selfish Passed to addStepping, control whether to show each group as unique level or
not. If set to FALSE, if two groups are not overlapped with each other, they will
probably be layout in the same level to save space.

Value

A 'Layer'.

Author(s)

Tengfei Yin

Examples

set.seed(1)
N <- 100
require(GenomicRanges)
## ==============================================================
## simmulated GRanges
## ==============================================================
gr <- GRanges(seqnames =
sample(c("chr1", "chr2", "chr3"),
size = N, replace = TRUE),
IRanges(
    start = sample(1:300, size = N, replace = TRUE),
    width = sample(70:75, size = N,replace = TRUE)),
strand = sample(c("+", "-", "*"), size = N,
```r
replace = TRUE),
value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
sample = sample(c("Normal", "Tumor"),
    size = N, replace = TRUE),
pair = sample(letters, size = N, replace = TRUE))
```

```r
## default
## -----------------------------------------
ggplot(gr) + geom_arrow()

# or
ggplot() + geom_arrow(gr)

## facetting and aesthetics
## -----------------------------------------
ggplot(gr) + geom_arrow(facets = sample ~ seqnames, aes(color = strand, fill = strand))

## stat:identity
## -----------------------------------------
ggplot(gr) + geom_arrow(stat = "identity", aes(y = value))

## stat:stepping
## -----------------------------------------
ggplot(gr) + geom_arrow(stat = "stepping", aes(y = value, group = pair))

## group:selfish
## -----------------------------------------
ggplot(gr) + geom_arrow(stat = "stepping", aes(y = value, group = pair), group.selfish = FALSE)

## other options to control arrow angle, density, ...
## -----------------------------------------
library(grid)
ggplot(gr) + geom_arrow(stat = "stepping", aes(y = value, group = pair),
    arrow.rate = 0.01, length = unit(0.3, "cm"), angle = 45,
    group.selfish = FALSE)
```

**geom_arrowrect**

Arrowrect geoms for GRanges object
**Description**

Show interval data as rectangle with a arrow head.

**Usage**

```r
## S4 method for signature 'GRanges'
geom_arrowrect(data, ..., xlab, ylab, main,
   facets = NULL, stat = c("stepping", "identity"),
   rect.height = NULL, arrow.head = 0.06,
   arrow.head.rate = arrow.head, arrow.head.fix = NULL,
   group.selfish = TRUE)
```

**Arguments**

- `data`: A `GRanges` object.
- `...`: Extra parameters such as `aes()` passed.
- `xlab`: Label for x
- `ylab`: Label for y
- `main`: Title for plot.
- `facets`: Faceting formula to use.
- `stat`: Character vector specifying statistics to use. "stepping" with randomly assigned stepping levels as y variable. "identity" allow users to specify y value in `aes`.
- `rect.height`: Half height of the arrow body.
- `arrow.head`: Arrow head to body ratio.
- `arrow.head.rate`: Arrow head to body ratio. same with `arrow.head`.
- `arrow.head.fix`: fixed length of arrow head.
- `group.selfish`: Passed to `addStepping`, control whether to show each group as unique level or not. If set to FALSE, if two groups are not overlapped with each other, they will probably be layout in the same level to save space.

**Value**

A 'Layer'.

**Author(s)**

Tengfei Yin

**Examples**

```r
set.seed(1)
N <- 100
require(GenomicRanges)
```
# Simulated GRanges

```r
gr <- GRanges(seqnames =
    sample(c("chr1", "chr2", "chr3"),
           size = N, replace = TRUE),
     IRanges(
           start = sample(1:300, size = N, replace = TRUE),
           width = sample(70:75, size = N, replace = TRUE),
           strand = sample(c("+", "-", "x"), size = N, replace = TRUE),
           value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
           sample = sample(c("Normal", "Tumor"),
                            size = N, replace = TRUE),
           pair = sample(letters, size = N, replace = TRUE))
```

## Default

```r
ggplot(gr) + geom_arrowrect()
goplot() + geom_arrowrect(gr)
```

## Facetting and aesthetics

```r
ggplot(gr) + geom_arrowrect(facets = sample ~ seqnames, aes(color = strand, fill = strand))
```

## Stat: identity

```r
ggplot(gr) + geom_arrowrect(stat = "identity", aes(y = value))
```

## Stat: stepping

```r
ggplot(gr) + geom_arrowrect(stat = "stepping", aes(y = value, group = pair))
```

## group.selfish controls when

```r
ggplot(gr) + geom_arrowrect(gr, stat = "stepping", aes(y = value, group = pair), group.selfish = FALSE)
```

---

**geom_bar**

*Segment geoms for GRanges object*
Description

Show interval data as vertical bar, width equals to interval width and use 'score' or specified 'y' as y scale.

Usage

## S4 method for signature 'ANY'
geom_bar(data, ...)
## S4 method for signature 'GRanges'
geom_bar(data, ..., xlab, ylab, main)

Arguments

data
Typically a GRanges or data.frame object.
...
Extra parameters such as aes() or color, size passed.
xlab
Label for x
ylab
Label for y
main
Title for plot.

Details

Useful for showing bed like files, when imported as GRanges, have a extra 'score' column, use it as default y, you could also specify y by using aes(y = ).

Value

A 'Layer'.

Examples

## load
library(GenomicRanges)

## simul
set.seed(123)
gr.b <- GRanges(seqnames = "chr1", IRanges(start = seq(1, 100, by = 10),
    width = sample(4:9, size = 10, replace = TRUE)),
    score = rnorm(10, 10, 3), value = runif(10, 1, 100))
gr.b2 <- GRanges(seqnames = "chr2", IRanges(start = seq(1, 100, by = 10),
    width = sample(4:9, size = 10, replace = TRUE)),
    score = rnorm(10, 10, 3), value = runif(10, 1, 100))
gr.b <- c(gr.b, gr.b2)
## default use score as y

## bar
ggplot(gr.b) + geom_bar(aes(fill = value))
## or
ggplot() + geom_bar(gr.b, aes(fill = value))
ggplot(gr.b) + geom_bar(aes(y = value))
## geom_chevron

Chevron geoms for GRanges object

### Description

Break normal intervals stored in GRanges object and show them as chevron, useful for showing model or splice summary.

### Usage

```r
# S4 method for signature 'GRanges'
geom_chevron(data, ..., xlab, ylab, main,
    offset = 0.1,
    facets = NULL,
    stat = c("stepping", "identity"),
    chevron.height.rescale = c(0.1, 0.8),
    group.selfish = TRUE)
```

### Arguments

- **data**: A GRanges object.
- **...**: Extra parameters passed to autoplot function.
- **xlab**: Label for x
- **ylab**: Label for y
- **main**: Title for plot.
- **offset**: A numeric value or characters. If it’s numeric value, indicate how much you want the chevron to wiggle, usually the rectangle for drawing GRanges is of height unit 1, so it’s better between -0.5 and 0.5 to make it nice looking. Unless you specify offset as one of those columns, this will use height of the chevron to indicate the columns. Of course you could use size of the chevron to indicate the column variable easily, please see the examples.
- **facets**: faceting formula to use.
- **stat**: character vector specifying statistics to use. "stepping" with randomly assigned stepping levels as y variable. "identity" allow users to specify y value in aes.
- **chevron.height.rescale**: A numeric vector of length 2. When the offset parameters is a character which is one of the data columns, this parameter rescale the offset.
- **group.selfish**: Passed to addStepping, control whether to show each group as unique level or not. If set to FALSE, if two groups are not overlapped with each other, they will probably be layout in the same level to save space.
Details

To draw a normal GRanges as Chevron, we need to provide a special geom for this purpose. Chevron is popular in gene viewer or genomoe browser, when they try to show isoforms or gene model. geom_chevron, just like any other geom_* function in ggplot2, you can pass aes() to it to use height of chevron or width of chevron to show statistics summary.

Value

A `Layer`.

Author(s)

Tengfei Yin

Examples

```r
set.seed(1)
N <- 100
require(GenomicRanges)

## ======================================================================
## simmulated GRanges
## ======================================================================
gr <- GRanges(seqnames =
sample(c("chr1", "chr2", "chr3"),
    size = N, replace = TRUE),
IRanges(
    start = sample(1:300, size = N, replace = TRUE),
    width = sample(70:75, size = N,replace = TRUE)),
strand = sample(c("+", "-", "*"), size = N,
    replace = TRUE),
value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
sample = sample(c("Normal", "Tumor"),
    size = N, replace = TRUE),
pair = sample(letters, size = N,
    replace = TRUE))

## ======================================================================
## default
## ======================================================================
ggplot(gr) + geom_chevron()
## or
ggplot() + geom_chevron(gr)

## ==============================================================
## facetting and aesthetics
## ==============================================================
ggplot(gr) + geom_chevron(facets = sample ~ seqnames, aes(color = strand))
```
```r
## stat:identity
ggplot(gr) + geom_chevron(stat = "identity", aes(y = value))

## stat:stepping
ggplot(gr) + geom_chevron(stat = "stepping", aes(group = pair))

## group.selfish controls when
ggplot(gr) + geom_chevron(stat = "stepping", aes(group = pair), group.selfish = FALSE,
                               xlab = "xlab", ylab = "ylab", main = "main")

p <- qplot(x = mpg, y = cyl, data = mtcars)

## offset
gr2 <- GRanges("chr1", IRanges(c(1, 10, 20), width = 5))
gr2.p <- gaps(gr2)
gr2.p <- resize(gr2.p, fix = "center", width = width(gr2.p)+2)

ggplot(gr2) + geom_rect() + geom_chevron(gr2.p)

## notice the rectangle height is 0.8
## offset = 0 just like a line
ggplot(gr2) + geom_rect() + geom_chevron(gr2.p, offset = 0)

## equal height
ggplot(gr2) + geom_rect() + geom_chevron(gr2.p, offset = 0.4)

## chevron.height
values(gr2.p)$score <- c(100, 200)

## normal chevron height
ggplot(gr2) + geom_rect() + geom_chevron(gr2.p, offset = "score")

## rescaled chevron height
ggplot(gr2) + geom_rect() + geom_chevron(gr2.p, offset = "score",
                                          chevron.height.rescale = c(0.4, 10))
```

34  geom_chevron
**Description**

Show interval data as rectangle.

**Usage**

```r
## S4 method for signature 'ANY'
geom_rect(data, ...)
## S4 method for signature 'GRanges'
geom_rect(data, ..., xlab, ylab, main,
             facets = NULL, stat = c("stepping", "identity"),
             rect.height = NULL,
             group.selfish = TRUE)
```

**Arguments**

data: Typically a GRanges or data.frame object. When it's data.frame, it's simply calling ggplot2::geom_rect.

...: Extra parameters such as aes() or color, size passed.

xlab: Label for x

ylab: Label for y

main: Title for plot.

facets: Faceting formula to use.

stat: Character vector specifying statistics to use. "stepping" with randomly assigned stepping levels as y variable. "identity" allow users to specify y value in aes.

rect.height: Half height of the arrow body.

group.selfish: Passed to addStepping, control whether to show each group as unique level or not. If set to FALSE, if two groups are not overlapped with each other, they will probably be layout in the same level to save space.

**Value**

A 'Layer'.

**Author(s)**

Tengfei Yin
Examples

```r
set.seed(1)
N <- 100
require(GenomicRanges)

## simmulated GRanges
gr <- GRanges(seqnames = sample(c("chr1", "chr2", "chr3"),
    size = N, replace = TRUE),
IRanges(
    start = sample(1:300, size = N, replace = TRUE),
    width = sample(70:75, size = N,replace = TRUE)),
strand = sample(c("+", "-", "*"), size = N,
    replace = TRUE),
value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
sample = sample(c("Normal", "Tumor"),
    size = N, replace = TRUE),
pair = sample(letters, size = N,
    replace = TRUE))
```

## data.frame call ggplot2::geom_rect
```r
ggplot() + geom_rect(data = mtcars, aes(xmin = mpg, ymin = wt, xmax = mpg + 10, ymax = wt + 0.2, fill = cyl))
```

## default
```r
ggplot(gr) + geom_rect()
```
```
```
## facetting and aesthetics
```r
ggplot(gr) + geom_rect(facets = sample ~ seqnames, aes(color = strand, fill = strand))
```

## stat:identity
```r
ggplot(gr) + geom_rect(stat = "identity", aes(y = value))
```
geom_segment

## Description
Show interval data as segments.

## Usage

```r
## S4 method for signature 'ANY'
geom_segment(data, ...)
geom_segment(data, ..., xlab, ylab, main, facets = NULL, stat = c("stepping", "identity"), group.selfish = TRUE)
```

## Arguments

- `data` A GRanges or data.frame object.
- `...` Extra parameters such as aes() or color, size passed.
- `xlab` Label for x
- `ylab` Label for y
- `main` Title for plot.
- `facets` Faceting formula to use.
- `stat` Character vector specifying statistics to use. "stepping" with randomly assigned stepping levels as y variable. "identity" allow users to specify y value in aes.
- `group.selfish` Passed to addStepping, control whether to show each group as unique level or not. If set to FALSE, if two groups are not overlapped with each other, they will probably be layout in the same level to save space.

## Value

A 'Layer'.
Author(s)

Tengfei Yin

Examples

```r
set.seed(1)
N <- 100
require(GenomicRanges)

## simmulated GRanges
gr <- GRanges(seqnames =
  sample(c("chr1", "chr2", "chr3"),
    size = N, replace = TRUE),
  IRanges(
    start = sample(1:300, size = N, replace = TRUE),
    width = sample(70:75, size = N,replace = TRUE)),
  strand = sample(c("+", "-", "*"), size = N,
    replace = TRUE),
  value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
  sample = sample("Normal", "Tumor"),
    size = N, replace = TRUE),
  pair = sample(letters, size = N,
    replace = TRUE))

## data.frame call ggplot2::geom_segment
ggplot() + geom_segment(data = mtcars, aes(x = mpg, y = wt, xend = mpg + 10, yend = wt + 0.2,
  color = cyl))

## default
##
## ggplot(gr) + geom_segment()
## or
ggplot() + geom_segment(gr)

## facetting and aesthetics
##
## ggplot(gr) + geom_segment(facets = sample ~ seqnames, aes(color = strand))
```

```
GGbio

## ggplot(gr) + geom_segment(stat = "identity", aes(y = value))

## ggplot(gr) + geom_segment(stat = "stepping", aes(y = value, group = pair))

## ggplot(gr) + geom_segment(stat = "stepping", aes(y = value, group = pair), group.selfish = FALSE)

---

GGbio  

class ggbio

Description

da sub class of ggplot and gg class defined in ggplot2 package, used for ggbio specific methods.

Usage

GGbio(ggplot = NULL, data = NULL, fetchable = FALSE, blank = FALSE, ...)

Arguments

ggplot       a ggplot or gg object.
data         raw data
fetchable    logical value, default FALSE, is there any fetch method available.
blank        logical value, default FALSE, is this plot a blank plot.
...          More properties passed to class like Cache.

Details

This class is defined to facilitate the ggbio-specific visualization method, especially when using ggplot to construct ggbio supported object, that will return a ggbio class. And internals tricks will help a lazy evaluation for following + method.

Value

a ggbio object.

Author(s)

Tengfei Yin
See Also

ggplot

Examples

p1 <- qplot()
g1 <- ggbio(p1)
class(g1)

Description

These methods extend ggplot to support several types of Bioconductor objects, as well as some base types like matrix. They return a ggbio object, which stores the original data object. Please check the corresponding method for mold to see how an object is coerced into a data.frame.

Usage

## S3 method for class 'Vector'
ggplot(data, mapping = aes(), ..., 
      environment = parent.frame())

## S3 method for class 'Seqinfo'
ggplot(data, mapping = aes(), ..., 
      environment = parent.frame())

## S3 method for class 'ExpressionSet'
ggplot(data, mapping = aes(), ..., 
      environment = parent.frame())

## S3 method for class 'RsamtoolsFile'
ggplot(data, mapping = aes(), ..., 
      environment = parent.frame())

## S3 method for class 'TxDbOREnsDb'
ggplot(data, mapping = aes(), ..., 
      environment = parent.frame())

## S3 method for class 'BSgenome'
ggplot(data, mapping = aes(), ..., 
      environment = parent.frame())

## S3 method for class 'matrix'
ggplot(data, mapping = aes(), ..., 
      environment = parent.frame())

## S3 method for class 'character'
ggplot(data, mapping = aes(), ..., 
      environment = parent.frame())

## S3 method for class 'SummarizedExperiment'
ggplot(data, mapping = aes(), 
      assay.id = 1L, ..., environment = parent.frame())
## S3 method for class 'GAlignments'
ggplot(data, mapping = aes(), ..., 
       environment = parent.frame())

## S3 method for class 'VCF'
ggplot(data, mapping = aes(), ..., 
       environment = parent.frame())

### Arguments

data
  original data object.

mapping
  the aesthetic mapping.

...
  other arguments passed to specific methods.

environment
  fall-back environment for evaluation of aesthetic symbols

assay.id
  index of assay you are using when multiple assays exist.

### Details

The biggest difference for objects returned by `ggplot` in ggbio from ggplot2, is we always keep the original data copy, this is useful because in ggbio, our starting point is not always data.frame, many special statistical transformation is computed upon original data objects instead of coerced data.frame. This is a hack to follow ggplot2’s API while allow our own defined components to trace back to original data copy and do the transformation. For objects supported by `mold` we transform them to data.frame stored along the original data set, for objects which not supported by `mold` method, we only store the original copy for ggbio specific graphics.

`ggplot()` is typically used to construct a plot incrementally, using the `+` operator to add layers to the existing `ggplot` object. This is advantageous in that the code is explicit about which layers are added and the order in which they are added. For complex graphics with multiple layers, initialization with `ggplot` is recommended. You can always call `qplot` in package ggplot2 or `autoplot` in ggbio for convenient usage.

There are three common ways to invoke `ggplot`:

- `ggplot(df, aes(x, y, <other aesthetics>))`
- `ggplot(df)`
- `ggplot()`

The first method is recommended if all layers use the same data and the same set of aesthetics, although this method can also be used to add a layer using data from another data frame. The second method specifies the default data frame to use for the plot, but no aesthetics are defined up front. This is useful when one data frame is used predominantly as layers are added, but the aesthetics may vary from one layer to another. The third method initializes a skeleton `ggplot` object which is fleshed out as layers are added. This method is useful when multiple data frames are used to produce different layers, as is often the case in complex graphics.

The examples below illustrate how these methods of invoking `ggplot` can be used in constructing a graphic.
Value

A return ggbio object, which is a subclass of ggplot defined in ggplot2 package, but that's more, a '.data' list entry is stored with the returned object.

Author(s)

Tengfei Yin

See Also

mold, ggbio

Examples

```r
set.seed(1)
N <- 100
library(GenomicRanges)
## GRanges
gr <- GRanges(seqnames =
  sample(c("chr1", "chr2", "chr3"),
     size = N, replace = TRUE),
IRanges(
    start = sample(1:300, size = N, replace = TRUE),
    width = sample(70:75, size = N,replace = TRUE)),
strand = sample(c("+", "-", "*"), size = N,
    replace = TRUE),
value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
sample = sample(c("Normal", "Tumor"),
    size = N, replace = TRUE),
pair = sample(letters, size = N,
    replace = TRUE))

## automatically facetting and assign y
## this must mean geom_rect support GRanges object
ggplot(gr) + geom_rect()

ggplot(gr) + geom_alignment()

## use pure ggplot2's geom_rect, no auto facet
ggplot(gr) + ggplot2::geom_rect(aes(xmin = start, ymin = score,
    xmax = end, ymax = score + 1))

## GRangesList
grl <- split(gr, values(gr)$pair)
ggplot(grl) + geom_alignment()

ggplot(grl) + geom_rect()

## use pure ggplot2's geom_rect, no auto facet
ggplot(grl) + ggplot2::geom_rect(aes(xmin = start, ymin = score,
    xmax = end, ymax = score + 1))

## IRanges
```
ir <- ranges(gr)
ggplot(ir) + geom_rect()
ggplot(ir) + layout_circle(geom = "rect")

## Seqinfo
seqlengths(gr) <- c(400, 500, 420)
ggplot(seqinfo(gr)) + geom_point(aes(x = midpoint, y = seqlengths))

## matrix
mx <- matrix(1:12, nrow = 3)
ggplot(mx, aes(x = x, y = y)) + geom_raster(aes(fill = value))
## row is the factor
ggplot(mx, aes(x = x, y = row)) + geom_raster(aes(fill = value))
colnames(mx)
colnames(mx) <- letters[1:ncol(mx)]
mx
## has extra 'colnames'
ggplot(mx, aes(x = x, y = row)) + geom_raster(aes(fill = colnames))
rownames(mx)
rownames(mx) <- LETTERS[1:nrow(mx)]
ggplot(mx, aes(x = x, y = row)) + geom_raster(aes(fill = rownames))
## please check autoplot, matrix for more control

## Views

## ExpressionSet
library(Biocbase)
data(sample.ExpressionSet)
sample.ExpressionSet
set.seed(1)
## select 50 features
idx <- sample(seq_len(dim(sample.ExpressionSet)[1]), size = 50)
eset <- sample.ExpressionSet[idx,]
ggplot(eset) + geom_tile(aes(x = x, y = y, fill = value))
## please check autoplot,matrix method which gives you more control
ggplot(eset) + geom_tile(aes(x = x, y = y, fill = sex))
ggplot(eset) + geom_tile(aes(x = x, y = y, fill = type))

## Rle
library(IRanges)
lambda <- c(rep(0.001, 4500), seq(0.001, 10, length = 500),
seq(10, 0.001, length = 500))
xVector <- rpois(1e4, lambda)
xRle <- Rle(xVector)
ggplot(xRle) + geom_tile(aes(x = x, y = y, fill = value))

## RleList
xRleList <- RleList(xRle, 2L * xRle)
xRleList
ggplot(xRleList) + geom_tile(aes(x = x, y = y, fill = value)) +
facet_grid(group~.)
names(xRleList) <- c("a", "b")
## RangedSummarizedExperiment

```r
library(SummarizedExperiment)
nrows <- 200;(ncols <- 6)
counts <- matrix(runif(nrows * ncols, 1, 1e4), nrows)
counts2 <- matrix(runif(nrows * ncols, 1, 1e4), nrows)
rowRanges <- GRanges(rep(c("chr1", "chr2"), c(50, 150)),
  IRanges(floor(runif(200, 1e5, 1e6)), width=100),
  strand=sample(c("+", "-"), 200, TRUE))
colData <- DataFrame(Treatment=rep(c("ChIP", "Input"), 3),
  row.names=LETTERS[1:6])
sset <- SummarizedExperiment(assays=SimpleList(counts=counts,
  counts2 = counts2),
  rowRanges=rowRanges, colData=colData)
ggplot(sset) + geom_raster(aes(x = x, y = y , fill = value))
```

### Description

ggsave is a convenient function for saving a plot. It defaults to saving the last plot that you displayed, and for a default size uses the size of the current graphics device. It also guesses the type of graphics device from the extension. This means the only argument you need to supply is the filename.

### Usage

ggsave(filename, plot = last_plot(),
  device = default_device(filename), path = NULL,
  scale = 1, width = par("din")[1],
  height = par("din")[2], units = c("in", "cm", "mm"),
  dpi = 300, limitsize = TRUE, ...)

### Arguments

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>filename</td>
<td>file name/filename of plot</td>
</tr>
<tr>
<td>plot</td>
<td>plot to save, defaults to last plot displayed</td>
</tr>
<tr>
<td>device</td>
<td>device to use, automatically extract from file name extension</td>
</tr>
<tr>
<td>path</td>
<td>path to save plot to (if you just want to set path and not filename)</td>
</tr>
<tr>
<td>scale</td>
<td>scaling factor</td>
</tr>
<tr>
<td>width</td>
<td>width (defaults to the width of current plotting window)</td>
</tr>
<tr>
<td>height</td>
<td>height (defaults to the height of current plotting window)</td>
</tr>
<tr>
<td>units</td>
<td>units for width and height when either one is explicitly specified (in, cm, or mm)</td>
</tr>
</tbody>
</table>
dpi
dpi to use for raster graphics

limitsize
when TRUE (the default), ggsave will not save images larger than 50x50 inches, to prevent the common error of specifying dimensions in pixels.

...
other arguments passed to graphics device

Details

ggsave currently recognises the extensions eps/ps, tex (pictex), pdf, jpeg, tiff, png, bmp, svg and wmf (windows only).

---

### Grob-class

Grob getter

---

**Description**

'Grob' class is a container for 'grob' based object defined with grid system. Generic function Grob gets grob object supported by grid system, and make an instance of subclass of class 'Grob'.

'GrobList' is a container of list of 'Grob' object.

**Usage**

```r
## S4 method for signature 'gg'
Grob(x)
## S4 method for signature 'gtable'
Grob(x)
## S4 method for signature 'trellis'
Grob(x)
## S4 method for signature 'lattice'
Grob(x)
## S4 method for signature 'GGbio'
Grob(x)
```

**Arguments**

- `x` object of class: gg, gtable, trellis, lattice, GGbio.

**Value**

A Grob object.

**Author(s)**

Tengfei Yin
## Ideogram

**Plot single chromosome with cytobands**

### Description

Plot single chromosome with cytobands.

### Usage

```r
plotIdeogram(obj, subchr = NULL, zoom.region = NULL, which = NULL, xlab, ylab, main, xlabel = FALSE, color = "red", fill = "red", alpha = 0.7, zoom.offset = 0.2, size = 1, cytobands = TRUE, aspect.ratio = 1/20, genome)
```

### Arguments

- **obj**
  - A GenomicRanges object, which include extra information about cytobands, check biovizBase::isIdeogram.
- **subchr**
  - A single character of chromosome names to show.
- **which**
  - GRanges object to subset and highlight the ideogram.
- **zoom.region**
  - A numeric vector of length 2 indicating zoomed region.
- **xlab**
  - Label for x
- **ylab**
  - Label for y
- **main**
  - Title for plot.
- **xlabel**
  - A logical value. Show the x label or not.
- **color**
  - color for highlight region.
- **fill**
  - fill color for highlight region.
- **alpha**
  - alpha for highlight region.
- **zoom.offset**
  - zoomed highlights region offset around chromosome plotting region.
- **size**
  - size for zoomed region rectangle boundary.
- **cytobands**
  - If FALSE, plot just blank chromosome without cytobands. default is TRUE. es
- **aspect.ratio**
  - aspect ratio for the chromosome ideogram plot, default is NULL.
- **genome**
  - genome character passed to `getIdeogram`
- **...**
  - passed to ggbio constructor.
**Details**

User could provide the whole ideogram and use subchr to point to particular chromosome.

**Value**

A ggplot object.

**Author(s)**

Tengfei Yin

**Examples**

```r
## Not run:
library(biovizBase)
p.ideo <- Ideogram(genome = "hg19")
p.ideo
library(GenomicRanges)
p.ideo + xlim(GRanges("chr2", IRanges(1e8, 1e8+10000)))
Ideogram(genome = "hg19", xlabel = TRUE)
## End(Not run)
```

---

**layout_circle**

Create a circle layout

**Description**

Create a circle layout.

**Usage**

```r
## S4 method for signature 'GRanges'
layout_circle(data, ..., geom = c("point", "line", "link", "ribbon", "rect", "bar", "segment", "hist", "scale", "heatmap", "ideogram", "text"), linked.to, radius = 10, trackWidth = 5, space.skip = 0.015, direction = c("clockwise", "anticlockwise"), link.fun = function(x, y, n = 30) bezier(x, y, evaluation = n), rect.inter.n = 60, rank, ylim = NULL, scale.n = 60, scale.unit = NULL, scale.type = c("M", "B", "sci"), grid.n = 5, grid.background = "gray70", grid.line = "white", grid = FALSE, chr.weight = NULL)

## S4 method for signature 'missing'
layout_circle(data, ...) circle(...)```

```
Arguments

- **data**: A GRanges object.
- **...**: Extra parameters such as aesthetics mapping in aes(), or color, size, etc. For circle function, it passed to layout_circle.
- **geom**: The geometric object to use display the data.
- **linked.to**: Character indicates column that specifying end of the linking lines, that column should be a GRanges object.
- **radius**: Numeric value indicates radius. Default is 10.
- **trackWidth**: Numeric value indicates the track width.
- **space.skip**: Numeric value indicates the ratio of skipped region between chunks(chromosomes in GRanges) to the whole track space.
- **direction**: Space layout orders.
- **link.fun**: Function used for interpolate the linking lines. Default is Hmisc::bezier.
- **rect.inter.n**: n passed to interpolate function in rectangle transformation(from a rectangle) to a section in circular view.
- **rank**: For default equal trackWidth, use rank to specify the circle orders.
- **ylim**: Numeric range to control y limits.
- **scale.n**: Approximate number of ticks you want to show on the whole space. used when scale.unit is NULL.
- **scale.unit**: Unit used for computing scale. Default is NULL.
- **scale.type**: Scale type used for grid
- **grid.n**: integer value indicate horizontal grid line number.
- **grid.background**: grid background color.
- **grid.line**: grid line color.
- **chr.weight**: numeric vectors which sum to <1, the names of vectors has to be matched with seqnames in seqinfo, and you can only specify part of the seqnames, other lengths of chromosomes will be assined proportionally to their seqlengths, for example, you could specify chr1 to be 0.5, so the chr1 will take half of the space and other chromosomes squeezed to take left of the space.

Value

A 'Layer'.

Author(s)

Tengfei Yin
**Examples**

```
N <- 100
library(GenomicRanges)

## ======================================================================
## simmulated GRanges
## ======================================================================
gr <- GRanges(seqnames = sample(c("chr1", "chr2", "chr3"), size = N, replace = TRUE),
             IRanges(
               start = sample(1:300, size = N, replace = TRUE),
               width = sample(70:75, size = N, replace = TRUE),
               strand = sample(c("+", "-", "*"), size = N, replace = TRUE),
               value = rnorm(N, 10, 3),
               score = rnorm(N, 100, 30),
               sample = sample(c("Normal", "Tumor"), size = N, replace = TRUE),
               pair = sample(letters, size = N, replace = TRUE)),
             value = rnorm(N, 10, 3),
             score = rnorm(N, 100, 30),
             sample = sample(c("Normal", "Tumor"), size = N, replace = TRUE),
             pair = sample(letters, size = N, replace = TRUE))

seqlengths(gr) <- c(400, 500, 700)
values(gr)$to.gr <- gr[sample(1:length(gr), size = length(gr))]

## don't pass gr to the ggplot
ggplot() + layout_circle(gr, geom = "ideo", fill = "gray70", radius = 7, trackWidth = 3) +
    layout_circle(gr, geom = "bar", radius = 10, trackWidth = 4, aes(fill = score, y = score)) +
    layout_circle(gr, geom = "point", color = "red", radius = 14,
                  trackWidth = 3, grid = TRUE, aes(y = score)) +
    layout_circle(gr, geom = "link", linked.to = "to.gr", radius = 6, trackWidth = 1)

## more formal API
ggplot(gr) + layout_circle(geom = "ideo", fill = "gray70", radius = 7, trackWidth = 3) +
    layout_circle(geom = "bar", radius = 10, trackWidth = 4, aes(fill = score, y = score)) +
    layout_circle(geom = "point", color = "red", radius = 14,
                  trackWidth = 3, grid = TRUE, aes(y = score)) +
    layout_circle(geom = "link", linked.to = "to.gr", radius = 6, trackWidth = 1)
```

**Description**

Create a karyogram layout.
## Usage

```r
## S4 method for signature 'GRanges'
layout_karyogram(data, ..., xlab, ylab, main,
                 facets = seqnames ~ ., cytobands = FALSE, geom = "rect",
                 stat = NULL, ylim = NULL, rect.height = 10)
```

### Arguments

- `data`: a GRanges object, which could contain extra information about cytobands. If you want an accurate genome mapping, please provide `seqlengths` with this GRanges object, otherwise it will emit a warning and use data space to estimate the chromosome space which is very rough.
- `...`: Extra parameters such as `aes()` or arbitrary `color` and `size`.
- `xlab`: character vector or expression for x axis label.
- `ylab`: character vector or expression for y axis label.
- `main`: character vector or expression for plot title.
- `facets`: faceting formula to use.
- `cytobands`: logical value indicate to show the cytobands or not.
- `geom`: The geometric object to use display the data.
- `stat`: character vector specifying statistics to use.
- `ylim`: limits for y axis, usually the chromosome spaces y limits are from 0 to `rect.height`, which 10, so if you wan to stack some data on top of it, you can set limits to like `c(10, 20)`.
- `rect.height`: numeric value indicate half of the rectangle plotting region, used for alignment of multiple layers.

### Value

A `Layer`.

### Author(s)

Tengfei Yin

### Examples

```r
### R code from vignette source 'karyogram.Rnw'

# code chunk number 1: loading
library(ggbio)
data(hg19IdeogramCyto, package = "biovizBase")
head(hg19IdeogramCyto)
# default pre-set color stored in
```
getOption("biovizBase")$cytobandColor

# code chunk number 2: default
 autoplot(hg19IdeogramCyto, layout = "karyogram", cytobands = TRUE)

# code chunk number 3: change-order
 library(GenomeInfoDb)
 hg19 <- keepSeqlevels(hg19IdeogramCyto, paste0("chr", c(1:22, "X", "Y")))
 head(hg19)
 autoplot(hg19, layout = "karyogram", cytobands = TRUE)

# code chunk number 4: cyto-normal
 library(GenomicRanges)
## it's a 'ideogram'
 biovizBase::isIdeogram(hg19)
## set to FALSE
 autoplot(hg19, layout = "karyogram", cytobands = FALSE, aes(fill = gieStain)) +
  scale_fill_giemsa()

# code chunk number 5: load-RNAediting
 data(darned_hg19_subset500, package = "biovizBase")
 dn <- darned_hg19_subset500
 head(dn)
## add seqLengths
## we have seqLengths information in another data set
 data(hg19Ideogram, package = "biovizBase")
 seqLengths(dn) <- seqLengths(hg19Ideogram)[names(seqLengths(dn))]
## now we have seqLengths
 head(dn)
## then we change order
 dn <- keepSeqlevels(dn, paste0("chr", c(1:22, "X")))
 autoplot(dn, layout = "karyogram")
## this equivalent to
## autoplot(seqinfo(dn))

# code chunk number 6: load-RNAediting-color
## since default is geom rectangle, even though it's looks like segment
## we still use both fill/color to map colors
autoplot(dn, layout = "karyogram", aes(color = exReg, fill = exReg))

##########################################################################
### code chunk number 7: load-RNAediting-color-NA
##########################################################################
## since default is geom rectangle, even though it's looks like segment
## we still use both fill/color to map colors
autoplot(dn, layout = "karyogram", aes(color = exReg, fill = exReg)) +
  scale_color_discrete(na.value = "brown")

##########################################################################
### code chunk number 8: load-RNAediting-color-fake
##########################################################################
dn2 <- dn
seqlengths(dn2) <- rep(max(seqlengths(dn2)), length(seqlengths(dn2)))
autoplot(dn2, layout = "karyogram", aes(color = exReg, fill = exReg))

##########################################################################
### code chunk number 9: plotKaryogram (eval = FALSE)
##########################################################################
## plotKaryogram(dn)
## plotKaryogram(dn, aes(color = exReg, fill = exReg))

##########################################################################
### code chunk number 10: low-default
##########################################################################
## plot ideogram
p <- ggplot(hg19) + layout_karyogram(cytobands = TRUE)
p
## equivalent autoplot(hg19, layout = "karyogram", cytobands = TRUE)

##########################################################################
### code chunk number 11: low-default-addon
##########################################################################
p <- p + layout_karyogram(dn, geom = "rect", ylim = c(11, 21), color = "red")
## commented line below won't work
## the cytoband fill color has been used already.
## p <- p + layout_karyogram(dn, aes(fill = exReg, color = exReg), geom = "rect")
p
##########################################################################
### code chunk number 12: edit-space
##########################################################################
## plot chromosome space
p <- autoplot(seqinfo(dn))
## make sure you pass rect as geom
## otherwise you just get background

`p <- p + layout_karyogram(dn, aes(fill = exReg, color = exReg), geom = "rect")`

`values(dn)$pvalue <- rnorm(length(dn))`

`p + layout_karyogram(dn, aes(x = start, y = pvalue), ylim = c(10, 30), geom = "line", color = "red")`

### code chunk number 13: sessionInfo

`sessionInfo()`

---

### Plot

#### Plot class

**Description**

genealize a graphic object to a Plot object.

**Usage**

```r
## S4 method for signature 'gg'
Plot(x)
```

```r
## S4 method for signature 'trellis'
Plot(x, mutable = FALSE)
```

```r
## S4 method for signature 'GGBio'
Plot(x)
```

```r
## S4 method for signature 'Ideogram'
Plot(x)
```

**Arguments**

- `x` object of gg, GGBio, trellis, Ideogram.
- `mutable` whether a plot repsonse to `+` method or not.

**Value**

A Plot object.

**Author(s)**

Tengfei Yin
plotFragLength

Plot estimated fragment length for paired-end RNA-seq data

Description

Plot estimated fragment length for paired-end RNA-seq data against single reduced data model.

Usage

```r
## S4 method for signature 'character,GRanges'
plotFragLength(data, model,
                gap.ratio = 0.0025,
                geom = c("segment", "point", "line"),
                type = c("normal", "cut"),
                heights = c(400, 100),
                annotation = TRUE)
```

Arguments

- `data`: A character indicate the bam file.
- `model`: A reduced model to compute estimated fragment length. please see details.
- `gap.ratio`: When type is set to "cut", it will provide a compact view, which cut the common gaps in a certain ratio.
- `geom`: One or all three geoms could be drawn at the same time. y value of "point" and "line" indicate the estimated fragment length. and if geom is set to "segment", the segment is from the left most position to paired right most position, should be equal to "isize".
- `type`: "normal" return a uncut view, loose but the coordinate is true genomic coordinates. "cut" cut the view in a compact way.
- `heights`: Numeric vector indicate the heights of tracks.
- `annotation`: A logical value. TRUE shows model, and FALSE shows only fragment length with labels.

Details

We use a easy way to define this estimated fragment length, we collect all paired reads and model, reduce model first, then find common gaps, remove common gaps between paired-end reads, and compute the new estimated fragment length.

Value

A ggplot object when annotation = FALSE and a frame grob if annotation = TRUE

Author(s)

Tengfei Yin
Examples

```r
## Not run:
data(genesymbol)
bamfile <- system.file("extdata", "SRR027894subRBM17.bam", package="biovizBase")
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
test <- exonsBy(txdb, by = "tx")
test.new <- subsetByOverlaps(test, genesymbol["RBM17"])
exons.rbm17 <- subsetByOverlaps(exons(txdb), genesymbol["RBM17"])
exons.new <- reduce(exons.rbm17)
plotFragLength(bamfile, exons.new, geom = "line")
plotFragLength(bamfile, exons.new, geom = c("point", "segment"), type = "cut",
              gap.ratio = 0.001)
## End(Not run)
```

---

**plotGrandLinear**

**Manhattan for GWAS**

**Description**

A Manhattan plot is a special scatter plot used to visualize data with a large number of data points, with a distribution of some higher-magnitude values. For example, in GWAS (genome-wide association studies). Here we mainly focus on GWAS Manhattan plots. X-axis is genomic coordinates and Y-axis is negative logarithm of the associated P-value for each single nucleotide polymorphism. So higher the value, more stronger the association they are.

**Usage**

```r
plotGrandLinear(obj, ..., facets, space.skip = 0.01, geom = NULL,
cutoff = NULL, cutoff.color = "red", cutoff.size = 1,
legend = FALSE, xlim, ylim, xlab, ylab, main,
highlight.gr = NULL, highlight.name = NULL,
highlight.col = "red", highlight.label = TRUE,
highlight.label.size = 5, highlight.label.offset = 0.05,
spaceline = FALSE)
```

**Arguments**

- **obj**
  - GRanges object which contains extra p value, before users pass this object, they need to make sure the p value has been changed to -log10(p).
- **...**
  - extra arguments passed. such as color, size, alpha.
- **facets**
  - facets formula, such as group ~ .
space.skip numeric value for skip ratio, between chromosome spaces. default is 0.01.
geom geometric object, default is "point".
cutoff A numeric vector which used as cutoff for Manhattan plot.
cutoff.color A character specifying the color used for cutoff. Default is "red".
cutoff.size A numeric value which used as cutoff line size.
legend A logical value indicate whether to show legend or not. Default is FALSE which disabled the legend.
xlim limits for x scale.
ylim limits for y scale.
xlab Label for x scale.
ylab Label for y scale.
main title.
highlight.gr a GRanges object, this will highlight overlapped region with provided intervals.
highlight.name if NULL, using rownames of GRanges object provided by argument highlight.gr, otherwise use character to indicate column used as labeled names.
highlight.col highlight colors.
highlight.label logical value, label the highlighted region of not.
highlight.label.size highlight label size.
highlight.label.offset highlight label offset.
highlight.label.col highlight label color.
spaceline show line between chromosomes.

Details
Please use seqlengths of the object and space.skip arguments to control the layout of the coordiant genome transformation.
aes(y = ...) is required.
aes(color = ) is used to mapping to data variables, if just pass "color" without aes(), then will recycle the color to represent each chromosomes. please see the example below.

Value
Return a ggplot object.

Author(s)
Tengfei Yin
Examples

```
## load
library(ggbio)
data(hg19IdeogramCyto, package = "biovizBase")
data(hg19Ideogram, package = "biovizBase")
library(GenomicRanges)

## simul_gr
library(biovizBase)
gr <- GRanges(rep(c("chr1", "chr2"), each = 5),
              IRanges(start = rep(seq(1, 100, length = 5), times = 2),
                      width = 50))
autoplot(gr)

## coord:genome
autoplot(gr, coord = "genome")
gr.t <- transformToGenome(gr)
head(gr.t)

## is
is_coord_genome(gr.t)
metadata(gr.t)$coord

## simul_snp
chrs <- as.character(levels(seqnames(hg19IdeogramCyto)))
seqlths <- seqlengths(hg19Ideogram)[chrs]
set.seed(1)
nchr <- length(chrs)
snps <- 100
gr.snp <- GRanges(rep(chrs, each=nsnps),
                  IRanges(start =
                        do.call(c, lapply(chrs, function(chr){
                                      N <- seqlths[chr]
                                      runif(nsnps,1,N)
                                })), width = 1),
                  SNP = sapply(1:(nchr*nsnps), function(x) paste("rs",x,sep='')),
                  pvalue = -log10(runif(nchr*nsnps)),
                  group = sample(c("Normal", "Tumor"), size = nchr*nsnps,
                     replace = TRUE)
)

## shorter
seqlengths(gr.snp)
nms <- seqnames(seqinfo(gr.snp))
nms.new <- gsub("chr", "", nms)
names(nms.new) <- nms
gr.snp <- renameSeqlevels(gr.snp, nms.new)
seqlengths(gr.snp)
```
## unorder
autoplot(gr.snp, coord = "genome", geom = "point", aes(y = pvalue), space.skip = 0.01)

## sort
gr.snp <- keepSeqlevels(gr.snp, c(1:22, "X", "Y"))
autoplot(gr.snp, coord = "genome", geom = "point", aes(y = pvalue), space.skip = 0.01)

## with_seql
names(seqlths) <- gsub("chr", "", names(seqlths))
seqlengths(gr.snp) <- seqlths[names(seqlengths(gr.snp))]
autoplot(gr.snp, coord = "genome", geom = "point", aes(y = pvalue), space.skip = 0.01)

## line
autoplot(gr.snp, coord = "genome", geom = "line", aes(y = pvalue, group = seqnames, color = seqnames))

## plotGrandLinear
plotGrandLinear(gr.snp, aes(y = pvalue))

## morecolor
plotGrandLinear(gr.snp, aes(y = pvalue, color = seqnames))
plotGrandLinear(gr.snp, aes(y = pvalue), color = c("green", "deepskyblue"))
plotGrandLinear(gr.snp, aes(y = pvalue), color = c("green", "deepskyblue", "red"))
plotGrandLinear(gr.snp, aes(y = pvalue), color = "red")

## cutoff
plotGrandLinear(gr.snp, aes(y = pvalue), cutoff = 3, cutoff.color = "blue", cutoff.size = 4)

## cutoff-low
plotGrandLinear(gr.snp, aes(y = pvalue)) + geom_hline(yintercept = 3, color = "blue", size = 4)

## longer
## let's make a long name
nms <- seqnames(seqinfo(gr.snp))
nms.new <- paste("chr00000", nms, sep = "")
names(nms.new) <- nms
gr.snp <- renameSeqlevels(gr.snp, nms.new)
seqlengths(gr.snp)

## rotate
plotGrandLinear(gr.snp, aes(y = pvalue)) + theme(axis.text.x = element_text(angle=-90, hjust=0))

## sessionInfo
sessionInfo()
Description

Plot GRanges object structure and linked to a even spaced paralell coordinates plot which representing the data in elementeMetadata.

Usage

```r
## S4 method for signature 'RangedSummarizedExperiment'
plotRangesLinkedToData(data, ..., stat.y = seq_len(ncol(data)), stat.ylab = names(assays(data)[stat.assay]), stat.assay = 1L)
```

```r
## S4 method for signature 'GenomicRanges_OR_GRangesList'
plotRangesLinkedToData(data, ..., stat.y = seq_len(ncol(mcols(data))), stat.ylab, sig, sig.col = c("black", "red"), stat.coord.trans = coord_trans(), annotation = list(), width.ratio = 0.8, theme.stat = theme_gray(), theme.align = theme_gray(), linetype = 3, heights)
```

Arguments

data: GRanges object with a DataFrame as elementMetadata.

...: Parameters passed to control lines in top plot.

stat.y: integer (variable position starting in DataFrame of data, start from 1) or strings (variable names) which indicate the column names.

stat.ylab: y label for stat track(the top track).

stat.assay: default 1L, element of assays.

sig: a character of element meta data column of logical value, indicates which row is significant. and will be shown in link lines and rectangle.

sig.col: colors for significant, valid when you specify "sig" argument, the first color indicates FALSE, non-significant, the second color indicates TRUE.

stat.coord.trans: transformation used for top plot.

annotation: A list of ggplot object.

width.ratio: Control the segment length of statistic layer.

theme.stat: top plot theme.

theme.align: alignment themes.

linetype: linetype

heights: Heights of each track.
plotRangesLinkedToData

Details

Inspired by some graphics produced in some other packages, for example in package DEXseq, the author provides graphics with gene models and linked to an even spaced statistics summary. This is useful because we always plot everything along the genomic coordinates, but genomic features like exons are not evenly distributed, so we could actually treat the statistics associated with exons like categorical data, and show them as “Paralell Coordinates Plots”. This is one special layout which represent the data in a nice manner and also keep the genomic structure information. With ability of tracks, it’s possible to generate such type of a graphic along with other annotations.

The data we want is a normal GRanges object, and make sure the intervals are not overlaped with each other(currently), and you may have multiple columns which store the statistics for multiple samples, then we produce the graphic we introduced above and users could pass other annotation track in the function which will be shown below the main linked track.

The reason you need to pass annotation into the function instead of binding them by tracks later is because binding manually with annotation tracks is tricky and this function doesn’t return a ggplot object.

Value

return a frame grob; side-effect (plotting) if plot=T.

Author(s)

Tengfei Yin

Examples

```r
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
library(ggbio)
data(genesymbol, package = "biovizBase")
taxdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
model <- exonsBy(txdb, by = "tx")
model17 <- subsetByOverlaps(model, genesymbol["RBM17"])
exons <- exons(txdb)
exon17 <- subsetByOverlaps(exons, genesymbol["RBM17"])
## reduce to make sure there is no overlap
## just for example
exon.new <- reduce(exon17)
## suppose
values(exon.new)$sample1 <- rnorm(length(exon.new), 10, 3)
values(exon.new)$sample2 <- rnorm(length(exon.new), 10, 10)
values(exon.new)$score <- rnorm(length(exon.new))
values(exon.new)$significant <- sample(c(TRUE,FALSE), size = length(exon.new),replace = TRUE)

plotRangesLinkedToData(exon.new, stat.y = c("sample1", "sample2"))
plotRangesLinkedToData(exon.new, stat.y = 1:2)
plotRangesLinkedToData(exon.new, stat.y = 1:2, size = 3, linetype = 4)
plotRangesLinkedToData(exon.new, stat.y = 1:2, size = 3, linetype = 4, sig = "significant")
plotRangesLinkedToData(exon.new, stat.y = 1:2, size = 3, linetype = 4, sig = "significant", sig.col = c("gray90","red"))
```
Description

Plot splice summary by simply counting overlapped junction read in weighted way or not.

Usage

## For character,GRangesList
## S4 method for signature 'character,GRangesList'
plotSpliceSum(data, model, ..., weighted = TRUE)

## For character,TxDB
## S4 method for signature 'character,TxDB'
plotSpliceSum(data, model, which, ..., weighted = TRUE)

## For character,EnsDb
## S4 method for signature 'character,EnsDb'
plotSpliceSum(data, model, which, ..., weighted = TRUE)

Arguments

data A character specifying the bam file path of RNA-seq data.
model A GRangesList which represting different isoforms, a TxDb or an EnsDb object. For the latter cases, users need to pass "which" argument which, for TxDb, is a GRanges object to specify the region and for EnsDb can be a GRanges object, an object extending AnnotationFilter, an AnnotationFilterList combining such filter objects or a filter expression in form of a formula.
which A GRanges object specifying the region you want to get model from the TxDb object. For EnsDb: can be a GRanges object, an object extending AnnotationFilter, an AnnotationFilterList combining such filter objects or a filter expression in form of a formula.
weighted If TRUE, weighted by simply add 1/cases matched to each model and if FALSE, simply add 1 to every case.
... Extra arugments passed to qplot function. such as, offset which control the height of chevron.

Details

Internally we use biovizBase:::spliceSummary for simple counting, but we encourage users to use their own robust way to make slicing summary and store it as GRangesList, then plot the summary by qplot function.

Value

A ggplot object.
Author(s)
   Tengfei Yin

See Also
   qplot

Examples
   ## Not run:
   bamfile <- system.file("extdata", "SRR027894subRBM17.bam", package="biovizBase")
   library(TxDb.Hsapiens.UCSC.hg19.knownGene)
   txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
   data(genesymbol)
   exons <- exonsBy(txdb, by = "tx")
   exons.rbm17 <- subsetByOverlaps(exons, genesymbol["RBM17"])
   plotSpliceSum(bamfile, exons.rbm17)
   plotSpliceSum(bamfile, exons.rbm17, weighted = FALSE, offset = 0.01)
   plotSpliceSum(bamfile, txdb, which = genesymbol["RBM17"])
   plotSpliceSum(bamfile, txdb, which = genesymbol["RBM17"], offset = 0.01)
   plotSpliceSum(bamfile, txdb, which = genesymbol["RBM17"],
                 show.label = TRUE,
                 label.type = "count")
   ## End(Not run)

plotStackedOverview  Plot stacked overview

Description
   Plot stacked overview for genome with or without cytobands. It's a wrapper around layout_karyogram.

Usage
   plotStackedOverview(obj, ..., xlab, ylab, main, geom = "rect",
                      cytobands = FALSE, rescale = TRUE,
                      rescale.range = c(0, 10))
   plotKaryogram(obj, ..., xlab, ylab, main, geom = "rect",
                  cytobands = FALSE, rescale = TRUE,
                  rescale.range = c(0, 10))

Arguments
   obj
      a GRanges object, which could contain extra information about cytobands. If it's missing, will ask user to provide species information and download proper data set from UCSC. If you want an accurate genome mapping, please provide seq_lengths with this GRanges object, otherwise it will emit a warning and use data space to estimate the chromosome space which is very rough.
arguments passed to graphic functions to control aesthetics. For example, if you
use geom "point", you need to provide "y" in aes(), and if can also pass color, fill, size etc. to control graphics.

xlab
label for x

ylab
label for y

main
title for plot.

geom
gem plotted on the stacked layout. Default is "rect", which showing interval
data as rectangles. It automatically figures out boundary so you don’t have to
provide information in aes, users could specify other supported geom works for
data.frame.

cytobands
logical value. Default is FALSE. If TRUE, plotting cytobands, this require your
data have arbitrary column as name and gieStain. the easiest way is to use
getIdeogram to get your data. Notice for this function, when cytobands is TRUE,
it will only plot cytobands without overlaying your data. If you really need to
overlay extra data on cytobands, please plus layout_karyogram for that pur-
pose.

rescale
logical value. Default is TRUE, which rescale your data into the rescale.range,
this make sure your data will not be plotted outside the stacked overview box.

rescale.range
Numeric range of length 2. Default is (0, 10), because stacked layout draws a
white background as chromosome space and this space is of height 10. We hide
the y-axis since we don’t need it for stacked overview. Sometime users may
want to leave some margin for their data, they can use this arguments to control
the rescale.

Details
Stacked overview is just a arbitrary layout for karyogram layout, it use facets seqnaems ~ . as
default to stack the genome. For accurate mapping, you need to provide seqlengths information
in your GRanges object. Otherwise, data space will be computed for stacked overview chromosome
background, this is _NOT_ the actual chromosome space!.

Value
A ggplot object.

Author(s)
Tengfei Yin

Examples
## Not run:
library(biovizBase)
data(hg19IdeogramCyto, package = "biovizBase")
library(GenomicRanges)

## you can also get ideogram by biovizBase::getIdeogram
## make shorter and clean labels
old.chrs <- seqnames(seqinfo(hg19IdeogramCyto))
new.chrs <- gsub("chr", ",", old.chrs)
## lst <- as.list(new.chrs)
names(new.chrs) <- old.chrs
new.ideo <- renameSeqlevels(hg19IdeogramCyto, new.chrs)
new.ideo <- keepSeqlevels(new.ideo, c(as.character(1:22), "X", "Y"))
new.ideo

## sample data
data(darned_hg19_subset500, package = "biovizBase")
idx <- is.na(values(darned_hg19_subset500)$exReg)
values(darned_hg19_subset500)$exReg[idx] <- "unknown"

## you need to add seqlengths for accurate mapping
chrnames <- unique(as.character(seqnames(darned_hg19_subset500)))
data(hg19Ideogram, package = "biovizBase")
seqlengths(darned_hg19_subset500) <- seqlengths(hg19Ideogram)[sort(chrnames)]

dn <- darned_hg19_subset500
values(dn)$score <- rnorm(length(dn))

## plotStackedOverview is a simple wrapper around this functions to create a stacked layout
plotStackedOverview(new.ideo, cytobands = TRUE)

plotStackedOverview(dn)
plotStackedOverview(dn, aes(color = exReg, fill = exReg))
## this will did the trick for you to rescale the space
plotStackedOverview(dn, aes(x = midpoint, y = score), geom = "line")
plotStackedOverview(dn, aes(x = midpoint, y = score), geom = "line", rescale.range = c(4, 6))
## no rescale
plotStackedOverview(dn, aes(x = midpoint, y = score), geom = "line", rescale = FALSE, xlab = "xlab", ylab = "ylab", main = "main") + ylab("ylab")

## no object? will ask you for species and query the data on the fly
plotStackedOverview()
plotStackedOverview(cytobands = TRUE)

## End(Not run)

---

rescale

rescale ggplot object

**Description**

Rescale a numeric vector or ggplot object, could be used for static zoom-in in ggbio.
rescale

Usage

## S4 method for signature 'numeric'
rescale(x, to = c(0, 1),
       from = range(x, na.rm = TRUE))

## S4 method for signature 'ggplot'
rescale(x, xlim, ylim, sx = 1, sy = 1)
## S4 method for signature 'gg'
rescale(x, xlim, ylim, sx = 1, sy = 1)

Arguments

x       A numeric object or ggplot object to be rescaled.
to      For numeric object. it's a vector of two numeric values, specifying the range to be rescale.
from    Range of x.
xlim    For ggplot object. This specify the new limits on x-scale.
ylim    For ggplot object. This specify the new limits on y-scale.
sx      Scale fold for x-scale. Default is 1, no change.
sy      Scale fold for y-scale. Default is 1, no change.

Details

When x is numeric value, it's just call scales::rescale, please refer to the manual page to check more details. If x is ggplot object, it first try to estimate current x limits and y limits of the ggplot object, then rescale based on those information.

Value

Return the object of the same class as x after rescaling.

Author(s)

Tengfei Yin

Examples

library(ggbi)
head(mtcars)
range(mtcars$mpg)
p <- ggplot(data = mtcars, x = mpg, y = disp, geom = "point")
p.new <- rescale(p, xlim = c(20, 25))
p.new
scale_fill_fold_change  

scale color for fold change values

---

Description

In biology, lots of data are scaled to value around 0, and people like to show them as blue-white-red scale color, where negative value are blue, 0 is white and positive value is red, and they are scaled for continuous variables.

Usage

scale_fill_fold_change()

Value

a list.

Author(s)

Tengfei Yin

Examples

p1 <- autoplot(volcano - 150)
p1
p1 + scale_fill_fold_change()

---

scale_fill_giemsa  
scale filled color to customized giemsa color.

---

Description

scale filled color to customized giemsa color.

Usage

scale_fill_giemsa(fill = getOption("biovizBase")$cytobandColor)

Arguments

fill  
a character vector to indicate colors, and names of vector mapped to gieStain name.

Value

a list.
scale_x_sequnit

Author(s)
Tengfei Yin

Examples
getOption("biovizBase")$cytobandColor
library(biovizBase)
data(hg19IdeogramCyto)
p1 <- autoplot(hg19IdeogramCyto, layout = "karyogram", aes(fill =
gieStain))
p1
p1 + scale_fill_giemsa()

scale_x_sequnit scale x by unit

Description
scale x by unit 'Mb', 'kb', 'bp'.

Usage
scale_x_sequnit(unit = c("Mb", "kb", "bp"), append = NULL)

Arguments
unit unit to scale x. Default is Mb.
append default NULL. If pass a character, it disalbe unit and arbitrarily append a text behind the original x scale numbers.

Value
'position_c'

Author(s)
Tengfei Yin

Examples
library(ggplot2)
p <- qplot(x = seq(1, to = 10000, length.out = 40), y = rnorm(40), geom = "point")
## default mb
p + scale_x_sequnit()
p + scale_x_sequnit("kb")
p + scale_x_sequnit("bp")
stat_aggregate  Generates summaries on the specified windows

Description

Generates summaries on the specified windows

Usage

```r
## S4 method for signature 'GRanges'
stat_aggregate(data, ..., xlab, ylab, main, by, FUN,
               maxgap=-1L, minoverlap=0L,
               type=c("any", "start", "end", "within", "equal"),
               select=c("all", "first", "last", "arbitrary"),
               y = NULL, window = NULL, facets = NULL,
               method = c("mean", "median", "max",
                          "min", "sum", "count", "identity"),
               geom = NULL)
```

Arguments

- **data**: A GRanges or data.frame object.
- **...**: Arguments passed to plot function, such as aes() and color.
- **xlab**: Label for x
- **ylab**: Label for y
- **main**: Title for plot.
- **by**: An object with 'start', 'end', and 'width' methods. Passed to aggregate.
- **FUN**: The function, found via 'match.fun', to be applied to each window of 'x'. Passed to aggregate.
- **maxgap, minoverlap, type**: Used in the internal call to findOverlaps() to detect overlaps. See ?findOverlaps in the IRanges package for a description of these arguments.
- **select**: It passed to findOverlaps.
  - When select is "all" (the default), the results are returned as a Hits object. When select is "first", "last", or "arbitrary" the results are returned as an integer vector of length query containing the first, last, or arbitrary overlapping interval in subject, with NA indicating intervals that did not overlap any intervals in subject.
  - If select is "all", a Hits object is returned. For all other select the return value depends on the drop argument. When select != "all" && !drop, an IntegerList is returned, where each element of the result corresponds to a space in query. When select != "all" && drop, an integer vector is returned containing indices that are offset to align with the unlisted query.
y

A character indicating the variable column for which aggregation is taken on, same as `aes(y = ).`

window

Integer value indicating window size.

facets

Faceting formula to use.

method

customized method for aggregating, if FUN is not provided.

geom

The geometric object to use to display the data.

Value

A `Layer`.

Author(s)

Tengfei Yin

Examples

```r
library(GenomicRanges)
set.seed(1)
N <- 1000
## --- workflow simulation ---
## --- simulating GRanges ---
gr <- GRanges(seqnames =
sample(c("chr1", "chr2", "chr3"), size = N, replace = TRUE),
IRanges(
  start = sample(1:300, size = N, replace = TRUE),
  width = sample(70:75, size = N, replace = TRUE)),
strand = sample(c("+", "-", "*"), size = N, replace = TRUE),
value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
sample = sample(c("Normal", "Tumor"), size = N, replace = TRUE),
pair = sample(letters, size = N, replace = TRUE))

# ggplot2

ggplot(gr) + stat_aggregate(aes(y = value))
# or

# ggplot2

ggplot(gr) + stat_aggregate(y = "value")
ggplot(gr) + stat_aggregate(aes(y = value), window = 36)
ggplot(gr) + stat_aggregate(aes(y = value), select = "first")
# Not run:
# no hits
# ggplot(gr) + stat_aggregate(aes(y = value), select = "first", type = "within")

# Not run

# ggplot2

ggplot(gr) + stat_aggregate(window = 30, aes(y = value), fill = "gray40", geom = "bar")
ggplot(gr) + stat_aggregate(window = 100, fill = "gray40", aes(y = value),
  method = "max", geom = "bar")
```
ggplot(gr) + stat_aggregate(aes(y = value), geom = "boxplot")
ggplot(gr) + stat_aggregate(aes(y = value), geom = "boxplot", window = 60)
## now facets need to take place inside stat_* geom_* for an accurate computation
ggplot(gr) + stat_aggregate(aes(y = value), geom = "boxplot", window = 30,
   facets = sample - seqnames)
## FIXME:
## autoplot(gr, stat = "aggregate", aes(y = value), window = 36)
## autoplot(gr, stat = "aggregate", geom = "boxplot", aes(y = value), window = 36)

---

stat_bin

### Binning method

#### Description

Binning method especially for Rle and RleList, for data.frame it’s just calling ggplot2::stat_bin.

#### Usage

```r
## S4 method for signature 'ANY'
stat_bin(data, ...)

## S4 method for signature 'Rle'
stat_bin(data, ..., binwidth, nbin = 30,
   xlab, ylab, main, geom = c("bar", "heatmap"),
   type = c("viewSums", "viewMins",
            "viewMaxs", "viewMeans"))

## S4 method for signature 'RleList'
stat_bin(data, ..., binwidth, nbin = 30,
   xlab, ylab, main,
   indName = "sample",
   geom = c("bar", "heatmap"),
   type = c("viewSums", "viewMins",
            "viewMaxs", "viewMeans"))
```

#### Arguments

- `data` Typically a data.frame or Rle or RleList object.
- `...` arguments passed to aesthetics mapping.
- `binwidth` width of the bins.
- `nbin` number of bins.
- `xlab` x label.
- `ylab` y label.
- `main` title.
- `indName` when faceted by a RleList, name used for labeling faceted factor. Default is 'sample'.
stat_bin

- **geom**: geometric types.
- **type**: statistical summary method used within bins, shown as bar height or heatmap colors.

**Value**

- a ggplot object.

**Author(s)**

- Tengfei Yin

**Examples**

```r
library(IRanges)
lambda <- c(rep(0.001, 4500), seq(0.001, 10, length = 500),
            seq(10, 0.001, length = 500))
xVector <- rpois(1e4, lambda)
xRle <- Rle(xVector)
xRleList <- RleList(xRle, 2L * xRle)

ggplot() + stat_bin(xRle)
ggplot(xRle) + stat_bin()
ggplot(xRle) + stat_bin(nbin = 100)
ggplot(xRle) + stat_bin(binwidth = 200)

p1 <- ggplot(xRle) + stat_bin(type = "viewMeans")
p2 <- ggplot(xRle) + stat_bin(type = "viewSums")
## y scale are different.
tracks(viewMeans = p1, viewSums = p2)

ggplot(xRle) + stat_bin(geom = "heatmap")
ggplot(xRle) + stat_bin(nbin = 100, geom = "heatmap")
ggplot(xRle) + stat_bin(binwidth = 200, geom = "heatmap")

## for RleList
ggplot(xRleList) + stat_bin()
ggplot(xRleList) + stat_bin(nbin = 100)
ggplot(xRleList) + stat_bin(binwidth = 200)

p1 <- ggplot(xRleList) + stat_bin(type = "viewMeans")
p2 <- ggplot(xRleList) + stat_bin(type = "viewSums")
## y scale are different.
tracks(viewMeans = p1, viewSums = p2)

ggplot(xRleList) + stat_bin(geom = "heatmap")
ggplot(xRleList) + stat_bin(nbin = 100, geom = "heatmap")
ggplot(xRleList) + stat_bin(binwidth = 200, geom = "heatmap")
```
stat_coverage  

**Calculate coverage**

**Description**

Calculate coverage.

**Usage**

```r
# for GRanges
## S4 method for signature 'GRanges'
stat_coverage(data, ..., xlim, xlab, ylab, main,
               facets = NULL, geom = NULL)

# for GRangesList
## S4 method for signature 'GRangesList'
stat_coverage(data, ..., xlim, xlab, ylab, main,
               facets = NULL, geom = NULL)

# for Bamfile
## S4 method for signature 'BamFile'
stat_coverage(data, ..., maxBinSize = 2^14,
               xlim, which, xlab, ylab,
               main, facets = NULL, geom = NULL,
               method = c("estimate", "raw"),
               space.skip = 0.1, coord = c("linear", "genome"))
```

**Arguments**

- `data`  A GRanges or data.frame object.
- `...`  Extra parameters such as aes() passed to geom_rect, geom_alignment, or geom_segment.
- `xlim`  Limits for x.
- `xlab`  Label for x
- `ylab`  Label for y
- `main`  Title for plot.
- `facets`  Faceting formula to use.
- `geom`  The geometric object to use display the data.
- `maxBinSize`  maxBinSize.
- `method`  'estimate' for parsing estimated coverage(fast), 'raw' is slow and parse the accurate coverage.
- `which`  GRanges which defines region to subset the results.
- `space.skip`  used for coordinate genome, skip between chromosomes.
- `coord`  coordinate system.
Value

A 'Layer'.

Author(s)

Tengfei Yin

Examples

```r
library(ggbio)
## ======================================================================
## simmulated GRanges
## ======================================================================
set.seed(1)
N <- 1000
library(GenomicRanges)
gr <- GRanges(seqnames =
  sample(c("chr1", "chr2", "chr3"),
    size = N, replace = TRUE),
  IRanges(
    start = sample(1:300, size = N, replace = TRUE),
    width = sample(70:75, size = N, replace = TRUE)),
  strand = sample(c("+", "-", "*"), size = N, replace = TRUE),
  value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
  sample = sample(c("Normal", "Tumor"),
    size = N, replace = TRUE),
  pair = sample(letters, size = N, replace = TRUE))

ggplot(gr) + stat_coverage()
```

```r
## for bam file
## TBD
```

---

**stat_gene**

*Calculate gene structure*

**Description**

Calculate gene structure.
Usage

## S4 method for signature 'TxDb'
stat_gene(data, ...)

Arguments

data A GRanges or data.frame object.
...
Extra parameters such as aes() passed to geom_alignment.

Value

A 'Layer'.

Author(s)

Tengfei Yin

See Also

geom_alignment

Examples

## Not run:
## loading package
## Deprecated
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
data(genesymbol, package = "biovizBase")
taxdb <- TxDb.Hsapiens.UCSC.hg19.knownGene

## made a track comparing full/reduce stat.
p1 <- ggplot() + geom_alignment(txdb, which = genesymbol["RBM17"])
p1 <- ggplot() + stat_gene(txdb, which = genesymbol["RBM17"])
## or
p1 <- ggplot(txdb) + stat_gene(which = genesymbol["RBM17"])  
p1 <- ggplot(txdb) + stat_gene(which = genesymbol["RBM17"])  
p2 <- ggplot(txdb) + stat_gene(which = genesymbol["RBM17"], stat = "reduce")  
p2 <- ggplot(txdb) + stat_gene(which = genesymbol["RBM17"], stat = "reduce")  
## ggplot(txdb) + geom_alignment(which = genesymbol["RBM17"], stat = "reduce")  
## ggplot(txdb) + geom_alignment(which = genesymbol["RBM17"] + stat_reduce()  
## ggplot(txdb) + geom_alignment(which = genesymbol["RBM17"])
tracks(full = p1, reduce = p2, heights = c(3, 1))

## change y labels
ggplot(txdb) + stat_gene(which = genesymbol["RBM17"], names.expr = "tx_id:::gene_id")

## End(Not run)
Transform the data to a suitable data.frame and then one could use multiple geom or even stat to re-plot the data.

Usage

```r
## S4 method for signature 'ANY'
stat_identity(data, ...)

## S4 method for signature 'GRanges'
stat_identity(data, ..., geom = NULL)

## S4 method for signature 'Rle'
stat_identity(data, ..., xlab, ylab, main, geom = NULL)

## S4 method for signature 'RleList'
stat_identity(data, ..., xlab, ylab, main,
             geom = NULL, indName = "sample")
```

Arguments

- `data` Typically a GRanges or data.frame object.
- `...` Extra parameters such as aes() passed to geom_rect, geom_alignment, or geom_segment.
- `geom` The geometric object to use display the data.
- `xlab` x label.
- `ylab` y label.
- `main` title of graphic.
- `indName` sample name.

Value

A 'Layer'.

Author(s)

Tengfei Yin
Examples

```r
## load
set.seed(1)
N <- 50

require(GenomicRanges)

## simul
## simmulated GRanges
##
## gr <- GRanges(seqnames =
## sample(c("chr1", "chr2", "chr3"),
## size = N, replace = TRUE),
## IRanges(
##  start = sample(1:300, size = N, replace = TRUE),
##  width = sample(70:75, size = N, replace = TRUE)),
##  strand = sample(c("+", "-", "*"), size = N,
##  replace = TRUE),
##  value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
##  sample = sample(c("Normal", "Tumor"),
##  size = N, replace = TRUE),
##  pair = sample(letters, size = N,
##  replace = TRUE))

## geom_point_start
ggplot() + stat_identity(gr, aes(x = start, y = value), geom = "point")
## or more formal
ggplot(gr) + stat_identity(aes(x = start, y = value), geom = "point")

## geom_point_midpoint
ggplot(gr) + stat_identity(aes(x = midpoint, y = value), geom = "point")

## geom_rect_all
ggplot(gr) + stat_identity(aes(xmin = start, xmax = end,
  ymin = value - 0.5, ymax = value + 0.5),
  geom = "rect")

## geom_rect_y
ggplot(gr) + stat_identity(aes(y = value), geom = "rect")

## geom_line
ggplot(gr) + stat_identity(aes(x = start, y = value), geom = "line")

## geom_segment
ggplot(gr) + stat_identity(aes(y = value), geom = "segment")

## Rle/RleList
library(IRanges)
lambda <- c(rep(0.001, 4500), seq(0.001, 10, length = 500),
    seq(10, 0.001, length = 500))
xVector <- rpois(1e4, lambda)
xRle <- Rle(xVector)
```
```r
xRleList <- RleList(xRle, 2L * xRle)

ggplot(xRle) + stat_identity(geom = "point")
ggplot(xRleList) + stat_identity(geom = "point")
```

---

### Description

Calculate mismatch summary

### Usage

```r
## for GRanges
## S4 method for signature 'GRanges'
stat_mismatch(data, ..., bsgenome, 
              xlab, ylab, main, 
              geom = c("segment", "bar"), 
              show.coverage = TRUE)
```

```r
## for BamFile
## S4 method for signature 'BamFile'
stat_mismatch(data, ..., bsgenome, which, 
              xlab, ylab, main, 
              geom = c("segment", "bar"), 
              show.coverage = TRUE)
```

### Arguments

- **data** A GRanges or BamFile object.
- **...** Extra parameters such as aes() passed to geom_rect, geom_alignment, or geom_segment.
- **bsgenome** BSgenome object.
- **which** GRanges object to subset the data.
- **xlab** Label for x
- **ylab** Label for y
- **main** Title for plot.
- **geom** The geometric object to use display the data.
- **show.coverage** whether to show coverage as background or not.

### Value

A 'Layer'.

### Author(s)

Tengfei Yin
stat_reduce

Reduce an object.

Description
Reduce GRanges, IRanges or TxDb object.

Usage
```r
## S4 method for signature 'GRanges'
stat_reduce(data, ..., 
    xlab, ylab, main, 
    drop.empty.ranges = FALSE, 
    min.gapwidth = 1L, 
    facets = NULL, geom = NULL)

## S4 method for signature 'IRanges'
stat_reduce(data, ..., 
    xlab, ylab, main, 
    drop.empty.ranges = FALSE, 
    min.gapwidth = 1L, 
    with.inframe.attrib=FALSE, 
    facets = NULL, geom = NULL)

## S4 method for signature 'TxDbOREnsDb'
stat_reduce(data, ...)
```

Arguments
- `data` : GRanges, IRanges or TxDb object.
- `...` : passed to aesthetics mapping.
- `xlab` : x label.
- `ylab` : y label.
- `main` : title.
- `drop.empty.ranges` : pass to `reduce` function.
- `min.gapwidth` : pass to `reduce` function.
- `with.inframe.attrib` : pass to `reduce` function.
- `facets` : pass to `reduce` function.
- `geom` : geometric type.

Value
- a ggplot object.
stat_slice

Author(s)

Tengfei Yin

See Also

reduce.

Examples

```r
set.seed(1)
N <- 1000
library(GenomicRanges)

gr <- GRanges(seqnames =
    sample(c("chr1", "chr2", "chr3"),
    size = N, replace = TRUE),
IRanges(
    start = sample(1:300, size = N, replace = TRUE),
    width = sample(70:75, size = N, replace = TRUE)),
strand = sample(c("+", "-", "*"), size = N, replace = TRUE),
value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
sample = sample(c("Normal", "Tumor"),
    size = N, replace = TRUE),
pair = sample(letters, size = N, replace = TRUE))

ggplot(gr) + stat_reduce()
autoplot(gr, stat = "reduce")
strand(gr) <- "*"
ggplot(gr) + stat_reduce()

library(TxDb.Hsapiens.UCSC.hg19.knownGene)
data(genesymbol, package = "biovizBase")
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
## made a track comparing full/reduce stat.
ggplot(txdb) + stat_reduce(which = genesymbol["RBM17"])
```

stat_slice

Slice Rle/RleList to view them as bar or heatmap.

Description

Slice Rle/RleList to different view by set lower or other parameters, then view summary for all those viewed region.
Usage

```r
## S4 method for signature 'Rle'
stat_slice(data, ..., 
          xlab, ylab, main, 
          na.rm = FALSE, 
          geom = NULL, 
          lower=-Inf, upper=Inf, 
          includeLower=TRUE, includeUpper=TRUE, 
          rangesOnly = FALSE, 
          type = c("viewSums","viewMins", 
                   "viewMaxs", "viewMeans"))
```

```r
## S4 method for signature 'RleList'
stat_slice(data, ..., 
          xlab, ylab, main, 
          indName = "sample", 
          na.rm = FALSE, 
          geom = NULL, 
          lower=-Inf, upper=Inf, 
          includeLower=TRUE, includeUpper=TRUE, 
          rangesOnly = FALSE, 
          type = c("viewSums","viewMins", 
                   "viewMaxs", "viewMeans"))
```

Arguments

- `data`: a `data.frame` or `Rle` or `RleList` object.
- `...`: arguments passed to aesthetics mapping.
- `xlab`: x label.
- `ylab`: y label.
- `main`: title.
- `indName`: when faceted by a `RleList`, name used for labeling faceted factor. Default is `sample`.
- `geom`: geometric types.
- `type`: statistical summary method used within bins, shown as bar height or heatmap colors.
- `na.rm`: logical value, default FALSE, passed to function like `viewMaxs` for statistical summary computation.
- `lower`: passed to `slice`.
- `upper`: passed to `slice`.
- `includeLower`: passed to `slice`.
- `includeUpper`: passed to `slice`.
- `rangesOnly`: passed to `slice`. 
stat_stepping

Calculate stepping levels

Description

Calculate stepping levels.
Usage

```r
## S4 method for signature 'GRanges'
stat_stepping(data, ..., xlab, ylab, main,
facets = NULL,
geom = c("rect", "alignment", "segment"))
```

Arguments

data A GRanges or data.frame object.

... Extra parameters such as aes() passed to geom_rect, geom_alignment, or geom_segment.

xlab Label for x

ylab Label for y

main Title for plot.

facets Faceting formula to use.

geom The geometric object used to display the data. For 'stepping', could be one of 'rect', 'alignment', 'segment'.

Value

A 'Layer'.

Author(s)

Tengfei Yin

Examples

```r
set.seed(1)
N <- 50

require(GenomicRanges)

## simul
## ======================================================================
## simmulated GRanges
## ======================================================================
gr <- GRanges(seqnames =
sample(c("chr1", "chr2", "chr3"),
size = N, replace = TRUE),
IRanges(
    start = sample(1:300, size = N, replace = TRUE),
    width = sample(70:75, size = N,replace = TRUE)),
strand = sample(c("+", "-", "x"), size = N,
replace = TRUE),
value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
sample = sample(c("Normal", "Tumor"),
size = N, replace = TRUE),
pair = sample(letters, size = N,
replace = TRUE))
```
## Usage

```r
## default
ggplot(gr) + stat_stepping()
## or
ggplot() + stat_stepping(gr)

## facet_aes
ggplot(gr) + stat_stepping(aes(color = strand, fill = strand), facets = sample ~ seqnames)

## geom_segment
ggplot(gr) + stat_stepping(aes(color = strand), geom = "segment", xlab = "Genomic coord", ylab = "y", main = "hello")

## geom_alignment
## ggplot(gr) + stat_stepping(geom = "alignment")

## geom_alignment_group
## ggplot(gr) + stat_stepping(aes(group = pair), geom = "alignment")
```

## Description

Tabulate a GRanges object

## Arguments

- `data`: A GRanges or data.frame object.
- `...`: Extra parameters such as `aes()` passed to `geom_rect`, `geom_alignment`, or `geom_segment`.
- `xlab`: Label for x
- `ylab`: Label for y
- `main`: Title for plot.
- `facets`: Faceting formula to use.
- `geom`: The geometric object to use display the data.
- `stat`: The geometric object to use display the data.
_theme_

Value

A 'Layer'.

Author(s)

Tengfei Yin

Examples

```r
## load
default
set.seed(1)
N <- 100
require(ggbio)
require(GenomicRanges)
## simul
## ======================================================================
## simmulated GRanges
## ======================================================================
gr <- GRanges(seqnames =
sample(c("chr1", "chr2", "chr3"),
size = N, replace = TRUE),
IRanges(
    start = sample(1:300, size = N, replace = TRUE),
    width = sample(70:75, size = N, replace = TRUE)),
strand = sample(c("+", "-", "x"), size = N,
    replace = TRUE),
value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
sample = sample(c("Normal", "Tumor"),
    size = N, replace = TRUE),
pair = sample(letters, size = N,
    replace = TRUE))
gr <- c(gr[seqnames(gr) == "chr1"][sample(1:10, size = 1e4, replace = TRUE)],gr)

## default
ggplot(gr) + stat_table()
ggplot(gr) + stat_table(geom = "segment", aes(y = ..score.., color = ..score..))
ggplot(gr) + stat_table(aes(color = score))
```

_theme_ in ggbio

Description

Theme defined in ggbio for plot or tracks.
Usage

theme_null()
theme_noexpand()
theme_alignment(ylabel = FALSE, base_size = 12, base_family = "", axis = TRUE, border = TRUE, grid = TRUE)
theme_pack_panels(strip.bg = FALSE, strip.text.y = TRUE)
theme_clear(grid.y = FALSE, grid.x.minor = FALSE, grid.x.major = FALSE, panel.background.fill = "white", panel.border.color = NA, axis.ticks.x = FALSE, axis.ticks.y = TRUE, grid.color = "gray95", axis.line.color = "gray80")
theme_tracks_sunset(bg = "#ffeebd", alpha = 1, ...)
theme_genome()

Arguments

alpha alpha blending from 0(transparent) to 1(solid).
axis logical value, show axis or not.
axis.line.color color for axis line.
axis.ticks.x show x ticks or not.
axis.ticks.y show y ticks or not.
base_family family for font.
base_size size for font.
bg background color for tracks.
border logical value, show border or not.
grid logical value, show background grid or not.
grid.color grid line color.
grid.x.major show x major grid line or not.
grid.x.minor show x minor grid line or not.
grid.y show y grid or not.
panel.background.fill panel background fill color.
panel.border.color panel border color.
strip.bg if strip background is removed.
strip.text.y if strip text is removed.
ylabel logical value. Show labels or not.
... passed to theme_clear.

details

Themes speciall designed for tracks, are named following naming schema theme_tracks_*
Tracked

Value
Return a theme.

Author(s)
Tengfei Yin

Examples
```r
## load
library(ggbio)
p <- qplot(data = mtcars, x = mpg, y = wt, facets = cyl ~ .)
p + theme_null()
p + theme_clear()
p + theme_pack_panels()
p + theme_alignment()
p1 <- qplot(data = mtcars, x = mpg, y = wt)
tracks(p1 = p, p2 = p1)
tracks(p1 = p, p2 = p1) + theme_tracks_sunset()
```

<table>
<thead>
<tr>
<th>Tracked</th>
<th>Tracked class</th>
</tr>
</thead>
</table>

Description
Create a tracked object, designed for tracks function.

Usage
```
Tracked(mutable = TRUE, fixed = FALSE, labeled = TRUE,
       hasAxis = FALSE, bgColor = "white", height = unit(1, "null"))
```

Arguments
- `mutable` logical value, default TRUE. To control whether a track is updatable by applying + on it.
- `fixed` logical value, default FALSE. To control whether the scale response to a xlim change or not.
- `labeled` logical value, default TRUE. To control whether to label it all not.
- `hasAxis` logical value, default FALSE. To control whether to show axis for that track or not.
- `bgColor` character to control background color of a track.
- `height` unit, to control track height.

Value
a Tracked object.
tracks

Author(s)

Tengfei Yin

tracks

Tracks for genomic graphics

Description

tracks is a convenient constructor for bindind graphics as trakcs. You dont’ have to worry about adjusting different graphics, tracks did that for you. It’s NOT just limited to bind genomic tracks, you can use this function to bind any tracks with the same definition of x axis, for example, sets of time series plots you made.

Tracks view is most common way to viewing genome features and annotation data and widely used by most genome browsers. Our assumption is that, most graphics you made with ggbio or by yourself using ggplot2, are almost always sitting on the genomic coordinates or the same x axis. And to compare annotation information along with genome features, we need to align those plots on exactly the same x axis in order to form your hypothesis. This function leaves you the flexibility to construct each tracks separately with worrying your alignments later.

Usage

tracks(..., heights, xlim, xlab = NULL, main = NULL, 
        title = NULL, theme = NULL, 
        track.plot.color = NULL, 
        track.bg.color = NULL, 
        main.height = unit(1.5, "lines"), 
        scale.height = unit(1, "lines"), 
        xlab.height = unit(1.5, "lines"), 
        padding = unit(-1, "lines"), 
        label.bg.color = "white", 
        label.bg.fill = "gray80", 
        label.text.color = "black", 
        label.text.cex = 1, 
        label.text.angle = 90, 
        label.width = unit(2.5, "lines"))

Arguments

... plots of class ggplot, generated from ggplot2 or ggbio.
heights numeric vector of the same length of passed graphic object to indicate the ratio of each track.
xlim limits on x. could be IRanges, GRanges, numeric value
xlab label for x axis.
main title for the tracks.
title title for the tracks, alias like main.
theme object used for building tracks, this will set to default, which could be reseted later.

track.plot.color
Vector of characters of length 1 or the same length of passed plots, background color for each track, default is white.

track.bg.color background color for the whole tracks.

main.height unit. Height to control the title track height.
scale.height unit. Height to control the scale track height.
xlab.height unit. Height to control the xlab track height.
padding single numeric value or unit, if numeric value, the unit would be "lines" by default.

label.bg.color track labeling background rectangle border color.

label.bg.fill track labeling background fill color.

label.text.color track labeling text color.

label.text.cex track labeling text size.

label.text.angle angle to rotate the track labels.

label.width track labeling size.

Details

tracks did following modification for passed plots.

- remove x-axis, ticks, xlab and tile for each track and add scales at bottom. We suppose a new xlab and title would be provided by the tracks function for the whole tracks, but we still keep individual’s y axis.
- align x-scale limits to make sure every plots sitting on exactly the same x scale.
- squeezing plots together to some extent.
- labeling tracks if names are provided, please check utilities section about labeled method.
- return a track object. This would allow many features introduced in this manual.

Value

A Tracks object.

Track class

constructor tracks will return a Tracks object, which has following slots.

grobs a ggplotGrobList object contains a list of ggplot object, which is our passed graphics.

backup a backup of all the slots for holding the original tracks, so users could edit it and reset it back at any time later, and backup method will reset the backumped copy.

ylim y limits for each plot.
labeled vector of logical value indicates whether a track is labeled or not, for labeled attributes please check utilities section.

mutable vector of logical value indicates whether a track is mutable for theme editing or not, for mutable attributes please check utilities section.

hasAxis vector of logical value indicates whether a track has axis or not, for hasAxis attributes please check utilities section.

heights, xlim, xlab, main, title, theme, fixed, track.plot.color, track.bg.color, main.height, scale.height, xlab.height, padding, label.bg.color, label.bg.fill, label.text.color, label.text.cex, label.text.angle, label.width those slots are described in arguments section for constructor.

Utilities

Please check examples for usage.

summary(object) summary information about tracks object.

fixed(x), fixed(x) <- value x is the ggplot object, this controls if a track has a fixed x scale or not, if the fixed attributes is TRUE, then when you pass this plot to a tracks, this plot won’t be re-aligned with other tracks and will keep the original x-axis, this allow you to pass some plot like ideogram. fixed function will return a logical value

labeled(x), labeled(x) <- value x is the ggplot object, if you pass named graphics into tracks, it will create the labels on the left for you. Several ways supported to name it. You can pass a list of graphics with names. Or you can use tracks('name1' = p1, 'name 2' = p2, ...) with quotes for complicated words or simply tracks(part1 = p1, part = p2, ...).

mutable(x), mutable(x) <- value x is the ggplot object, this controls whether a plot in the tracks mutable to theme changing or not, when you use + method for Tracks object, add-on edit will only be applied to the the mutable plots.

bgColor(x), bgColor(x) <- value x is the ggplot object, this change the background color for single plot shown in the tracks.

xlim(x), xlim(x) <- value when x is the numeric value, it calls ggplot2::coord_cartesian(xlim = ...) method, we doesn’t use ggplot2::xlim() for the reason it will cut data outside the range, and we believe the best behavior would be zoom-in/out like most browser. when x is IRanges, GRanges, it get the range and passed to ggplot2::coord_cartesian function.

when x is Tracks object, xlim(x) will return x limits for that tracks. xlim(x) <- value replace method only works for Tracks object. value could be numeric, IRanges, GRanges object. This will change the x limits associated with tracks.

+ xlim(obj):obj is the numeric range, or IRanges, GRanges object.
+ coord_cartesian(): please read manual in ggplot2, this controls both xlim an ylim, only accept numerical range.

+ The most nice features about Tracks object is the one inherited from ggplot2’s components additive features, with + method you can use any theme object and utilities in ggplot2 package, to add them on a Tracks object, for example, if x is our Tracks object, x + theme would apply theme to any plots in the tracks except those are immutable.

as(x, "grob") Coerces a Tracks object to a grob for embedding in a larger figure.
Backup and reset

reset(obj) obj is the Tracks object, this reset the tracks back to original or backuped version.

backup(obj) obj is the Tracks object, this clear previous backup and use current setting for a new backup.

Author(s)
Tengfei Yin

See Also
align.plots

Examples

```r
## make a simulated time series data set
df1 <- data.frame(time = 1:100, score = sin((1:100)/20)*10)
p1 <- qplot(data = df1, x = time, y = score, geom = "line")
df2 <- data.frame(time = 30:120, score = sin((30:120)/20)*10, value = rnorm(120-30 + 1))
p2 <- ggplot(data = df2, aes(x = time, y = score)) +
  geom_line() + geom_point(size = 4, aes(color = value))
## check p2
p1
## check p2
p2

## binding
tracks(p1, p2)

## or
tks <- tracks(p1, p2)
tks

## combine
c(tks, tks)
tks + tks

cbind(tks, tks)
rbind(tks, tks) ## different wth c()!
library(grid)
x <- as(tks, "grob")
grid.draw(cbind(x, x))

## labeling: default labeling a named graphic
## simply pass a name with it
tracks(time1 = p1, time2 = p2)
## or pass a named list with it
lst <- list(time1 = p1, time2 = p2)
tracks(lst)
```
## more complicated case please use quotes

```r
tracks(time1 = p1, "second time" = p2)
```

## set heights

```r
tracks(time1 = p1, time2 = p2, heights = c(1, 3))
```

## if you want to disable label arbitrarily

### default label is always TRUE

```r
labeled(p2)
labeled(p2) <- FALSE
```

### set labeled to FALSE, remove label even the plot has a name

```r
tracks(time1 = p1, time2 = p2)
labeled(p2) <- TRUE
```

## fix a plot, not synchronize with other plots

```r
p3 <- p1
## default is always FALSE
fixed(p3)
## set to TRUE
fixed(p3) <- TRUE
fixed(p3)
```

```r
tracks(time1 = p1, time2 = p2, "time3(fixed)" = p3)
```

```r
fixed(p3) <- FALSE
## otherwise you could run
```

## control axis

```r
hasAxis(p1)
hasAxis(p1) <- TRUE
# ready for weird looking
tracks(time1 = p1, time2 = p2)
# set it back
hasAxis(p1) <- FALSE
```

## mutable

```r
mutable(p1)
tracks(time1 = p1, time2 = p2) + theme_bw()
mutable(p1) <- FALSE
## mutable for "+" method
tracks(time1 = p1, time2 = p2) + theme_bw()
mutable(p1) <- TRUE
```

## bgColor

```r
bgColor(p1)
tracks(time1 = p1, time2 = p2)
bgColor(p1) <- "brown"
## mutable for "+" method
tracks(time1 = p1, time2 = p2)
```
# set it back
bgColor(p1) <- "white"

## apply a theme to each track
tks <- tracks(time1 = p1, time2 = p2) + theme_bw()
tks
reset(tks)

## store it with tracks
#tks <- tracks(time1 = p1, time2 = p2, theme = theme_bw())
tks
## reset will be introduced later
reset(tks)

## apply a pre-defined theme for tracks!
tracks(time1 = p1, time2 = p2) + theme_tracks_sunset()
tracks(p1, p2) + theme_tracks_sunset()

## change limits
tracks(time1 = p1, time2 = p2) + xlim(c(1, 40))
tracks(time1 = p1, time2 = p2) + xlim(1, 40)
tracks(time1 = p1, time2 = p2) + coord_cartesian(xlim = c(1, 40))

# change y
tracks(time1 = p1, time2 = p2) + xlim(1, 40) + ylim(0, 10)
library(GenomicRanges)
gr <- GRanges("chr", IRanges(1, 40))
# GRanges
tracks(time1 = p1, time2 = p2) + xlim(gr)
# IRanges
tracks(time1 = p1, time2 = p2) + xlim(ranges(gr))
tks <- tracks(time1 = p1, time2 = p2)
xlim(tks)
xlim(tks) <- c(1, 35)
xlim(tks) <- gr
xlim(tks) <- ranges(gr)

## xlab, title
tracks(time1 = p1, time2 = p2, xlab = "time")
tracks(time1 = p1, time2 = p2, main = "title")
tracks(time1 = p1, time2 = p2, title = "title")
tracks(time1 = p1, time2 = p2, xlab = "time", title = "title") + theme_tracks_sunset()

## backup and restore
#tks <- tracks(time1 = p1, time2 = p2)
tks
tks <- tks + xlim(1, 40)
tks
reset(tks)
tks <- tks + xlim(1, 40)
tks
## Description

A set of simple navigation API apply to ggbio object, let you move along the genome and zoom in/out.

## Usage

```r
zoom(fac = 1/2)
zoom_in(fac = 1/2)
zoom_out(fac = 2)
nextView(unit = c("view", "gene", "exon", "utr"))
prevView(unit = c("view", "gene", "exon", "utr"))
```

## Arguments

- **fac**
  - numeric value to indicate zoom factor, multiple of current view width. If it’s smaller than 1, then it’s zoom-in operation; if it’s bigger than 1, then it’s zoom-out operation.

- **unit**
  - only support 'view' unit now.

## Details

- `zoom_in` and `zoom_out` are just simple wrapper around `zoom` function.
  - For more convenient, gene features based jumping we will support it in the future.

## Value

A special class of navigation.

## Author(s)

Tengfei Yin
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