Package 'mimager'

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Title mimager: The Microarray Imager

Description Easily visualize and inspect microarrays for spatial

artifacts.

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LazyData TRUE

Depends Biobase

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URL https://github.com/aaronwolen/mimager

BugReports https://github.com/aaronwolen/mimager/issues

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Author Aaron Wolen [aut, cre, cph]

Maintainer Aaron Wolen <aaron@wolen.com>

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 $a rank \hspace{3.1in} Array \hspace{1.1in} rank$

Description

Determines the rank of values within each matrix of a three-dimensional array.

Usage

```
arank(x, na.last = TRUE, ties.method = "first")
```

Arguments

x a three-dimensional array of matrices

na.last for controlling the treatment of NAs. If TRUE, missing values in the data are put last; if FALSE, they are put first; if NA, they are removed; if "keep" they are kept with rank NA.

ties.method a character string specifying how ties are treated, see 'Details'; can be abbreviated.

Value

an array with the same dimensions as x

See Also

rank

Other array.transformations: arle

Examples

```
# microarray visualization
if (require(affydata, quietly = TRUE)) {
  data("Dilution", package = "affydata")
  x <- arank(marray(Dilution, transpose = TRUE))
}</pre>
```

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arle

Array relative log expression

Description

The relative log expression (RLE) quantifies the extent to which each sample in a dataset differs from a "reference" sample, which represents each probe's median value across all samples.

Usage

```
arle(x, log2 = TRUE, normalize = TRUE)
```

Arguments

```
    x a three-dimensional array of matrices
    log2 should values be $log_2$ transformed
    normalize should values be quantile normalized
```

Value

an array with the same dimensions as x

See Also

RLE

Other array.transformations: arank

Examples

```
# microarray visualization
if (require(affydata, quietly = TRUE)) {
  data("Dilution", package = "affydata")
  x <- arle(marray(Dilution, transpose = TRUE))
}</pre>
```

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marray Microarray array

Description

Convert S4 microarray data structures into a three-dimensional array of matrices, where each matrix corresponds to an individual sample's microarray with values arranged to reflect the physical position of the corresponding feature (i.e., probe) on the microarray surface.

Usage

```
marray(object, type = NULL, select = NULL, transpose = NULL)
## S4 method for signature 'AffyBatch'
marray(object, type = "pm", select = NULL,
    transpose = FALSE)
## S4 method for signature 'PLMset'
marray(object, type = "residuals", select = NULL,
    transpose = FALSE)
## S4 method for signature 'FeatureSet'
marray(object, type = "pm", select = NULL,
    transpose = FALSE)
## S4 method for signature 'oligoPLM'
marray(object, type = "residuals", select = NULL,
    transpose = FALSE)
```

Arguments

object	a valid Bioconductor microarray data structure
type	for microarray objects type refers to <i>probe type</i> ; for objects containing probelevel models (e.g., PLMsets) type refers to the <i>value type</i> (i.e, "residuals" or "weights"). See probe type section for more information.
select	a numeric, character or logical vector indicating samples to include
transpose	TRUE (the default), ensures the reconstructed microarrays are vertically oriented, as is typically expected. Set to FALSE to return an array in the orientation strictly specified by the platform coordinates

Value

three-dimensional array

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Probe types

For microarray data structures the type argument determines the *type* of probe that should be included. The following table provides a list of valid values for each supported microarray class:

```
AffyBatch
                       "all"
                               "pm"
ExpressionFeatureSet
                       "all"
                                      "mm"
                               "pm"
                                             "bg"
      GeneFeatureSet
                       "all"
                               "pm"
      ExonFeatureSet "all"
                                      "mm"
                               "pm"
                                             "bg"
       SnpFeatureSet "all"
                               "pm"
                                      "mm"
```

Examples

```
if (require(affydata, quietly = TRUE)) {
  data("Dilution", package = "affydata")
  dilution.array <- marray(Dilution, select = c("20A", "10A"))
}</pre>
```

mimage

Microarray image

Description

Visualize microarray probe intensities arranged by their physical location on the array. A false color image is produced for each sample in the microarray object and arranged in a grid.

Usage

```
mimage(object, type = NULL, select = NULL, colors = NULL,
  legend.label = NULL, nrow = NULL, ncol = NULL, fixed = FALSE,
  empty.rows = "fill", empty.thresh = 0.6, transform = NULL,
  trim = 0.01, fontsize = 12)

## S4 method for signature 'AffyBatch'
mimage(object, type = "pm", select = NULL,
  colors = NULL, legend.label = "Intensity", nrow = NULL, ncol = NULL,
  fixed = FALSE, empty.rows = "fill", empty.thresh = 0.6,
  transform = log2, trim = 0.01, fontsize = 12)

## S4 method for signature 'PLMset'
mimage(object, type = "residuals", select = NULL,
  colors = NULL, legend.label = type, nrow = NULL, ncol = NULL,
  fixed = FALSE, empty.rows = "fill", empty.thresh = 0.6,
  transform = identity, trim = 0.01, fontsize = 12)

## S4 method for signature 'FeatureSet'
```

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```
mimage(object, type = "pm", select = NULL,
    colors = NULL, legend.label = "Intensity", nrow = NULL, ncol = NULL,
    fixed = FALSE, empty.rows = "fill", empty.thresh = 0.6,
    transform = log2, trim = 0.01, fontsize = 12)

## S4 method for signature 'oligoPLM'
mimage(object, type = "residuals", select = NULL,
    colors = NULL, legend.label = type, nrow = NULL, ncol = NULL,
    fixed = FALSE, empty.rows = "fill", empty.thresh = 0.6,
    transform = identity, trim = 0.01, fontsize = 12)

## S4 method for signature 'array'
mimage(object, type = NULL, select = NULL,
    colors = NULL, legend.label = "Values", nrow = NULL, ncol = NULL,
    fixed = FALSE, empty.rows = "ignore", empty.thresh = 1,
    transform = identity, trim = 0, fontsize = 12)
```

Arguments

object	a valid Bioconduc	ctor microarray d	lata structure

type for microarray objects type refers to *probe type*; for objects containing probe-

level models (e.g., PLMsets) type refers to the *value type* (i.e, "residuals" or

"weights"). See probe type section for more information.

select a numeric, character or logical vector indicating samples to include

colors a vector of colors used to represent probe values

legend.label Legend label

nrow optional, number of rows in grid layout
ncol optional, number of columns in grid layout

fixed Force images to assume a fixed aspect ratio corresponding to their physical di-

mensions

empty.rows Should empty rows be filled with values from neighboring rows (the default,

"fill"), should they be dropped ("drop") entirely, or should they be left alone

("ignore")

empty.thresh what proportion of features must be missing from a row to consider that row

empty

transform a function to be applied to the values prior to visualizatio

trim a percentile (default = 0.02) or range or 2 values see **trimming** section for

details

fontsize font size for labels and legend

Value

invisibly a gtable matrix of grobs

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Trimming

By default, a 98% winsorization is performed prior to visualization, pulling in values outside of the 1st and 99th percentiles to their respective endpoints. This can be modified using the trim argument to provide either a new percentile, or a range of 2 values defining the min/max of the trimmed endpoints. Set trim = 0 to avoid trimming entirely.

Empty Rows

As a result of platform design, the presence unindexed probes or probe selection by the user (e.g., including only "pm" probes), the matrix-representation of a microarray can contain numerous rows comprised entirely (or mostly) of missing values, which may produce undesirable rasterization artifacts in the microarray image. To avoid this, empty rows are filled with values from a neighboring row. The threshold for what constitutes an empty row can be tweaked with the empty.thresh argument.

Probe types

For microarray data structures the type argument determines the *type* of probe that should be included. The following table provides a list of valid values for each supported microarray class:

```
AffyBatch
                        "all"
                                 "ma"
                                        "mm"
ExpressionFeatureSet
                        "all"
                                "pm"
                                        "mm"
      GeneFeatureSet
                        "all"
                                "pm"
                                               "bg"
                        "all"
                                "pm"
                                               "bg"
      ExonFeatureSet
                                        "mm"
                       "all"
       SnpFeatureSet
                                "pm"
                                        "mm"
```

Examples

```
# standard array visualization
mimage(iris3)

# microarray visualization
if (require(affydata, quietly = TRUE)) {
  data("Dilution", package = "affydata")
  mimage(Dilution, select = c("20A", "10A"))
}
```

mimager

mimager: The Microarray Imager

Description

mimager simplifies the creation of microarray images (sometimes called "pseudo-images") for the purpose of identifying problematic regional aberrations. Notable features include support for many of Bioconductor's core microarray data structures, providing compatibility with a variety of common microarray platforms, and the ability to visualize multiple microarrays simultaneously.

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Details

The following Bioconductor microarray data structures are currently supported:

- AffyBatch-class for Affymetrix GeneChip probe level data
- PLMset-class for probe-level linear models fitted to Affymetrix GeneChip probe level data
- FeatureSet-class for storing Expression/Exon/SNP data from a variety of oligonucleotide platforms
- oligoPLM-class for probe-level linear models fitted to any of the FeatureSet-like classes

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