Package ‘mogsa’

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Type Package
Title Multiple omics data integrative clustering and gene set analysis
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Author Chen Meng
Maintainer Chen Meng <mengchen18@gmail.com>
Description This package provide a method for doing gene set analysis
based on multiple omics data.
License GPL-2
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Multiple omics clustering and gene set analysis

Description

Modern "omics" technologies enable quantitative monitoring of the abundance of various biological molecules in a high-throughput manner, accumulating an unprecedented amount of quantitative information on a genomic scale. Gene set analysis is a particularly useful method in high throughput data analysis since it can summarize single gene level information into the biological informative gene set levels. This package provide a method do the gene set analysis based on multiple omics data that describing the same set of observations/samples.
Details

Package: mogsa
Type: Package
Version: 1.3.1
Date: 2016-01-19
License: GPL-2
Depends: methods

The main function in the package is "mogsa", see the function help manu for more details.

Author(s)

Chen Meng Maintainer: Chen Meng <chen.meng@tum.de>

References

Chen Meng, Dominic Helm, Martin Frejno, and Bernhard Kuster. moCluster: Identifying Joint Patterns Across Multiple Omics Data Sets. Journal of Proteome Research 2016.

Examples

# library(mogsa)
# loading gene expression data and supplementary data
data(NCI60_4array_supdata)
data(NCI60_4arrays)

# using a list of data.frame as input
mgsa1 <- mogsa(x = NCI60_4arrays, sup=NCI60_4array_supdata, nf=9,
               proc.row = "center_ssq1", w.data = "inertia", statis = TRUE)

# using moa as input
ana <- moa(NCI60_4arrays, proc.row = "center_ssq1", w.data = "inertia", statis = TRUE)
smoa <- sup.moa(ana, sup=NCI60_4array_supdata, nf=3)
mgsa2 <- mogsa(x = ana, sup=NCI60_4array_supdata, nf=9)
mgsa3 <- mogsa(x = ana, sup=smoa)

annotate.gs

Summary annotation information of a gene set

Description

Retrieve variables/features (genes) mapped to the annotated data sets in a gene set. Also returns the information about presence and absence of a feature for a specific data set.

Usage

annotate.gs(mgsa, gs)

Arguments

mgsa An object of class mogsa-class or moa.sup-class.
gs The name of a geneset
bootMbpca

Value

This function returns a data.frame. The first column shows the name of features. The last column is for the count of how many data sets has the corresponding features. Columns in the middle contains logical value indicating whether a feature is presented in a particular data set.

Author(s)

Chen Meng

See Also

see GIS

Examples

```r
# library(mogsa)
# loading gene expression data and supplementary data
data(NCI60_4array_supdata)
data(NCI60_4arrays)
mgsa <- mogsa(x = NCI60_4arrays, sup=NCI60_4array_supdata, nf=9,
               proc.row = "center_ssq!", w.data = "inertia", statis = TRUE)
allgs <- colnames(NCI60_4array_supdata[[1]])
annotate.gs(mgsa, allgs[1])
```

bootMbpca

*Bootstrap mbpca to estimate the coherence of different data sets*

Description

Bootstrap mbpca to estimate the coherence of different data sets and estimate the number of components should be included in an analysis.

Usage

```r
bootMbpca(moa, mc.cores = 1, B = 100, replace = TRUE,
           resample = c("sample", "gene", "total"),
           log = "y", ncomp = NULL, method = NULL,
           maxiter = 1000, svd.solver = c("svd", "fast.svd", "propack"), plot = TRUE)
```

Arguments

- **moa**: An object of `moa` returned by `mbpca`.
- **mc.cores**: Integer; number of cores used in bootstrap. This value is passed to function `mclapply`.
- **B**: Integer; number of bootstrap.
- **replace**: Logical; sampling with or without replacement.
- **resample**: Could be one of "sample", "gene" or "total". "sample" and "gene" means sample-wise and variable-wise resampling, repectively. "total" means total resampling.
- **log**: Could be "x", "y" or "xy" for plot log axis.
- **ncomp**: Passed to function `mbpca`. In most of cases, user do not need to specify this argument because it could be inferred from `moa`. 


method Passed to function \texttt{mbpca}. In most of cases, user do not need to specify this argument because it could be inferred from \texttt{moa}.

\texttt{maxiter} Passed to function \texttt{mbpca}. In most of cases, user do not need to specify this argument because it could be inferred from \texttt{moa}.

\texttt{svd.solver} Passed to function \texttt{mbpca}. In most of cases, user do not need to specify this argument because it could be inferred from \texttt{moa}.

\texttt{plot} Logical; whether the result should be plotted.

Details

update details.

Value

It returns a matrix, columns are eigenvalues for different components. Each rows is a bootstrap sample.

Author(s)

Chen Meng

Examples

\begin{verbatim}
# see examples in \code{\link{mbpca}}
\end{verbatim}

\texttt{bootMbpcaK} \hspace{1cm} \textit{An internal function called by \texttt{bootMbpca}.}

\textbf{Description}

An internal function called by \texttt{bootMbpca}.

\textbf{Usage}

\begin{verbatim}
bootMbpcaK(data, replace, B = 100, mc.cores = 1, resample = c("sample", "total", "gene"), ncomp, method, k, center = FALSE, scale = FALSE, option = "uniform", maxiter = 1000, svd.solver = c("svd", "fast.svd", "propack"))
\end{verbatim}

\textbf{Arguments}

\begin{itemize}
\item \texttt{data} A list of matrix to bootstrap.
\item \texttt{replace} A logical variable to indicate sampling with or without replacement.
\item \texttt{B} Integer; number of bootstrap.
\item \texttt{mc.cores} Integer; number of cores used in bootstrap. This value is passed to function \texttt{mclapply}.
\item \texttt{resample} Could be one of "sample", "gene" or "total". "sample" and "gene" means sample-wise and variable-wise resampling, repectively. "total" means total resampling.
\item \texttt{ncomp} passed to \texttt{mbpca}.
\item \texttt{method} passed to \texttt{mbpca}.
\end{itemize}
box.gs.feature

k passed to mbpca.
center passed to mbpca.
scale passed to mbpca.
option passed to mbpca.
maxiter passed to mbpca.
svd.solver passed to mbpca.

Value

A matrix of mbpca eigenvalues resulted from bootstrap samples

Author(s)

Chen Meng

See Also

bootMbpca

box.gs.feature  boxplot of gene set variables across all samples.

Description

boxplot to show the variables (e.g. gene expression) of a gene set across all samples.

Usage

box.gs.feature(x, gs, moa = NULL, col = 1, layout = NULL, plot = TRUE, obs.order = NULL, ...)

Arguments

x An object of class mgsa-class or moa.sup-class
gs Gene set want to be explored
moa An object of class moa. It is required if x is an object of class moa.sup-class
col The color code for samples
layout The layout control, see examples.
plot A logical indicates whether the result should be plotted. If FALSE, a list of expression matrix of the gene set genes is returned. Otherwise nothing returned.
obs.order Can be used to reorder the matrix, could be used when clustering result is available.
... The arguments passed to boxplot

Details

This is a convenient function used to explore the expression of a set of features/genes
Value

Do not return anything (plot=TRUE) or return a list of matrix (plot=FALSE) depends on plot argument.

Author(s)
Chen meng

Examples

```r
# library(mgsa)
# loading gene expression data and supplementary data
data(NCI60_4array_supdata)
data(NCI60_4arrays)
mgsa <- mgsa(x = NCI60_4arrays, sup=NCI60_4array_supdata, nf=9,
  proc.row = "center_ssq1", w.data = "inertia", statis = TRUE)

allgs <- colnames(NCI60_4array_supdata[[1]])
colcode <- as.factor(sapply(strsplit(colnames(NCI60_4arrays$agilent), split="\."), "[", 1))
a <- box.gs.feature(x=mgsa, gs=allgs[5], type=3, col=colcode, plot=FALSE)
box.gs.feature(x=mgsa, gs=allgs[5], type=3, col=colcode, plot=TRUE, layout=matrix(1:4, 2, 2))
```
Examples

```r
# library(mogsa)
data(NCI60_4array_supdata)
data(NCI60_4arrays)
# split gene set annotation into two sets.
sup1 <- lapply(NCI60_4array_supdata, function(x) x[, 1:10])
sup2 <- lapply(NCI60_4array_supdata, function(x) x[, -(1:10)])
# project two sets of annotation
mgsa1 <- mogsa(x = NCI60_4arrays, sup=sup1, nf=9,
                proc.row = "center_ssq1", w.data = "inertia", statis = TRUE)
mgsa2 <- mogsa(x = NCI60_4arrays, sup=sup2, nf=9,
                proc.row = "center_ssq1", w.data = "inertia", statis = TRUE)
# combine two independent mgsa sets
mgsa_comb <- combine(mgsa1, mgsa2)
dim(getmgsa(mgsa1, "score"))
dim(getmgsa(mgsa2, "score"))
dim(getmgsa(mgsa_comb, "score"))
```

### decompose.gs.group

**Data-wise or PC-wise decomposition of gene set scores for all observations.**

**Description**

Data-wise or PC-wise decomposition of gene set scores (GSS) across all observations. The predefined group/cluster information should be given so that the mean decomposed GSSs for each group are returned and plotted.

**Usage**

```r
decompose.gs.group(x, gs, group, decomp = "data", nf = 2, x.legend = "bottomleft",
y.legend = NULL, plot = TRUE, main = NULL, ...)
```

**Arguments**

- `x`: An object of class `mgsa-class` or `moa.sup-class`
- `gs`: The gene set want to exam.
- `group`: An vector or factor to indicate the group of observations, such as clusters. See examples.
- `decomp`: A character string either "data" or "pc" to indicate how the gene set scores should be decomposed (with respect to data or PC).
- `nf`: The number of axes/PCs to be calculated and plotted.
- `x.legend`: Used to control the position of legends.
- `y.legend`: Used to control the position of legends.
- `plot`: A logical indicates if a plot should be drawn.
- `main`: The main title of plot.
- `...`: Other arguments passed to `barplot`.
Details

This function could be used when the number of observation is large and there are cluster/group information is available. In this case, the means of decomposed gene set scores over each group is calculated. The vertical bar on the end of each bar indicates the 95% confident interval of the means.

Value

Return nothing or a matrix depends on how argument plot is set.

Author(s)

Chen Meng

References

TBA

See Also

See Also `decompose.gs.ind`

Examples

```r
# library(mogsa)
# loading gene expression data and supplementary data
data(NCI60_4array_supdata)
data(NCI60_4arrays)

# using a list of data.frame as input
mgsa <- mogsa(x = NCI60_4arrays, sup=NCI60_4array_supdata, nf=9,
              proc.row = "center_ssq!", w.data = "inertia", statis = TRUE)
colcode <- as.factor(sapply(strsplit(colnames(NCI60_4arrays$agilent), split="\."), "[", 1))
decompose.gs.group(x = mgsa, gs = 2, group = colcode, decomp = "data", plot = TRUE)
decompose.gs.group(x = mgsa, gs = 2, group = colcode, decomp = "pc", nf = 3, plot = TRUE)
```
Arguments

- **x**: An object of class `mgsa-class` or `moa.sup-class`
- **gs**: The gene set want to exam.
- **obs**: The observations want to exam.
- **type**: Which type of plot. `type=1` - the data-pc mode; `type=2` - the pc-data mode; `type=3` - both. See detail.
- **nf**: The number of axes/PCs to be calculated and plotted.
- **plot**: A logical indicates if a plot should be drawn
- **col.data**: The bar color of datasets
- **col.pc**: The bar color of PCs
- **legend**: A logical if legend should be shown

Details

type=1 (the data-pc mode), the axes/PCs are represented as the narrow bars with different colors and the background wide bars behind narrow bars are gene set scores for datasets, which is calculated from the sum of all underlying individual axes/PC scores. When type=2 (the pc-data mode) the interpretation of narrow and wide bars are in the other way around. If type=3, both are shown.

This function could only be used to check the decomposition of gene set scores of a single observation. So the function is not efficient when the number of observation is large. Another function `decompose.gs.group`, could be used in this case, particularly when the cluster information of the observation panel is available.

Value

Return nothing or a matrix depends on how argument `plot` is set.

Author(s)

Chen Meng

References

TBA

See Also

See Also as `decompose.gs.group`

Examples

```r
# library(mogsa)
# loading gene expression data and supplementary data
data(NCI60_4array_supdata)
data(NCI60_4arrays)
mgsa <- mogsa(x = NCI60_4arrays, sup=NCI60_4array_supdata, nf=9,
              proc.row = "center_ssq!", w.data = "inertia", statis = TRUE)
allgs <- colnames(NCI60_4array_supdata[[1]])
# plot
decompose.gs.ind(x=mgsa, gs=allgs[5], obs="BR.MDA_MB_231", type=2, nf=5)
# or
decompose.gs.ind(x=getmgsa(mgsa, "sup"), gs=allgs[5], obs="BR.MDA_MB_231", type=3, nf=5)
```
deflat

deflat function used by mbpca

Description
An internal function called by mbpca.

Usage

deflat(x, t, tb, pb, method = "globalScore")

Arguments

x
A list of matrix want to deflat

t
The global scores returned by msvd or nipalsSoftK

tb
The block scores returned by msvd or nipalsSoftK

pb
The block loadings returned by msvd or nipalsSoftK

method
A charater to specify the deflation strateg, could be one of c("globalScore", "blockLoading", "blockScore").

Value
A list of deflated matrix

Author(s)
Chen Meng

distMoa
Calculate the distance matrix from an object of class moa-class.

Description
A convenient function to calculate the distance matrix from an object of class moa-class.

Usage
distMoa(x, nf = NA, tol = 1e-05, method = "euclidean", diag = FALSE, upper = FALSE, p = 2)

Arguments

x
An object of class moa-class.

nf
Integer; the number of component used to calculate the distance. Default setting (NA) will keep all the axes.

tol
Numerical; the tolerance of component with low variance.

method
passed to function dist
diag
passed to function dist
upper
passed to function dist
p
passed to function dist
getmgsa

Value

An object of class dist, see function "dist".

Author(s)

Chen Meng

Examples

# see examples in \code{\link{mbpca}}

data("NCI60_4arrays")
moa <- mbpca(NCI60_4arrays, ncomp = 10, k = "all", method = "globalScore", option = "lambda1",
center=TRUE, scale=FALSE)
dst <- distMoa(moa)

getmgsa(mgsa, value)

Arguments

mgsa An object of class mgsa-class.
value The name of the value want to extract from "mgsa". See detail for options.

Details

if value in c("call", "moa", "sup"), the function equal function slot.
if value in c("eig", "tau", "partial.eig", "eig.vec", "loading", "fac.scr", "partial.fs", "ctr.obs", "ctr.var", "ctr.tab", "RV"), the function extract corresponding value from moa-class.
if value in c("data", "coord.sep", "coord.comb", "score", "score.data", "score.pc", "score.sep", "p.val"), the function extract value from moa.sup-class.

Value

The function return the selected value in "mgsa".

Author(s)

Chen Meng
Examples

```r
# library(mogsa)
data(NCI60_4array_supdata)
data(NCI60_4arrays)
mgas <- mogsa(x = NCI60_4arrays, sup = NCI60_4array_supdata, nf = 9,
               proc.row = "center_ssq1", w.data = "inertia", statis = TRUE)
part.eig <- getmgsa(mgas, "partial.eig")
barplot(as.matrix(part.eig))
```

GIS (abbreviation: GIS)

calculate gene influential scores of genes in a gene set.

Description

Calculate the gene influential score of individual feature to the overall variance of GS score. Using a leave-one-out procedure (See detail).

Usage

```r
GIS(x, geneSet, nf = NA, barcol = NA, topN = NA, plot = TRUE, Fvalue = FALSE, ff = NA, cor = FALSE)
```

Arguments

- `x`: An object of class `mgsa-class`.
- `geneSet`: A character string or number to indicated the gene sets under conserderation.
- `nf`: The number of PCs used in the calculation of gene set scores. The default is NA, which means using all the PCs in the mogsa. This should work for most of the cases.
- `barcol`: The color of the bars, which is used to distinguish features/genes from different datasets, so its length should be the same as the number of data sets.
- `topN`: An positive integer specify the number of top influencers that should to returned.
- `plot`: A logical indicate if the result should be plotted.
- `Fvalue`: A logical indicate if the GIS should be calculated in a supervised manner.
- `ff`: The vector indicates the group of columns for calculating the F-ratio when Fvalue=TRUE.
- `cor`: A logical indicates whether use correlation between reconstructed expression with GSS. This is faster than the standard GIS.
Details

The evaluation of the importance of a single feature is calculated in the supervised or unsupervised manner.

In the unsupervised manner, the value is calculated by:

\[
\log_2(\frac{\text{var}(\text{GS}_-i)}{\text{var}(\text{GS})})
\]

where GS is the gene set score, and the GS_-i is a recalculate of gene set score without i’th feature.

var() is the variance.

In the supervised manner, the value is calculated as the F-ratio over a class vector:

\[
\log_2(\frac{\text{F}(\text{GS}_-i)}{\text{F}(\text{GS})})
\]

Where F() is the calculation of F-ratio. The unsupervised GIS is encouraged since it works better for most of the cases in practice.

Value

An object of class data.frame contains three columns. The first column is the feature name, the second columns is the gene influential score. The third columns indicates from where the feature/gene is selected.

Author(s)

Chen Meng

References

TBA

See Also

see annotate.gs

Examples

```r
# library(mogsa)
# loading gene expression data and supplementary data
data(NCI60_4array_supdata)
data(NCI60_4arrays)
mgsa <- mogsa(x = NCI60_4arrays, sup=NCI60_4array_supdata, nf=9,
              proc.row = "center_ssq1", w.data = "inertia", statis = TRUE)
algs <- colnames(NCI60_4array_supdata[[1]])

# unsupervised measurement
GIS(mgsa, algs[1], topN = 5)

# supervised measurement

tissueType <- as.factor(sapply(strsplit(colnames(NCI60_4arrays$agilent), split="\."), ",", 1))
GIS(mgsa, algs[1], topN = 5, Fvalue = TRUE, ff = tissueType)

# more PCs to calculate
GIS(mgsa, algs[1], nf = 20, topN = 5, Fvalue = TRUE, ff = tissueType)
```
**matpower**

*compute the power of a matrix*

**Description**

the power of a matrix

**Usage**

```r
matpower(x, n, nf = min(dim(x)), tol = 1e-07)
```

**Arguments**

- `x`: a numerical matrix object that the power of which should be calculated
- `n`: The matrix to the power of
- `nf`: The number of axes kept in the calculation of SVD and reconstruction
- `tol`: The tolerance of the axis, singular vectors with singular value lower than tol will be ignored in the reconstruction.

**Details**

The power of a matrix is calculated in two steps: decomposition step: \( x = UDV' \) and the reconstruction step: \( x^n = U D^n V' \). In the reconstruction, the singular vectors with a singular value more than tol are kept.

**Value**

A matrix \( x^n \)

**Note**

Called by the wsvd function.

**Author(s)**

Chen Meng

**See Also**

See Also `wsvd`

**Examples**

```r
cat("set.seed(56)\n")
m <- matrix(rnorm(15), 5, 3)
s <- matpower(m, 2)
s <- matpower(m, -2)
```
Extension of PCA to analyze multiple data sets

Description

Three approaches are supplied in this function, consensus PCA (CPCA), generalized CCA (GCCA) and multiple co-inertia analysis (MCIA).

Usage

mbpca(x, ncomp, method, k = "all", center = TRUE, scale = FALSE, option = "uniform", maxiter = 1000, moa = TRUE, verbose = TRUE, svd.solver = c("svd", "fast.svd", "propack"))

Arguments

x A list of matrix or data.frame, where rows are variables and columns are samples. The columns among the matrices need to be match but the variables do not need to be.
ncomp An integer; the number of components to calculate. To calculate more components requires longer computational time.
method A character string could be one of c("globalScore", "blockScore", "blockLoading"). The "globalScore" approach equals consensus PCA; The "blockScore" approach equals generalized canonical correlation analysis (GCCA); The "blockLoading" approach equals multiple co-inertia analysis (MCIA);
k The absolute number (if k >= 1) or the proportion (if 0<k<1) of non-zero coefficients for the variable loading vectors. It could be a single value or a vector has the same length as x so the sparsity of individual matrix could be different.
center Logical; if the variables should be centered
scale Logical; if the variables should be scaled
option A character string could be one of c("lambda1", "inertia", "uniform") to indicate how the different matrices should be normalized. If "lambda1", the matrix is divided by its the first singular value, if "inertia", the matrix is divided by its total inertia (sum of square), if "uniform", none of them would be done.
maxiter Integer; Maximum number of iterations in the algorithm
moa Logical; whether the output should be converted to an object of class moa-class
verbose Logical; whether the process (# of PC) should be printed
svd.solver A character string could be one of c("svd", "fast.svd", "propack"). The default "fast.svd" has a good compromise between the robustness and speed. "propack" is the fastest but may failed to converge in practice.

Details
details need to update
Value

An object of class `moa-class` (if `moa=TRUE`) or an list object contains the following elements:
- `tb` - the block scores
- `pb` - the block loadings
- `t` - the global scores
- `w` - the weights of block scores to construct the global score

Note

no note now

Author(s)

Chen Meng

References

reference need to be updated

See Also

see `moa` for non-iterative algorithms for multi-block PCA.

Examples

data("NCI60_4arrays")
tumorType <- sapply(strsplit(colnames(NCI60_4arrays$agilent), split="\."), "[", 1)
colcode <- as.factor(tumorType)
levels(colcode) <- c("red", "green", "blue", "cyan", "orange",
                   "gray25", "brown", "gray75", "pink")
colcode <- as.character(colcode)

moa <- mbpca(NCI60_4arrays, ncomp = 10, k = "all", method = "globalScore", option = "lambda1",
center=TRUE, scale=FALSE)
plot(moa, value="eig", type=2)
r <- bootMbpca(moa, mc.cores = 1, B=6, replace = FALSE, resample = "sample")
moas <- mbpca(NCI60_4arrays, ncomp = 3, k = 0.1, method = "globalScore", option = "lambda1",
center=TRUE, scale=FALSE)

scr <- moaScore(moa)
scrs <- moaScore(moas)
diag(cor(scr[, 1:3], scrs))

layout(matrix(1:2, 1, 2))
plot(scrs[, 1:2], col=colcode, pch=20)
legend("topright", legend = unique(tumorType), col=unique(colcode), pch=20)
plot(scrs[, 2:3], col=colcode, pch=20)

gap <- moGap(moas, K.max = 12, cluster = "hcl")
```r
gap$nClust

hcl <- hclust(dist(scrs))
cis <- cutree(hcl, k=4)
cisColor <- as.factor(cis)
levels(cisColor) <- c("red", "blue", "orange", "pink")
cisColor <- as.character(cisColor)

heatmap(t(scrs[hcl$order, ]), ColSideColors = clsColor[hcl$order], Rowv = NA, Colv=NA)
heatmap(t(scrs[hcl$order, ]), ColSideColors = clsColor[hcl$order], Rowv = NA, Colv=NA)

genes <- moaCoef(moas)
genes$nonZeroCoef$agilent.V1.neg
```

---

### mgsa-class  
_CLASS "mgsa"

**Description**

mgsa class here.

**Objects from the Class**

Objects can be created by calls of the form `new("mgsa", ...)`.  

**Slots**

- `call`: call
- `moa`: Object of class moa
- `sup`: Object of class moa.sup

**Methods**

- `combine` _signature_(x = "mgsa", y = "mgsa") To combine two objects of class "mgsa". This function could only be used to combine two "mgsa" objects, using "Reduce" function to combine more.

- `show` _signature_(x = "moa", y = "missing"): show the "mgsa" result.

**Author(s)**

Chen Meng

**See Also**

moa and moa.sup
Examples

```r
showClass("mgsa")
# library(mgsa)
# loading gene expression data and supplementary data
data(NCI60_4array_supdata)
data(NCI60_4arrays)
# split gene set annotation into two sets.
sup1 <- lapply(NCI60_4array_supdata, function(x) x[, 1:10])
sup2 <- lapply(NCI60_4array_supdata, function(x) x[, -(1:10)])
# project two sets of annotation
mgsa1 <- mogsa(x = NCI60_4arrays, sup=sup1, nf=9,
                proc.row = "center_ssq1", w.data = "inertia", statis = TRUE)
mgsa2 <- mogsa(x = NCI60_4arrays, sup=sup2, nf=9,
                proc.row = "center_ssq1", w.data = "inertia", statis = TRUE)
# combine two independent mgsa sets
mgsa_comb <- combine(mgsa1, mgsa2)
dim(getmgsa(mgsa1, "fac.scr"))
dim(getmgsa(mgsa2, "fac.scr"))
dim(getmgsa(mgsa_comb, "fac.scr"))
```

---

**moa**

*Multiple omics data analysis using MFA or STATIS*

**Description**

Analysis multiple omics data using MFA or STATIS. The input multiple tables are in a form that columns are samples and rows are variables/features.

**Usage**

```r
moa(data, proc.row="center_ssq1", w.data="inertia", w.row=NULL, statis=FALSE)
```

**Arguments**

- `data` A list of data.frame or matrix that contains the input datas, the columns in all datasets should be samples/observations (which need to be matched) and rows should be variables.

- `proc.row` Preprocessing of rows of datasets, should be one of `none` - no preprocessing, `center` - center only, `center_ssq1` - center and scale (sum of squared values equals 1), `center_ssqN` - center and scale (sum of squared values equals the number of columns), `center_ssqNm1` - center and scale (sum of squared values equals the number of columns - 1) MFA corresponds to "proc.row=center_ssq1" and "w.data="lambda1"

- `w.data` The weights of each separate dataset, should be one of `uniform` - no weighting, `lambda1` - weighted by the reverse of the first eigenvalue of each individual dataset or `inertia` - weighted by the reverse of the total inertia. See detail.

- `w.row` If it is not null, it should be a list of positive numerical vectors, the length of which should be the same with the number of rows of each dataset to indicated the weight of rows of datasets.

- `statis` A logical indicates whether STATIS method should be used. See details.
Details

Different methods employs different precessing of row and datasets. For multipple factorial analysis (MFA), the rows of each dataset are first centered and scaled, then each dataset is weighted by the reverse of its first eigenvalue (proc.row=center_ssq1, w.data="lambda1"). This algorithm does not have a well defined criterion to be optimized (see reference).

If statis=TRUE, the statis algorithm will be used, that is, each dataset will be further weighted so that datasets closer to the overall structure will receive a higher weight.

Value

An object of class moa-class.

Author(s)

Chen Meng

References


See Also

sup.moa, mogsa. More about plot see moa-class.

Examples

```r
# library(mogsa)
# loading data
data(NCI60_4arrays)
# run analysis
ana <- moa(NCI60_4arrays, proc.row = "center_ssq1", w.data = "inertia", statis = TRUE)
# plot
# plot eigen value
plot(ana, value = "eig", type = 2)
# plot the normalized (percentage) eigen value
plot(ana, value = "tau", type = 2)
# ploting the observations
colcode <- as.factor(sapply(strsplit(colnames(NCI60_4arrays$agilent), split="\."), "]", 1))
plot(ana, type = 1, value = "obs", col=colcode)
plot(ana, type = 2, value = "obs", col=colcode, data.pch=1:4)
# plot variables/features in each data sets
plot(ana, value = "var", layout=matrix(1:4, 2, 2))
# plot the RV coefficients for the data sets
plot(ana, value = "RV")
```
moa-class

Class "moa"

Description

moa class object

Objects from the Class

Objects can be created by calls of the form `new("moa", ...)`. 

Slots

eig: eigen values

tau: The percentage of explained variance by each datasets sparately.

partial.eig: matrix, rows indicate the partial eigenvalues from each data.

eig.vec: a matrix, eigenvectors.

loading: the coordinate of variables/features.

fac.scr: factor score of observations.

partial.fs: partial factor score.

ctr.obs: contribution of each observation to the total factor score.

ctr.var: contribution of each variables to the total variance.

ctr.tab: contribution of each data to the total variance.

RV: pairwise RV coefficients

w.row: weight of rows

w.data: weight of datasets

data: the original input data

tab.dim: the dimension of each input data

call: call

Methods

plot signature(x = "moa", y = "missing"): Argument "value" sould be one of "eig", "tau", "obs", "var" and "RV"

if value = "eig", the eigenvalue would be plotted as scree plot. The following arguments could be set:

type=1 - The type of plot to show eigenvalues. (type=1: the eigenvalue are plotted; type=2: partial eigenvalue shown as concatenated bars; type=3: partial eigenvalue shown as bars side by side; type=4: matplot view of eigenvales, lty need to be set; type=5; the two dimensional plot of partial eigenvalues, axes and pch need to be set in this case.)

axes=NULL - The axes selected to plot

n=NULL - Top n eigenvalues to be drawn

tol=1e-5 - The tolerance of eigenvalue, eigenvalues lower than this value will not be shown.

legend=NULL - legend to put, a character string as calling legend function

col=NULL - The color of partial eigenvalues from each data set
moa-class

- The line type used in the matplot, used when type = 4
- The pch to draw 2D partial eigen plot, when type = 5 used
- The position of legend
- Position argument passed to function "legend"
- Other arguments passed to functions
  - if value = "tau", the same with eig, but in the eigenvalues are scaled to 1
  - if value = "obs", the observation space will be shown, the following argument could be set:
    - Which axes should be draw
    - Which type, see below (for type=1: the center points draw; type=2: the separate factor scores linked by lines; ... will be passed to function "points")
    - the pch of dataset, if type=1, the first one is used
    - the color of observations, recycled used by data.frame
    - A logical indicates if labels should be shown
    - Position of legend
    - Position of legend
    - The x limit
    - The y limit
    - the cex of text
    - the separate gene view, layout can be specified
    - the heatmap of RV coefficients

  show signature(x = "moa", y = "missing"): show "moa" object

Author(s)

Chen Meng

References


Herve Abdi, Lynne J. Williams, Domininique Valentin. Multiple factor analysis: principal component analysis for multitable and multiblock data sets. WIREs Comput Stat 2013

Examples

```r
showClass("moa")
# load("R/mogs/data/NCI60_4arrays.rda")
data(NCI60_4arrays)
a <- moa(NCI60_4arrays, proc.row = "center_ssq!", w.data = "inertia", statis = TRUE)
plot(a, value="eig")
plot(a, value="tau", type=2)
```
### Description

moa.sup class desc.

### Objects from the Class

Objects can be created by calls of the form `new("moa.sup", ...).

### Slots

- **sup**: Object of class "list", the matrix of supplementary data.
- **coord.sep**: The projection of geneset information on each separate data.
- **coord.comb**: The projection of geneset information on total dataset.
- **score**: the gene set-sample pathway score
- **score.data**: the gene set-sample pathway score, data separate
- **score.pc**: the gene set-sample pathway score, PC separate
- **score.sep**: the gene set-sample pathway score, separate.
- **p.val**: the p value matrix have the same dimension with score matrix.

### Methods

There is no generic function for objects of "moa.sup", but have specific function, including: - decompose.gs.ind - box.gs.feature - plotGS - decompose.gs.group

### Author(s)

Chen Meng

### See Also

objects to See Also as `decompose.gs.ind, box.gs.feature, plotGS, decompose.gs.group`

### Examples

```r
showClass("moa.sup")
data(NCI60_4array_supdata)
data(NCI60_4arrays)
sapply(NCI60_4array_supdata, dim)
ana <- moa(NCI60_4arrays, proc.row = "center_ssq1", w.data = "inertia", statis = TRUE)
plot(ana, value="eig")
smoa <- sup.moa(ana, sup=NCI60_4array_supdata, nf=5)
```
moaCoef

Extract the loadings/coefficients from an object of class \texttt{moa-class}.

**Description**

Extract the loadings/coefficients from an object of class \texttt{moa-class}.

**Usage**

\texttt{moaCoef(moa)}

**Arguments**

\texttt{moa} An object of class \texttt{moa-class}.

**Value**

It returns a list consist of two components:

- \texttt{coefMat} - the loading matrix
- \texttt{nonZeroCoef} - it is a list of \texttt{data.frame} to list the non-zero coefficient variable in each of loading vectors and data sets. The element names are in a format as "xxxx.yy.zzz"
  
  \texttt{xxxx} - are the data names, tells the data set where a variable is from
  
  \texttt{yy} - the number of Axes, for example, "V1" indicate the variable has a non-zero coefficient in the first loading vector.
  
  \texttt{zzz} - could be either "pos" (coefficient \textgreater{} 0) or "neg" (coefficient \textless{} 0)

The \texttt{data.frame} has two columns, the first column is the ID of a variable the second column is the coefficient/loading.

**Author(s)**

Chen Meng

**See Also**

\texttt{moaScore}

**Examples**

# see examples in \code{\link{mbpca}}

data("NCI60_4arrays")
moa <- mbpca(NCI60_4arrays, ncomp = 10, k = "all", method = "globalScore", option = "lambda1",
            center=TRUE, scale=FALSE)

genes <- moaCoef(moa)
scr <- moaScore(moa)
**moaScore**

Extract global scores from an object of class `moa-class`.

**Description**

Extract global scores from an object of class `moa-class`.

**Usage**

```r
moaScore(moa)
```

**Arguments**

- `moa`: An object of class `moa-class`

**Value**

A matrix of global score

**Author(s)**

Chen Meng

**See Also**

`moaCoef`

**Examples**

```r
# see examples in \
\code{\link{mbpca}}

data("NCI60_4arrays")
moa <- mbpca(NCI60_4arrays, ncomp = 10, k = "all", method = "globalScore", option = "lambda1",
            center=TRUE, scale=FALSE)
genes <- moaCoef(moa)
scr <- moaScore(moa)
```

---

**moGap**

Gap statistic for clustering latent variables in `moa-class`.

**Description**

Gap statistic is a measurement of goodness of clustering result. This is a convenient function to calculate the gap statistic of clustering "moa".
Usage

moGap(x, K.max, B = 100, cluster = c("kmeans", "hclust"), plot = TRUE,
      dist.method = "euclidean", dist.diag = FALSE, dist.upper = FALSE, dist.p = 2,
      hcl.method = "complete", hcl.members = NULL,
      km.iter.max = 10, km.nstart = 10,
      km.algorithm = c("Hartigan-Wong", "Lloyd", "Forgy", "MacQueen"), km.trace = FALSE)

Arguments

x       An object of class moa-class returned by mbpca.
K.max   The maximum number of clusters to consider, passed to clusGap
B       The number of bootstrap, passed to clusGap
cluster A character string could be either "kmeans" or "hclust" to specify the clustering
         algorithm.
plot    Logical; whether return the gap statistic plot.
dist.method Distance measurement, passed to function "dist".
dist.diag Passed to function "dist".
dist.upper Passed to function "dist".
dist.p   Passed to function "dist".
hcl.method Hierarchical clustering method, passed to "hclust"
hcl.members Passed to "hclust"
k.m.iter.max Maximum number of iteration in kmeans, passed to "kmeans".
k.m.nstart An integer to specify how many random sets should be chosen. passed to
            "kmeans".
k.m.algorithm Kmeans algorithm, passed to "kmeans".
k.m.trace  See function "kmeans".

Value

It returns a list consists of five components:
"Tab", "n", "B", "FUNcluster" - see clusGap
"nClust" - the estimated number of clusters using different method, see maxSE

Author(s)

Chen Meng

References

Tibshirani, R., Walther, G. and Hastie, T. (2001). Estimating the number of data clusters via the


See Also

Function "clusGap" in "cluster" package Function "dist", "hclust", "kmeans"
Examples

```r
# see examples in \code{\link{mbpca}}

data("NCI60_4arrays")
moa <- mbpca(NCI60_4arrays, ncomp = 10, k = "all", method = "globalScore", option = "lambda1",
            center=TRUE, scale=FALSE)
gap <- moGap(moa, K.max = 12, cluster = "hcl")

genes <- moaCoef(moa)
scr <- moaScore(moa)
```

mogsa

multiple omics data integration and gene set analysis

Description

The main function called by users, omics data analysis and gene set annotation. A wrapper function of `moa` and `sup.moa`.

Usage

```r
mogsa(x, sup, nf=NULL, proc.row=NULL, w.data=NULL, w.row=NULL, statis=FALSE, ks.stat=FALSE, ks.B = 1000, ks.cores = NULL)
```

Arguments

- `x`: An object of class `list` or `moa-class`. A list would be a list of data frame.
- `sup`: An object of class `list` or `moa.sup-class`. A list would be a list of supplementary data.
- `nf`: The number of principal components used to reconstruct, only used when `x` is an object of list.
- `proc.row`: Preprocessing of rows. If `x` is a object of list, it is passed `moa`.
- `w.data`: Weights of datasets. If `x` is a object of list, it is passed `moa`.
- `w.row`: Weight of row. If `x` is a object of list, it is passed `moa`.
- `statis`: A logical indicates if statis algrithm should be used. If `x` is a object of list, it is passed `moa`.
- `ks.stat`: The logical indicates if the p-value should be calculated using K-S statistic (the method used in "ssgsea" in GSVA package). Default is FALSE, which means using the z-score method. See `sup.moa`.
- `ks.B`: An integer to indicate the number of bootstrapping samples to calculated the p-value of KS statistic.
- `ks.cores`: An integer indicate the number of cores to be used in bootstrapping. It is passed to function `mclapply` in the parallel package.

Details

A wrapper function of `moa` and `sup.moa`.
msvd

Value

An object of class mgsa-class.

Note

This function will be changed to a generic function for "S4-style" programming.

Author(s)

Chen Meng

References


See Also

moa and sup.moa

Examples

# library(mogsa)
# loading gene expression data and supplementary data
data(NCI60_4array_supdata)
data(NCI60_4arrays)

# using a list of data.frame as input
mgsa1 <- mogsa(x = NCI60_4arrays, sup=NCI60_4array_supdata, nf=9,
proc.row = "center_ssq1", w.data = "inertia", statis = TRUE)
# using moa as input
ana <- moa(NCI60_4arrays, proc.row = "center_ssq1", w.data = "inertia", statis = TRUE)
smoa <- sup.moa(ana, sup=NCI60_4array_supdata, nf=3)
mgsa2 <- mogsa(x = ana, sup=NCI60_4array_supdata, nf=3)
mgsa3 <- mogsa(x = ana, sup=smoa)

msvd

SVD based algorithm to calculate block Score and global scores for mbpca.

Description

An internal function called by mbpca. It returns the result comparable with nipalsSoftK, but way faster since it uses the SVD algorithm. No sparse opertors in this function.

Usage

msvd(x, svd.sol = svd)
NCI60_4arrays

Arguments

- **x**
  - The input matrix, rows are observations, columns are variables
- **svd.sol**
  - A function object to specify the preferred SVD solver, default is svd.

Value

- an list object contains the following elements:
  - **tb** - the block scores
  - **pb** - the block loadings
  - **t** - the global scores
  - **w** - the weights of block scores to construct the global score

Author(s)

Chen Meng

See Also

nipalsSoftK

NCI60_4arrays

Microarray gene expression profiles of the NCI 60 cell lines from 4 different platforms

Description

The 60 human tumour cell lines are derived from patients with leukaemia, melanoma, lung, colon, central nervous system, ovarian, renal, breast and prostate cancers. The cell line panel is widely used in anti-cancer drug screen. In this dataset, a subset of microarray gene expression of the NCI 60 cell lines from four different platforms are combined in a list, which could be used as input to mcia directly.

Usage

data(NCI60_4arrays)

Format

The format is: List of 4 data.frames

- **\$agilent**: data.frame containing 300 rows and 60 columns. 300 gene expression log ratio measurements of the NCI60 cell lines, by Agilent platform.
- **\$hgu133**: data.frame containing 298 rows and 60 columns. 298 gene expression log ratio measurements of the NCI60 cell lines, by H-GU133 platform.
- **\$hgu133p2**: data.frame containing 268 rows and 60 columns. 268 gene expression log ratio measurements of the NCI60 cell lines, by H-GU133 plus 2.0 platform.
- **\$hgu95**: data.frame containing 288 rows and 60 columns. 288 gene expression log ratio measurements of the NCI60 cell lines, by H-GU95 platform.
**Value**

`NCI60_4arrays` will be loaded in your working space.

**Source**


**References**


---

**NCI60_4array_supdata**

Supp data for Microarray gene expression profiles of the NCI 60 cell lines from 4 different platforms

**Description**

Supplementary to `NCI60_4arrays`.

**Usage**

data(`NCI60_4arrays`)

**Format**

The format is: List of 4 matrix

- \$agilent:matrix containing 300 rows and 60 columns. 300 gene expression log ratio measurements of the NCI60 cell lines, by Agilent platform.
- \$hgu133:matrix containing 298 rows and 60 columns. 298 gene expression log ratio measurements of the NCI60 cell lines, by H-GU133 platform.
- \$hgu133p2:matrix containing 268 rows and 60 columns. 268 gene expression log ratio measurements of the NCI60 cell lines, by H-GU133 plus 2.0 platform.
- \$hgu95:matrix containing 288 rows and 60 columns. 288 gene expression log ratio measurements of the NCI60 cell lines, by H-GU95 platform.

**Value**

`NCI60_4array_supdata` will be loaded in your working space.
nipalsSoftK

NIPALS algorithm with soft thresholding operator

Description

An internal function called by mbpca.

Usage

nipalsSoftK(x, maxiter, k)

Arguments

- x: The input matrix, rows are observations, columns are variables
- maxiter: # of maximum iteration the algorithm can run
- k: The number (>=1) or proportion (<1) of variables want to keep. It could be a single value or a vector has the same length as x so the sparsity of individual matrix could be different.

Value

An list object contains the following elements:
- tb: the block scores
- pb: the block loadings
- t: the global scores
- w: the weights of block scores to construct the global score.

Author(s)

Chen Meng

See Also

msvd

pairwise.rv

pairwise RV coefficients.

Description

Calculating pairwise RV coefficients for a list of matrices or data.frame.

Usage

pairwise.rv(data.list, match="col")
Arguments

- **data.list**: A list of data.frame or matrix, either rows or columns in each data set should be matched.
- **match**: Whether columns or rows of data.frame/matrix should be matched.

Details

The RV coefficient for each pair of matrices is calculated as \( R_v = \frac{\text{trace}(XX'YY')}{\sqrt{\text{trace}(XX'XX')\text{trace}(YY'YY')}} \)

Value

The function will return a matrix containing the pairwise RV coefficients.

Note

The variable in matrices are not automatically centered or scaled in this function. So these step may need to be performed before calling this function.

Author(s)

Chen Meng

References


Examples

```r
data(NCI60_4arrays)
pairwise.rv(NCI60_4arrays)
```

---

**plot-methods**

Methods for function `plot`

**Description**

Methods for function `plot`

**Methods**

```r
signature(x = "moa", y = "missing") plot "moa" object
```

Argument "value" sould be one of "eig", "tau", "obs", "var" and "RV"

if value = "eig", the eigenvalue would be plotted as scree plot. The following arguments could be set:

- **type=1** - The type of plot to show eigenvalues. (type=1: the eigenvalue are plotted; type=2: partial eigenvalue shown as concatenated bars; type=3: partial eigenvalue shown as bars side by side; type=4: matplot view of eigenvales, lty need to be set; type=5: the two dimensional plot of partial eigenvalues, axes and pch need to be set in this case.)
- **axes=NULL** - The axes selected to plot
- **n=NULL** - Top n eigenvalues to be drawn
- **tol=1e-5** - The tolerance of eigenvalue, eigenvalues lower than this value will not be shown.
- **legend=NULL** - legend to
put, a character string as calling legend function \col=NULL - The color of partial eigenvalues from each data set \lty=1 - The line type used in the matplot, used when type =4 \pch=NULL - the pch to draw 2D partial eigen plot, when type = 5 used \lg.x="topright" - The position of legend \lg.y=NULL - Poistion argument passed to function "legend"  \... - other arguemnts passed to functions \ 
if value = "tau", the same with eig, but in the eigenvalues are scaled to 1 \if value = "obs", the observation space will be shown, the following argument could be set:\axes=1:2 - Which axes should be draw\type=1 - Which type, see below (for type=1: the center points draw; type=2: the separate factor scores linked by lines; ... will be passed to function "points")\data.pch=20 - the pch of dataset, if type=1, the first one is used\col=1 - the color of observations, recycled used by data.frame\label=FALSE - A logical indicates if labels should be shown\lg.x="topright" - Position of legend \lg.y=NULL - Position of legend \xlim=NULL - The x limit \ylim=NULL - The y limit \label.cex=1 - the cex of text \... 
var - the separate gene view, layout can be specified \RV - the heatmap of RV coefficients

---

**plotGS**

*Plot the gene set space*

**Description**

Plot the gene set space of objects of "moa" and "mgsa"

**Usage**

```r
plotGS(x, axes=1:2, center.only=FALSE, topN=1, data.pch=20, data.col=1, highlight.col = 2, label=NULL, label.cex=1, layout=NULL, ...)
```

**Arguments**

- `x` An object of class `mgsa-class` or `moa.sup-class`
- `axes` An integer vector in the length 2 to indicate the axes to be drawn.
- `center.only` A logical to indicate whether the separate gene set spaces from each of the data set should be plotted. Default is FALSE.
- `topN` An integer specify N gene set from the most positive and negative end of axes to be labeled
- `data.pch` The shape for plotting each data set. This argument is passed to points function, so only used when separate gene set spaces are plotted (i.e. center.only = FALSE).
- `data.col` The col for plotting each data set. This argument is passed to points function, so only used when separate gene set spaces are plotted (i.e. center.only = FALSE).
- `highlight.col` The color used to highlight the selected gene sets
- `label` Either a character vector or NULL (default). The character vector should be the name of some gene sets want ot be labeled.
- `label.cex` Passed to `text` function to adjust the the labels
- `layout` A matrix passed to the layout function.
- `...` Other arguments passed to `points`
**prepGraphite**

Details

This is a convenience function to explore the gene set space so not very flexible. For customized plot, please use the object of `data@coord.comb` and `data@coord.sep`.

Value

If assign to variable, A list of selected/highlighted gene set at the (positive and negative) end of each axis will be returned.

Author(s)

Chen Meng

Examples

```r
# library(mogsa)
# loading gene expression data and supplementary data
data(NCI60_4array_supdata)
data(NCI60_4arrays)
mgsa <- mogsa(x = NCI60_4arrays, sup=NCI60_4array_supdata, nf=9,
              proc.row = "center_ssq!", w.data = "inertia", statis = TRUE)

plotGS(mgsa, center.only = TRUE, topN=5)
res <- plotGS(mgsa, center.only = FALSE, data.pch=1:4, data.col=1:4)
res
```

---

**Description**

Prepare pathway gene sets from "graphite" package, which could be passed to "prepSupMoa" function.

**Usage**

```r
prepGraphite(db, id = c("entrez", "symbol"))
```

**Arguments**

- `db` The database to be used, an object of class either 'PathwayList' create by "pathways" function.
- `id` Which identifier for output, either "entrez" or "symbol".

**Details**

Only support "entrez" or "symbol" output currently.
This function returns an object of list containing gene set information, which could be further processed by function "prepSupMoa" to convert to the object that can be used as input of "sup.moa" or "mogsa".

Author(s)
Chen Meng

References

See Also
See Also as prepMsigDB and prepSupMoa.

Examples
library(graphite)
keggdb <- prepGraphite(db = pathways("hsapiens", "kegg")[1:3], id = "entrez")

prepMsigDB
Convert gmt format file to a list

Description
Convert a gmt file (Could be downloaded from MSigDB) to a list of gene sets information.

Usage
prepMsigDB(file)

Arguments
file The directory and file name of the gmt file.

Value
This function returns an object of list containing gene set information, which could be further processed by function "prepSupMoa" to convert to the object that can be used as input of "sup.moa" or "mogsa".

Author(s)
Chen Meng

See Also
See Also as prepGraphite and prepSupMoa.
prepSupMoa

Prepare supplementary tables for projection by sup.moa or mogsa.

Description

Convert a list of gene set information to a set of supplementary tables that can be used as input of function "sup.moa" or "mogsa".

Usage

prepSupMoa(X, geneSets, minMatch = 10, maxMatch = 500)

Arguments

X
A matrix/data.frame or a list of matrix/data.frame or a list of character vector. If it is a list of matrix/data.frame, row names of matrix/data.frame will be used to create the projection matrix. Otherwise the character vectors will used to create the supplementary matrix.

geneSets
Gene sets list or an object of class "GeneSet" or "GeneSetCollection". A gene set list could be returned by prepGraphite or prepMolsigDB.

minMatch
The minimum match of geneset.

maxMatch
The maximum match genesets.

Details

Details here

Value

A list of matrix could used as supplementary tables by "sup.moa" or "mogsa".

Author(s)

Chen Meng

See Also

See Also as prepGraphite and prepMolsigDB.
Examples

```r
library(graphite)
data(NCI60_4arrays)
kegg <- pathways(species = "hsapiens", "kegg")
pw <- c("Purine metabolism", "Endocrine resistance", "MAPK signaling pathway")
gss <- prepGraphite(db = kegg[pw], id="symbol")
gss <- lapply(gss, function(x) sub("SYMBOL: ", ", ", x))
sup_data1 <- prepSupMoa(NCI60_4arrays, geneSets=gss)
gene_list <- lapply(NCI60_4arrays, rownames)
sup_data2 <- prepSupMoa(gene_list, geneSets=gss)
```

Description

Methods for function print

Methods

- `signature(object = "moa")`: print "moa" class
- `signature(object = "moa.sup")`: print "sup.moa" class
- `signature(object = "mgsa")`: print "mgsa" class

**processOpt**

Preprocessing of input data in `mbpca`.

Description

An internal function called by `mbpca`.

Usage

```r
processOpt(x, center = TRUE, scale = FALSE, option = c("lambda1", "inertia", "uniform"))
```

Arguments

- `x`: A list of matrices, rows are observations and columns are variables
- `center`: A logical variable indicates whether columns should be centered
- `scale`: A logical variable indicates whether columns should be scaled
- `option`: A character string could be one of c("lambda1", "inertia", "uniform") to indicate how the different matrices should be normalized. If "lambda1", the matrix is divided by its the first singular value, if "inertia", the matrix is divided by its total inertia (sum of square), if "uniform", none of them would be done.

Value

A list of normalized matrix.

Author(s)

Chen Meng
**softK**

**Description**

Methods for function `show`

**Methods**

- `signature(object = "moa")` show "moa" class
- `signature(object = "moa.sup")` show "sup.moa" class
- `signature(object = "mgsa")` show "mgsa" class

**softK**

*Soft-thresholding operator*

**Description**

Soft-thresholding operator, which is called by `mbpca`.

**Usage**

`softK(x, k)`

**Arguments**

- `x` A numerical vector
- `k` Number of non-zero elements want to keep

**Value**

A numerical vector

**Author(s)**

Chen Meng

**Examples**

```r
v <- rnorm(10)
softK(v, k = 2)
```
summary-methods

Methods for function `summary`

Description

Methods for function `summary`

Methods

- `signature(object = "moa")` summary "moa" class
- `signature(object = "moa.sup")` summary "sup.moa" class
- `signature(object = "mgsa")` summary "mgsa" class

sup.moa

Projecting supplementary tables on object of class moa-class.

Description

Projecting supplementary tables on `moa-class`

Usage

```r
sup.moa(X, sup, nf = 2, ks.stat=FALSE, ks.B = 1000, ks.cores = NULL)
```

Arguments

- `X` An object of class `moa-class`
- `sup` A list of data.frames contains supplementary data.
- `nf` The number of principal components used in the projection.
- `ks.stat` The logical indicates if the p-value should be calculated using K-S statistic (the method used in "ssgsea" in GSVA package). Default is FALSE, which means using the z-score method.
- `ks.B` An integer to indicate the number of bootstrapping samples to calculated the p-value of KS statistic.
- `ks.cores` An integer indicate the number of cores to be used in bootstrapping. It is passed to function `mclapply` in the parallel package.

Details

Projecting supplementary tables on `moa-class`, for details see reference.

Value

An object of class `moa.sup-class`.

Author(s)

Chen Meng
References


Examples

```r
# library(mogsa)
# loading gene expression data and supplementary data
data(NCI60_4array_supdata)
data(NCI60_4arrays)
# check the dimension of each supplementary data to see how many gene set annotated the data
sapply(NCI60_4array_supdata, dim)
# run analysis
ana <- moa(NCI60_4arrays, proc.row = "center_ssq!", w.data = "inertia", statis = TRUE)
plot(ana, value="eig")
# projectin supplementary data
smoa <- sup.moa(ana, sup=NCI60_4array_supdata, nf=3)
# heatmap visualize the gene set scores
heatmap(slot(smoa, "score"))
```

---

toMoa

**convert mbpca result to moa-class**

**Description**

An internal function called by mbpca.

**Usage**

`toMoa(data, x, call)`

**Arguments**

- `data` The preprocessed data in mbpca
- `x` The object calculated in mbpca
- `call` The call of mbpca

**Value**

An object of moa-class.

**Author(s)**

Chen Meng
Description

The weighted version of singular value decomposition.

Usage

\[
\text{wsvd}(X, D1 = \text{diag}(1, \text{nrow}(X)), D2 = \text{diag}(1, \text{ncol}(X)))
\]

Arguments

- **X**: A numeric matrix whose wSVD decomposition is to be computed.
- **D1**: A square matrix or vector. The left constraint/weight matrix (symmetric and positive in diagonal). The dimension of D1 should be the same with the number of rows in X. A vector input will be converted to a diagonal matrix.
- **D2**: A square matrix or vector. The right constraint/weight matrix (symmetric, positive in diagonal). The dimension of D1 should be the same with the number of columns in X. A vector input will be converted to a diagonal matrix.

Details

The weighted version of generalized singular value decomposition (SVD) of matrix \( A = UDV' \) with the constraints \( U'D1U = I \) and \( V'D2V = I \). \( D1 \) and \( D2 \) are two matrices express constraints imposed on the rows and the columns of matrix \( A \).

Value

- **d**: singular values
- **u**: left singular vectors
- **v**: right singular vectors
- **D1**: the left weight matrix (directly from input)
- **D2**: the right weight matrix (directly from input)

Author(s)

Chen Meng

References


See Also

svd
Examples

```r
set.seed(56)
m <- matrix(rnorm(15), 5, 3)
w1 <- rnorm(5)
wr <- runif(3)
s <- wsvd(X=m, D1=w1, D2=wr)
# t(s$u) %*% diag(w1) %*% s$u
# t(s$v) %*% diag(wr) %*% s$v
# all.equal(m, as.matrix(s$u) %*% diag(s$d) %*% t(s$v))
```
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