Package ‘netSmooth’

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Type Package

Title Network smoothing for scRNAseq

Version 1.22.0

Description netSmooth is an R package for network smoothing of single cell RNA sequencing data. Using bio networks such as protein-protein interactions as priors for gene co-expression, netsmooth improves cell type identification from noisy, sparse scRNAseq data.

biocViews Network, GraphAndNetwork, SingleCell, RNASeq, GeneExpression, Sequencing, Transcriptomics, Normalization, Preprocessing, Clustering, DimensionReduction

URL https://github.com/BIMSBbioinfo/netSmooth

BugReports https://github.com/BIMSBbioinfo/netSmooth/issues

License GPL-3

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LazyData true

Depends R (>= 3.5), scater (>= 1.15.11), clusterExperiment (>= 2.1.6)

Imports entropy, SummarizedExperiment, SingleCellExperiment, Matrix, cluster, data.table, stats, methods, DelayedArray, HDF5Array (>= 1.15.13)

Suggests knitr, testthat, Rtsne, biomaRt, igraph, STRINGdb, NMI, pheatmap, ggplot2, BiocStyle, rmarkdown, BiocParallel, uwot

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R topics documented:

calc2DEntropy ........................................ 2
clusterExperimentWorkflow  ................................ 3
clusterOne ........................................ 4
dimReduce ........................................ 4
human.ppi ........................................ 5
l1NormalizeColumns ................................ 5
l1NormalizeRows .................................. 6
mouse.ppi ......................................... 6
netSmooth,matrix-method  ............................ 7
pickDimReduction,matrix-method  ................. 9
projectFromNetworkRecombine,matrix-method ...... 10
projectOnNetwork,matrix-method  .................. 10
randomWalkByIterations  ............................ 11
randomWalkByMatrixInv,matrix-method ............. 12
randomWalkBySolve,matrix-method ................ 12
robustClusters,SummarizedExperiment-method .... 13
scoreSmoothing ...................................... 14
smallPPI .......................................... 14
smallscRNAseq .................................... 15
smoothAndRecombine,matrix-method ............... 15

Index  17

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calc2DEntropy  Calculate entropy in 2D data

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Description

Calculate entropy in 2D data

Usage

calc2DEntropy(x, numBins1 = 20, numBins2 = 20)

Arguments

x  the 2D data to get entropy from
numBins1  the number of bins along the first dimension to discretize data into
numBins2  the number of bins along the second dimension to discretize data into
**clusterExperimentWorkflow**

**Value**

The Shannon entropy in the 2D data \( x \)

---

Performs clustering workflow using ‘clusterExperiment’ functions

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**Description**

Performs clustering workflow using ‘clusterExperiment’ functions

**Usage**

```r
clusterExperimentWorkflow(
  se,
  dimReduceFlavor = c("pca", "tsne", "dm", "umap"),
  cluster.ks = 5:10,
  cluster.function = "pam",
  nVarDims = c(100, 500, 1000),
  makeConsensusProportion = 0.7,
  makeConsensusMinSize = 4,
  runMergeClusters = TRUE,
  is.counts = TRUE,
  random.seed = 1
)
```

**Arguments**

- **se** SummarizedExperiment object
- **dimReduceFlavor** algorithm for reduced dimension embedding step
- **cluster.ks** range of Ks to cluster over
- **cluster.function** clustering algorithm to use for all clusterings
- **nVarDims** numbers of variable genes to perform clusterings over
- **makeConsensusProportion** proportion of times samples need to be co-clustered for co-clustering step
- **makeConsensusMinSize** minimum cluster size
- **runMergeClusters** logical: merge similar clusters
- **is.counts** logical: is data counts
- **random.seed** passed to clusterExperiment. set to NULL in order to not set a random seed.

**Value**

cluster assignments
**clusterOne**

*Run one clustering using kmeans o PAM*

**Description**

Run one clustering using kmeans o PAM

**Usage**

clusterOne(x, algorithm = c("kmeans", "pam"), k = 5)

**Value**

kmeans or PAM cluster assignments

---

**dimReduce**

*Get lower dimension embedding*

**Description**

Get lower dimension embedding

**Usage**

dimReduce(
  x,
  flavor = c("pca", "tsne", "umap"),
  k = 2,
  is.counts = TRUE,
  ntop = 500
)

**Arguments**

- `x`: gene expression matrix [GENES x SAMPLES]
- `flavor`: the algorithm to use to obtain the dimensionality reduction must be in c(‘pca’, ‘tsne’, ‘umap’)
- `k`: the number of dimensions in the reduced dimension representation
- `is.counts`: logical: is ‘x’ counts data
- `ntop`: number of most variable genes to use for dimensionality reduction

**Value**

reduced dimensionality representation
human.ppi

Human Protein-Protein interaction graph

Description
An adjacency matrix of the 10 percent highest confidence interactions between human proteins on STRINGdb.

Usage
human.ppi

Format
A square matrix where A_ij=1 if gene i interacts with gene j

Details
See the script in ‘system.file(package="netSmooth", "data-raw", "make_ppi_from_string.R")’ for full details of how this object was made.

Source
http://www.string-db.org/

l1NormalizeColumns

Column-normalize a sparse, symmetric matrix (using the l1 norm) so that each column sums to 1.

Description
Column-normalize a sparse, symmetric matrix (using the l1 norm) so that each column sums to 1.

Usage
l1NormalizeColumns(A)

Arguments
A matrix

Value
column-normalized sparse matrix object
l1NormalizeRows  
Row-normalize a sparse, symmetric matrix (using the l1 norm) so that each row sums to 1.

Description
Row-normalize a sparse, symmetric matrix (using the l1 norm) so that each row sums to 1.

Usage
l1NormalizeRows(A)

Arguments
A  matrix

Value
row-normalized sparse matrix object

mouse.ppi  
Mouse Protein-Protein interaction graph

Description
An adjacency matrix of the 10 percent highest confidence interactions between mouse proteins on STRINGdb.

Usage
mouse.ppi

Format
A square matrix where A_ij=1 if gene i interacts with gene j

Details
See the script in ‘system.file(package="netSmooth", "data-raw", "make_ppi_from_string.R")’ for full details of how this object was made.

Source
http://www.string-db.org/
netSmooth, matrix-method

Perform network smoothing of gene expression or other omics data

Description

Perform network smoothing of gene expression or other omics data

Usage

```r
## S4 method for signature 'matrix'
netSmooth(
  x,
  adjMatrix,
  alpha = "auto",
  normalizeAdjMatrix = c("rows", "columns"),
  autoAlphaMethod = c("robustness", "entropy"),
  autoAlphaRange = 0.1 * (seq_len(9)),
  autoAlphaDimReduceFlavor = "auto",
  is.counts = TRUE,
  bpparam = BiocParallel::SerialParam(),
  ...
)

## S4 method for signature 'SummarizedExperiment'
netSmooth(x, ...)

## S4 method for signature 'SingleCellExperiment'
netSmooth(x, ...)

## S4 method for signature 'Matrix'
netSmooth(
  x,
  adjMatrix,
  alpha = "auto",
  normalizeAdjMatrix = c("rows", "columns"),
  autoAlphaMethod = c("robustness", "entropy"),
  autoAlphaRange = 0.1 * (seq_len(9)),
  autoAlphaDimReduceFlavor = "auto",
  is.counts = TRUE,
  bpparam = BiocParallel::SerialParam(),
  ...
)

## S4 method for signature 'DelayedMatrix'
netSmooth(
  x,
  ...
)
```
adjMatrix,
alpha = "auto",
normalizeAdjMatrix = c("rows", "columns"),
autoAlphaMethod = c("robustness", "entropy"),
autoAlphaRange = 0.1 * (seq_len(9)),
autoAlphaDimReduceFlavor = "auto",
is.counts = TRUE,
bpparam = BiocParallel::SerialParam(),
filepath = NULL,
...
)

Arguments

x
testMatrix
alpha
numeric in [0,1] or 'auto'. if 'auto', the optimal value for alpha will be automatically chosen among the values specified in 'autoAlphaRange', using the strategy specified in 'autoAlphaMethod'
normalizeAdjMatrix
how to normalize the adjacency matrix possible values are 'rows' (in-degree) and 'columns' (out-degree)
autoAlphaMethod
if 'robustness', pick alpha that gives the highest proportion of samples in robust clusters if 'entropy', pick alpha that gives highest Shannon entropy in 2D PCA embedding
autoAlphaRange
if 'alpha='optimal'', search these values for the best alpha
autoAlphaDimReduceFlavor
algorithm for dimensionality reduction that will be used to pick the optimal value for alpha. Either the 2D embedding to calculate the Shannon entropy for (if 'autoAlphaMethod='entropy''), or the dimensionality reduction algorithm to be used in robust clustering (if 'autoAlphaMethod='robustness'')
is.counts
logical: is the assay count data
bpparam
instance of bpparam, for parallel computation with the 'alpha='auto' option. See the BiocParallel manual.
...
arguments passed on to 'robustClusters' if using the robustness criterion for optimizing alpha
filepath
String: Path to location where hdf5 output file is supposed to be saved. Will be ignored when regular matrices or SummarizedExperiment are used as input.

Value

network-smoothed gene expression matrix or SummarizedExperiment object
Examples

```
x <- matrix(rnbinom(12000, size=1, prob = .1), ncol=60)
rownames(x) <- paste0('gene', seq_len(dim(x)[1]))

adj_matrix <- matrix(as.numeric(rnorm(200*200)>0.8), ncol=200)
rownames(adj_matrix) <- colnames(adj_matrix) <- paste0('gene', seq_len(dim(x)[1]))
x.smoothed <- netSmooth(x, adj_matrix, alpha=0.5)
```

Description

Pick the dimensionality reduction method for a dataset that gives the 2D embedding with the highest entropy.

Usage

```r
## S4 method for signature 'matrix'
pickDimReduction(x, flavors = c("pca", "tsne", "umap"), is.counts = TRUE)
```

Arguments

- `x`: matrix or SummarizedExperiment object [GENES x SAMPLES]
- `flavors`: list of dimensionality reduction algorithms to try. Currently the options are "pca", "tsne" and "umap"
- `is.counts`: logical: is exprs count data

Value

name of dimensionality reduction method that gives the highest 2d entropy

Examples

```
x <- matrix(rnbinom(60000, size=1, prob = .1), ncol=100)
pickDimReduction(x)
```
projectFromNetworkRecombine, matrix-method
Combine gene expression from smoothed space (that of the network) with the expression of genes that were not smoothed (not present in network)

Description
Combine gene expression from smoothed space (that of the network) with the expression of genes that were not smoothed (not present in network)

Usage
## S4 method for signature 'matrix'
projectFromNetworkRecombine(original_expression, smoothed_expression)

Arguments
original_expression
the non-smoothed expression
smoothed_expression
the smoothed gene expression, in the space of the genes defined by the network
filepath
String: Path to location where hdf5 output file is supposed to be saved. Will be ignored when regular matrices or SummarizedExperiment are used as input.

Value
a matrix in the dimensions of original_expression, where values that are present in smoothed_expression are copied from there.

projectOnNetwork, matrix-method
Project the gene expression matrix onto a lower space of the genes defined in the smoothing network

Description
Project the gene expression matrix onto a lower space of the genes defined in the smoothing network

Usage
## S4 method for signature 'matrix'
projectOnNetwork(gene_expression, new_features, missing.value = 0)
**randomWalkByIterations**

**Arguments**
- `gene_expression` (gene expression matrix)
- `new_features` (the genes in the network, on which to project the gene expression matrix)
- `missing.value` (value to assign to genes that are in network, but missing from gene expression matrix)

**Value**
- the gene expression matrix projected onto the gene space defined by `new_features`

---

**randomWalkByIterations**

*Smooth data on graph by computing iterations*

---

**Description**
Smooth data on graph by computing iterations

**Usage**

```r
randomWalkByIterations(
  f0,
  adjMatrix,
  alpha,
  normalizeAdjMatrix = c("rows", "columns"),
  tol = 1e-06,
  max.iter = 100
)
```

**Arguments**
- `f0` (initial data matrix [NxM])
- `adjMatrix` (adjacency matrix of graph to network smooth on will be column-normalized)
- `alpha` (smoothing coefficient (1 - restart probability of random walk))
- `tol` (the tolerance (stopping criterion))
- `max.iter` (the maximum number of iterations before terminating)

**Value**
- network-smoothed gene expression
randomWalkByMatrixInv, matrix-method

Smooth data on graph by computing the closed-form steady state distribution of the random walk with restarts process.

Description

The closed-form solution is given by $f_{ss} = (1 - \alpha) \cdot (I - \alpha \cdot A)^{-1} \cdot f_0$ and is computed by matrix inversion in this function.

Usage

```r
## S4 method for signature 'matrix'
randomWalkByMatrixInv(
  f0,  
  adjMatrix, 
  alpha, 
  normalizeAdjMatrix = c("rows", "columns")
)
```

Arguments

- **f0**: initial data matrix [NxM]
- **adjMatrix**: adjacency matrix of graph to network smooth on will be column-normalized.
- **alpha**: smoothing coefficient (1 - restart probability of random walk)

Value

network-smoothed gene expression

randomWalkBySolve, matrix-method

Smooth data on graph by solving the linear equation $(I - \alpha A) \cdot E_{sm} = E \cdot (1-\alpha)$

Description

Smooth data on graph by solving the linear equation $(I - \alpha A) \cdot E_{sm} = E \cdot (1-\alpha)$

Usage

```r
## S4 method for signature 'matrix'
randomWalkBySolve(E, A, alpha, normalizeAdjMatrix = c("rows", "columns"))
```
robustClusters, SummarizedExperiment-method

Arguments

- **E** initial data matrix [NxM]
- **A** adjacency matrix of graph to network smooth on will be column-normalized.
- **alpha** smoothing coefficient (1 - restart probability of random walk)

Value

network-smoothed gene expression

Description

Perform robust clustering on dataset, and calculate the proportion of samples in robust clusters

Usage

```r
## S4 method for signature 'SummarizedExperiment'
robustClusters(x, dimReduceFlavor = "auto", is.counts = TRUE, ...)

## S4 method for signature 'matrix'
robustClusters(x, ...)
```

Arguments

- **x** matrix or SummarizedExperiment object
- **dimReduceFlavor** algorithm for dimensionality reduction step of clustering procedure. May be ‘pca’, ‘tsne’, ‘dm’, ‘umap’ or ‘auto’, which uses shannon entropy to pick the algorithm.
- **is.counts** logical: is the data counts
- **...** arguments passed on to ‘clusterExperimentWorkflow’

Value

list(clusters, proportion.robust)

Examples

```r
data("smallscRNAseq")
robustClusters(smallscRNAseq, dimReduceFlavor='pca')
```
scoreSmoothing  

*Calculate a score for a smoothing result, for picking the best alpha value*

**Description**

Calculate a score for a smoothing result, for picking the best alpha value

**Usage**

`scoreSmoothing(x, method = c("entropy", "robustness"), is.counts = TRUE, ...)`

**Arguments**

- `x`: the network-smoothed expression matrix
- `method`: the scoring method. ‘entropy’ calculates shannon entropy in a 2D PCA of the data. ‘robustness’ performs robust clustering and reports the proportion of samples in robust clusters

**Value**

the score

---

smallPPI  

*A small human Protein-Protein interaction graph for use in examples.*

**Description**

Contains a synthetic PPI of human genes.

**Usage**

`smallPPI`

**Format**

An object of class `matrix` with 611 rows and 611 columns.
smallscRNAseq

A small single cell RNA-seq dataset for use in examples.

Description

Contains scRNAseq profiles of human blastomeres.

Usage

smallscRNAseq

Format

SingleCellExperiment

Source


smoothAndRecombine, matrix-method

Perform network smoothing on network when the network genes and
the experiment genes aren’t exactly the same.

Description

The gene network might be defined only on a subset of genes that are measured in any experiment. Further, an experiment might not measure all genes that are present in the network. This function projects the experiment data onto the gene space defined by the network prior to smoothing. Then, it projects the smoothed data back into the original dimensions.

Usage

## S4 method for signature 'matrix'
smoothAndRecombine(
  gene_expression,
  adj_matrix,
  alpha,
  smoothing.function = randomWalkBySolve,
  normalizeAdjMatrix = c("rows", "columns")
)
**Arguments**

- `gene_expression`: gene expression data to be smoothed [N_genes x M_samples]
- `adj_matrix`: adjacency matrix of network to perform smoothing over. Will be column-normalized. Rownames and colnames should be genes.
- `alpha`: network smoothing parameter (1 - restart probability in random walk model).
- `smoothing.function`: must be a function that takes in data, adjacency matrix, and alpha. Will be used to perform the actual smoothing.
- `normalizeAdjMatrix`: which dimension (rows or columns) should the adjacency matrix be normalized by. rows corresponds to in-degree, columns to out-degree.
- `filepath`: String: Path to location where hdf5 output file is supposed to be saved. Will be ignored when regular matrices or SummarizedExperiment are used as input.

**Value**

matrix with network-smoothed gene expression data. Genes that are not present in smoothing network will retain original values.
Index

* datasets
  human.ppi, 5
  mouse.ppi, 6
  smallPP1, 14
  smallscRNAseq, 15

* internal
  calc2DEntropy, 2
  clusterExperimentWorkflow, 3
  clusterOne, 4
  dimReduce, 4
  l1NormalizeColumns, 5
  l1NormalizeRows, 6
  projectFromNetworkRecombine, matrix-method, 10
  projectOnNetwork, matrix-method, 10
  randomWalkByIterations, 11
  randomWalkByMatrixInv, matrix-method, 12
  randomWalkBySolve, matrix-method, 12
  scoreSmoothing, 14
  smoothAndRecombine, matrix-method, 15
  netSmooth, Matrix-method (netSmooth, matrix-method), 7
  netSmooth, matrix-method, 7
  netSmooth, SingleCellExperiment-method (netSmooth, matrix-method), 7
  netSmooth, SummarizedExperiment-method (netSmooth, matrix-method), 7
  pickDimReduction (pickDimReduction, matrix-method), 9
  pickDimReduction, DelayedMatrix-method (pickDimReduction, matrix-method), 9
  pickDimReduction, Matrix-method (pickDimReduction, matrix-method), 9
  robustClusters (robustClusters, SummarizedExperiment-method), 13
  robustClusters, matrix-method (robustClusters, SummarizedExperiment-method), 13
  robustClusters, SummarizedExperiment-method, 13
scoreSmoothing, 14
smallPPI, 14
smallscRNAseq, 15
smoothAndRecombine, matrix-method, 15