Package ‘scviR’

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Title experimental interface from R to scvi-tools
Version 1.2.0
Description This package defines interfaces from R to scvi-tools. A vignette works through the totalVI tutorial for analyzing CITE-seq data. Another vignette compares outputs of Chapter 12 of the OSCA book with analogous outputs based on totalVI quantifications. Future work will address other components of scvi-tools, with a focus on building understanding of probabilistic methods based on variational autoencoders.
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       MatrixGenerics
Suggests knitr, testthat, reshape2, ggplot2, rhdf5, BiocStyle
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adtProfiles produce a heatmap from a specialized CITE-seq SingleCellExperiment

Description
produce a heatmap from a specialized CITE-seq SingleCellExperiment

Usage
adtProfiles(x, lb = -3, ub = 3, do_z = FALSE)

Arguments
x SingleCellExperiment instance that has an 'se.averaged' component in its metadata
lb numeric(1) lower bound on 'breaks' sequence for ComplexHeatmap::pheatmap, defaults to -3
ub numeric(1) upper bound on 'breaks' sequence for ComplexHeatmap::pheatmap, defaults to 3
do_z logical(1) if TRUE, divide the residuals by their standard deviation across clusters, defaults to false
Value

ComplexHeatmap::pheatmap instance
side effect of pheatmap::pheatmap call

Note

See the OSCA book ch12.5.2 for the application.

Examples

ch12sce <- getCh12Sce()
adProfiles(ch12sce)
adProfiles(ch12sce, do_z = TRUE)
cacheCiteseq5k10kPbmcs

grab scvi-tools-processed PBMC CITE-seq data in anndata format (gzipped) from Open Storage Network

Description

grab scvi-tools-processed PBMC CITE-seq data in anndata format (gzipped) from Open Storage Network

Usage

`cacheCiteseq5k10kPbmcs()`

Value

invisibly, the path to the .h5ad file

Note

Original h5ad files obtained using scvi-tools 0.18.0 scvi.data.pbmcs_10x_cite_seq, then processed according to steps in the scviR vignette, which follow the [scvi-tools tutorial](https://colab.research.google.com/github/scverse/scvi-tutorials/blob/0.18.0/totalVI.ipynb) by Gayoso et al.

It may be advantageous to set `options(timeout=3600)` or to allow an even greater time for internet downloads, if working at a relatively slow network connection.

Examples

```r
h5path <- cacheCiteseq5k10kPbmcs()
cmeta <- rhdf5::h5ls(h5path)
dim(cmeta)
head(cmeta, 17)
```

---

cacheCiteseq5k10kTutvae

grab scvi-tools VAE instance built on the PBMC datasets following the tutorial

Description

grab scvi-tools VAE instance built on the PBMC datasets following the tutorial

Usage

`cacheCiteseq5k10kTutvae()`
clusters.adt

Value

invisibly, the path to the .zip file holding the fitted VAE and associated data

Note


It may be advantageous to set `options(timeout=3600)` or to allow an even greater time for internet downloads, if working at a relatively slow network connection.

Examples

```r
zpath <- cacheCiteseq5k10kTutvae()
td <- tempdir()
utils::unzip(zpath, exdir = td)
vaedir <- paste0(td, "/vae2_ov")
scvi <- scviR()
adm <- anndataR()
hpath <- cacheCiteseq5k10kBmcsrc()
data <- adm$read(hpath)
mod <- scvi$model$"_totalvi"$TOTAL$load(vaedir, adata, use_gpu = FALSE)
mod
```

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Description

ADT-based cluster labels for 7472 cells in OSCA chapter 12 analysis

Usage

```
clusters.adt
```

Format

```
factor
```
exploreSubcl

clusters.rna  mRNA-based cluster labels for 7472 cells in OSCA chapter 12 analysis

Description
mRNA-based cluster labels for 7472 cells in OSCA chapter 12 analysis

Usage
clusters.rna

Format
factor

exploreSubcl  app to explore diversity in RNA-subclusters within ADT clusters

Description
app to explore diversity in RNA-subclusters within ADT clusters

Usage
exploreSubcl(sce, inlist, adtcls)

Arguments
sce  a SingleCellExperiment with altExp with ADT quantification
inlist  list of SingleCellExperiments (SCEs) formed by scran::quickSubCluster
adtcls  vector of ADT cluster assignments

Value
shinyApp instance

Note
TSNE should already be available in `altExp(sce)`; follow OSCA book 12.5.2. If using example, set `ask=FALSE`.

Examples
sce <- getCh12Sce()
all.sce <- getCh12AllSce()
data(clusters.adt)
runApp(exploreSubcl(sce, all.sce, clusters.adt)) # trips up interactive pkgdown?)
getCh12AllSce

get list of cluster-specific SCE for 10k PBMC annotated as in OSCA book chapter 12

Description
get list of cluster-specific SCE for 10k PBMC annotated as in OSCA book chapter 12

Usage
getCh12AllSce()

Value
SimpleList of SingleCellExperiment instances

Note
This is a list of SingleCellExperiment instances with data on a total of 7472 cells from a 10x CITE-seq experiment. An altExp component in each list element includes antibody-derived tag (ADT) counts on 17 proteins. The data are acquired and processed as described in ch 12 of the OSCA book, circa February 2023. List elements correspond to mRNA-based sub-clusters of ADT-based clusters.

Examples
ch12_allSce <- getCh12AllSce()
vapply(ch12_allSce, ncol, numeric(1))

getCh12Sce

get SCE for 10k PBMC annotated as in OSCA book chapter 12

Description
get SCE for 10k PBMC annotated as in OSCA book chapter 12

Usage
getCh12Sce(clear_cache = FALSE)

Arguments
clear_cache logical(1) will delete relevant entries in available cache before continuing, defaults to FALSE
Value

SingleCellExperiment instance

Note

This is a SingleCellExperiment instance with data on 7472 cells from a 10x CITE-seq experiment. An altExp component includes antibody-derived tag (ADT) counts on 17 proteins. The data are acquired and processed as described in ch 12 of the OSCA book, circa February 2023. A metadata element (se.averaged) includes the result of averaging protein abundance estimates within ADT-based clusters, as is done to give rise to Figure 12.8 of the OSCA book.

Examples

```r
ch12sce <- getCh12Sce()
ch12sce
```

---

getciteseq5k10kPbmcs  

helper to get the processed anndata for CITE-seq PBMCs from scvi-tools tutorial

Description

helper to get the processed anndata for CITE-seq PBMCs from scvi-tools tutorial

Usage

```r
getciteseq5k10kPbmcs()
```

Value

python reference to anndata

Note

It may be advantageous to set \texttt{options(timeout=3600)} or to allow an even greater time for internet downloads, if working at a relatively slow network connection.

Examples

```r
getciteseq5k10kPbmcs()
```
getCiteseqTutvae

helper to get the tutorial VAE for PBMCs from scvi-tools tutorial

Description

helper to get the tutorial VAE for PBMCs from scvi-tools tutorial

Usage

getCiteseqTutvae(use_gpu = FALSE)

Arguments

use_gpu logical(1), defaulting to FALSE, passed to TOTALVI.load

Value

python reference to anndata

Examples

getCiteseqTutvae()

getPro5k10kAdata

generate an anndata reference to 5k10k protein after totalVI from tutorial

Description

generate an anndata reference to 5k10k protein after totalVI from tutorial

Usage

getPro5k10kAdata()

Value

python reference to anndata

Note

It may be advantageous to set ‘options(timeout=3600)’ or to allow an even greater time for internet downloads, if working at a relatively slow network connection.

Examples

getPro5k10kAdata()
getSubclLM

get lmFit for heterogeneity across subclusters

Description

get lmFit for heterogeneity across subclusters

Usage

getSubclLM(inlist, clname)

Arguments

inlist               list of SingleCellExperiments (SCEs) formed by scran::quickSubCluster
clname               character(1) name of cluster SCE to assess

Value

limma::lmFit output

Note

It is assumed that 'logcounts' is an assay element, and that 'subcluster' is a colData element of each SCE in inlist

Examples

all.sce <- getCh12AllSce()
lm3 <- getSubclLM(all.sce, "3")
names(lm3)

getSubclusteringFeatures

get lmFit F-stat based collection of n genes most varying in mean across subclusters

Description

get lmFit F-stat based collection of n genes most varying in mean across subclusters

Usage

getSubclusteringFeatures(inlist, clname, n = 20)
Arguments

inlist  list of SingleCellExperiments (SCEs) formed by scran::quickSubCluster
clname  character(1) name of cluster SCE to assess
n       numeric(1) number to preserve

Value

list with two elements, feat = rowData corresponding to variable genes, stats = topTable result

Note

Symbol will be taken from feat and placed in stats component if available

Examples

all.sce <- getCh12AllSce()
scl <- getSubclusteringFeatures(all.sce, "3", 10)
names(scl)

getTotalVI5k10kAdata
get anndata reference to full totalVI processing of 5k10k data

Description

get anndata reference to full totalVI processing of 5k10k data

Usage

g getTotalVI5k10kAdata()

Value

python reference to anndata

Examples

full <- getTotalVI5k10kAdata()
full
**getTotalVINormalized5k10k**

get matrices of normalized quantifications from full totalVI 5k10k from tutorial

**Description**

get matrices of normalized quantifications from full totalVI 5k10k from tutorial

**Usage**

g getTotalVINormalized5k10k()

**Value**

list of matrices

**Examples**

```r
nmlist <- getTotalVINormalized5k10k()
vapply(nmlist, dim, numeric(2))
```

---

**pyHelp2**

helper to get text from python help utility – may need handling through basilisk

**Description**

helper to get text from python help utility – may need handling through basilisk

**Usage**

```r
pyHelp2(object)
```

**Arguments**

- `object` a reference to a python module typically with class `python.builtin.module`

**Value**

character vector of lines from python help result
scanpyHelper

**Description**

shiny app that helps access documentation on python-accessible components

**Usage**

scanpyHelper()

**Value**

shinyApp instance

---

scanpyR

**Description**

basic interface

**Usage**

scanpyR()

**Value**

basiliskRun result with import from reticulate, typically a Module

**Examples**

```r
sc <- scanpyR()
sc
sc$pp
```
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<tr>
<th>scviHelper</th>
<th>shiny app that helps access documentation on python-accessible components</th>
</tr>
</thead>
</table>

**Description**

shiny app that helps access documentation on python-accessible components

**Usage**

```r
scviHelper()
```

**Value**

shinyApp instance

---

<table>
<thead>
<tr>
<th>scviR</th>
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</tr>
</thead>
</table>

**Description**

basic interface

**Usage**

```r
scviR()
```

**Value**

basiliskRun result with import from reticulate, typically a Module

**Examples**

```r
scvi <- scviR()
scki
scvi
scvi$model
```
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