Package ‘snapcount’

March 28, 2024

Type  Package
Title  R/Bioconductor Package for interfacing with Snaptron for rapid querying of expression counts
Version  1.14.0
Description  snapcount is a client interface to the Snaptron webservices which support querying by gene name or genomic region. Results include raw expression counts derived from alignment of RNA-seq samples and/or various summarized measures of expression across one or more regions/genes per-sample (e.g. percent spliced in).
Depends  R (>= 4.0.0)
Imports  R6, httr, rlang, purrr, jsonlite, assertthat, data.table, Matrix, magrittr, methods, stringr, stats, IRanges, GenomicRanges, SummarizedExperiment
Suggests  BiocManager, bit64, covr, knitcitations, knitr (>= 1.6), devtools, BiocStyle (>= 2.5.19), rmarkdown (>= 0.9.5), testthat (>= 2.1.0)
License  MIT + file LICENSE
Encoding  UTF-8
LazyData  False
RoxygenNote  7.1.1
Roxygen  list(markdown = TRUE, roclets = c(``rd``, ``namespace``, ``collate``))
URL  https://github.com/langmead-lab/snapcount
BugReports  https://github.com/langmead-lab/snapcount/issues
biocViews  Coverage, GeneExpression, RNASeq, Sequencing, Software, DataImport
VignetteBuilder  knitr
ByteCompile  true
git_url  https://git.bioconductor.org/packages/snapcount
git_branch  RELEASE_3_18
Description

snapcount is a client interface to the Snaptron webservice which supports querying by gene name or genomic region.

Details

Results include raw expression counts derived from alignment of RNA-seq samples and/or various summarized measures of expression across one or more regions/genes per-sample (e.g. percent spliced in).

To learn more about snapcount, check out the vignette: `browseVignettes(package = "snapcount")`
Package options

snapcount.host Change the host that snapcount uses when connecting to Snaptron. Default: snaptron.cs.jhu.edu

snapcount.port Change the port that snapcount uses when connecting to Snaptron. Default: 80

Author(s)

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See Also

Useful links:

• https://github.com/langmead-lab/snapcount
• Report bugs at https://github.com/langmead-lab/snapcount/issues

Compilation

Enum for Snaptron compilations

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<tr>
<td>The variants for this enum will be populated dynamically after the package has been loaded. If the package cannot connect to the internet the variants will default to:</td>
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<td>An object of class environment of length 21.</td>
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<td>• tcga</td>
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</tbody>
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See Also

http://snaptron.cs.jhu.edu/data.html for more information about Snaptron compilations.

Examples

qb <- QueryBuilder(compilation = Compilation$gtex, regions = "KCNIP4")
query_jx(qb)
Coordinates

<table>
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<th>Coordinates</th>
<th>Enum for Snaptron Coordinate modifiers</th>
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**Description**

Enum for Snaptron Coordinate modifiers

**Usage**

Coordinates

**Format**

An object of class `environment` of length 4.

**Fields**

- **Exact** Return junctions whose start and end coordinates match the boundaries of the region requested.
- **Within** Return junctions whose start and end coordinates are within the boundaries of the region requested.
- **StartIsExactOrWithin** Return junctions whose start coordinate matches, or is within, the boundaries of the region requested.
- **EndIsExactOrWithin** Return junctions whose end coordinate matches, or is within, the boundaries of the region requested.

**Examples**

```r
define qb <- QueryBuilder(compilation = "gtex", regions = "CD99")
define qb <- set_coordinate Modifier(qb, Coordinates$Exact)
define qb
```

**Description**

Constructs a QueryBuilder object from the given url

**Usage**

```r
from_url(url)
```
get_column_filters

Arguments

url

a well-formed url preferably obtained from a call to the uri_of_last_successful_request function

Value

Returns a QueryBuilder object with attributes set from the parsed url.

Examples

sb <- from_url("http://snaptron.cs.jhu.edu/gtex/snaptron?regions=CD99")
get_regions(sb)
get_compilation(sb)

Description

Get or set sample-related contraints for query

Usage

get_column_filters(qb)

set_column_filters(qb, ...)

Arguments

qb

a QueryBuilder object constructed using the QueryBuilder function

... one or more boolean predicates as either strings or unevaluated expressions

Value

get_column_filters returns the current filters as a list of strings. set_column_filters returns a new QueryBuilder object with the column filters set to the value of column_filters.

Examples

qb <- QueryBuilder(compilation = "gtex", regions = "CD99")
# column filters set using a string
qb <- set_column_filters(qb, "SMTS == Brain")
get_column_filters(qb)
# column filters set using unevaluated expression
qb <- set_column_filters(qb, SMTS == "Spleen")
get_column_filters(qb)
get_compilation

Get or set query compilation

Description

Get or set query compilation

Usage

get_compilation(qb)

set_compilation(qb, compilation)

Arguments

qb A QueryBuilder object constructed using the QueryBuilder function.
compilation A single string containing the name of the Snaptron data source. Any variant of the Compilation enum can also be passed as an argument.

Value

get_compilation returns the current compilation as string. set_compilation returns a new QueryBuilder object with the compilation set to the value of compilation.

Examples

qb <- QueryBuilder(compilation = "gtex", regions = "CD99")
get_compilation(qb)
qb <- set_compilation(qb, Compilation$tcga)
get_compilation(qb)

get_coordinate_modifier

Get or set coordinate modifiers for the query.

Description

Get or set coordinate modifiers for the query.

Usage

get_coordinate_modifier(qb)

set_coordinate_modifier(qb, coordinate_modifier)
get_regions

Arguments

\texttt{qb} \quad \text{a QueryBuilder object constructed using the \texttt{QueryBuilder} function.}

\texttt{coordinate\_modifier} \quad \text{any of the variants of the \texttt{Coordinates} enum.}

Value

\texttt{get\_coordinate\_modifier} \text{ returns the current coordinate modifier as a string.} \texttt{set\_coordinate\_modifier} \text{ returns a new QueryBuilder object with the coordinate modifier set to the value of \texttt{coordinate\_modifier}.}

Examples

\begin{verbatim}
qb <- QueryBuilder(compilation = "gtex", regions = "CD99")
qb <- set\_coordinate\_modifier(qb, Coordinates\$Within)
get\_coordinate\_modifier(qb)
\end{verbatim}

---

get\_regions \quad \textit{Get or set query regions}

Description

Get or set query regions

Usage

\begin{verbatim}
get\_regions(qb)
set\_regions(qb, regions)
\end{verbatim}

Arguments

\texttt{qb} \quad \text{A QueryBuilder object constructed using the \texttt{QueryBuilder} function.}

\texttt{regions} \quad \text{Either a list of 1 more HUGO gene names as strings e.g. "BRCA1" or a Granges class object containing one or more geneomic intervals (e.g. "chr1:1-1000").}

Value

\texttt{get\_regions} \text{ returns the current regions as a list of strings.} \texttt{set\_regions} \text{ returns a new QueryBuilder object with the regions set to the value of \texttt{regions}.}

Examples

\begin{verbatim}
qb <- QueryBuilder(compilation = "gtex", regions = "CD99")
get\_regions(qb)
qb <- set\_regions(qb, "chr1:1-1000")
get\_regions(qb)
qb <- set\_regions(qb, GenomicRanges::GRanges("chr1", "1-1000"))
get\_regions(qb)
\end{verbatim}
get_row_filters  Get or set range-related contraints for query

Description
Get or set range-related contraints for query

Usage
get_row_filters(qb)
set_row_filters(qb, ...)

Arguments
qb a QueryBuilder object constructed using the QueryBuilder function.
... one or more boolean predicates as either strings or unevaluated expressions.

Value
get_row_filters returns the current row filters as list of strings. set_row_filters returns a new
QueryBuilder object with the row filters set to the value of row_filters.

Examples
qb <- QueryBuilder(compilation = "gtex", regions = "CD99")
# row filters set as a string
qb <- set_row_filters(qb, "strand == +")
get_row_filters(qb)
# row filters set using unevaluated expression
qb <- set_row_filters(qb, strand == "+")
get_row_filters(qb)

get_sids  Get or set query sample ids

Description
Get or set query sample ids

Usage
get_sids(qb)
set_sids(qb, sids)
**junction_inclusion_ratio**

**Arguments**

- qb: a QueryBuilder object constructed using the `QueryBuilder` function.
- sids: a vector or 1 or more whole numbers to filter results on.

**Value**

- `get_sids` returns the current sample ids as a vector of integers. `set_sids` returns a new `QueryBuilder` object with the sample ids set to the value of `sids`.

**Examples**

```r
qb <- QueryBuilder(compilation = "gtex", regions = "CD99")
qb <- set_sids(qb, c(1, 2, 3))
get_sids(qb)
```

---

**junction_inclusion_ratio**

*Relative measure of splice variant usage similar to PSI that allows for 2 arbitrarily defined groups of junctions (not limited to cassette exons).*

**Description**

Calculates a coverage summary statistic per sample of the normalized coverage difference between two sets of separate junctions defined by at least two basic queries and organized into two groups.

**Usage**

```r
junction_inclusion_ratio(group1, group2, group_names = NULL)
```

**Arguments**

- group1, group2: Each group is a list of 1 or more `QueryBuilder` objects
- group_names: Optional vector of strings representing the group names

**Details**

The summary statistic is as follows: If the coverage of the first group is "A" and the second is "B":

\[ \text{JIR}(A, B) = \frac{(A - B)}{(A + B + 1)} \]

This is calculated for every sample that occurs in one or the other (or both) groups results.

**Value**

A DataFrame of samples, with their JIR score and metadata, which had > 0 coverage in at least one resulting row in at least one of the groups.
junction_intersection

Examples

sb1 <- QueryBuilder(compilation = "srav2", regions = "chr2:29446395-30142858")
sb1 <- set_coordinate_modifier(sb1, Coordinates$Within)
sb1 <- set_row_filters(sb1, strand == "-")

sb2 <- QueryBuilder(compilation = "srav2", regions = "chr2:29416789-29446394")
sb2 <- set_coordinate_modifier(sb2, Coordinates$Within)
sb2 <- set_row_filters(sb2, strand == "-")

junction_inclusion_ratio(list(sb1), list(sb2))

junction_intersection(...) Get the intersection of junctions from 2 or more compilations which are on the same reference

Description

This function operates similar to the junction_union() function, i.e. it is cross compilation and merging of the same junction from multiple compilations will be handled exactly the same way. But instead of every junction which appears in at least one compilation, only the junctions which appear in every compilation will be returned.

Usage

junction_intersection(...)

Arguments

... One or more QueryBuilder objects

Value

A RangedSummarizedExperiment of junctions common across compilations

See Also

junction_union()

Examples

# Using query builder wrappers
sb1 <- QueryBuilder(compilation = "gtex", regions = "chr1:1879786-1879786")
sb1 <- set_coordinate_modifier(sb1, Coordinates$EndIsExactOrWithin)
sb1 <- set_row_filters(sb1, strand == "-")

sb2 <- QueryBuilder(compilation = "tcga", regions = "chr1:1879786-1879786")
sb2 <- set_coordinate_modifier(sb2, Coordinates$EndIsExactOrWithin)
sb2 <- set_row_filters(sb2, strand == "-")

junction_intersection(sb1, sb2)
junction_union

Get the union of junctions from 2 or more compilations which are on the same reference

Description
This function queries 2 or more compilations which are on the same reference version (e.g. hg38) and merges the resulting junctions into a single output table, unioning the sample coverage columns and the snaptron_id (jx ID) columns (the latter delimiter will be ":"). All sample IDs will be disjoint between compilations.

Usage
junction_union(...) 

Arguments
... One or more QueryBuilder objects

Details
Union is based on the following fields (combined into a comparison key):

• group
• chromosome
• start
• end
• strand

The goal is to have a single list of junctions where every junction occurs in at least one compilation and if a junction occurs in > 1 compilations it still only has a single row representing all the samples across compilations that it appears in. Sample aggregate statistics will be recalculated for junctions which are merged across all samples from all compilations:

• sample_count
• coverage_sum
• coverage_avg
• coverage_median

Value
A RangedSummarizedExperiment of junctions appearing in at least one compilation

See Also
junction_intersection()
Examples

# Using query builder wrappers
sb1 <- QueryBuilder(compilation = "gtex", regions = "chr1:1879786-1879786")
sb1 <- set_coordinate_modifier(sb1, Coordinates$EndIsExactOrWithin)
sb1 <- set_row_filters(sb1, strand == "-"

sb2 <- QueryBuilder(compilation = "tcga", regions = "chr1:1879786-1879786")
sb2 <- set_coordinate_modifier(sb2, Coordinates$EndIsExactOrWithin)
sb2 <- set_row_filters(sb2, strand == "-"

junction_union(sb1, sb2)

percent_spliced_in

Relative measure of splice variant usage, limited currently to cassette exon splice variants

Description

Similar to the JIR, this calculates Percent Spliced In (PSI) statistics for the definition of 2 different groups: inclusion and exclusion. Currently this function only supports the cassette exon use case.

Usage

percent_spliced_in(
inclusion_group1, inclusion_group2, exclusion_group,
min_count = 20,
group_names = NULL
)

Arguments

inclusion_group1, inclusion_group2, exclusion_group
  Where each is a list of 1 or more QueryBuilder objects
min_count    minimum total count (denominator) required to not be assigned -1
group_names  Optional vector of strings representing the group names

Details

Inclusion typically defines 2 basic queries, one for the junction preceding the cassette exon, and the second for the junction following the cassette exon. The exclusion group contains one basic query which defines the junction which skips the cassette exon.

The PSI itself is implemented as:

$$\text{PSI}(\text{inclusion1}, \text{inclusion2}, \text{exclusion}) = \frac{\text{mean}(\text{inclusion1}, \text{inclusion2})}{\text{mean}(\text{inclusion1}, \text{inclusion2}) + \text{exclusion}}$$

where each term denotes the coverage of junctions that resulted from the basic queries in that group in the current sample.
**Value**

A DataFrame of samples, with their PSI score and metadata, which had > 0 coverage in at least one resulting row in at least one of the groups.

**Examples**

```r
in1 <- QueryBuilder(compilation = "srav2", regions = "chr1:94468008-94472172")
in1 <- set_coordinate_modifier(in1, Coordinates$Exact)
in1 <- set_row_filters(in1, strand == "+")

in2 <- QueryBuilder(compilation = "srav2", regions = "chr1:94468008-94472172")
in2 <- set_coordinate_modifier(in2, Coordinates$Exact)
in2 <- set_row_filters(in2, strand == "+")

ex <- QueryBuilder(compilation = "srav2", regions = "chr1:94468008-94475142")
ex <- set_coordinate_modifier(ex, Coordinates$Exact)
ex <- set_row_filters(ex, strand == "+")

percent_spliced_in(list(in1), list(in2), list(ex))
```

---

**QueryBuilder**

*Construct a QueryBuilder object given a compilation and one or regions.*

**Description**

Construct a QueryBuilder object given a compilation and one or regions.

**Usage**

`QueryBuilder(compilation, regions)`

**Arguments**

- **compilation**: A single string containing the name of the Snaptron data source. Any variant of the Compilation enum can also be passed an argument.
- **regions**: Either a list of 1 more HUGO gene names as strings e.g. "BRCA1" or a Granges class object containing one or more genomic intervals (e.g. "chr1:1-1000").

**Value**

A QueryBuilder object.
Examples

```r
# construct a query builder for GTEX data source and BRCA1 gene
qb <- QueryBuilder(compilation = Compilation$gtex, regions = "BRCA1")

# construct a query builder for TCGA data source and chromosome region
qb <- QueryBuilder(compilation = "tcga", regions = "chr1:1-1000")

# construct a query builder for TCGA data source using GRanges object
library(GenomicRanges)
qb <- QueryBuilder(compilation = "tcga", regions = GRanges("chr1", "1-1000"))
```

**Description**

Given one or more gene names or genomic range intervals it will return a list of 0 or more genes, junctions, or exons (depending on which query form is used) which overlap the ranges.

**Usage**

```r
query_jx(sb, return_rse = TRUE, split_by_region = FALSE)
query_gene(sb, return_rse = TRUE, split_by_region = FALSE)
query_exon(sb, return_rse = TRUE, split_by_region = FALSE)
```

**Arguments**

- **sb** A SnaptronQueryBuilder object
- **return_rse** Should the query data be returned as a simple data frame or converted to a RangedSummarizedExperiment.
- **split_by_region** By default the results from multiple queries will be returned in a RangedSummarizedExperiment object with a rowData entry for each, labeling each result row according to the query it resulted from. However, if this is set to TRUE, the result will be a list of RangedSummarizedExperiment objects, one per original interval/gene. This latter option may be useful, but it requires a separate copy of the sample metadata for each original interval/gene.

**Value**

Functions will return either a RangedSummarizedExperiment or data.table depending on whether the return_rse parameter is set to TRUE or FALSE.
Examples

# Construct a QueryBuilder object
qb <- QueryBuilder(compilation = "gtex", regions = "chr1:1-100000")
qb <- set_row_filters(qb, samples_count >= 20)
query_jx(qb)

qb <- set_row_filters(qb, NULL)
qb <- set_column_filters(qb, SMTS == "Brain")
query_gene(qb)

shared_sample_counts

Shared Sample Count (SSC): counts total number of samples in which
2 different junctions both occur in.

Description

This produces a list of user-specified groups and the read coverage of the junctions in all the samples
which were shared across all the basic queries occurring in each group.

Usage

shared_sample_counts(..., group_names = NULL)

Arguments

... One or more lists of QueryBuilder objects

 group_names Optional vector of character strings representing group names

Details

Example: User defines a single group of junctions "GroupA" made up of 2 separate regions (two
basic queries).
An SSC query will return a single line for GroupA which will have the total number of samples
which had at least one junction which was returned from both basic queries. It will also report a
summary statistic of the total number of groups which had one or more samples that were shared
across the basic queries, in this case it would be 1. Also, it will report the number of groups which
had at least one shared sample and which had matching junctions (from the query) which were fully
annotated.

This function can be used to determine how much cross-sample support there is for a particular
junction configuration (typically a cassette exon).

Value

A DataFrame of results based on the list of groups passed in via "group_names". Each group
is reported with the # of unique samples which occurred in all of its defined set of related basic
queries (e.g. two inclusion basic queries in a cassette exon scenario).
tissue_specificity

Examples

g1 <- QueryBuilder(compilation = "gtex", regions = "chr1:1879786-1879786")
g1 <- set_coordinate_modifier(g1, Coordinates$EndIsExactOrWithin)
g1 <- set_row_filters(g1, strand == "-"")

g2 <- QueryBuilder(compilation = "gtex", regions = "chr1:1879903-1879903")
g2 <- set_coordinate_modifier(g2, Coordinates$StartIsExactOrWithin)
g2 <- set_row_filters(g2, strand == "-"")

ssc<-shared_sample_counts(list(g1, g2))

Description

Lists the number of samples labeled with a specific tissue type. Samples are filtered for ones which have junctions across all the user-specified groups. That is, if a sample only appears in the results of some of the groups (from their basic queries) it will be assigned a 0, otherwise if it is in all of the groups’ results it will be assigned a 1. This is similar to the SSC high level query type, but doesn’t sum the coverage.

Usage

tissue_specificity(..., group_names = NULL)

Arguments

... One or more QueryBuilder objects

group_names Optional vector of strings representing the group names

Details

The samples are then grouped by their tissue type (e.g. Brain). This is useful for determining if there’s an enrichment for a specific tissue in the set of junctions queried. Results from this can be fed to a statistical test, such as the Kruskal-wallis non-parametric rank test. This query is limited to GTEx only, due to the fact that GTEx is one of the few compilations that has consistent and complete tissue metadata.

Value

A DataFrame of all samples in the compilation with either a 0 or 1 indicating their occurrence and shared status (if > 1 group passed in). Occurrence here is if the sample has at least one result with > 0 coverage, and further, if > 1 group is passed in, then if it occurs in the results of all groups. Also includes the sample tissue type and sample_id.
Examples

```r
in1 <- QueryBuilder(compilation = "gtex", regions = "chr4:20763023-20763023")
in1 <- set_coordinate_modifier(in1, Coordinates$EndIsExactOrWithin)
in1 <- set_row_filters(in1, strand == "-")

in2 <- QueryBuilder(compilation = "gtex", regions = "chr4:20763098-20763098")
in2 <- set_coordinate_modifier(in2, Coordinates$StartIsExactOrWithin)
in2 <- set_row_filters(in2, strand == "-")

tissue_specificity(list(in1, in2))
```

---

**uri_of_last_successful_request**

*Return the URI of the last successful request to Snaptron*

---

**Description**

This function can be paired with the `from_url` method from the `QueryBuilder` class, allowing users to share sources of data from Snaptron.

**Usage**

```r
uri_of_last_successful_request()
```

**Value**

URI of last successful request to Snaptron or NULL if there have not been any successful requests.

**Examples**

```r
qb <- QueryBuilder(compilation = "gtex", regions = "CD99")
query_jx(qb)
uri_of_last_successful_request()
```
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