Package ‘spicyR’

April 4, 2024

Type Package

Title Spatial analysis of in situ cytometry data

Version 1.14.3

Description The spicyR package provides a framework for performing inference on changes in spatial relationships between pairs of cell types for cell-resolution spatial omics technologies. spicyR consists of three primary steps: (i) summarizing the degree of spatial localization between pairs of cell types for each image; (ii) modelling the variability in localization summary statistics as a function of cell counts and (iii) testing for changes in spatial localizations associated with a response variable.

License GPL (>=2)

LazyData true

biocViews SingleCell, CellBasedAssays, Spatial

Encoding UTF-8

Depends R (>= 4.1)

VignetteBuilder knitr

BugReports https://github.com/SydneyBioX/spicyR/issues

URL https://ellispatrick.github.io/spicyR/

https://github.com/SydneyBioX/spicyR

Imports ggplot2, concaveman, BiocParallel, spatstat.explore, spatstat.geom, lmerTest, BiocGenerics, S4Vectors, methods, mgcv, pheatmap, rlang, grDevices, IRanges, stats, data.table, dplyr, tidyr, scam, SingleCellExperiment, SpatialExperiment, SummarizedExperiment, ggforce, ClassifyR, tibble

Suggests BiocStyle, knitr, rmarkdown, pkgdown, imcRtools

RoxygenNote 7.2.3

git_url https://git.bioconductor.org/packages/spicyR

git_branch RELEASE_3_18

git_last_commit 79d608c

git_last_commit_date 2024-01-08
Accessors

Repository  Bioconductor 3.18
Date/Publication  2024-04-03
Author  Nicolas Canete [aut],
        Ellis Patrick [aut, cre]
Maintainer  Ellis Patrick <ellis.patrick@sydney.edu.au>

R topics documented:

Accessors

<table>
<thead>
<tr>
<th>Topic</th>
<th>Page</th>
</tr>
</thead>
<tbody>
<tr>
<td>as.data.frame.SegmentedCells</td>
<td>4</td>
</tr>
<tr>
<td>bind</td>
<td>5</td>
</tr>
<tr>
<td>colTest</td>
<td>6</td>
</tr>
<tr>
<td>convPairs</td>
<td>6</td>
</tr>
<tr>
<td>diabetesData</td>
<td>7</td>
</tr>
<tr>
<td>diabetesData_SCE</td>
<td>8</td>
</tr>
<tr>
<td>getPairwise</td>
<td>8</td>
</tr>
<tr>
<td>getProp</td>
<td>10</td>
</tr>
<tr>
<td>plot.SegmentedCells,ANY-method</td>
<td>10</td>
</tr>
<tr>
<td>SegmentedCells-class</td>
<td>11</td>
</tr>
<tr>
<td>show-SegmentedCells</td>
<td>13</td>
</tr>
<tr>
<td>signifPlot</td>
<td>14</td>
</tr>
<tr>
<td>spicyBoxPlot</td>
<td>15</td>
</tr>
<tr>
<td>SpicyResults-class</td>
<td>16</td>
</tr>
<tr>
<td>spicyTest</td>
<td>18</td>
</tr>
<tr>
<td>topPairs</td>
<td>18</td>
</tr>
</tbody>
</table>

Index  20

Accessors  Accessors for SegmentedCells

Description

Methods to access various components of the 'SegmentedCells' object.

Usage

```
cellSummary(x, imageID = NULL, bind = TRUE)

cellSummary(x, imageID = NULL) <- value

cellMarks(x, imageID = NULL, bind = TRUE)

cellMarks(x, imageID = NULL) <- value

cellMorph(x, imageID = NULL, bind = TRUE)
```
Accessors

\[
\begin{align*}
cellMorph(x, \text{imageID} = \text{NULL}) & \leftarrow \text{value} \\
imagePheno(x, \text{imageID} = \text{NULL}, \text{bind} = \text{TRUE}, \text{expand} = \text{FALSE}) & \leftarrow \text{value} \\
imagePheno(x, \text{imageID} = \text{NULL}) & \leftarrow \text{value} \\
imageID(x, \text{imageID} = \text{NULL}) & \\
cellID(x, \text{imageID} = \text{NULL}) & \\
cellID(x) & \leftarrow \text{value} \\
imageCellID(x, \text{imageID} = \text{NULL}) & \\
imageCellID(x) & \leftarrow \text{value} \\
cellType(x, \text{imageID} = \text{NULL}) & \\
cellType(x, \text{imageID} = \text{NULL}) & \leftarrow \text{value} \\
filterCells(x, \text{select}) & \\
cellAnnotation(x, \text{variable}, \text{imageID} = \text{NULL}) & \\
cellAnnotation(x, \text{variable}, \text{imageID} = \text{NULL}) & \leftarrow \text{value}
\end{align*}
\]

Arguments

- **x**: A ‘SegmentedCells’ object.
- **imageID**: A vector of imageIDs to specifically extract.
- **bind**: When false outputs a list of DataFrames split by imageID
- **expand**: Used to expand the phenotype information from per image to per cell.
- **value**: The relevant information used to replace.
- **select**: A logical vector of the cells to be kept.
- **variable**: A variable to add or retrieve from cellSummary.

Value

DataFrame or a list of DataFrames

Descriptions

- **‘cellSummary’**: Retrieves the DataFrame containing ‘x’ and ‘y’ coordinates of each cell as well as ‘cellID’, ‘imageID’ and ‘cellType’. imageID can be used to select specific images and bind=FALSE outputs the information as a list split by imageID.
- **‘cellMorph’**: Retrieves the DataFrame containing morphology information.
‘cellMarks‘: Retrieves the DataFrame containing intensity of gene or protein markers.
‘imagePheno‘: Retrieves the DataFrame containing the phenotype information for each image. Using expand = TRUE will produce a DataFrame with the number of rows equal to the number of cells.

Examples

### Something that resembles cellProfiler data

```r
set.seed(51773)

n = 10

cells <- data.frame(row.names = seq_len(n))
cells$ObjectNumber <- seq_len(n)
cells$ImageNumber <- rep(1:2, c(n/2, n/2))
cells$AreaShape_Center_X <- runif(n)
cells$AreaShape_Center_Y <- runif(n)
cells$AreaShape_round <- rexp(n)
cells$AreaShape_diameter <- rexp(n, 2)
cells$Intensity_Mean_CD8 <- rexp(n, 10)
cells$Intensity_Mean_CD4 <- rexp(n, 10)

cellExp <- SegmentedCells(cells, cellProfiler = TRUE)

### Cluster cell types

intensities <- cellMarks(cellExp)
kM <- kmeans(intensities, 2)
cellType(cellExp) <- paste('cluster', kM$cluster, sep = '')
cellSummary(cellExp, imageID = 1)
```

Description

Function to coerce a SegmentedCells object to a data frame.

Usage

```r
## S3 method for class 'SegmentedCells'
as.data.frame(x, ...)
```

Arguments

- x: A SegmentedCells object.
- ...: Other arguments.
Value

A data.frame

## Generate toy data set.seed(51773) x <- round(c(runif(200),runif(200)+1,runif(200)+2,runif(200)+3, runif(200)+3,runif(200)+2,runif(200)+1,runif(200)),4) y <- round(c(runif(200),runif(200)+1,runif(200)+2,runif(200)+3, runif(200),runif(200)+1,runif(200)+2,runif(200)+3,4) cellType <- factor(paste('c',rep(c(1:2),rep(200,2)),4),sep = "") imageID <- rep(c('s1', 's2'),c(800,800)) cells <- data.frame(x, y, cellType, imageID)

## Store data in SegmentedCells object cellExp <- SegmentedCells(cells, cellTypeString = 'cell-Type')

## Generate LISA cellsDF <- as.data.frame(cellExp)

NULL

---

bind Produces a dataframe showing L-function metric for each imageID entry.

Description

Produces a dataframe showing L-function metric for each imageID entry.

Usage

bind(results, pairName = NULL)

Arguments

results Spicy test result obtained from spicy.

pairName A string specifying the pairwise interaction of interest. If NULL, all pairwise interactions are shown.

Value

A data.frame containing the colData related to the results.

Examples

data(spicyTest)
df <- bind(spicyTest)
**colTest**

*Perform a simple wilcoxon-rank-sum test or t-test on the columns of a data frame*

**Description**

Perform a simple wilcoxon-rank-sum test or t-test on the columns of a data frame

**Usage**

```
colTest(df, condition, type = NULL, feature = NULL, imageID = "imageID")
```

**Arguments**

- `df`: A data.frame or SingleCellExperiment, SpatialExperiment
- `condition`: The condition of interest
- `type`: The type of test, "wilcox", "ttest" or "survival".
- `feature`: Can be used to calculate the proportions of this feature for each image
- `imageID`: The imageID’s if presenting a SingleCellExperiment

**Value**

Proportions

**Examples**

```r
# Test for an association with long-duration diabetes
# This is clearly ignoring the repeated measures...
data("diabetesData")
props <- getProp(diabetesData)
condition <- imagePheno(diabetesData)$stage
names(condition) <- imagePheno(diabetesData)$imageID
condition <- condition[condition %in% c("Long-duration", "Onset")]
test <- colTest(props[names(condition), ], condition)
```

---

**convPairs**

*Converts colPairs object into an abundance matrix based on number of nearby interactions for every cell type.*

**Description**

Converts colPairs object into an abundance matrix based on number of nearby interactions for every cell type.
**Usage**

`convPairs(cells, colPair, cellType = "cellType", imageID = "imageID")`

**Arguments**

- **cells**: A `SingleCellExperiment` that contains objects in the `colPairs` slot.
- **colPair**: The name of the object in the `colPairs` slot for which the dataframe is constructed from.
- **cellType**: The cell type if using `SingleCellExperiment`.
- **imageID**: The image ID if using `SingleCellExperiment`.

**Value**

Matrix of abundances

**Examples**

```r
data("diabetesData_SCE")

diabetesData_SPE <- SpatialExperiment::SpatialExperiment(diabetesData_SCE, 
  colData = SingleCellExperiment::colData(diabetesData_SCE))
SpatialExperiment::spatialCoords(diabetesData_SPE) <- data.frame(
  SingleCellExperiment::colData(diabetesData_SPE)$x, 
  SingleCellExperiment::colData(diabetesData_SPE)$y) |> 
  as.matrix()

SpatialExperiment::spatialCoordsNames(diabetesData_SPE) <- c("x", "y")

diabetesData_SPE <- imcRtools::buildSpatialGraph(diabetesData_SPE, 
  img_id = "imageID", 
  type = "knn", 
  k = 20, 
  coords = c("x", "y")

pairAbundances <- convPairs(diabetesData_SPE, 
  colPair = "knn_interaction_graph")
```

---

**diabetesData**

*Diabetes IMC data*

---

**Description**

This is a subset of the Damond et al 2019 imaging mass cytometry dataset. The data contains cells in the pancreatic islets of individuals with early onset diabetes and healthy controls. The object contains single-cell data of 160 images from 8 subjects, with 20 images per subject.
getPairwise

Usage

diabetesData

Format

diabetesData a SegmentedCells object

diabetesData_SCE Diabetes IMC data in SCE format.

Description

This is a subset of the Damond et al 2019 imaging mass cytometry dataset. The data contains cells in the pancreatic islets of individuals with early onset diabetes and healthy controls. The object contains single-cell data of 160 images from 8 subjects, with 20 images per subject.

Usage

diabetesData_SCE

Format

diabetesData_SCE a SingleCellExperiment object

Details

Converted into a SingleCellExperiment format.

getPairwise Get statistic from pairwise L curve of a single image.

Description

Get statistic from pairwise L curve of a single image.

Usage

getPairwise(
cells,
from = NULL,
to = NULL,
window = "convex",
window.length = NULL,
Rs = c(20, 50, 100),
sigma = NULL,
getPairwise = 0.05,
edgeCorrect = TRUE,
includeZeroCells = TRUE,
BPPARAM = BiocParallel::SerialParam(),
imageID = "imageID",
cellType = "cellType",
spatialCoords = c("x", "y")
)

Arguments

cells A SegmentedCells or data frame that contains at least the variables x and y, giving the location coordinates of each cell, and cellType.

from The "from" cellType for generating the L curve.

to The "to" cellType for generating the L curve.

window Should the window around the regions be 'square', 'convex' or 'concave'.

window.length A tuning parameter for controlling the level of concavity when estimating concave windows.

Rs A vector of the radii that the measures of association should be calculated.

sigma A numeric variable used for scaling when fitting inhomogeneous L-curves.

minLambda Minimum value for density for scaling when fitting inhomogeneous L-curves.

edgeCorrect A logical indicating whether to perform edge correction.

includeZeroCells A logical indicating whether to include cells with zero counts in the pairwise association calculation.

BPPARAM A BiocParallelParam object.

imageID The imageID if using a SingleCellExperiment or SpatialExperiment.

cellType The cellType if using a SingleCellExperiment or SpatialExperiment.

spatialCoords The spatialCoords if using a SingleCellExperiment or SpatialExperiment.

Value

Statistic from pairwise L curve of a single image.

Examples

data("diabetesData")
pairAssoc <- getPairwise(diabetesData[1, 1])
### getProp

*Get proportions from a SegmentedCells, SingleCellExperiment, SpatialExperiment or data.frame.*

**Description**

Get proportions from a SegmentedCells, SingleCellExperiment, SpatialExperiment or data.frame.

**Usage**

```r
getProp(cells, feature = "cellType", imageID = "imageID")
```

**Arguments**

- **cells**
  - SegmentedCells, SingleCellExperiment, SpatialExperiment or data.frame
- **feature**
  - The feature of interest
- **imageID**
  - The imageID’s

**Value**

Proportions

**Examples**

```r
data("diabetesData")
prop <- getProp(diabetesData)
```

### plot,SegmentedCells,ANY-method

*A basic plot for SegmentedCells object*

**Description**

This function generates a basic x-y plot of the location coordinates and cellType data.

**Usage**

```r
## S4 method for signature 'SegmentedCells,ANY'
plot(x, imageID = NULL)
```

**Arguments**

- **x**
  - A SegmentedCells object.
- **imageID**
  - The image that should be plotted.
SegmentedCells-class

Value

A ggplot object.

usage

'plot(x, imageID = NULL)'

Examples

### Something that resembles cellProfiler data

set.seed(51773)

n = 10

cells <- data.frame(row.names = seq_len(n))
cells$ObjectNumber <- seq_len(n)
cells$ImageNumber <- rep(1:2, c(n/2, n/2))
cells$AreaShape_Center_X <- runif(n)
cells$AreaShape_Center_Y <- runif(n)
cells$AreaShape_round <- rexp(n)
cells$AreaShape_diameter <- rexp(n, 2)
cells$Intensity_Mean_CD8 <- rexp(n, 10)
cells$Intensity_Mean_CD4 <- rexp(n, 10)

cellExp <- SegmentedCells(cells, cellProfiler = TRUE)

### Cluster cell types

markers <- cellMarks(cellExp)
kM <- kmeans(markers, 2)
cellType(cellExp) <- paste('cluster', kM$cluster, sep = '')

#plot(cellExp, imageID=1)

---

SegmentedCells-class  The SegmentedCells class

Description

The SegmentedCells S4 class is for storing data from segmented imaging cytometry and spatial omics data. It extends DataFrame and defines methods that take advantage of DataFrame nesting to represent elements of cell-based experiments with spatial orientation that are commonly encountered. This object is able to store information on a cell’s spatial location, cellType, morphology, intensity of gene/protein markers as well as image level phenotype information.
Usage

```r
SegmentedCells(
  cellData,
  cellProfiler = FALSE,
  spatialCoords = c("x", "y"),
  cellTypeString = "cellType",
  intensityString = "intensity_",
  morphologyString = "morphology_",
  phenotypeString = "phenotype_",
  cellIDString = "cellID",
  cellAnnotations = NULL,
  imageCellIDString = "imageCellID",
  imageIDString = "imageID",
  verbose = TRUE
)
```

Arguments

- **cellData**: A data frame that contains at least the columns `x` and `y` giving the location coordinates of each cell.
- **cellProfiler**: A logical indicating that `cellData` is in a format similar to what `cellProfiler` outputs.
- **spatialCoords**: The column names corresponding to spatial coordinates. eg. `x`, `y`, `z`...
- **cellTypeString**: The name of the column that contains cell type calls.
- **intensityString**: A string which can be used to identify the columns which contain marker intensities. (This needs to be extended to take the column names themselves.)
- **morphologyString**: A string which can be used to identify the columns which contains morphology information.
- **phenotypeString**: A string which can be used to identify the columns which contains phenotype information.
- **cellIDString**: The column name for `cellID`.
- **cellAnnotations**: A vector of variables that provide additional annotation of a cell.
- **imageCellIDString**: The column name for `imageCellID`.
- **imageIDString**: The column name for `imageID`.
- **verbose**: logical indicating whether to output messages.

Value

A `SegmentedCells` object
Examples

### Something that resembles cellProfiler data

```r
set.seed(51773)

n = 10

cells <- data.frame(row.names = seq_len(n))
cells$ObjectNumber <- seq_len(n)
cells$ImageNumber <- rep(seq_len(2),c(n/2,n/2))
cells$AreaShape_Center_X <- runif(n)
cells$AreaShape_Center_Y <- runif(n)
cells$AreaShape_round <- rexp(n)
cells$AreaShape_diameter <- rexp(n, 2)
cells$Intensity_Mean_CD8 <- rexp(n, 10)
cells$Intensity_Mean_CD4 <- rexp(n, 10)

cellExp <- SegmentedCells(cells, cellProfiler = TRUE)

### Cluster cell types
intensities <- cellMarks(cellExp)
kM <- kmeans(intensities,2)
cellType(cellExp) <- paste('cluster',kM$cluster, sep = '')
cellSummary(cellExp)
```

show-SegmentedCells  Show SegmentedCells

Description

This outputs critical information about a SegmentedCells.

Arguments

object  A SegmentedCells.

Value

Information of the number of images, cells, intenisties, morphologies and phenotypes.

usage

‘show(object)’
### Examples

```
### Something that resembles cellProfiler data

set.seed(51773)

n = 10

cells <- data.frame(row.names = seq_len(n))
cells$ObjectNumber <- seq_len(n)
cells$ImageNumber <- rep(1:2, c(n/2, n/2))
cells$AreaShape_Center_X <- runif(n)
cells$AreaShape_Center_Y <- runif(n)
cells$AreaShape_round <- rexp(n)
cells$AreaShape_diameter <- rexp(n, 2)
cells$Intensity_Mean_CD8 <- rexp(n, 10)
cells$Intensity_Mean_CD4 <- rexp(n, 10)

cellExp <- SegmentedCells(cells, cellProfiler = TRUE)

### Cluster cell types

markers <- cellMarks(cellExp)
kM <- kmeans(markers, 2)
cellType(cellExp) <- paste('cluster', kM$cluster, sep = '')

cellExp
```

---

**signifPlot**

Plots result of signifPlot.

**Description**

Plots result of signifPlot.

**Usage**

```
signifPlot(
  results,
  fdr = FALSE,
  type = "bubble",
  breaks = NULL,
  comparisonGroup = NULL,
  colours = c("#4575B4", "white", "#D73027"),
  marksToPlot = NULL,
  cutoff = 0.05
)
```
spicyBoxPlot

Arguments

results Data frame obtained from spicy.
fdr TRUE if FDR correction is used.
type Where to make a bubble plot or heatmap.
breaks Vector of 3 numbers giving breaks used in pheatmap. The first number is the minimum, the second is the maximum, the third is the number of breaks.

comparisonGroup A string specifying the name of the outcome group to compare with the base group.
colours Vector of colours to use in pheatmap.
marksToPlot Vector of marks to include in pheatmap.
cutoff significance threshold for circles in bubble plot

Value

a pheatmap object

Examples

data(spicyTest)
signifPlot(spicyTest, breaks = c(-3, 3, 0.5))

spicyBoxPlot Plots boxplot for a specified cell-cell relationship

Description

Plots boxplot for a specified cell-cell relationship

Usage

spicyBoxPlot(results, from = NULL, to = NULL, rank = NULL)

Arguments

results Data frame obtained from spicy.
from Cell type which you would like to compare to the to cell type.
to Cell type which you would like to compare to the from cell type.
rank Ranking of cell type in terms of p-value, the smaller the p-value the higher the rank.

Value

a ggplot2 boxplot
SpicyResults-class

Performs spatial tests on spatial cytometry data.

Description

Performs spatial tests on spatial cytometry data.

Usage

spicy(
cells,
condition = NULL,
subject = NULL,
covariates = NULL,
from = NULL,
to = NULL,
alternateResult = NULL,
verbose = TRUE,
weights = TRUE,
weightsByPair = FALSE,
weightFactor = 1,
window = "convex",
window.length = NULL,
BPPARAM = BiocParallel::SerialParam(),
sigma = NULL,
Rs = NULL,
minLambda = 0.05,
edgeCorrect = TRUE,
includeZeroCells = FALSE,
imageID = "imageID",
cellType = "cellType",
spatialCoords = c("x", "y"),
...
)

Arguments

cells A SegmentedCells or data frame that contains at least the variables x and y, giving the location coordinates of each cell, and cellType.
condition  Vector of conditions to be tested corresponding to each image if cells is a data frame.
subject    Vector of subject IDs corresponding to each image if cells is a data frame.
covariates Vector of covariate names that should be included in the mixed effects model as fixed effects.
from       vector of cell types which you would like to compare to the to vector
to         vector of cell types which you would like to compare to the from vector
alternateResult An pairwise association statistic between each combination of cell types in each image.
verbose    logical indicating whether to output messages.
weights    logical indicating whether to include weights based on cell counts.
weightsByPair logical indicating whether weights should be calculated for each cell type pair.
weightFactor numeric that controls the convexity of the weight function.
window     Should the window around the regions be 'square', 'convex' or 'concave'.
window.length A tuning parameter for controlling the level of concavity when estimating concave windows.
BPPARAM    A BiocParallelParam object.
sigma      A numeric variable used for scaling when fitting inhomogeneous L-curves.
Rs          A vector of radii that the measures of association should be calculated.
minLambda   Minimum value for density for scaling when fitting inhomogeneous L-curves.
edgeCorrect A logical indicating whether to perform edge correction.
includeZeroCells A logical indicating whether to include cells with zero counts in the pairwise association calculation.
imageID     The image ID if using SingleCellExperiment.
cellType    The cell type if using SingleCellExperiment.
spatialCoords The spatial coordinates if using a SingleCellExperiment.
...         Other options.

Value
Data frame of p-values.

Examples

data("diabetesData")

# Test with random effect for patient on a pairwise combination of cell types.
spicy(diabetesData, 
  condition = "stage", subject = "case", 
  from = "Tc", to = "Th"
)
# Test all pairwise combinations of cell types without random effect of patient.
# spicyTest <- spicy(diabetesData, condition = "stage", subject = "case")

# Test all pairwise combination of cell types with random effect of patient.
# spicy(diabetesData, condition = "condition", subject = "subject")

---

### spicyTest

**Results from spicy for diabetesData**

**Description**

Results from the call: `spicyTest <- spicy(diabetesData, condition = "condition", subject = "subject")`

**Usage**

`spicyTest`

**Format**

`spicyTest` a spicy object

---

### topPairs

**A table of the significant results from spicy tests**

**Description**

A table of the significant results from spicy tests

**Usage**

`topPairs(x, coef = NULL, n = 10, adj = "fdr", cutoff = NULL, figures = NULL)`

**Arguments**

- `x` The output from spicy.
- `coef` Which coefficient to list.
- `n` Extract the top n most significant pairs.
- `adj` Which p-value adjustment method to use, argument for p.adjust().
- `cutoff` A p-value threshold to extract significant pairs.
- `figures` Round to ‘figures’ significant figures.
Value

A data.frame

Examples

data(spicyTest)
topPairs(spicyTest)
Index

* datasets
  - diabetesData, 7
  - diabetesData_SCE, 8
  - spicyTest, 18

Accessors, 2
as.data.frame.SegmentedCells, 4
bind, 5

cellAnnotation (Accessors), 2
cellAnnotation,SegmentedCells-method (Accessors), 2
cellAnnotation<-,SegmentedCells-method (Accessors), 2
cellID (Accessors), 2
cellID,SegmentedCells-method (Accessors), 2
cellID<-, (Accessors), 2
cellMarks (Accessors), 2
cellMarks,SegmentedCells-method (Accessors), 2
cellMarks<-, (Accessors), 2
cellMarks<-,SegmentedCells-method (Accessors), 2
cellMorph (Accessors), 2
cellMorph,SegmentedCells-method (Accessors), 2
cellMorph<-, (Accessors), 2
cellMorph<-,SegmentedCells-method (Accessors), 2
cellType (Accessors), 2
cellType,SegmentedCells-method (Accessors), 2
cellType<-, (Accessors), 2
cellType<-,SegmentedCells-method (Accessors), 2
colTest, 6
cnvPairs, 6
diabetesData, 7
diabetesData_SCE, 8
filterCells (Accessors), 2
filterCells,SegmentedCells-method (Accessors), 2
getPairwise, 8
getProp, 10
imageCellID (Accessors), 2
imageCellID,SegmentedCells-method (Accessors), 2
imageCellID<-, (Accessors), 2
imageCellID<-,SegmentedCells-method (Accessors), 2
imageID (Accessors), 2
imageID,SegmentedCells-method (Accessors), 2
imagePheno (Accessors), 2
imagePheno,SegmentedCells-method (Accessors), 2
imagePheno<-, (Accessors), 2
imagePheno<-,SegmentedCells-method (Accessors), 2
plot (plot,SegmentedCells,ANY-method), 10
plot,SegmentedCells (plot,SegmentedCells,ANY-method), 10
plot,SegmentedCells,ANY-method, 10
SegmentedCells (SegmentedCells-class), 11
SegmentedCells,SegmentedCells-method (SegmentedCells-class), 11
SegmentedCells-class, 11
show (show-SegmentedCells), 13
show-SegmentedCells, 13
signifPlot, 14
spicy (SpicyResults-class), 16
spicy,spicy-method (SpicyResults-class), 16
spicyBoxPlot, 15
SpicyResults,list,ANY-method (SpicyResults-class), 16
SpicyResults-class, 16
spicyTest, 18
topPairs, 18
topPairs,SpicyResults-method (topPairs), 18