Package ‘trigger’

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Imports qvalue, methods, graphics, sva

Description This R package provides tools for the statistical analysis of integrative genomic data that involve some combination of: genotypes, high-dimensional intermediate traits (e.g., gene expression, protein abundance), and higher-order traits (phenotypes). The package includes functions to: (1) construct global linkage maps between genetic markers and gene expression; (2) analyze multiple-locus linkage (epistasis) for gene expression; (3) quantify the proportion of genome-wide variation explained by each locus and identify eQTL hotspots; (4) estimate pair-wise causal gene regulatory probabilities and construct gene regulatory networks; and (5) identify causal genes for a quantitative trait of interest.

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plot

Description

Graphical display of genomewide linkage map, multi-locus linkage or eQTL variation

Usage

```r
## S4 method for signature 'trigger,missing'
plot(x, y, type = c("link", "mlink", "eqtl"),
cutoff = 3.3e-4, qcut = 0.1, bin.size = NULL)
```

Arguments

- `x`: An object of class `trigger`.
- `y`: Ignore option, not used.
- `type`: An argument describing the type of plot. Select from `link` (default) for genomewide linkage map, `eqtl.R2` for graphical display of eQTL-R^2 contribution or `mlink` for display of genome-wide epistasis effect.
- `cutoff`: Threshold value for `link`. The measures below the threshold are called significant and are plotted.
- `qcut`: Q-value threshold for `mlink`. The joint multi-locus linkage probabilities with q-values below the threshold are called significant and are plotted.
- `bin.size`: Optional for `mlink`. If not `NULL`, each chromosome will be divided into several bins, each with size `bin.size`. Markers within a bin will be considered as at a same position.
trigger-class

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See Also
trigger.link, trigger.mlink and trigger.eigenR2

Examples

```r
## Not run:
data(yeast)
attach(yeast)
triggerobj <- trigger.build(marker=marker, exp=exp,
marker.pos=marker.pos, exp.pos=exp.pos)
triggerobj <- trigger.link(triggerobj, gender=NULL, norm=TRUE)
plot(triggerobj, type = "link", cutoff=1e-5)
triggerobj <- trigger.eigenR2(triggerobj, adjust=FALSE)
plot(triggerobj, type = "eigenR2")
triggerobj<- trigger.mlink(triggerobj, B=5, seed=123)
plot(triggerobj, qcut=0.1, bin.size=NULL)
detach(yeast)

## End(Not run)
```

trigger-class

A class to store and analyze data for Transcriptional Regulation Inference from Genetics of Gene Expression

Description

trigger is a class of objects to store and analyze data for Integrative Genomic Analysis. Use trigger.build to generate new objects of the class from input data.

Details

The positions in marker-pos and exp-pos matrix should be in the same units (e.g., base pair, kb, or cM).

Value

An object of S4 class trigger containing the marker genotype matrix (a matrix of 1,2 for haploid genotypes, or 1,2,3 for diploid genotypes), expression matrix, marker position matrix and gene/trait position matrix with ordered coordinates in respective slots. Use slot(objectname, varname) to retrieve individual variables from the object. Use print to see the first 10 rows and columns of the expression and marker matrix.
**Slots**

- `exp`: A numeric matrix with $m$ rows and $n$ columns, containing the gene expression (or intermediate trait) data.
- `exp.pos`: A matrix with $m$ rows and 3 columns containing the chromosome number, gene start and gene end for all the genes in the gene expression matrix. The rows of `exp.pos` should match those of `exp`.
- `marker`: A matrix with $p$ rows and $n$ columns, containing genotyping information.
- `marker.pos`: A matrix with $p$ rows and 2 columns containing the chromosome number and SNP position for all the genes in the gene expression matrix. The rows of `exp.pos` should match those of `exp`.
- `stat`: A matrix of pair-wise likelihood ratio statistics for linkage analysis, with genes in rows and markers in columns.
- `pvalue`: A matrix of parametric pvalues corresponding to statistics in the `stat` matrix.
- `mlink`: A list containing the results of Multi-locus linkage analysis. See `trigger.mlink` for details.
- `eqtl.R2`: A vector containing the proportion of genome-wide variation explained by each observed locus (eQTL). See `trigger.eigenR2` for details.
- `loc.obj`: A list containing the results of local-linkage probability estimation. See `trigger.loclink` for details.

**Author(s)**

Lin S. Chen <lschen.stat@gmail.com>, Dipen P. Sangurdekar <dps@genomics.princeton.edu> and John D. Storey <jstorey@princeton.edu>

**See Also**

`trigger.build`, `trigger.link`, `trigger.mlink`, `trigger.eigenR2`, `trigger.net` and `trigger.trait`

---

**trigger.build**

*Format the input data and create an Trigger object*

**Description**

This function takes high-dimensional expression data and genotype data with each of their position data in the genome and creates a `trigger` object for subsequent analysis.

**Usage**

```r
trigger.build(exp = exp, exp.pos = exp.pos, marker = marker, marker.pos = marker.pos)
```
**Arguments**

- **exp**
  A gene (or intermediate trait) by individual matrix of expression data.

- **exp.pos**
  A matrix containing the position information for genes (intermediate traits). The first column is the chromosome name of the gene. The second column is the starting coordinate of the gene, and the third column is the ending coordinate. Each row corresponds to one gene/trait in the exp matrix.

- **marker**
  A marker genotype by individual matrix.

- **marker.pos**
  A matrix containing the position information for markers. The first column is the chromosome name of the marker. We recommend to use integers for autosomal chromosomes and "X" for sex chromosome. The second column is the position of the marker on the chromosome. Each row corresponds to one marker in the marker matrix.

**Details**

The positions in marker.pos and exp.pos matrix should be in the same units (e.g., base pair, kb, or cM).

**Value**

An object of S4 class `trigger` containing the marker genotype matrix (a matrix of 1,2 for haploid genotypes, or 1,2,3 for diploid genotypes), expression matrix, marker position matrix and gene/trait position matrix with ordered coordinates in respective slots. Use `slot(objectname, varname)` to retrieve individual variables from the object. Use `print` to see the first 10 rows and columns of the expression and marker matrix.

**Author(s)**

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**See Also**

`trigger.link`, `trigger.mlink`, `trigger.eigenR2`, `trigger.net` and `trigger.trait`

**Examples**

```r
## Not run:
data(yeast)
attach(yeast)
triggerobj <- trigger.build(marker = marker, exp = exp,
                             marker.pos = marker.pos, exp.pos = exp.pos)
print(triggerobj)
## End(Not run)
```
trigger.eigenR2-methods

Estimate the proportion of genome-wide variation explained by each eQTL.

Description

Estimate eqtl-R2, the proportion of genome-wide variation explained by each eQTL and identify linkage hotspots.

Usage

```r
## S4 method for signature 'trigger'
trigger.eigenR2(triggerobj, adjust = FALSE, meanR2 = FALSE)
```

Arguments

- `triggerobj`: An object of class `trigger`.
- `adjust`: Logical. If TRUE, the estimated R-square for each locus will be adjusted for small sample size effect. Recommend to use when sample size is less than 100.
- `meanR2`: Logical. If TRUE, the function computes the mean of R-squares of genome-wide gene expression for each locus.

Value

An updated object of class `trigger` with a slot `loc.obj` containing the proportion of genome-wide variation explained by each observed locus (eQTL). Use `slot(triggerobj, "eigenR2")` to retrieve the eqtl-R2 values as a vector.

Author(s)

Lin S. Chen <lschen.stat@gmail.com>, Dipen P. Sangurdekar <dps@genomics.princeton.edu> and John D. Storey <jstorey@princeton.edu>

References


See Also

plot
Examples

```r
## Not run:
data(yeast)
attach(yeast)
triggerobj <- trigger.build(marker = marker, exp = exp,
marker.pos = marker.pos, exp.pos = exp.pos)
triggerobj <- trigger.eigenR2(triggerobj, adjust = FALSE)
plot(triggerobj, type = "eigenR2")
eqtlR2 <- slot(triggerobj, "eigenR2")
detach(yeast)
```

## End(Not run)

trigger.export2cross-methods

Export Trigger data to R/qtl's cross class object

Description

trigger.export2cross exports trigger data from triggerobj to a cross format for Trait-Trigger analysis. See trigger.trait for details.

Usage

```r
## S4 method for signature 'trigger'
trigger.export2cross(triggerobj, plotarg = TRUE, verbose = TRUE, warning = FALSE)
```

Arguments

- `triggerobj`: An object of class trigger.
- `plotarg`: Logical. If TRUE, the function plots the default plot from the R/qtl package while reading in the genotype data.
- `verbose`: Logical. If TRUE, the function lists the default output from the R/qtl package while reading in the genotype data.
- `warning`: Logical. If FALSE, the function suppresses warnings output from the R/qtl package while reading in the genotype data.

Details

The trigger.export2cross command writes a csv format file "geno_trait_data.csv" to the working directory and reads it using the read.cross command.

Value

An object of class cross from the R/qtl package.
**Author(s)**

Lin S. Chen <lschen.stat@gmail.com>, Dipen P. Sangurdekar <dps@genomics.princeton.edu> and John D. Storey <jstorey@princeton.edu>

**References**


**See Also**

`trigger.trait`

**Examples**

```r
data(yeast)
attach(yeast)
triggerobj <- trigger.build(marker = marker, exp = exp,
marker.pos = marker.pos, exp.pos = exp.pos)
crossfile <- trigger.export2cross(triggerobj, plotarg = TRUE, verbose = TRUE, warning = FALSE)
tt.pval <- trigger.trait(triggerobj, trait = "DSE1", cross = crossfile)
causal.reg <- names(which(p.adjust(tt.pval, method = "fdr")<.05))
detach(yeast)
```

---

**Description**

A method of class `trigger` for genomewide Expression-trait QTL analysis. This function estimates the linkage statistic and parametric p-value for each gene expression to every locus in the genome.

**Usage**

```r
## S4 method for signature 'trigger'
trigger.link(triggerobj, gender = NULL, norm = TRUE)
```

**Arguments**

- `triggerobj` An object of class `trigger`.
- `gender` Optional. When computing linkage statistics involving markers on sex chromosome, gender of each sample should be specified.
- `norm` Logical. If TRUE, each row of expression matrix `exp` in the `triggerobj` will be transformed to follow a standard normal distribution, based on the rank of value.
Value

An updated object of class trigger containing slots:

- **stat**: A matrix of pair-wise likelihood ratio statistics for linkage analysis, with genes in rows and markers in columns.
- **pvalue**: A matrix of parametric pvalues corresponding to statistics in the stat matrix.

Use `slot(triggerobj, "stat")` and `slot(triggerobj, "pvalue")` to retrieve the values.

Author(s)

Lin S. Chen <lschen.stat@gmail.com>, Dipen P. Sangurdekar <dps@genomics.princeton.edu> and John D. Storey <jstorey@princeton.edu>

See Also

- `plot` and `trigger.mlink`

Examples

```r
# Not run:
data(yeast)
attach(yeast)
triggerobj <- trigger.build(marker = marker, exp = exp,
  marker.pos = marker.pos, exp.pos = exp.pos)
triggerobj <- trigger.link(triggerobj, gender = NULL, norm = TRUE)
plot(triggerobj, type = "link", cutoff = 1e-5)
stat = slot(triggerobj, "stat"); pvalue = slot(triggerobj, "pvalue")
detach(yeast)

# End(Not run)
```

---

**Description**

A method of class **trigger** to identify the best local-linkage marker for each gene and compute the local linkage probabilities.

**Usage**

```r
# S4 method for signature 'trigger'
trigger.loclink(triggerobj, gender = NULL, window.size = 30000)
```
Arguments

triggrobject An object of class trigger.
gender Optional. When computing linkage statistics involving markers on sex chromosome, gender of each sample should be specified.
window.size Optional. The size of a window that places the putative regulator gene in the center. Every marker within the window is a candidate marker for local-linkage to the regulator gene.

Value

An updated object of class trigger containing a slot loc.obj with fields:

prob.loc The estimated local-linkage probability for each putative regulator gene.
loc.idx The indices of the best local marker for each putative regulator gene.

Use slot(triggerobj, "loc.obj") to retrieve the list.

Author(s)

Lin S. Chen <lschen.stat@gmail.com>, Dipen P. Sangurdekar <dps@genomics.princeton.edu> and John D. Storey <jstorey@princeton.edu>

References


See Also

trigger.trait

Examples

```r
## Not run:
data(yeast)
attach(yeast)
triggrobject <- trigger.build(marker = marker, exp = exp,
    marker.pos = marker.pos, exp.pos = exp.pos)
triggrobject <- trigger.loclink(triggrobject, window.size = 30000)
triggrobject <- trigger.net(triggrobject, Bsec = 100)
detach(yeast)

## End(Not run)
```
Multi-Locus Linkage (Epistasis) Analysis

Description

Multi-locus linkage (epistasis) analysis.

Usage

```r
## S4 method for signature 'trigger'
trigger.mlink(triggerobj, prob.cut = 0.9,
             gender = NULL, idx = NULL, B = 5, seed = 123)
```

Arguments

- `triggerobj`: An object of class `trigger`.
- `prob.cut`: Probability threshold for primary linkage.
- `gender`: Optional. When computing linkage statistics involving markers on sex chromosome, gender of each sample should be specified.
- `idx`: The indices for genes to be computed for multi-locus linkage.
- `B`: The number of null iterations to perform.
- `seed`: Optional. A numeric seed for reproducible results.

Details

When data set is large, one can the option `idx` to select a subset of genes in each computation and parallel-computes the genome-wide multi-locus linkage. Since the function computes the linkage probability by borrowing information across genes, at least more than 100 genes should be selected in applying this function. If `idx=NULL`, all the genes in the input data will be computed for multi-locus linkage.

The current version of the function could only compute two-locus joint linkage (epistasis).

Value

An updated object of class `trigger` containing a slot `trigger.mlink` with fields:

- `qtl`: The major and secondary QTLs for each selected gene.
- `prob`: The posterior probability of linkage for major QTL, secondary QTL, and the joint posterior probability of multi-locus linkage.
- `qvalue`: Q-value estimates for joint multi-locus linkage probabilities.

Use `slot(triggerobj, "mlink")` to retrieve the list.

Author(s)

Lin S. Chen <lschen.stat@gmail.com>, Dipen P. Sangurdekar <dps@genomics.princeton.edu> and John D. Storey <jstorey@princeton.edu>
References


See Also

*trigger.link* and *plot*

Examples

```r
## Not run:
data(yeast)
attach(yeast)
triggerobj <- trigger.build(marker = marker, exp = exp,
marker.pos = marker.pos, exp.pos = exp.pos)
## Genome-wide multiple locus linkage analysis
triggerobj <- trigger.mlink(triggerobj, B = 10, idx = NULL, seed = 123)

plot(triggerobj, type = "trigger.mlink", qcut=0.1, bin.size=NULL)
mlink = slot(triggerobj, "trigger.mlink")
detach(yeast)
## End(Not run)
```

### trigger.net-methods

**Network-Trigger analysis**

**Description**

Network-Trigger analysis estimates the joint posterior probability of causal regulation for each pair of genes in the genome. These probabilities can further be used to construct a gene regulatory network.

**Usage**

```r
## S4 method for signature 'trigger'
trigger.net(triggerobj, gender = NULL, idx = NULL,
Bsec = 100, prob.cut = 0.7, include.loc = TRUE, seed = 123, inputfile = NULL)
```

**Arguments**

- `triggerobj`: An object of class *trigger* containing slot *loc.obj* with local-linkage probabilities and marker indices of the best local-linkage markers for genes in the genome. See *trigger* and *trigger.loclink* for details.

- `gender`: Optional. When computing statistics involving markers on sex chromosome, gender of each sample should be specified.
idx Optional. One can specify the indices of selected genes as putative regulators. By default, all the genes will be selected as putative regulators.

Bsec Number of iterations to perform when estimating null statistics for secondary-linkage and conditional independence.

prob.cut Probability threshold. The joint regulatory probabilities of a regulator to all the other genes will be set to zero if the local-linkage probability of the regulator is below the threshold; default prob.cut = 0.7.

include.loc Logical. If TRUE, the estimated posterior probability of regulation is more conservative.

seed Optional. A numeric seed for reproducible results.

inputfile Optional. If provided, reads in the probability matrix from working directory.

Details

The option idx contains the indices of putative regulator genes. When the data set is large, one can use this option by selecting a subset of genes as putative regulators in one computation and parallel-computes the genome-wide regulatory probability. If idx=NULL, all the genes will be computed for probability of regulation to other genes in the data.

If include.loc = TRUE, the joint posterior probability of regulation is the product of local-linkage, secondary-linkage and conditional independence. Otherwise, it is the product of secondary-linkage and conditional independence. The local-linkage is not a necessary condition for calculating regulation probability. If the probability of local-linkage is considered, the joint probability of regulation is more conservative. See references for details.

Value

A matrix of genome-wide regulatory probabilities with putative regulators in rows and regulated genes in columns. Note that the matrix is not symmetric. If gene i is estimated to be causal for gene j with high probability, the reverse is not true.

Author(s)

Lin S. Chen <lschen.stat@gmail.com>, Dipen P. Sangurdekar <dps@genomics.princeton.edu> and John D. Storey <jstorey@princeton.edu>

References


See Also

trigger.loclink, trigger.netPlot2ps and trigger.trait
Examples

## Not run:
data(yeast)
attach(yeast)
triggerobj <- trigger.build(marker = marker, exp = exp,
marker.pos = marker.pos, exp.pos = exp.pos)
triggerobj <- nettrig.loc(triggerobj, window.size = 30000)
trig.prob <- trigger.net(triggerobj, Bsec = 100)
netPlot2ps(trig.prob)
detach(yeast)

## End(Not run)

trigger.netPlot2ps-methods

Write the network from a trigger probability matrix to a postscript file

Description

Write the network from a trigger probability matrix to a postscript file.

Usage

## S4 method for signature 'trigger'
trigger.netPlot2ps(triggerobj, trig.prob, filenam = NULL, pcut = 0.95,
layout = c("radial", "energy-minimized", "circular","hierarchical"),
node.color = NULL, edge.color = NULL, node.shape = NULL, nreg = 20)

Arguments

triggerobj An object of class trigger.
trig.prob A network-Trigger regulatory probability matrix with putative regulator genes in rows and putative regulated genes in columns. See trigger.net for details.
filenam The output file name, without extension. If the name is not specified, the network will be write to the files temp.ps and temp.dot at the current directory.
pcut Threshold value for regulatory probabilities. The probabilities above the threshold are called significant and the corresponding regulatory relationships are plotted.
layout The layout of the output network. One can choose from "radial" (default), "energy-minimized", "circular" or "hierarchical" layouts. You can specify just the initial letter.
node.color The color of the nodes (genes). The default color is green.
edge.color The color of the edges. The default color is blue.
node.shape The shape of nodes (genes) if the number of regulatory relationships is below 1000. If that number is above 1000, the shape of nodes will be dot.
nreg The number of top regulators to be selected. These selected top regulators will be plotted in red ellipses with their gene names labeled inside.
Details

To use this function, please install the software Graphviz, which is available at http://www.graphviz.org/.
For large networks, layout "radial" or "energy-minimized" is recommended. If the total number
of significant regulatory relationships (directed edges) of the network is below 1000, we plot each
node (gene) as a "box" with its name labeled inside. Otherwise, we plot each gene as a "dot" without
name labeled to facilitate visualization. The top nreg (by default nreg = 20) regulators will be
plotted in red ellipses labeled with their names.

See manual of Graphviz for other available colors and shapes of nodes.

Author(s)

Lin S. Chen <lschen.stat@gmail.com>, Dipen P. Sangurdekar <dps@genomics.princeton.edu>
and John D. Storey <jstorey@princeton.edu>

See Also

trigger.link and trigger.mlink

Examples

## Not run:
  data(yeast)
  attach(yeast)
  triggerobj <- trigger.build(marker = marker, exp = exp,
  marker.pos = marker.pos, exp.pos = exp.pos)
  triggerobj <- trigger.loclink(triggerobj, window.size = 30000)
  trig.prob <- trigger.net(triggerobj, Bsec = 100)

  trigger.netPlot2ps(trig.prob, pcut = 0.95, layout = "e", filenam = "net95", nreg = 20)

  detach(yeast)
  ## End(Not run)
Arguments

- `triggerobj` : An object of class `trigger`. See `trigger` for details.
- `trait` : Trait for which causal regulator is to be found. It can either be a gene-name for a gene expression trait present in `triggerobj` or a vector of values for the individuals present in `triggerobj`.
- `cross` : An object of class `cross` obtained from `trigger.export2cross`. See `R/qtl` for more details.
- `thr` : LOD threshold to search for locally linked putative causal genes (default 3).
- `n.sv` : Number of surrogate variables used to model the local heterogeneity. If not set, it is computed from the expression data.
- `addplot` : If `TRUE`, a plot of the LOD scores from a genome-scan for a single-QTL model from package `R/qtl`.

Value

A vector of p-values associated with each tested causal regulator.

Author(s)

Lin S. Chen <lschen.stat@gmail.com>, Dipen P. Sangurdekar <dps@genomics.princeton.edu> and John D. Storey <jstorey@princeton.edu>

References


See Also

- `trigger.loclink` and `trigger.export2cross`

Examples

```r
## Not run:
data(yeast)
attach(yeast)
triggerobj <- trigger.build(marker = marker, exp = exp, 
 marker.pos = marker.pos, exp.pos = exp.pos)
crossfile <- trigger.export2cross(triggerobj)
tt.pval <- trigger.trait(triggerobj, trait = "DSE1", cross = crossfile)
causal.reg <- names(which(p.adjust(tt.pval, method = "fdr")<.05))
detach(yeast)
## End(Not run)
```
Description

A yeast data set for integrative genomic analysis.

Details


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